

Carbohydrate Research 307 (1998) 217-232

CARBOHYDRATE RESEARCH

Synthesis and antitumor activity of the 7-O-(2,6dideoxy-2-fluoro- α -L-talopyranosyl)daunomycinone derivatives modified at C-3' or C-4'

Yasushi Takagi, Naoki Kobayashi, Min Sun Chang, Geun-Jho Lim, Tsutomu Tsuchiya *

Institute of Bioorganic Chemistry, 3-34-17 Ida, Nakahara-ku, Kawasaki 211, Japan

Received 26 August 1997; accepted with revisions 18 December 1997

Abstract

As a part of a study to exploit anthracycline glycosides effective against resistant tumor cells, the 3'-O-methyl (3), 4'-O-methyl (4), 3'-deoxy (6), 3'-deoxy-3'-fluoro (7), and 3'-deoxy-3'-iodo (8) derivatives of 7-O-(2,6-dideoxy-2-fluoro- α -L-talopyranosyl)daunomycinone have been prepared by coupling suitably protected glycosyl bromides with daunomycinone. The doxorubicin-type analog (5) of 4 was also prepared. Among the compounds prepared, 5 showed the highest antitumor activity. Relationships between chemical structures of the synthetic products and antitumor activities, together with the degree of resistance were discussed. © 1998 Elsevier Science Ltd. All rights reserved

Keywords: Anthracycline glycoside; Antitumor activity; Multidrug resistance; 7-O-(2,6-Dideoxy-2-fluoro- α -L-talopyranosyl)daunomycinone

1. Introduction

Anthracycline antibiotics as exemplified by daunorubicin (DNR) and doxorubicin (DOX) are antitumor agents widely used in cancer chemotherapy. Their clinical use is, however, limited by undesirable side-effects such as drug-cumulative cardiotoxicity and myelosuppression, as well as the appearance of multidrug resistance (MDR) in tumor cells. To circumvent these drawbacks, many chemical and biosynthetic modifications have been undertaken [1,2]. In previous papers [3,4], we reported the synthesis of 7-O-(2,6-dideoxy-2fluoro- α -L-talopyranosyl)-daunomycinone (1) and -adriamycinone (2), both of which showed higher antitumor activities than DNR and DOX in murine leukemia L1210 *in vivo*; moreover they showed stronger cytotoxicity against P388 and its DOXresistant cell lines (P388/DOX) than DNR and DOX (this paper; Table 6). Another notable fact was that they had lower toxicity than DNR and DOX. From the standpoint of chemical structure, 1 and 2 have a fluorine atom at C-2' and a hydroxyl group at C-3' instead of the 3'-amino or 3'-alkylamino group present in conventional anthracycline glycosides; this replacement by a hydroxyl group, first reported by Horton and co-workers [5], is

^{*} Corresponding author.

important in view of altering the resistance factor [6]. We further synthesized the 14-hemipimelate [7] of **2** to make **2** soluble in water, and found that this compound exhibited stronger antitumor activity than the parent compound **2** against sensitive P388, but showed slightly lower activity against the P388/ DOX cell line ([8] and unpublished data). These results stimulated us to exploit further derivatization of **1** or **2** in the hope of obtaining compounds more effective against MDR cells.

Studies on the mechanism of MDR have shown considerable recent advances. A large proportion of MDR cells has been shown to over-express P-glycoprotein, an ATP-dependent drug-transport protein [9,10], in the plasma membrane, which causes such lipophilic drugs as doxorubicin to remain outside the cells, resulting in low intracellular drug-accumulation. Another interesting feature of MDR cells is their high Ca^{2+} content inside the cells as compared to the normal cells [11]. It has been reported that coadministration of verapamil, a calcium channel blocker, or its analogs [12], or calmodulin inhibitors, with DOX reversed the MDR activity [13]. Other types of MDR and the circumvention of resistance have been reviewed [14].

In spite of these findings, however, the relationship between drug structure and MDR is still obscure, except for the lipophilicity of the drug molecules [15]. For this reason, determination of the direction required for improving the biological properties of the drugs (in terms of antitumor activity, MDR, and side effects) is not easy. In this paper, we describe the activity-changes brought about by altering the functional groups at C-3' or 4' in 1 and 2 with several other groups, in the hope of finding a clue to this MDR problem in anthracycline glycosides [16].

2. Results and discussion

Synthesis.—At first, the 3'-O- (3) and 4'-Omethyl derivatives (4) of 1 were prepared. Conventional methylation (MeI, NaH in DMF) of methyl 4-*O*-benzyl-2,6-dideoxy-2-fluoro-α-L-talopyranoside 9 [4] (to give 10) with subsequent catalytic debenzylation gave the 4-ol 11 (Scheme 1). Its 19 F NMR spectrum showed a through-space coupling $(\sim 5 \text{ Hz})$ between OH-4 and F-2, supporting the ${}^{1}C_{4}(L)$ conformation of **11** (the conformation is not necessarily distinct if judged only from the $J_{\rm H,H}$ values) (Tables 1 and 2). Acetylation of 11 with Ac_2O in MeNO₂ in the presence of a catalytic amount of H_2SO_4 gave a mixture (~11:1) of 1,4-di-*O*-acetyl- α -L (12) and - β -L anomers 13. Their anomeric configurations were assigned based on the small $(J_{\text{H-1,F-2}} 8.5 \text{ Hz for } 12)$ and large $(J_{\text{H-1,F-2}}$ 20 Hz for 13) coupling constants. The mixture was converted into the α -L-talopyranosyl bromide 14 by treatment with TiBr₄, and 14 was used for the next coupling. The corresponding 4-O-methyl-α-Ltalopyranosyl bromide 19 was prepared also from 9. A first attempt at applying a sequence of reactions involving 3-O-acetylation, catalytic debenzylation, and methylation (MeI, Ag₂O in DMF) failed, giving a mixture of 3-O- and 4-O-methyl derivatives, possibly by acetyl migration in the final step. The hydroxyl group of 9 was therefore first protected with 3,4-dihydro-2H-pyran and the 3-O-tetrahydropyranyl derivative 15 was hydrogenolyzed, and the resulting 4-ol 16 was



Scheme 1.

Table 1 ^{1}H and ^{19}F chemical shifts (δ (ppm)) and multiplicity of fluoro sugar derivatives in CDCl_3

Multiplicity	H-1 dd ^a	H-2 ddt ^a	H-3 dt ^a	H-4	H-5 dq ^a	H ₃ C-5 d ^a	CH ₃ O s	Ac s	¹⁹ F ddd ^a
10	4.94	4.68	3.46	3.61	3.83	1.24	3.37		-204.0
11	4.90 br.d	4.68 br.d	3.46	3.81 br s	3.86	1.36	3.40 3.50		b
12	6.33	4.59	3.56	5.37 dt	4.10	1.22	3.48	2.13 2.18	-202.9
13	5.65	4.78	3.41 ddd	5.33 dt	3.80	1.29	3.47	2.18 2.19	-220.0
14	6.57	4.86	3.97 ddd	5.43 dt	4.24	1.27	3.49	2.17	-180.9
15	4.90 4.92	4.62 4.69	3.93 4.02	3.57 dt 3.61 dt	3.87 3.90	1.23 1.27	3.36 3.37		-204.0 (0.44 F) -202.7 (0.56 F)
16	4.89 4.90	4.63 4.69 br d	÷	-3.75-4.0-	\rightarrow	1.35	3.40		-201.8 ddt -200.4 ddt
17	4.90 4.87	4.57 (0.38H) 4.63	3.97 3.92	~3.36	~3.91	1.32	3.37(3 H) 3.58 3.64		-204.1 (0.38 F) -202.9
18	6.29	(0.62H) 4.56	5.12	3.45	4.09	1.34	3.56	2.11	(0.62 F) -202.2
19	6.54	4.84	5.44	dt ∼3.55	4.22	1.38	3.57	2.20	-180.2
21	4.73	4.41	5.06	3.61 dd	4.15	1.27	3.45		-194.7 ddda
22	4.83	4.46 br ddd	4.58	3.52 m	4.08	1.25	3.39		-185.1
23	4.79 br d	4.43 dddt	1.85 ddt 2.39 dddt	3.29 m	3.94	1.28	3.40		-187.0 dddd
25	4.86 br d	4.49 dddt	2.22 ddt 2.40 dddt	5.06 m	4.15	1.26	3.46		-186.5 ddt
26	6.29 br d	4.52 br d	2.26 ddt 2.52	5.12 br s	4.26	1.27		2.15	-185.1 ddt
27	5.79	4.71 br dt	2.14 dddd 2.65 ddt	5.12 m	4.08	1.34		2.23	-202.2 dddd
28	6.61 br d	4.81 br d	2.66 ddt 2.48 dddd	5.19 m	4.37	1.32			-163.2 dddd
29	4.94 ddd	4.72 dddt	4.74 ddt	3.71 ddt	3.86 tq	1.26 dd	3.37		-205.5(F-2) dddd -2.02.9(F-3) br dda
31	6.34 ddd	4.69 ddddd	4.84 br ddt	5.41 br ddt	4.14 tq	1.24 dd		2.13 2.19	-204.6(F-2) dddd -206.2(F-3) br dda
32	5.67 br dd	4.87 dddt	4.68 dddd	5.38 ddt	3.83 tq	1.31 br d		2.19 2.20	-220.7(F-2) dddd -202.6(F-3) br dddd
33	6.53 br dd	4.97 dddt	5.21 dddd	5.49 ddt	4.28 br q	1.29 dd		2.18	-182.8(F-2) dddd

(continued)

	Table 1—contd										
Multiplicity	H-1 dd ^a	H-2 ddt ^a	H-3 dt ^a	H-4	H-5 dq ^a	H ₃ C-5 d ^a	CH ₃ O s	Ac s	¹⁹ F ddd ^a		
35 40	4.78 br d 4.91	4.44 dddd 4.53	5.07 ddt 4.74	~3.45 m 5.37	4.20 4.29	1.30 1.22	3.43 3.46	2.10	-207.4(F-3) br ddq -191.9 dddd -185.6		
41	6.32	4.53 br.d	4.75	5.44 m	4.39	1.22		2.18	-183.7		
42	6.58	4.81	5.13	5.49 m	4.51	1.28			-163.0		

^a Unless otherwise stated.

^b See Experimental section.

Table 2 1 H- 1 H and 1 H- 19 F coupling constants (*J*, Hz) of fluoro sugar derivatives in CDCl₃

	$J_{1,2}$	$J_{2,3}$	$J_{2,4}$	$J_{3,4}$	$J_{4,5}$	$J_{5,\mathrm{CH}_{3-5}}$	$J_{1,\mathrm{F}}$	$J_{2,\mathrm{F}}$	$J_{3,\mathrm{F}}$
10	1.5	3	1.5	3	1.5	6.5	9.5	49.5	33
11		3		3	~ 1.5	6.5	9	50	32.5
12	2	3	~ 1	3.5	1.5	6.5	8.5	48.5	31.5
13	≤ 1	2.5	~ 1	3.5	1.5	6.5	20	51	30.5
14	~ 1	3	1.5	3.5	~ 1.5	6.5	11.5	49.5	30
15	1.5	3	1.5	3	1.5	6.5	9	49.5	33
16	1.5	3	1.5			6.5	~ 7	49.5	33
17	1.5	3	1.5	3		6.5	9.5	49.5	33.5
18	2	3	~ 1.5	3	~ 1.5	6.5	8.5	49	31.5
19	1.5	3	1.5	3	~ 1.5	6.5	11.5	50.5	30.5
21	4 5	65	~ 0	65	4 5	6.5	9	48	12
21	1.5	3	0	3	~1.5	6.5	85	46.5	37.5
73 a,b	2	35	a.1	35	2	6.5	10	40.5	15
25	2	(I)	/01	(I)	2	0.5	10	,	ч <i>J</i>
		$(J_{2,3ax})$		$(J_{3ax,4})$					$(J_{3ax,F})$
		(I)		(\mathbf{I})					(I)
35 a.c	1	$(J_{2,3eq})$	1	$(J_{3eq,4})$	15	6.5	0.5	16 5	(J _{3eq,F})
25	~ 1	(I)	~ 1	3.3	1.5	0.5	9.5	40.3	45.5
		$(J_{2,3ax})$		$(J_{3ax,4})$					$(J_{3ax,F})$
		2.5		2.5					14.5
acd		$(J_{2,3eq})$		$(J_{3eq,4})$		<i>.</i> -	o -		$(J_{3eq,F})$
26 ^d		3.5		3.5	1.5	6.5	8.5	45.5	45
		$(J_{2,3ax})$		$(J_{3ax,4})$					$(J_{3ax,F})$
		3.5		2.5					13.5
		$(J_{2,3eq})$		$(J_{3eq,4})$					$(J_{3eq,F})$
27 ^d	1	3		4	2	6.5	21	48.5	43.5
		$(J_{2,3ax})$		$(J_{3ax,4})$					$(J_{3ax,F})$
		~ 3		~ 3					11.5
		$(J_{2,3eq})$		$(J_{3eq,4})$					$(J_{3eq,F})$
28 ^d		$\sim 3^{-1}$		$\sim 3^{\circ}$	1.5	6.5	11	47	43.5
		$(J_{2,3ax})$		$(J_{3ax,4})$					$(J_{3ax,F})$
		2.5		4.5					14
		$(J_{2,3eq})$		$(J_{3eq,4})$					$(J_{3eq,F})$
29 ^{e,f,g,h}	1.5	3	1.5	3	1.5	6.5	9 (F-2)	49.5 (F-2)	31.5 (F-2)
							7 (F-3)	7 (F-3)	44 (F-3)
31 ^{i,f,j,k}	2	3	1.5	4	1.5	6.5	8 (F-2)	49 (F-2)	30 (F-2)
							6 (F-3)	6.5 (F-3)	43 (F-3)
32 ^{1,f,m}	1	3	1	4	1.5	6.5	19 (F-2)	52 (F-2)	28 (F-2)
							2 (F-3)	7 (F-3)	42.5 (F-3)
33 ^{e,n,h}	1.5	3	1.5	4	1.5	6.5	11.5 (F-2)	50 (F-2)	28 (F-2)
		-		-			6.5 (F-3)	5 (F-3)	42.5 (F-3)
35°	2	3.5	1	3.5	2	6.5	13	45	11
40	1.5	2.5	1	3.5	1.5	6.5	7.5	46.5	36.5
41	1.5	3		3	1.5	6.5	7.5	46	35.5
42	1.5	3	1	35	~ 1.5	6.5	10	46 5	34
• •	1	5	1	5.5	-1.5	0.5	10	10.5	51

 $\begin{array}{c} \hline a & J_{1,3eq} & 1.5 \, \text{Hz.} \ ^{\text{b}} & J_{3ax,3eq} & 15.5 \, \text{Hz.} \ ^{\text{c}} & J_{3ax,3eq} & 16 \, \text{Hz.} \ ^{\text{d}} & J_{3ax,3eq} & 16.5 \, \text{Hz.} \ ^{\text{e}} & J_{4,F-3} & 6 \, \text{Hz.} \ ^{\text{f}} & J_{5,F-3} & 1.5 \, \text{Hz.} \ ^{\text{g}} & J_{6,F-3} & 0.8 \, \text{Hz.} \ ^{\text{h}} & J_{F-2,F-3} & 14 \, \text{Hz.} \ ^{\text{h}} & J_{4,F-3} & 5 \, \text{Hz.} \ ^{\text{g}} & J_{6,F-3} & 1 \, \text{Hz.} \ ^{\text{h}} & J_{F-2,F-3} & 13.5 \, \text{Hz.} \ ^{\text{h}} & J_{F-2,F-3} & 13 \, \text{Hz.} \ ^{\text{h}} & J_{6,F-3} & 0.7 \, \text{Hz.} \ ^{\text{o}} & J_{1,3} & 1 \, \text{Hz.} \end{array}$

methylated to give the 4-*O*-methyl derivative 17. Compound 17 was then acetolyzed (Ac₂O, AcOH, cat. H₂SO₄) and the resulting 1,3-di-*O*-acetyl- α -L-talopyranose (18) was brominated with TiBr₄ to give the α -1-bromide 19 in high yield.

Coupling of 12 with daunomycinone was first attempted according to the procedure reported [17] (CF₃SO₃SiMe₃ and molecular sieves 4A in CH₂Cl₂-Et₂O), but the desired product was not obtained. Coupling of the bromide 14 with daunomycinone under Koenigs-Knorr conditions, however, gave the α -L- (43) and β -L-glycosides 44 in 38 and 8% yields, respectively. The anomeric configurations of 43 and 44 were again determined by their $J_{\text{H-1',F-2'}}$ coupling constants (9.5 Hz for 43 and 19 Hz for 44). Previously, we reported [4] that the coupling of 3,4-di-O-acetyl-2,6-dideoxy-2-fluoro- α -L-talopyranosyl bromide (an analog of 14) with daunomycinone gave the 3',4'-di-O-acetyl derivative of 1 in high yield (82%) without accompaniment of the undesirable β -L anomer; the present poor yield of **43** (confirmed by repeated experiments), therefore, must be ascribed to the difference in substituent at C-3 in **14**. Alkaline treatment of **43** gave the desired 7-*O*-(2,6-dideoxy-2-fluoro-3-*O*-methyl- α -L-talopyranosyl)daunomycinone (**3**). The structure was confirmed by the ¹H (Table 3), ¹⁹F, and ¹³C NMR spectra (Table 4). In its ¹⁹F NMR spectrum, through-space coupling $J_{OH-4',F-2'}$ (9 Hz) was observed.

Next, 4-*O*-methyl- α -L-talopyranosyl bromide **19** was coupled with daunomycinone as described for **43**, giving the α -L-glycoside **45** in 50% yield. Alkaline hydrolysis of **45** gave 7-*O*-(2,6-dideoxy-2-fluoro-4-*O*-methyl- α -L-talopyranosyl)daunomycinone **4** in good yield. The daunomycin-type structure of **4** was then transformed into a doxorubicin-type structure by applying, basically, a sequence of reactions reported by Arcamone et al. [18]; bromination of **4** with Br₂ in the presence of HC(OMe)₃ gave the 14-bromo-13-dimethylacetal, the acetal portion being then deblocked by treatment with

Table 3

Selected ¹H NMR chemical shifts (δ (ppm)) and coupling constants (J, Hz) of **3–8** and **43–51** in CDCl₃

Compound	H-7	OMe-4	H-14	H-1′	H-2′	H-3'ax	H-4′	OMe- 3' or 4'	J _{7,8ax}	$J_{7,8eq}$	$J_{1',2'}$	J _{2',3'ax}	<i>J</i> _{3'<i>ax</i>,4'}	J _{1',F-2'}	$J_{2',\mathrm{F-}2'}$	$J_{3'ax,{ m F-2'}}\ (J_{3'eq,{ m F-2'}})$
3	5.27	4.03	2.41	5.61	4.73	3.32	3.84	3.44	4.5	1.5	~ 1	3	3	10	~ 50	32.5
	dd	S	s	dd	br d	dt	br d	S								
4	5.26	4.07	2.40	5.58	4.50	3.69	3.33	3.60	4	~ 2	*	${\sim}4$	4	10	49.5	31.5
	dd	S	s	br d	br d	ddt ^a	br d	S								
5	5.34	4.09	4.74	5.62	4.47	3.65	3.34	3.61	4	2	1.5	~ 3	~ 4	10	49	31
	dd	S	br s	dd	br d	ddt ^a	br d	S								
6	5.35	4.08	2.41	5.50	4.56	1.92	3.55		4	2	*	3	3	10	47	49
	dd	S	s	br d	br d	ddt ^b	br d ^c									(13)
7	5.28	4.09	2.40	5.65	4.83	4.57	3.92		4.5	2	2	3	3	9.5	50.5	31.5
_	dd	S	S	ddd	dddta	ddt	m									
8	5.30	4.09	2.41	5.53	4.56	4.42	3.59		4.5	1.5	1.5	3	3	8.5	46.5	38
	dd	S	S	dd	br d	dt	br d ^e			-				~ -		
43	5.30	4.09	2.41	5.63	4.62	3.38	5.34	3.39	4.5	2	1.5	~ 3	~ 3	9.5	49	32.5
	dd	S	s	dd	br d	dt	m	S			0					2 0 -
44	5.57	4.09	2.43	4.95	4.79	3.31	5.18	3.45	3.5	~ 2.5	~ 0	2.5	4	19	51	30.5
	br t	S A OT	S	d	br dd	ddd	br dd	S S S S S S		•	1 -	2	2	10	50	22.5
45	5.26	4.07	2.41	5.59	4.59	4.90	3.47	3.55	4	~ 2	1.5	3	3	10	50	32.5
16	dd	S 1 OO	S 2 12	dd	br d	dt	m	S		1.7	*	2	2	10	16	16
46	5.39	4.09	2.43	5.60	4.52	2.08	5.10		4	1.5	~	~ 3	~ 3	10	46	46
47		S	S	br d	br d		br s		2.5	2	*	4	4	20	40.5	(13)
4/	5.63	4.09	2.45	5.11	4.68	2.05	4.96		3.5	3		~ 4	~ 4	20	48.5	43.5
40	aa 5 29	S	2 40	br a	br d		br s		4.5	2	15	2.5	2.5	0.5	50	(12)
40	3.28 dd	4.09	2.40	3.00 444	4./3	4.0/	5.39		4.5	2	1.5	3.3	3.3	9.5	50	30
40	5 50	s 4 00	5 2 4 4	5 00	1 95	4 59	5 22		25	2	0	2.5	4	10	50	28 5
49	5.59	4.09	2.44	3.00 AA	4.63 ddd	4.30 dddd	3.23		5.5	\sim 3	~ 0	2.3	4	10	32	28.3
50	เ 5 3 ว	5 4 10	3 2 1 2	5.62	4 57	4.61	5 30		4	15	*	- 3	- 3	8	15 5	37
50	3.32 dd	4.10	2.4Z	5.02 br.d	$\frac{4.5}{brd}$	4.01 dt	5.59 br s		4	1.5		\sim s	\sim 3	0	45.5	57
51	5.62	× 1 00	2 16	5 21	1 72	4.48	5 22		35		*	3	3	10.5	18	34.5
51	br t	4.09 S	∠.+0 S	br d	$\frac{1}{1}$	dt	br s		5.5	,~2.5		5	5	17.5	40	54.5
	21 0	5	Ŭ				21 0									

^a J_{3',OH-3'} 11.5 Hz. ^b J_{3'ax,3'eq} 16 Hz. ^c J_{4',OH-4'} 11 Hz. ^d J_{2',4'} 2 Hz. ^e J_{4',OH-4'} 10 Hz.

* Not detected by broadening.

С	3	4	5	6	7 ^a	8
1	119.9	119.7	119.9	119.8	119.9	119.9
2	135.8	135.7	135.8	135.4	135.9	135.9
3	118.6	118.5	118.6	118.5	118.6	118.6
4	161.2	161.1	161.2	161.1	161.2	161.2
4a	120.9	120.8	120.9	120.8	120.9	120.9
5	187.0 ^ь	186.8 ^ь	187.0 ^ь	186.9 ^b	187.1	187.1 ^ь
5a	111.6 °	111.54 °	111.7	111.4	111.7	111.7 °
6	156.3 ^d	155.7 ^d	156.0 °	156.6 °	156.2	156.2 ^d
6a	133.2 °	133.1 ^e	132.7 ^d	133.4 ^d	132.9	133.0 °
7	71.4	71.0	70.8	70.9	71.4	71.4
8	35.2	35.0	35.6	35.1	35.3	35.4
9	76.5	76.4	76.6	76.6	76.3	76.3
10	33.1	33.1	33.9	33.2	33.1	33.1
10a	134.3 ^e	134.0 ^e	133.4 ^d	134.4 ^d	134.1	134.1 ^e
11	155.6 ^d	155.1 ^d	155.4 °	155.3 °	155.5	155.6 ^d
11a	111.5 °	111.48 °	111.7	111.4	111.6	111.6 °
12	186.7 ^b	186.6 ^b	186.8 ^b	186.7 ^b	186.9	186.8 ^b
12a	135.5 °	135.4 ^e	135.5 ^d	135.8 ^d	135.5	135.5 °
13	211.1	211.3	213.4	211.5	211.0	211.0
14	24.6	24.6	65.4	24.7	24.5	24.5
OMe-4	56.7	56.6	56.7	56.7	56.7	56.7
OMe-3'	55.9					
OMe-4'		62.9	63.0			
1′	101.4 d (31.5)	101.3 d (32.2)	101.1 d (32.2)	100.3 d (34.0)	101.3 dd (30.5, 6.8)	100.5 d (34.4)
2'	86.3 d (176.9)	87.3 d (177.0)	87.1 d (177.6)	86.5 d (166.7)	87.0 dd (178.2, 17.1)	89.9 d (176.9)
3'	74.5 d (14.8)	66.0 d (17.1)	66.0 d (16.9)	30.8 d (17.6)	85.7 dd (192.3, 14.7)	25.8 d (16.9)
4′	68.7	81.0	80.9	66.5	70.4 d (17.4)	72.3
5'	68.0	67.5	67.8	67.9	67.8 d (5.9)	68.2
6'	16.5	16.7	16.8	16.9	16.1 d (2.1)	17.4

¹³C NMR chemical shifts (δ (ppm)) and coupling constants ($J_{C,F}$, Hz, in parentheses) for compounds **3–8** in CDCl₃

^a Measured at 125.8 MHz and confirmed by the HMQC followed by HMBC method.

^{b, c, d, e} Figures in the same column may be interconvertible.

acetone. The resulting 14-bromo-13-oxo derivative was hydrolyzed with sodium formate in aq acetone giving a mixture of 5 and its 14-O-formyl derivative; thorough deblocking of the mixture with NH₄OH in CHCl₃-MeOH gave the doxorubicintype derivative 5.

Next, the 3'-deoxy (6), 3'-deoxy-3'-fluoro (7), and 3'-deoxy-3'-iodo derivatives (8) of 1 were prepared. The synthetic routes for the glycosyl bromides are shown in Scheme 2. Treatment of methyl 4-O-benzyl-2,6-dideoxy-2-fluoro-α-L-idopyranoside **20** [4] with $(CF_3SO_2)_2O$ gave the unstable 3-triflate **21**, whose ¹⁹F NMR spectrum showed interspace coupling (6 Hz) between F-2 and CF_3 . It is worth mentioning that 21 showed somewhat different $J_{\rm H,H}$ values from those for similar α -L-*ido* derivatives (20 [4], 34 [4], 35), suggesting the presence of some influence of the CF₃SO₂-3 group on its conformation. Iodination (NaI in DMF, 90°C) of 21 gave the 3-deoxy-3-iodo derivative 22. The L-talo structure was confirmed by the large coupling constant ($J_{H-3,F-2}$ 37.5 Hz); if L-*ido* had been the structure, the $J_{\text{H-3,F-2}}$ value should be small, irrespective of the conformations. Its ¹³C NMR spectrum showed a doublet ($J_{\text{C-3,F-2}}$ 15.7 Hz) at high field (δ 24.2), indicating the presence of I-3. Radical deiodination of **22** with Bu₃SnH gave the 3-deoxy derivative **23** quantitatively. Hydrogenolysis of **23** (to give **24**) followed by *p*-nitrobenzoylation afforded the 4-*p*-nitrobenzoate **25**. Acetylation of **25** in the manner described for **12** gave a mixture of 1-*O*-acetyl- α -L (**26**) and - β -L anomers **27**, the ratio being 6:1. Compound **26** was then converted into the α -L-*lyxo*-hexopyranosyl bromide **28** by treatment with HBr in AcOH.

Preparation of 3-deoxy-3-fluoro- α -L-talopyranosyl bromide **33** was attempted initially by treatment of **20** with Et₂NSF₃ (DAST), but it failed, giving only a complex mixture. Then the 3triflate **21** was treated with (Me₂N)₃S⁺ (Me₃SiF₂)⁻ (TASF) [19] and the resulting 2,3,6-trideoxy-2,3difluoro- α -L-talopyranoside **29** was hydrogenolysed (to give **30**) and acetolyzed as described for **18** to give a mixture of 1,3-di-*O*-acetyl- α -L (**31**)

Table 4



Scheme 2.

and $-\beta$ -L anomers **32**, the ratio being 2.5:1. Bromination of **31** as described for **28** gave the crystalline α -L-talopyranosyl bromide **33**.

The 3-deoxy-3-iodo- α -L-talopyranosyl bromide 42 was prepared from methyl 3-O-acetyl-4-O-benzyl-2,6-dideoxy-2-fluoro- α -L-idopyranoside 34 [4]. Successive debenzylation of 34 (to give 35), protection of the OH-4 group with a tetrahydropyranyl group (to give 36), deacetylation of 36 (to give 37), and triflation gave the unstable 3triflate 38. Iodination of 38 with NaI in DMF gave the 3-deoxy-3-iodo- α -L-talopyranoside 39, the tetrahydropyranyl group in 38 being removed during the reaction. After *p*-nitrobenzoylation, the resulting ester 40 was acetolyzed (to give the 1-acetate 41), and brominated to give the α -L-talopyranosyl bromide 42.

Characteristic features of the 1-bromides prepared in the ¹⁹F NMR spectra are that the shiftvalues for F-2 always appear downfield by ~20 ppm compared to those for the parent methyl α -L-glycosides or α -L-glycosyl acetates (compare **11** (or **12**) and **14**; **17** (or **18**) and **19**; **25** (or **26**) and **28**; **29** (or **31**) and **33**; and **40** (or **41**) and **42**; Table 1).



Coupling of the respective glycosyl bromides **28**, **33**, and **42** with daunomycinone was performed in the manner described for **43** to give the corresponding coupling products: α -L (**46**, 43%) and β -L anomers (**47**, 41%) from **28**; α -L (**48**, 28%) and β -L anomers (**49**, 16%) from **33**; α -L (**50**, 22%) and β -L anomers (**51**, 62%) from **42**, the ratios of α -L: β -L anomers being ~1:1, 1.8:1, and 1:2.8, in the foregoing order. In these coupling reactions, the α selectivity observed [4] in **1** was completely lost and the undesirable β -L anomers were sometimes major products. The anomeric configurations of the products were determined by their $J_{H-1',F-2'}$ values in the ¹H and ¹⁹F NMR spectra. Alkaline treatment

Compound		D	ose (mg kg ⁻¹ day	^{y-1})				C ₁₅₀ ^c	Ec
	10	5	2.5	1.25	0.6	0.3	0.15		
1		184	217	171	125	105	105	0.9	5.7
2		>740	> 352	275	185	182	127	0.2	6.0
3	174	109	112	100	91	94		8.2	12.3
4		> 314 ^d	260	146	124	117	111	1.25	5.1
5	102 ^d	124 ^d	>761 ^d	> 501	279	152		0.3	6.0
6		121	113	113	96	110	107	> 5	> 12.5
7		148	135	114	117	108	105	5	18
8		116	97	106	103	97	97	> 5	>16

Table 5 Antitumor activities^a $(T/C^{b} (\%))$ of 1–8 against the murine L1210 cell line

^a Leukemia L1210 cells (10^5) were inoculated into CDF₁ mice (20 ± 1 g) intraperitoneally. Drugs were administered daily, starting 24 h after inoculation, from day 1 to 9 intraperitoneally. Survival studies were continued up to 60 days; a part of the data were reported [3,16].

^b Mean survival days of treated mice/mean survival days of control mice ×100.

^c See text portion.

^d More than 10% weight decrease in the treated mice was observed.

of 46, 48, and 50 gave the respective desired products 6, 7, and 8 in good yields.

Antitumor activity.—Antitumor activities of 3-8 against L1210 murine leukemia in vivo (Table 5) showed that only 4 and 5 exhibited strong antitumor activity (5 was much stronger). This result suggests, together with the activities [3] of the parent compounds 1 and 2, that those modifications that remove the exchangeable hydrogen from the HO-3 group (as in 3, 6–8) cause total loss or great decreases in the activity inherent in the parent anthracycline antibiotics. However, in view of drug resistance against P388/DOX (Table 6), there was a considerable difference between 4 and 5 (4 showed partial drug-resistance against P388/DOX). Judging from the resistant factor (RF)-values (Table 6), it appears that HO-14 (in 2, 5, and DOX) induced resistance (possibly by MDR), and absence of the HO-3' (or H_2N-3') group reversed the resistance. However, these hydroxyl groups, especially HO-3', seem to be the groups essential for manifesting antitumor activity in these anthracycline glycosides. It appears, therefore, that the more active compounds suffer the stronger resistance. In this context, a 3',4'-di-O-methyl analog of DNR, SO-075R1 [20] was expected to show only weak antitumor activity. However, we set a parameter (E) as a measure for estimating the antitumor activity and resistance (to P388/DOX) concurrently, by multiplying the RF-value in Table 6 and the dose (C_{150}) needed to attain the value of T/C 150% in Table 5 (for example, in 4: $4.1 \times 1.25 = 5.1$; the values are shown in the last column in Table 5). Based on this parameter, the antitumor activities of 4 and 5 (as well as 1 and 2) are estimated as being almost equal, and superior to those of the other derivatives. Since the Ca^{2+} ion is considered to play an important role in MDR, the pursuit of the relationship between Ca-chelating ability of our synthetic products and RF (or E) value will be an interesting problem in future.

3. Experimental

General methods.—Melting points were determined on a Kofler block, and are uncorrected. Optical rotations were measured with a Perkin– Elmer 241 polarimeter. ¹H NMR (at 250 MHz) and ¹⁹F NMR (at 235.7 MHz) spectra were recorded with a Bruker WM 250 spectrometer. ¹³C NMR (at 62.9 or 125.8 MHz) spectra were recorded with a Bruker WM 250 or Bruker AMX 500 spectrometer. Chemical shifts (δ) for ¹H, ¹³C, and ¹⁹F spectra were measured, downfield from internal Me₄Si, downfield from Me₄Si with aid of internal 1,4-dioxane ($\delta = \delta_{1,4-dioxane}+67.4$), and downfield from internal Freon 11 (CFCl₃), respectively. TLC was performed on Kieselgel 60 F₂₅₄ (Merck), and column chromatography on Wakogel C-200.

Methyl 4-O-benzyl-2,6-dideoxy-2-fluoro-3-Omethyl- α -L-talopyranoside (10).—A mixture of NaH (170 mg, 60% NaH in mineral oil, 4.3 mmol) and 9 [4] (460 mg, 1.7 mmol) in dry DMF (4.6 ml) was stirred under an atmosphere of N₂. After hydrogen evolution had ceased, MeI (1 mL, 10.8 mmol) was added, and the stirring was continued for 1 h at room temperature. After addition





Compound		Subs	tituent		IC	D Ep	
	R ¹	\mathbb{R}^2	R ³	R ⁴	P388	P388/DOX	- RF ⁶
1	Н	F	OH	OH	0.008	0.05	6.3
2	OH	F	OH	OH	0.003	0.09	30
3	Н	F	OMe	OH	0.066	0.10	1.5
4	Н	F	OH	OMe	0.032	0.13	4.1
5	OH	F	OH	OMe	0.004	0.08	20
6	Н	F	Н	OH	0.150	0.37	2.5
7	Н	F	F	OH	0.073	0.26	3.6
8	Н	F	Ι	OH	0.130	0.42	3.2
DNR	Н	Н	NH_2	OH	0.020	0.34	17
DOX	OH	Н	NH_2^2	OH	0.014	> 0.50	> 36

 a IC₅₀ values (50% inhibition concentration) were determined on day-3 culture.

^b Resistant factor (RF): IC_{50} for resistant subline/ IC_{50} for sensitive subline.

of AcOH (0.2 mL) followed by CHCl₃, the solution was washed with aq NaCl (saturated), dried (Na₂SO₄), and concentrated with occasional addition of xylene. The residue was chromatographed (9:1 toluene–EtOAc) to give **10** as a syrup, 409 mg (84%), TLC (10:1 toluene–EtOAc): R_f 0.27 (*cf* **9**: R_f 0.17), $[\alpha]_D^{24}$ –38° (*c* 1, CHCl₃). Anal. Calcd for C₁₅H₂₁FO₄: C, 63.36; H, 7.44; F, 6.68. Found: C, 63.37; H, 7.41; F, 6.73.

1,4-Di-O-acetyl-2,6-dideoxy-2-fluoro-3-O-methylα- (12) and -β-L-talopyranose (13).—To a solution of 10 (379 mg, 1.3 mmol) and AcOH (1.9 mL) in 1,4dioxane—H₂O (8:1, 21 mL) was gently introduced H₂ in the presence of palladium black for 1 h at room temperature. Filtration followed by concentration with aid of toluene gave 11 as a syrup, which was chromatographically homogeneous, 260 mg (quant), R_f 0.2 (TLC, 30:1 CHCl₃–acetone); ¹⁹F NMR (CDCl₃) δ –201.6 (br ddd, which collapsed to sharp ddd in addition of D₂O; $J_{OH-4,F}$ ~5 Hz; Table 2). To an ice-cold solution of the syrup in dry MeNO₂ (8 mL) were added Ac₂O (1.3 mL, 13.8 mmol) and H₂SO₄ (8 μL), and the solution was kept for 2.5 h at room temperature. TLC (30:1 CHCl₃-acetone) showed two spots at R_f 0.3 (12, major) and 0.2 (13, minor). After pouring into NaHCO₃ (4.7 g) suspended in water (13.7 mL), and stirring for 1 h, the mixture was extracted with CHCl₃. The solution was washed with aq 10%NaCl, dried (Na₂SO₄), and concentrated. Chromatography (30:1 CHCl₃-acetone) of the residue gave 12 as crystals, 236 mg (67%), and a mixture of 12 and 13 (74 mg). Rechromatography of the latter with the same solvent system gave an additional crop of **12**, 38 mg (11%), and **13** as a syrup, 25 mg (7.2%). Compound 12, mp 139-140°C (prisms from CHCl₃–diisopropyl ether); $[\alpha]_{\rm D}^{21}$ –159° (c 1, CHCl₃). Anal. Calcd for C₁₁H₁₇FO₆: C, 50.00; H, 6.48; F, 7.19. Found: C, 49.90; H, 6.72; F, 6.93. Compound **13** had $[\alpha]_{D}^{23} - 28^{\circ}$ (*c* 1, CHCl₃). Anal. Calcd for C₁₁H₁₇FO₆: C, 50.00; H, 6.48. Found: C, 50.20; H, 6.51.

4-O-Acetyl-2,6-dideoxy-2-fluoro-3-O-methyl- α -Ltalopyranosyl bromide (14).—A mixture of 12 (329 mg, 1.2 mmol) and TiBr₄ (1.14 g, 3.8 mmol) in dry CH₂Cl₂-EtOAc (9:1, 7.3 mL) was stirred for 16 h at room temperature. TLC (3:1 hexane-acetone) of the deep-brown mixture showed a single spot at R_f 0.35. After dilution with dry MeCN (13 mL), anhydrous NaOAc (2.88 g) was added at 0 °C and the mixture was stirred until the color faded to pale yellow. The mixture was filtered through Celite with aid of toluene, and the organic solution was concentrated. Extraction of the residue with toluene followed by concentration gave 14 as a pale-yellow syrup (314 mg, 88%), which was used without further purification.

Methyl 4-O-benzyl-2,6-dideoxy-2-fluoro-3-O $tetrahydropyran-2-yl-\alpha-L-talopyranoside$ (15).—A mixture of 9 (536 mg, 2.0 mmol), 3,4-dihydro-2H-pyran (0.53 mL, 6 mmol), and pyridinium p-toluenesulfonate (57 mg, 0.23 mmol) in dry CH_2Cl_2 (8 mL) was kept for 5 h at room temperature. The solution, after pouring into aq NaHCO₃ (saturated), was extracted with CHCl₃ and the extract was concentrated. Chromatography (20:1 toluene-acetone) of the residue gave a diastereoisomeric mixture (1:0.8 by ¹⁹F NMR) of **15** as a syrup, 571 mg (81%), TLC (20:1 toluene–acetone): $R_f \ 0.25, \ [\alpha]_{\rm D}^{22} \ -47^{\circ} \ (c \ 1, \ {\rm CHCl}_3); \ {}^1{\rm H} \ {\rm NMR}$ (CDCl₃) (signals not listed in Table 1): δ 7.2–7.5 (5 H, m, Ph), 4.81–4.87 (1 H, H-2'), 4.71, 4.75, 4.91, 5.01 (each d, 2 H in total, J 12 Hz, PhCH₂), \sim 3.9 (1 H, H-6'b), 3.48-3.55 (1 H, H-6'a), 1.5–2.0 (6 H, H-3'a,b, 4'a,b, 5'a,b of THP), Anal. Calcd for C₁₉H₂₇FO₅: C, 64.39; H, 7.68; F, 5.36. Found: C, 64.31; H, 7.66; F, 5.11.

Methyl 2,6-dideoxy-2-fluoro-3-O-tetrahydropyran-2-yl- α -L-talopyranoside (16).—Compound 15 (666 mg, 1.9 mmol) was hydrogenolyzed in 1,4dioxane–H₂O (16:1, 34 mL) for 2 h as described for 11, and the products were chromatographed (10:1 CHCl₃–EtOAc) to give 16 as a diastereoisomeric mixture of semi-crystalline syrup, 411 mg (83%). ¹H NMR (CDCl₃) (signals not listed in Table 1): δ 4.85–4.94 (1 H, H-2'), 3.75–4.0 (4 H, H-3, 4, 5, 6'b), 3.54 (m, 1 H, H-6'a), 2.39 (t, 0.86 H, $J_{4,OH-4}=J_{OH-4,F}$ 7 Hz, OH-4 of the major isomer), 2.28 (t, 0.14 H, $J_{4,OH-4}=J_{OH-4,F}$ 8.5 Hz, OH-4 of the minor isomer), 1.5–2.0 (6 H, H-3'a,b, 4'a,b, 5'a,b). Anal. Calcd for C₁₂H₂₁FO₅: C, 54.53; H, 8.01; F, 7.19. Found: C, 54.62; H, 8.01; F, 6.88.

Methyl 2,6-dideoxy-2-fluoro-4-O-methyl-3-Otetrahydropyran-2-yl- α -L-talopyranoside (17).— Compound 16 (434 mg, 1.6 mmol) in DMF (4.3 mL) was treated with NaH (188 mg, 60% in mineral oil, 4.7 mmol) and MeI (0.4 mL, 6.4 mmol; 2h) as described for 10. Column chromatography (10:1 CHCl₃-EtOAc) of the products gave a diastereoisomeric mixture (1:0.6 by ¹⁹F NMR) of 17 as a syrup, 338 mg (74%), TLC (10:1 CHCl₃– EtOAc): R_f 0.3, $[\alpha]_D^{25}$ –103° (*c* 1, CHCl₃); ¹H NMR (CDCl₃) (signals not listed in Table 1): δ 4.78–4.90 (1 H, H-2'), 3.86–3.96 (2 H, H-5, 6'b), 3.3–3.6 (2 H, H-4, 6'a), 1.5–2.0 (6 H, H-3'a,b, 4'a,b, 5'a,b). Anal. Calcd for C₁₃H₂₃FO₅: C, 56.10; H, 8.33; F, 6.83. Found: C, 55.94; H, 8.27; F, 6.96.

1,3-Di-O-acetyl-2,6-dideoxy-2-fluoro-4-O-methyl- α -L-talopyranose (18).—A mixture of 17 (425 mg, 1.5 mmol), Ac₂O (4.2 mL), AcOH (4.2 mL), and H₂SO₄ (85 μ L, 1.6 μ mol) was kept for 1 h at 0 °C, then for 1.5 h at room temperature. The darkbrown solution resulted was poured into ice-cold aq NaHCO3 (saturated, 40 mL) and the mixture was stirred for 1.5 h. Extraction of the product with CHCl₃ and concentration gave a residue, which was chromatographed (30:1 CHCl₃–EtOAc) to give 18 as crystals, 260 mg (64%), mp 92–93 °C (prisms from EtOAc–hexane), $[\alpha]_D^{22} -97^\circ$ (*c* 1, CHCl₃). Anal. Calcd for C₁₁H₁₇FO₆: C, 50.00; H, 6.48; F, 7.19. Found: C, 50.12; H, 6.48; F, 7.20.

3-O-Acetyl-2,6-dideoxy-2-fluoro-4-O-methyl- α -Ltalopyranosyl bromide (**19**).—A mixture of **18** (230 mg, 0.87 mmol) and TiBr₄ (800 mg, 2.2 mmol) in dry CH₂Cl₂–EtOAc (10:1, 5 mL) was stirred for 16 h at room temperature. The deep-brown mixture was treated as described for **14** to give **19** as a paleyellow syrup, 246 mg (quant), which was used without further purification.

Methyl 4-O-benzyl-2,6-dideoxy-2-fluoro-3-O-trifluoromethylsulfonyl- α -L-idopyranoside (21).—To a cold (-20 °C) solution of 20 [4] (2.00 g, 7.4 mmol) in pyridine (3.6 mL)–CH₂Cl₂ (20 mL) was added (CF₃SO₂)₂O (1.9 mL, 11 mmol) and the solution was kept for 1 h at 0 °C. After addition of MeOH (10 mL) followed by dilution with CHCl₃, the solution was washed successively with aq 20% KHSO₄, aq NaHCO₃ (saturated), and water, dried (Na₂SO₄), and concentrated to give 21 as a paleyellow syrup, 2.83 g (95%) which was used without further purification; ¹⁹F NMR (CDCl₃): δ –194.7 (dddq, 1 F, F-2; collapsed to ddd by irradiation at δ –75.1), –75.1 (d, 3 F, J_{CF_3} F-2 6 Hz, CF₃).

Methyl 4-O-benzyl-2,3,6-trideoxy-2-fluoro-3-iodo- α -L-talopyranoside (22).—A mixture of 21 (2.83 g, 7.0 mmol) and NaI (5.28 g, 32 mmol) in DMF (29 mL) was stirred the dark place for 3 h at 90 °C. Dilution of the solution with CHCl₃ gave a precipitate, which was filtered and washed with CHCl₃. The filtrates combined were washed with aq 20% Na₂S₂O₃ and water, dried (Na₂SO₄), and concentrated with aid of xylene. Chromatography (50:1 toluene–EtOAc) of the residue gave **22** as an amorphous solid, 2.01 g (75%), $[\alpha]_{D}^{23}$ –74° (*c* 1, CHCl₃); ¹³C NMR (CDCl₃): δ 127.7, 128.1, 128.3, 137.4 (Ph), 98.3 (d, $J_{C-1,F}$ 31.3 Hz, C-1), 87.9 (d, $J_{C-2,F}$ 188.4 Hz, C-2), 78.9 (C-4), 76.3 (PhCH₂), 67.3 (C-5), 55.2 (OCH₃), 24.2 (d, $J_{C-3,F}$ 15.7 Hz, C-3). Anal. Calcd for C₁₄H₁₈FIO₃; C, 44.23; H, 4.77; F, 5.00; I, 33.38. Found: C, 44.56; H, 4.93; F, 4.84; I, 33.56.

Methyl 4-O-benzyl-2,3,6-trideoxy-2-fluoro- α -Llyxo-hexopyranoside (23).—A mixture of 22 (1.99 g, 5.2 mmol), Bu₃SnH (8.4 mL, 31.3 mmol), and 2,2'-azobisisobutyronitrile (43 mg, 0.26 mmol) in dry 1,4-dioxane was stirred for 1 h at 80 °C under the atmosphere of N₂. After concentration, the residue was chromatographed (1:1 toluene– hexane→CHCl₃) to give 23 as a syrup, 1.32 g (quant), TLC (50:1 toluene–EtOAc) R_f 0.25 (cf 22 : R_f 0.35), $[\alpha]_D^{23}$ –31° (c 2, CHCl₃). Anal. Calcd for C₁₄H₁₉FO₃: C, 66.12; H, 7.53; F, 7.47. Found: C, 65.85; H, 7.64; F, 7.61.

Methyl 2,3,6-trideoxy-2-fluoro-4-O-p-nitrobenzoyl- α -L-lyxo-hexopyranoside (25).—A solution of 23 (50 mg, 0.20 mmol) in 1,4-dioxane-H₂O-AcOH (10:1:1, 1 mL) was hydrogenated in the presence of Pd black under gentle bubbling of H_2 for 2h at room temperature. In TLC (3:1 hexane-acetone), the solution showed a single spot at $R_f 0.4$ of 24 (cf 23: R_f 0.5). After filtration followed by dilution with CH₂Cl₂, the organic solution was washed with aq NaHCO₃ (saturated), dried (MgSO₄), and concentrated. To the residue in CH₂Cl₂ (1.5 mL) pnitrobenzoyl chloride (110 mg, 0.59 mmol) and pyridine (0.7 mL) were added, and the mixture was stirred for 11h at room temperature. Water (0.05 mL) was added, and stirring was continued for 1 h. After addition of large amount of CHCl₃, the organic solution was washed successively with aq 20% KHSO₄, aq NaHCO₃ (saturated), and water, dried (Na_2SO_4), and concentrated to give 25 as needles (58 mg, 94% based on 23), mp 107- $108 \,^{\circ}\text{C}, \, [\alpha]_{\text{D}}^{23} - 56^{\circ} \, (c \ 1, \text{ CHCl}_3), \text{ IR (KBr): } 1720$ (C=O), 1530 cm⁻¹ (NO₂). Anal. Calcd for C₁₄H₁₆FNO₆: C, 53.68; H, 5.15; F, 6.06; N, 4.47. Found: C, 53.97; H, 5.12; F, 6.09; N, 4.44.

1-O-Acetyl-2,3,6-trideoxy-2-fluoro-4-O-p-nitrobenzoyl-α- (**26**) and -β-L-lyxo-hexopyranose (**27**).— A solution of **25** (420 mg, 1.3 mmol), Ac₂O (1.0 mL, 10.8 mmol), and H₂SO₄ (7.2 μ L) in dry MeNO₂ (9 mL) was kept for 1 h at room temperature. After pouring into cold water (13 mL) containing NaHCO₃ (3.6 g), the mixture was stirred for 1 h, and the products were extracted with CHCl₃. The solution was washed with water, dried (Na₂SO₄), and concentrated. Chromatography (50:1 CHCl₃–EtOAc) of the residue gave **26** as crystals, 356 mg (78%), TLC: R_f 0.35 (3:1 hexane-acetone) and **27** as crystals, 58 mg (13%), R_f 0.25. Compound **26**, mp 126–127.5 °C, $[\alpha]_D^{23}$ –62° (*c*, 1.5, CHCl₃). Anal. Calcd for C₁₅H₁₆FNO₇: C, 52.79; H, 4.73; F, 5.57; N, 4.10. Found: C, 52.96; H, 4.65; F, 5.30; N, 4.04. Compound **27**, mp 189.5–190.5 °C, $[\alpha]_D^{23}$ –5° (*c* 1, CHCl₃). Anal. Calcd for C₁₅H₁₆FNO₇: C, 52.79; H, 4.73; F, 5.57; N, 4.10. Found: C, 52.96; H, 4.63; F, 5.40; N, 3.83.

2,3,6-Trideoxy-2-fluoro-4-O-p-nitrobenzoyl- α -Llyxo-hexopyranosyl bromide (28).—A solution of 26 (329 mg, 0.97 mmol) in 30% HBr in AcOH (3.3 mL) was kept for 1.5 h at room temperature. CHCl₃ (60 mL) was added, and the solution was washed with cold aq NaHCO₃ (saturated) and water, dried (MgSO₄), and concentrated to give 28 as a pale-yellow syrup, 349 mg (quant), which was used without purification; TLC (3:1 hexane–acetone) R_f 0.45.

Methyl 4-O-benzyl-2,3,6-trideoxy-2,3-difluoro-a-L-talopyranoside (29).—A mixture of 21 (2.70 g, tris(dimethylamino)sulfonium $6.7 \,\mathrm{mmol}$ and difluorotrimethylsilicate [19] (6.67 g, 24.2 mmol) in dry CH₂Cl₂ (54 mL) was stirred for 15 h under an atmosphere of N₂ at room temperature. After dilution with CHCl₃, the solution was washed with aq NaHCO₃ (saturated) and water, dried (Na₂SO₄), and concentrated. The residue was chromatographed (30:1 toluene–EtOAc) to give 29 as a solid, 1.20 g (66%). Recrystallization from CHCl₃-hexane gave prisms, mp 51.5–52.5 °C, $[\alpha]_{D}^{25}$ –41° (c 1, CHCl₃). Anal. Calcd for C₁₄H₁₈F₂O₃: C, 61.75; H, 6.66; F, 13.95. Found: C, 61.84; H, 6.85; F, 13.75.

1,4-Di-O-acetyl-2,3,6-trideoxy-2,3-difluoro-α- (**31**) and -β-L-talopyranose (**32**).—A solution of **29** (1.09 g, 4.0 mmol) in 1,4-dioxane–H₂O–AcOH (10:1:1, 22 mL) was hydrogenated in the presence of Pd black under gentle bubbling of H₂ for 3.5 h at room temperature. After filtration, the filtrate diluted with CH₂Cl₂ was washed with aq NaHCO₃ (saturated) and water, dried (MgSO₄), and concentrated to give **30** as a syrup, 0.63 g (86%), TLC (30:1 CHCl₃–EtOAc) R_f 0.3 (cf **29**: R_f 0.6). To an ice-cold solution of **30** (0.62 g, 3.4 mmol) in AcOH (6.2 mL) were added Ac₂O (9.3 mL) and H₂SO₄ (0.06 mL), and the solution was kept for 4 h at room temperature. Processing as described for **12** followed by chromatography (30:1 CHCl₃–EtOAc) gave the α-L anomer **31** as prisms, 0.44 g (52%) and the β-L anomer **32** as plates, 0.17 g (20%). Compound **31**, TLC (30:1 CHCl₃–EtOAc) R_f 0.3, mp 143.5-144.5 °C, $[\alpha]_D^{25}$ –120° (*c* 1, CHCl₃). Anal. Calcd for C₁₀H₁₄F₂O₅: C, 47.62; H, 5.60; F, 15.07. Found: C, 47.76; H,5.80; F, 14.79. Compound **32**, TLC (30:1 CHCl₃–EtOAc) R_f 0.2, mp 108–109 °C, $[\alpha]_D^{23}$ –20° (*c* 1, CHCl₃). Anal. Calcd for C₁₀H₁₄F₂O₅: C, 47.62; H, 5.60; F, 15.07. Found: C, 47.43; H, 5.73; F, 14.83.

4-O-Acetyl-2,3,6-trideoxy-2,3-difluoro-α-L-talopyranosyl bromide (**33**).—A solution of **31** (320 mg, 1.3 mmol) in 30% HBr in AcOH (3.2 mL) was kept for 2 h at room temperature. Working up as described for **28** gave **33** as crystals, 346 mg (quant). Analytical samples were prepared by recrystallization (CHCl₃-hexane) as prisms, mp 57.5–59 °C, $[\alpha]_D^{19}$ –260° (*c* 1, CHCl₃). Anal. Calcd for C₈H₁₁BrF₂O₃: C, 35.19; H, 4.06; Br, 29.26; F, 13.91. Found: C, 35.46; H, 4.30; Br, 28.99; F, 13.99.

Methyl 3-O-*acetyl*-2,6-*dideoxy*-2-*fluoro*-α-L*idopyranoside* (**35**).—A solution of **34** [4] (108 mg, 0.35 mmol) in 1,4-dioxane–H₂O–AcOH (10:1:1, 2.5 mL) was hydrogenated as described for **11** to give **35** as a pale yellow syrup, 77 mg (quant), $[\alpha]_{D}^{23}$ -104° (*c* 1.8, CHCl₃). Anal. Calcd for C₉H₁₅FO₅: C, 48.65; H, 6.80; F, 8.55. Found: C, 48.69; H, 6.86; F, 8.85.

Methyl 3-O-acetyl-2,6-dideoxy-2-fluoro-4-O-tetrahydropyran-2-yl- α -L-idopyranoside (36).—A solution of 35 (71 mg, 0.32 mmol), 3,4-dihydro-2Hpyran (88 μ L, 0.96 mmol), and pyridinium *p*-toluenesulfonate (8 mg, 0.03 mmol) in dry CH₂Cl₂ (1.5 mL) was kept overnight at room temperature. After working up as described for 15, the products were chromatographed (10:1 toluene-EtOAc) to give a diastereoisomeric mixture (1:2 by ¹⁹F NMR) of **36** as a syrup, 95 mg (97%), $[\alpha]_D^{23} - 92^\circ$ (*c* 3, CHCl₃); ¹H NMR (CDCl₃) (only selected signals): δ 5.33 and 5.24 (each dt, 2:1 in strength, 1 H in total, H-3), 4.35 (ddd, 0.67 H, H-2), 3.444 and 3.436 (each s, 3 H in total, OMe), 2.10 (s, 3 H, Ac), 1.38 and 1.28 (each d, 1:2 in strength, 3 H in total, Me-5); $J_{1,2}$ 3.5, $J_{2,3} = J_{3,4}$ 5.5 Hz. ¹⁹F NMR (CDCl₃): δ –194.8 (ddd, 0.33 F, J_{1,F} 10.5, J_{2,F} 46.5, J_{3,F} 12.5 Hz), -195.9 (dt, 0.67 F, $J_{1,F} = J_{3,F}$ 11.5, $J_{2,F}$ 47 Hz). Anal. Calcd for C₁₄H₂₃FO₆: C, 54.89; H, 7.57; F, 6.20. Found: C, 55.25; H, 7.69; F, 6.19.

Methyl 2,6-dideoxy-2-fluoro-4-O-tetrahydropyran-2-yl- α -L-idopyranoside (37).—Zemplén deacetylation of 36 (122 mg, 0.40 mmol) in MeOH (0.1 mL) gave, after conventional post-treatment, **37** as a pale-yellow syrup, 103 mg (98%), $[\alpha]_D^{23}$ -89° (*c* 3, CHCl₃); ¹H NMR (CDCl₃) (only selected signals): δ 3.47 and 3.46 (each s, 3 H in total, OMe), 1.37 and 1.28 (each d, 1:2 in strength, 3 H in total, $J_{5,6}$ 6.5 Hz, Me-5). ¹⁹F NMR (CDCl₃): δ -196.2 (br dt, 0.33 F, $J_{1,F} = J_{3,F} \sim 10$, $J_{2,F}$ 47 Hz), -197.9 (ddd, 0.67 F, $J_{1,F}$ 9.5, $J_{2,F}$ 49.5, $J_{3,F}$ 13 Hz). Anal. Calcd for C₁₂H₂₁FO₅: C, 54.53; H, 8.01; F, 7.19. Found: C, 54.36; H, 7.96; F, 7.47.

Methyl 2,3,6-trideoxy-2-fluoro-3-iodo-4-O-p-nitrobenzoyl-a-L-talopyranoside (40).-To an ice-cold solution of 37 (315 mg, 1.2 mmol) in pyridine (0.6 mL, 7.3 mmol)-CH₂Cl₂ (4.4 mL) was added (CF₃SO₂)₂O (0.3 mL, 2.1 mmol) and the solution was kept for 1 h in the cold. MeOH (0.4 mL) was added, and after dilution with CHCl₃, the solution was washed successively with cold aq 20% KHSO₄, aq NaHCO₃ (saturated), and water, dried (Na_2SO_4) , and concentrated to give **38** (442 mg, 94%) as an unstable syrup, TLC (4:1 toluene-EtOAc) R_f 0.65. A mixture of the syrup (430 mg, 1.1 mmol) and NaI (820 mg, 5.5 mmol) in dry DMF (4.5 mL) was stirred for 3 h at 90 °C. In TLC (4:1 toluene-EtOAc), the mixture showed a main spot at R_f 0.5 (39). After cooling to room temperature. *p*-nitrobenzoyl chloride (610 mg. 3.3 mmol) and 4-dimethylaminopyridine (400 mg, 3.3 mmol) were added, and the mixture was stirred for 2.5 h at room temperature. Water (0.2 mL) was added, and after dilution with CHCl₃, the organic solution was washed successively with aq 20% Na₂S₂O₃, aq 20% KHSO₄, aq NaHCO₃ (saturated), and water, dried (Na₂SO₄), and concentrated. Chromatography (toluene) of the residue gave 40 as an amorphous solid, 276 mg (58% based on 38), TLC (4:1 toluene–EtOAc) R_f 0.7, $[\alpha]_{D}^{22}$ -117° (c 1.8, CHCl₃). ¹³C NMR (CDCl₃): δ 164.2 (C=O), 150.9, 134.8, 131.5, 123.7 (Ph), 98.1 (d, J_{C-1.F} 32.8 Hz, C-1), 87.7 (d, J_{C-2.F} 183.2 Hz, C-2), 72.8 (C-4), 65.8 (C-5), 55.5 (OCH₃), 21.1 (d, J_{C-3,F} 18.6 Hz, C-3), 17.3 (Me-5). Anal. Calcd for C₁₄H₁₅FINO₆: C, 38.29; H, 3.44; F, 4.33; I, 28.89; N, 3.19. Found: C, 38.44; H, 3.54; F, 4.40; I, 28.95; N, 3.16.

1-O-*Acetyl*-2,3,6-*trideoxy*-2-*fluoro*-3-*iodo*-4-O-p*nitrobenzoyl*-α-L-*talopyranose* (**41**).—A solution of **40** (170 mg, 0.39 mmol), Ac₂O (0.29 mL, 3.1 mmol), and H₂SO₄ (7 μL) in dry MeNO₂ (3.8 mL) was kept for 2 h at room temperature. Working up as described for **26** gave a solid, which was chromatographed (20:1 toluene–EtOAc) to give **41** as an amorphous solid, 130 mg (72%), $[\alpha]_{\rm D}^{22}$ –147° (*c* 2.7, CHCl₃). Anal. Calcd for C₁₅H₁₅FINO₇: C, 38.56; H, 3.24; F, 4.07; I, 27.16; N, 2.30. Found: C, 38.64; H, 3.49; F, 3.67; I, 27.33; N, 2.52.

2,3,6-Trideoxy-2-fluoro-3-iodo-4-O-p-nitrobenzoyl- α -L-talopyranosyl bromide (42).—A solution of 41 (200 mg, 0.43 mmol) in 30% HBr in AcOH (2 mL) was kept for 2 h at room temperature. Working up as described for 28 gave 42 as an amorphous solid, 209 mg (99%), TLC (20:1 toluene–EtOAc) R_f 0.7 (cf 41: R_f 0.3), $[\alpha]_D^{22}$ –223° (c 1.5, CHCl₃). Anal. Calcd for C₁₃H₁₂BrFINO₅: C, 31.99; H, 2.48; F, 3.89; N, 2.87. Found: C, 31.77; H, 2.64; F, 3.59; N, 2.58.

7-O-(4-O-Acetyl-2,6-dideoxy-2-fluoro-3-O-methyl- α - (43) and - β -L-talopyranosyl) daunomycinone (44). A mixture of daunomycinone (349 mg, 0.88 mmol), yellow HgO (1.26 g, 5.8 mmol), HgBr₂ (360 mg, 1.0 mmol), and powdered molecular sieves 3A $(3.34 \text{ g}, \text{ activated at } 350 \,^{\circ}\text{C} \text{ under a stream of } N_2)$ in dry CH_2Cl_2 (58 mL) was stirred for 30 min at room temperature. After 14 (304 mg, 1.1 mmol) in dry CH_2Cl_2 (2.4 mL) was added, the mixture was refluxed for 17h in a dark place. In TLC (10:1 CHCl₃-acetone), the mixture showed spots at R_f 0.33 (43, major), 0.22 (daunomycinone, slight), and 0.14 (44, minor) along with several minor spots. The mixture was filtered through a Celite bed, the bed being washed repeatedly with CHCl₃. The filtrate and washings combined were washed with aq 30% KI, aq NaHCO₃ (saturated), and water, dried (Na₂SO₄), and concentrated. Triplicate chromatographies (10:1 CHCl₃-acetone \rightarrow 1:1 CHCl₃-EtOAc \rightarrow 1:1 toluene–EtOAc) of the residue gave 43, 206 mg (38%) and 44, 42 mg (8%) as reddishorange solids. Analytical samples were prepared by reprecipitation from CHCl₃-hexane. Compound **43**, $[\alpha]_{D}^{24}$ +175° (*c* 0.02, CHCl₃); ¹⁹F NMR (CDCl₃): δ -202.2 (ddd). Anal. Calcd for C₃₀H₃₁ FO₁₂·0.5 H₂O: C, 58.92; H, 5.27; F, 3.11. Found: C, 59.19; H, 5.07; F, 2.70. Compound 44, $[\alpha]_{D}^{26} + 426^{\circ}$ $(c 0.02, CHCl_3)$; ¹⁹F NMR (CDCl_3): δ –220.9 (ddd). Anal. Calcd for C₃₀H₃₁FO₁₂·0.5 H₂O: C, 58.92; H, 5.27; F, 3.11. Found: C, 58.62; H, 5.10; F, 2.98.

7-O-(2,6-Dideoxy-2-fluoro-3-O-methyl- α -L-talopyranosyl)daunomycinone (3).—To an ice-cold solution of **43** (53 mg, 0.09 mmol) in 2:1 CHCl₃-MeOH (6.3 mL) was added 0.3 M methanolic NaOH (2.1 mL), and the mixture was stirred for 4 h at 0 °C under the atmosphere of N₂. After neutralization of the deep-purple mixture with aq 0.5 M HCl, the mixture was extracted with CHCl₃. The extract was washed with water, dried (Na₂SO₄), and concentrated. Chromatography of the residue (6:1 CHCl₃–acetone) gave **3** as a reddish-orange solid, 34 mg (69%), TLC (6:1 CHCl₃– acetone) R_f 0.3, $[\alpha]_D^{24}$ +176° (*c* 0.02, CHCl₃); ¹⁹F NMR (CDCl₃): δ –199.6 (ddt; $J_{OH-4',F} \sim 9$ Hz; the signals collapsed to ddd on addition of D₂O). Anal. Calcd for C₂₈H₂₉FO₁₁·0.5 H₂O: C, 59.05; H, 5.31; F,3.34. Found: C, 59.29; H, 5.23; F, 3.39.

7-O-(3-O-Acetyl-2,6-dideoxy-2-fluoro-4-O-methylα-L-talopyranosyl) daunomycinone (45).—A mixture of daunomycinone (292 mg, 0.73 mmol), yellow 3.3 mmol), HgBr₂ $(212 \,\mathrm{mg},$ HgO (713 mg, 0.59 mmol), and powdered molecular sieves 3A (2.84 g) in dry CH₂Cl₂ (48 mL) was stirred for 30 min at room temperature. A solution of 19 (254 mg, 0.89 mmol) in dry CH_2Cl_2 (1.9 mL) was added, and the mixture was refluxed for 19 h in the dark place. In TLC (15:1 CHCl₃-acetone), the mixture showed spots at R_f 0.26 (45, major), 0.18 (trace), 0.13 (slight), and 0.09 (slight). Working up as described for 43 gave a solid, which was doubly chromatographed (15:1 CHCl₃-acetone) to give 45 as a reddish-orange solid, 220 mg (50%). An analytical sample was prepared by reprecipitation from CHCl₃-Et₂O-hexane, $[\alpha]_{D}^{22} + 251^{\circ}$ (c, 0.02, CHCl₃); ¹⁹F NMR (CDCl₃): δ –201.3 (ddd). Anal. Calcd for C₃₀H₃₁FO₁₂: C, 59.80; H, 5.19; F, 3.15. Found: C, 59.89; H, 5.13; F, 2.98.

7-O-(2,6-Dideoxy-2-fluoro-4-O-methyl-α-L-talopyranosyl)daunomycinone (4).—To an ice-cold solution of 45 (115 mg, 0.19 mmol) in oxolane (7.3 mL) was added aq 1 M NaOH (1.7 mL), and the mixture was stirred for 1 h in the cold. The resultant deep-purple solution was neutralized with aq 1 M HCl, concentrated to a small volume, and after addition of CHCl₃, the solution was washed with aq 20% NaCl, dried (Na₂SO₄), and concentrated. The residue was passed through a short column of silica gel with 5:1 CHCl3-acetone giving 4 as a reddish-orange solid, 103 mg (96%). An analytical sample was prepared by reprecipitation from CHCl₃-Et₂O-hexane, TLC (5:1 CHCl₃-acetone): R_f 0.6, $[\alpha]_D^{22}$ +159° (*c*, 0.02, CHCl₃); ¹⁹F NMR (CDCl₃): δ -204.2 (slightly br ddd, which sharpened on addition of D₂O). Anal. Calcd for C₂₈H₂₉FO₁₁·0.5 H₂O: C, 59.05; H, 5.31; F, 3.34. Found: C, 58.96; H, 5.19; F, 3.35.

7-O-(2,6-Dideoxy-2-fluoro-4-O-methyl- α -L-talopyranosyl)adriamycinone (5).—To a solution of 4 (83 mg, 0.15 mmol) and HC(OMe)₃ (0.11 mL, 1.0 mmol) in dry 2:3 MeOH–1,4-dioxane (5 mL) was added Br₂ (35 mg, 0.22 mmol) in CH₂Cl₂

(0.35 mL), and the solution was kept for 0.5 h at 0° C, and then for 3h at room temperature. TLC (2:1 benzene-acetone) of the solution showed a main spot at $R_f 0.35$. Addition of diisopropyl ether followed by hexane gave a precipitate, which was collected and washed with hexane to give a red solid (mainly the 14-bromo-13-dimethylacetal derivative). A suspension of the solid in acetone (15 mL) was shaken for 1.5 h at room temperature, and concentrated to give a dark-red solid (mainly the deacetalated 14-bromo derivative). A mixture of the solid and HCO₂Na (169 mg, 2.5 mmol) in 4:1 acetone-H₂O (10 mL) was stirred vigorously for 15 h at room temperature, whereupon, in TLC (2:1 benzene-acetone), two spots of $R_f 0.28$ (the 14-Oformyl derivative of 5) and 0.15 (5) appeared. After concentration to a small volume followed by addition of water, the products were extracted with CHCl₃. The organic solution was washed with water, dried (Na_2SO_4) , and concentrated. A suspension of the dark-red solid (83 mg) in 4:4:1 CHCl₃–MeOH–aq 1 M NH₃ (9 mL) was shaken for 45 min at 0 °C, when a substance with R_f 0.28 disappeared and 5 became the major one. After dilution with water, products were extracted with CHCl₃, and the solution was washed with aq 0.05 M HCl and aq 20% NaCl, dried (Na₂SO₄), and concentrated. The residue was chromatographed on a short column with 3:2 CHCl₃-acetone to give 5 as a red solid, 64 mg (75%). An analytical sample was prepared by reprecipitation from CHCl₃-MeOH-Et₂O-hexane, $[\alpha]_{D}^{23} + 155^{\circ}$ (c 0.02, CHCl₃); ¹⁹F NMR (CDCl₃): δ –204.0 (ddd). Anal. Calcd for C₂₈H₂₉FO₁₂·0.25 H₂O: C, 57.88; H, 5.12; F, 3.27. Found: C, 57.91; H, 5.19; F, 2.93.

7-O-(2,3,6-Trideoxy-2-fluoro-4-O-p-nitrobenzoyl- α - (**46**) and - β -L-lyxo-hexopyranosyl) daunomycinone (47). A mixture of 28 (349 mg, 0.97 mmol), daunomycinone (298 mg, 0.73 mmol), yellow HgO (729 mg, 3.37 mmol), HgBr₂ (216 mg, 0.60 mmol), and powdered molecular sieves 3A (1.2g) in dry CH₂Cl₂ (65 mL) was refluxed under stirring for 17h in a dark place. Post-treatment as described for 43 gave a residue, which showed, in TLC (30:1 CHCl₃-acetone), three spots at $R_f 0.03$ (daunomycinone), 0.15 (47), and 0.2 (46). Chromatography (10:1 CHCl₃-EtOAc) of the residue gave 46, 217 mg (43%) and 47, 207 mg (41%) as red solids. Compound 46, $[\alpha]_{D}^{23} - 119^{\circ}$ (c 0.1, CHCl₃). ¹⁹F NMR (CDCl₃): δ –184.3 (ddt). Anal. Calcd for C₃₄H₃₀FNO₁₃·H₂O: C, 58.54; H, 4.62; F, 2.72; N, 2.01. Found: C, 58.33; H, 4.43; F, 3.05; N, 1.76.

Compound **47**, $[\alpha]_{D}^{23}$ +601° (*c* 0.1, CHCl₃). ¹⁹F NMR (CDCl₃): δ -205.4 (ddd). Anal. Calcd for C₃₄H₃₀FNO₁₃·H₂O: C, 58.54; H, 4.62; F, 2.72; N, 2.01. Found: C, 58.84; H, 4.49; F, 2.92; N, 1.87.

7-O-(2,3,6-Trideoxy-2-fluoro-α-L-lyxo-hexopyranosyl)daunomycinone (6).—A mixture of **46** (15 mg, 0.022 mmol) in 0.2 M NaOH in oxolane–H₂O (4:1, 1.9 mL) was stirred for 15 h at 0 °C. Conventional work-up as described for **3** gave a solid, which was chromatographed (15:1 CHCl₃–acetone) and purified by reprecipitation from CHCl₃–Et₂O–hexane gave **6** as a red solid, 10.1 mg (76%), $[\alpha]_D^{23} + 72^\circ$ (*c* 0.06, CHCl₃). ¹⁹F NMR (CDCl₃): δ –180.8 (dddd; $J_{OH-4',F}$ 11 Hz). Anal. Calcd for C₂₇H₂₇FO₁₀·0.5 H₂O: C, 60.11; H, 5.23; F, 3.52. Found: C, 60.23; H, 5.18; F, 3.21.

7-O-(4-O-Acetyl-2,3,6-trideoxy-2,3-difluoro-a-(48) and $-\beta$ -L-talopyranosyl) daunomycinone (49).—A mixture of 33 (296 mg, 1.1 mmol), daunomycinone 1.1 mmol), yellow (424 mg, HgO (693 mg, 3.2 mmol), HgBr₂ (194 mg, 0.54 mmol), and powdered molecular sieves 3A (2.31 g) in dry CH₂Cl₂ (71 mL) was refluxed under stirring for 19h in a dark place. Additional yellow HgO (700 mg, 3.2 mmol), HgBr₂ (145 mg, 0.40 mmol), and powdered molecular sieves 3A (567 mg) were added and the refluxing was continued for further 21 h. Post-treatment as described for 43 gave a red solid, which showed, in TLC (6:1 toluene-acetone), three spots at R_f 0.15 (daunomycinone), 0.25 (49), and 0.4 (48). Chromatography (6:1 toluene-acetone) of the solid gave 48, 175 mg (28%) and 49, 99 mg (16%). An analytical sample of **48** was prepared by recrystallization from CHCl₃-acetone-Et₂O as red needles, and the sample of 49 by reprecipitation from CHCl₃-Et₂O as a red solid. Compound 48, mp 222.5–225 °C, $[\alpha]_{D}^{24}$ + 85° (c 0.02, CHCl₃). ¹⁹F NMR (CDCl₃): δ -206.1 (br ddq, 1 F, $J_{1',F-3'} = J_{2',F-3}$ $_{3'} = J_{4',F-3'} \sim 6, J_{3',F-3'} 43, J_{F-2',F-3'} 14 \text{ Hz F-3'}, -203.9$ (dddd, 1 F, F-2'). Anal. Calcd for $C_{29}H_{28}F_2O_{11}\cdot 0.5$ H₂O: C, 58.10; H, 4.88; F, 6.34. Found: C, 58.16; H, 4.71; F, 6.24. Compound **49**, $[\alpha]_{D}^{22} + 431^{\circ}$ (c 0.02, CHCl₃), ¹⁹F NMR (CDCl₃): δ –221.4 (br dddd, 1 F; $J_{\text{F-2',F-3'}}$ 13 Hz, F-2'), -202.4 (br dddd, 1 F, $J_{2',\text{F-3'}}$ 7.5, $J_{3',F-3'}$ 43, $J_{4',F-3'}$ 5, $J_{F-2',F-3'}$ 13 Hz, F-3'). Anal. Calcd for C₂₉H₂₈F₂O₁₁·0.5 H₂O: C, 58.10; H, 4.88; F, 6.34. Found: C, 58.28; H, 5.03; F, 6.42.

7-O-(2,3,6-Trideoxy-2,3-difluoro- α -L-talopyranosyl)daunomycinone (7).—To a cold solution of **48** (30 mg, 0.05 mmol) in 3:2 CHCl₃–MeOH (1.2 mL) was added 0.3 M NaOH in MeOH (1.5 mL) and the mixture was stirred for 3 h at -5 °C under an atmosphere of N₂. Working up as described for **3** gave a red solid, which was chromatographed (7:1 CHCl₃-acetone) to give **7**, 22 mg (78%), $[\alpha]_{D}^{24}$ + 111° (*c* 0.02, CHCl₃). ¹⁹F NMR (CDCl₃): δ -205.2 (br ddq, 1 F, $J_{1',F-3'}=J_{2',F-3'}=J_{4',F-3'}$ 6, $J_{3',F-3'}$ 43.5, $J_{F-2',F-3'}$ 16Hz, F-3'), -200.8 (br dddt, 1 F, $J_{OH-4',F-2'}$ 9.5, $J_{F-2',F-3'}$ 16Hz, F-2'). Anal. Calcd for C₂₇H₂₆F₂O₁₀·0.25 H₂O: C, 58.64; H, 4.83; F, 6.87. Found: C, 58.72; H, 4.82; F, 7.16.

7-O-(2,3,6-Trideoxy-2-fluoro-3-iodo-4-O-p-nitrobenzoyl- α - (50) and - β -L-talopyranosyl)dauno*mycinone* (51).—A mixture of 42 (209 mg, 0.43 mmol), daunomycinone (142 mg, 0.36 mmol), yellow HgO (348 mg, 1.6 mmol), HgBr₂ (103 mg, 0.29 mmol), and powdered molecular sieves 3A (568 mg) in dry CH₂Cl₂ (29 mL) was refluxed for 14h in the dark place. Additional yellow HgO (348 mg, 1.6 mmol) and HgBr₂ (103 mg, 0.29 mmol) were added and the refluxing was continued for further 24 h. Working up as described for 43 gave a red solid, which was chromatographed (30:1 CHCl₃-acetone) to give 50, 63 mg (22%), R_f 0.35 in TLC (30:1 CHCl₃-acetone), and 51, 178 mg (62%), R_f 0.25. Compound **50**, $[\alpha]_D^{22} + 20^\circ$ (c 0.1, CHCl₃). ¹⁹F NMR (CDCl₃): δ –183.4 (ddd). Anal. Calcd for C₃₄H₂₉FINO₁₃·H₂O: C, 49.59; H, 3.79; F, 2.31; I, 15.41; N, 1.70. Found: C, 49.75; H, 3.73; F, 2.66; I, 15.63; N, 1.78. Compound **51**, $[\alpha]_{D}^{22}$ $+427^{\circ}$ (c 0.1, CHCl₃). ¹⁹F NMR (CDCl₃): δ -203.7 (ddd). Anal. Calcd for C₃₄H₂₉FINO₁₃·1.5 H₂O: C, 49.05; H, 3.87; F, 2.28; I, 15.24; N, 1.68. Found: C, 48.96; H, 3.62; F, 2.54; I, 15.18; N, 1.82.

7-O-(2,3,6-Trideoxy-2-fluoro-3-iodo- α -L-talopyranosyl)daunomycinone (8).—A mixture of 50 (33 mg, 0.04 mmol) in 0.1 M NaOH in oxolane– H₂O (1:1, 3.3 mL) was stirred for 19 h at 0 °C. Working up as described for **3** gave a red solid, which was chromatographed (20:1 CHCl₃–acetone) and reprecipitated from CHCl₃–Et₂O–hexane to give **8**, 18 mg (68%), $[\alpha]_D^{23} + 203^\circ$ (*c* 0.1, CHCl₃). ¹⁹F NMR (CDCl₃): δ –180.6 (apparently ddt; *J*_{OH-4',F} 9 Hz). Anal. Calcd for C₂₇H₂₆FIO₁₀·H₂O: C, 48.09; H, 4.18; F, 2.82; I, 18.82. Found: C, 48.05; H, 4.21; F. 3.18; I, 18.49.

Acknowledgements

The authors are grateful to the Pharmaceutical Research Center of Meiji Seika Co., Ltd. for carrying out the bioassay for Table 6. We also express deep thanks to Dr. Tomio Takeuchi (the director), Ms. Chisato Nosaka, and Keiko Komuro of Institute of Microbial Chemistry (IMC) for antitumor assay for Table 5, and to Ms. Hiroko Hino of IMC for elemental analysis.

References

- Reviews: (a) F. Arcamone, The Development of New Antitumour Anthracyclines, in J.M. Cassady and J.D. Douros (Eds.) Anticancer Agents Based on Natural Product Models, Academic Press, New York, 1980, pp 1–41; (b) F. Arcamone, Doxorubicin, Academic Press, New York, 1981; (c) H.S. El Khadem (Ed.), Anthracycline Antibiotics, Academic Press, New York, 1982; (d) J.W. Lown (Ed.), Anthracycline and Anthracenedione-Based Anticancer Agents, Elsevier, Amsterdam, 1988; (e) A. Suarato, F. Angelucci and A. Bargiotti, Chimicaoggi, 8 (1990) 9–19; (f) W. Priebe (Ed.), Anthracycline Antibiotics, ACS Symposium Series. 574, Am. Chem. Soc., Washington, DC, 1995.
- [2] (a) J. Igarashi and M. Sunagawa, Bioorg. Med. Chem. Lett., 5 (1995) 2923-2928; (b) N. Aligiannis, N. Pouli, P. Marakos, A.-L. Skaltsounis, S. Leonce, A. Pierre, and G. Atassi, Bioorg. Med. Chem. Lett., 6 (1996) 2473-2476; (c) F. Kratz, U. Beyer, P. Schumacher, M. Krüger, H. Zahn, T. Roth, H.H. Fiebig, and C. Unger, Bioorg. Med. Chem. Lett., 7 (1997) 617-622; (d) T. Kunnari, J. Niemi, K. Ylihonko, P. Mäntsälä, and J. Hakala, Bioorg. Med. Chem. Lett., 7 (1997) 725-726; (e) B.A. Schweitzer and T.H. Koch, J. Am. Chem. Soc., 115 (1993) 5440-5445; (f) A. Cherif and D. Farquhar, J. Med. Chem., 35 (1992) 3208-3214; (g) V.M. Vrudhula, H.P. Svensson, and P.D. Senter, J. Med. Chem., 38 (1995) 1380-1385; (h) J.B. Chaires, F. Leng, T. Przewloka, I. Fokt, Y.-H. Ling, R. Perez-Solar, and W. Priebe, J. Med. Chem., 40 (1997) 261-266; (i) G. Capranico, R. Supino, M. Binaschi, L. Capolongo, M. Grandi, A. Suarato, and F. Zunino, Mol. Pharmacol., 45 (1994) 908–915; (j) A. Nagy, P. Armatis, and A.V. Schally, Proc. Natl. Acad. Sci. USA, 93 (1996) 2464–2469; (k) F. Pasqui, F. Canfarini, A. Giolitti, A. Guidi, V. Pestellini, and F. Arcamone, *Tetrahedron*, 52 (1996) 185–198; (1) S. Castillon, A. Dessinges, R. Faghih, G. Lukacs, A. Olesker, and T.T. Thang, J. Org. Chem., 50 (1985) 4913–4917; (m) H.H. Baer and L. Siemsen, Can. J. *Chem.*, 66 (1988) 187–190; (n) Y. Takagi, H. Park, T. Tsuchiya, S. Umezawa, T. Takeuchi, K. Komuro, and C. Nosaka, J. Antibiot., 42 (1989) 1315-1317; (o) H.H. Baer and F. Hernández Mateo, Can. J. Chem., 68 (1990) 2055-2059.
- [3] T. Tsuchiya, Y. Takagi, K. Ok, S. Umezawa, T. Takeuchi, N. Wako, and H. Umezawa, J. Antibiot., 39 (1986) 731–733.

- [4] K. Ok, Y. Takagi, T. Tsuchiya, S. Umezawa, and H. Umezawa, *Carbohydr. Res.*, 169 (1987) 69–81.
- [5] (a) E.-F. Fuchs, D. Horton, and W. Weckerle, *Carbohydr. Res.*, 57 (1977) C36–C39; (b) D. Horton, W. Priebe, and O. Varela, *J. Antibiot.*, 37 (1984) 853–858.
- [6] (a) M.-N. Borrel, M. Fiallo, W. Priebe, and A. Garnier-Suillerot, *FEBS Lett.*, 356 (1994) 287–290;
 (b) L. Lothstein, T.W. Sweatman, and W. Priebe, *Bioorg. Med. Chem. Lett.*, 5 (1995) 1807–1812.
- [7] T. Tsuchiya, Y. Takagi, S. Umezawa, T. Takeuchi, K. Komuro, C. Nosaka, H. Umezawa, S. Fukatsu, and T. Yoneta, J. Antibiot., 41 (1988) 988–991.
- [8] T. Tsuruo, K. Yusa, Y. Sudo, R. Takamori, and Y. Sugimoto, *Cancer Res.*, 49 (1989) 5537–5542.
- [9] J.A. Endicott and V. Ling, Annu. Rev. Biochem., 58 (1989) 137–171.
- [10] A.B. Shapiro and V. Ling, J. Biol. Chem., 270 (1995) 16167–16175.
- [11] T. Tsuruo, H. Ida, H. Kawabata, S. Tsukagoshi, and Y. Sakurai, *Cancer Res.*, 44 (1984) 5095–5099.
- [12] E. Pereira, E. Teodori, S. Dei, F. Gualtieri, and A. Garnier-Suillerot, *Biochem. Pharmacol.*, 50 (1995) 451–457.

- [13] T. Tsuruo, H. Ida, S. Tsukagoshi, and Y. Sakurai, *Cancer Res.*, 42 (1982) 4730–4733.
- [14] M. Bakker, W.T.A. van der Graaf, H.J.M. Groen, E.F. Smit, and E.G.E. De Vries, *Curr. Pharm. Des.*, 1 (1995) 133–144.
- [15] H.S. Mülder, H. Dekker, H.M. Pinedo, and J. Lankelma, *Biochem. Pharmacol.*, 50 (1995) 967–974.
- [16] Brief summary: (a) Y. Takagi, T. Tsuchiya, T. Miyake, T. Takeuchi, and S. Umezawa, J. Synthetic Org. Chem., Jpn., 50 (1992) 131–142; (b) T. Tsuchiya and Y. Takagi, Synthesis and Biological Activities of Florinated Daunorubicin and Doxorubicin Analogues, in W. Priebe (Ed.), Anthracycline Antibiotics, ACS Symposium Series 547, Am. Chem. Soc., Washington DC, 1995. pp 100–114.
- [17] Y. Kimura, M. Suzuki, T. Matsumoto, R. Abe, and S. Terashima, *Bull. Chem. Soc. Jpn*, 59 (1986) 423–431.
- [18] F. Arcamone, L. Bernardi, P. Giardino, and A. Di Marco, Ger. offen. 2.652, 391, May 26, 1977; *Chem. Abstr.*, 87 (1977) 152, 522f.
- [19] W. J. Middleton, Org. Synth., 64 (1985) 221-225.
- [20] A. Maeda, K. Yazawa, Y. Mikami, M. Ishibashi, and J. Kobayashi, J. Antibiot., 45 (1992) 1848–1852.