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MMP-13 selective isonipecotamide α -sulfone hydroxamates

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ABSTRACT

A series of *N*-aryl isonipecotamide α -sulfone hydroxamate derivatives has been prepared utilizing a combination of solution-phase and resin-bound library technologies to afford compounds that are potent and highly selective for MMP-13.

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Matrix metalloproteinases (MMPs) are zinc-dependent enzymes that are responsible for remodeling and degradation of all components of the extracellular matrix,^{1,2} yet excessive activity of MMPs has been implicated in numerous disease states including cancer,^{3,4} arthritis,⁵ and cardiovascular disease.^{6–9} MMP inhibitors (MMPi's) have therefore been explored as therapeutic treatments to halt progression of various diseases.^{10–12} The MMP family of enzymes includes at least 24 distinct mammalian isozymes, but MMP-13 in particular has been identified as a significant target since its upregulation has been implicated in cancer, osteoarthritis, and cardiovascular disease.

Treatment of patients with broad-spectrum MMPi's gives rise to stiffening of the joints referred to as musculoskeletal syndrome⁹ (MSS). Inhibition of MMP-1 has been hypothesized to be the cause of MSS observed clinically with broad-spectrum inhibitors, and the broad-spectrum inhibitor marimastat induces musculoskeletal side effects in rats.¹³ MMP-1 has long been suspected as a culprit whose inhibition plays a role in MSS. In addition, MT-1 MMP (MMP-14) knockout mice suffer connective tissue disease due to inadequate collagen turnover¹⁴ and impaired endochondral ossification¹⁵ reminiscent of joint lesions in MSS. We have therefore concentrated our efforts on potentially inhibiting MMP-13 while sparing other MMPs to achieve joint safety, in particular MMP-1 and MMP-14, which we refer to as the dual-sparing hypothesis. MMP-13 selective α -carboxylic acids have been reported by Wyeth

researchers.^{16–18} Moderately selective pyrimidinetrione MMP-13 inhibitors have been reported that gave rise to fibroplasia in a 14-day rat study, but MMP-14 data was not reported.¹⁹

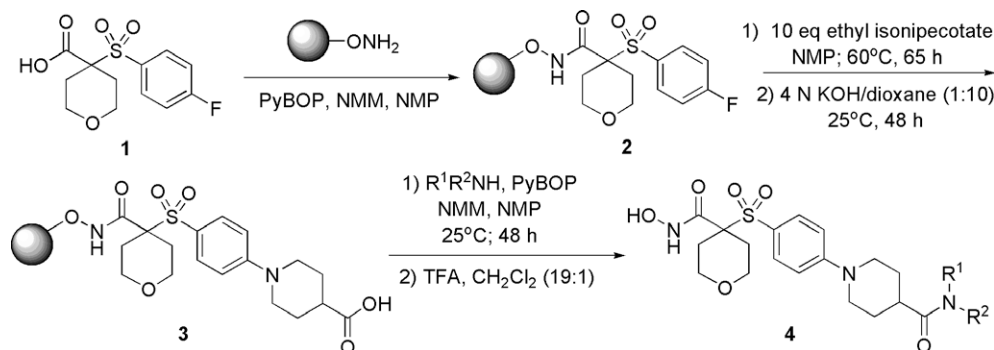
We previously described the synthesis and MMP inhibitory activity of β -sulfone hydroxamates^{20,21} and aryl-linked isosteres^{22,23} that potently inhibit MMP-2 and MMP-13 but spare MMP-1, and discovered that α -sulfone hydroxamates including **SC-276** are superior to the β -sulfones in both MMP-1 sparing enzyme profiles and ADME properties, and exhibit excellent oral antitumor efficacy *in vivo*.²⁴ MMP-1 sparing α -sulfone hydroxamates have also been reported by the Wyeth group through modification of P1' substituents,^{25,26} and Wyeth researchers have also employed β -sulfones to attain potent and selective TACE inhibitors.^{27,28} Zhang et al. of J&J have employed α -sulfone carboxylic acids as MMP-1 sparing gelatinase (MMP-2/9) inhibitors.²⁹ Our work in exploring modifications in the P' region toward further enhancing MMP-13 selectivity through interaction with the S'₁ pocket has afforded a series of aryl piperidines and isonipecotamide derivatives that are highly selective for MMP-13 and sparing of both MMP-1 and MMP-14 as we report herein.

Isonipecotamide sulfone hydroxamates **4** in the α -tetrahydropyran series were prepared as outlined in Scheme 1. Carboxylic acid **1**²⁴ was coupled with the hydroxamate-containing modified Wang resin of Floyd³⁰ employing benzotriazol-1-yl-oxytripyrroli-dinophosphonium hexafluorophosphate (PyBOP) as the coupling agent with *N*-methylmorpholine (NMM) in *N*-methylpyrrolidinone (NMP) to give polymer-bound aryl fluoride **2**. Nucleophilic aromatic substitution with a 10-fold excess of ethyl isonipecotate in NMP, and subsequent hydrolysis of the ethyl ester gave resin-bound carboxylic acid **3**. The polymer-bound acid was activated

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Scheme 1. Synthesis of isonipecotamide sulfone hydroxamates in the α -tetrahydropyran series.

with PyBOP and reacted with the requisite amine to give the corresponding polymer-bound amides, which were liberated from the resin with TFA to afford isonipecotamides **4**.

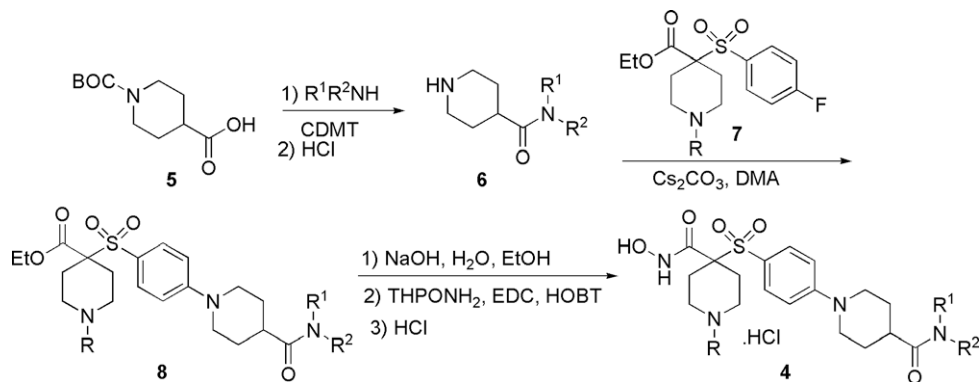
Isonipecotamides **4** in the α -piperidine sulfone series were prepared by traditional solution-phase methodologies as outlined in Scheme 2. Ethyl isonipecotatate *N*-*tert*-butylcarbamate **5**³¹ was coupled with the requisite amine using 2-chloro-4,6-dimethoxy-1,3,5-triazine (CDMT) as a coupling reagent followed by deprotection with HCl to afford piperidine **6**. Nucleophilic aromatic displacement of aryl fluoride **7**²⁴ gave the aryl piperidine sulfone **8**. Hydrolysis of the ethyl ester, coupling with THP-protected hydroxylamine using EDC and HOBT followed by acidic deprotection afforded the hydroxamates **4** as the α -piperidine hydrochloride salts.

The inhibitory potencies of α -tetrahydropyranyl and α -piperidine sulfone hydroxamates **4a–w** versus MMP-2 and MMP-13 are summarized in Table 1 wherein the isonipecotic acid amide moiety was varied. Also shown in Table 1 is a selectivity ratio derived from dividing the IC_{50} at MMP-2 by that of MMP-13. Moderate potencies for MMP-13 were maintained, and single-digit nanomolar potency was attained for several analogs. All compounds had IC_{50} values of >10,000 nM for MMP-1 (not shown), thus selectivities for MMP-13 versus MMP-1 varied from >100X (**4n**) to >2000X. Selectivity ratios versus MMP-2 were generally in a range of 50–500, and as high as 1659 for **4f**. Allyl and propargyl derivatives **4a** and **4b** were moderately potent for MMP-13 with selectivities versus MMP-2 of approximately 85X. Selectivities rose for aralkyl substituted derivatives **4c**, **4d**, and **4e** to nearly 400X for **4e**. 3,5-Dimethylpiperidine amide **4f** (mixture of *cis* and *trans* isomers) distinguished itself as the most potent for MMP-13 (IC_{50} = 4.4 nM) and the most selective versus MMP-2 as well (1659X). The corresponding α -piperidine *N*-methoxyethyl **4g** analog was prepared to improve aqueous solubility and ADME properties relative to **4f** (X = O). Surprisingly the MMP-13 potency for **4g** dropped to an IC_{50} of 50 nM, although

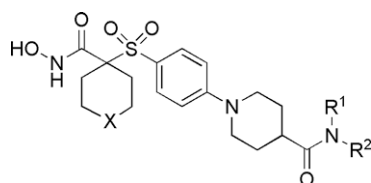
α -piperidines were as potent as α -tetrahydropyrans in the broader-spectrum, MMP-1 sparing series,²⁴ while the potency for MMP-2 increased modestly to 1700 nM resulting in a 50-fold drop in selectivity versus MMP-2. *cis*-Dimethylmorpholine **4h** was 4X less potent than **4f**, suggesting that the *trans* isomer may be the more potent isomer in **4f**. Piperazine amides **4i–4n** suffered a loss of potency for MMP-13, particularly with the introduction of a basic amine leading to the least potent analog **4n**. *N*-Aryl piperazine amides **4o–4w** in general were more potent for MMP-13 with good selectivities versus MMP-2. Fluoro analogs **4o** and **4q** were among the most potent analogs (IC_{50} = 6.7 and 6.0 nM, respectively), along with 4-acyl derivative **4r**. The 2,4-dimethylphenyl analog **4s** maintained decent potency for MMP-13 (IC_{50} = 12.2 nM) and was less potent at MMP-2 leading to a selectivity of 460X. MMP-13 tolerated heterocyclic analogs **4t–4w** with a nitrogen in the 2-position of **4t** and **4u** (IC_{50} = 10.7 and 6.4 nM, respectively) with good selectivities (330X and 300X, respectively), whereas a nitrogen in the 3- or 4-position led to a loss of some potency and selectivity (**4v** and **4w**).

Table 2 summarizes inhibitory potency for 2,3-dimethylphenylpiperidine amides **4x**, **4y**, and **4z**. The α -tetrahydropyranyl (X = O) compound **4x** distinguished itself as both the most potent and selective of the isonipecotic amides, with an IC_{50} for MMP-13 of 4.0 nM and selectivity of 40X versus MMP-3, 1500X versus MMP-2, and >2500X versus MMPs-1, 8, 9, and 14. Unfortunately, this compound was below the detection level when dosed orally in rats. The corresponding *N*-cyclopropyl and *N*-methoxyethyl piperidine analogs **4y** and **4z** were thus prepared, but the MMP-13 inhibitory potency for these compounds dropped 7X and 17X, respectively.

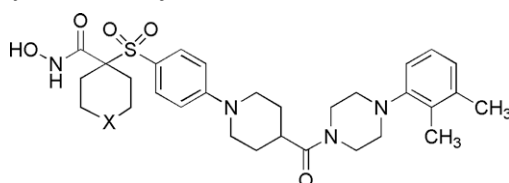
Table 3 shows the MMP inhibitory and rat PK data for aniline amide **4aa**, which had good potency for MMP-13 (IC_{50} = 9.0 nM) and very good selectivities versus both MMP-1 and MMP-14



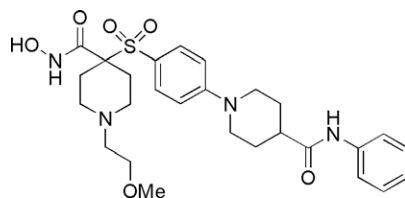
Scheme 2. Synthesis of isonipecotamide sulfone hydroxamates in the α -piperidine series.

Table 1MMP inhibitory potency of isonipecotamide α -tetrahydropyranyl and α -piperidine sulfone hydroxamates **4a–w**^a

Compd	X	NR ¹ R ²	IC ₅₀ (nM) at MMP-X ^a		Selectivity ratio 2/13
			2	13	
4a	O	Allyl(methyl)amino	2900	35	83
4b	O	Methyl(prop-2-ynyl)amino	2100	24.5	86
4c	N-Cyclopropyl	Benzyl(methyl)amino	3500	20	175
4d	O	3,4-Dihydroisoquinolin-2(1H)-yl	2100	9	233
4e	O	6,7-Dimethoxy-3,4-dihydroisoquinolin-2(1H)-yl	2400	6.2	387
4f	O	3,5-Dimethylpiperidin-1-yl	7300	4.4	1659
4g	N-CH ₂ CH ₂ OMe	3,5-dimethylpiperidin-1-yl	1700	50	34
4h	O	cis-2,6-Dimethylmorpholin-4-yl	>10K	18.1	552
4i	O	4-Acetylpiperazin-1-yl	2500	50	50
4j	O	4-Isopropylpiperazin-1-yl	5500	28	196
4k	O	4-(2-Methoxyethyl)piperazin-1-yl	7000	45	156
4l	O	4-Phenethylpiperazin-1-yl	900	25.1	36
4m	O	4-(2-Hydroxyethyl)piperazin-1-yl	6000	40	150
4n	O	4-(2-(Dimethylamino)ethyl)piperazin-1-yl	>10,000	90	>111
4o	O	4-(2-Fluorophenyl)piperazin-1-yl	1400	6.7	209
4p	O	4-(2-Methoxyphenyl)piperazin-1-yl	4500	18.0	250
4q	O	4-(4-Fluorophenyl)piperazin-1-yl	1600	6.0	267
4r	O	4-(4-Acetylphenyl)piperazin-1-yl	700	6.7	104
4s	O	4-(2,4-Dimethylphenyl)piperazin-1-yl	5600	12.2	459
4t	O	4-(Pyridin-2-yl)piperazin-1-yl	3600	10.7	336
4u	O	4-(Pyrimidin-2-yl)piperazin-1-yl	1600	6.4	250
4v	O	4-(Pyridin-4-yl)piperazin-1-yl	1750	30	58
4w	O	4-(Pyrazin-2-yl)piperazin-1-yl	900	26.8	34

^a MMP-1 IC₅₀ >10,000 nM for all compounds.**Table 2**MMP inhibitory potency of isonipecotamide α -sulfone hydroxamates **4x**, **4y**, and **4z**

Compd	X	IC ₅₀ (nM) at MMP-X							
		1	2	3	8	9	13	14	
4x	O	>10,000	6000	160	>10,000	>10,000	4.0	>10,000	
4y	N-Cyclopropyl	>10,000	6350	550	9400	>10,000	27.7	>10,000	
4z	NCH ₂ CH ₂ OMe	>10,000	400	—	—	—	70	—	

Table 3MMP inhibitory potency and rat PK data of isonipecotamide α -sulfone hydroxamate **4aa**

Compd	clog P	IC ₅₀ (nM) at MMP-X								Rat PK			
		1	2	3	8	9	13	14		C _{max}	C _{6h}	t _{1/2} (h)	BA (%)
4aa	1.0	>10,000	400	370	>10,000	1230	9.0	>10,000		1490	20	0.83	4.2

(>1100X). However, exposure and half life in the rat were very poor after oral dosing, with a half life of less than 1 h, and a BA of only 4%. Isonipecotamide hydroxamates described herein have demonstrated double-digit to single-digit potency for MMP-13 combined with very good selectivity versus MMP-1 (110–2500) and versus MMP-2 ranging from 30X to 1500X. Compound **4x** exhibits >2500X selectivity for MMP-13 versus both MMP-1 and MMP-14, hence we refer to this profile as dual-sparing (e.g., MMP-13 potency while sparing both MMP-1 and MMP-14). Yet rat PK for **4aa** was disappointing, but not surprising with a high molecular weight of 544 a.u.³² We therefore turned our attention to lower molecular weight species, while applying our learnings about P1' manipulations toward optimizing MMP-13 selectivity and ultimately to MMP-1/14 dual-sparing profiles with lower MW and fewer reduce rotatable bonds as described in the subsequent publication.³³

References and notes

- Brinckerhoff, C. E.; Matrisian, L. M. *Nat. Rev. Mol. Cell Biol.* **2002**, 3, 207.
- Nagase, H.; Woessner, J. F., Jr. *J. Biol. Chem.* **1999**, 274, 21491.
- Corbitt, C. A.; Lin, J.; Lindsey, M. L. *Recent Pat. Anti-Cancer Drug Discov.* **2007**, 2, 135.
- Fisher, J. F.; Mobashery, S. *Cancer Metastasis Rev.* **2006**, 25, 115.
- Martel-Pelletier, J.; Welsch, D. J.; Pelletier, J. *Best Pract. Res. Clin. Rheumatol.* **2001**, 15, 805.
- Hudson, M. P.; Armstrong, P. W.; Ruzyllo, W.; Brum, J.; Cusmano, L.; Krzeski, P.; Lyon, R.; Quinones, M.; Theroux, P.; Sydlowski, D.; Kim, H. E.; Garcia, M. J.; Jaber, W. A.; Weaver, W. D. *J. Am. Coll. Cardiol.* **2006**, 48, 15.
- MacFadyen, R. J. *Curr. Opin. Pharmacol.* **2007**, 7, 171.
- Lindsey, M. L. *Heart Fail. Rev.* **2004**, 9, 7.
- Peterson, J. T. *Heart Fail. Rev.* **2004**, 9, 63.
- Doherty, T. M.; Asotra, K.; Pei, D.; Uzui, H.; Wilkin, D. J.; Shah, P. K.; Rajavashisth, T. B. *Expert Opin. Ther. Pat.* **2002**, 12, 665.
- Jacobsen, F. E.; Lewis, J. A.; Cohen, S. M. *ChemMedChem* **2007**, 2, 152.
- Skiles, J. W.; Gonnella, N. C.; Jeng, A. Y. *Curr. Med. Chem.* **2004**, 11, 2911.
- Renkiewicz, R.; Qiu, L.; Lesch, C.; Sun, X.; Devalaraja, R.; Cody, T.; Kaldjian, E.; Welgus, H.; Baragi, V. *Arthritis Rheum.* **2003**, 48, 1742.
- Holmbeck, K.; Bianco, P.; Caterina, J.; Yamada, S.; Kromer, M.; Kuznetsov, S. A.; Mankani, M.; Robey, P. G.; Poole, A. R.; Pidoux, I.; Ward, J. M.; Birkedal-Hansen, H. *Cell (Cambridge, Mass.)* **1999**, 99, 81.
- Zhou, Z.; Apte, S. S.; Soininen, R.; Cao, R.; Baaklini, G. Y.; Rauser, R. W.; Wang, J.; Cao, Y.; Tryggvason, K. *Proc. Natl. Acad. Sci. U.S.A.* **2000**, 97, 4052.
- Hu, Y.; Xiang, J. S.; DiGrandi, M. J.; Du, X.; Ipek, M.; Laakso, L. M.; Li, J.; Li, W.; Rush, T. S.; Schmid, J.; Skotnicki, J. S.; Tam, S.; Thomason, J. R.; Wang, Q.; Levin, J. I. *Bioorg. Med. Chem.* **2005**, 13, 6629.
- Li, J.; Rush, T. S.; Li, W.; DeVincentis, D.; Du, X.; Hu, Y.; Thomason, J. R.; Xiang, J. S.; Skotnicki, J. S.; Tam, S.; Cunningham, K. M.; Chockalingam, P. S.; Morris, E. A.; Levin, J. I. *Bioorg. Med. Chem. Lett.* **2005**, 15, 4961.
- Wu, J.; Rush, T. S.; Hotchandani, R.; Du, X.; Geck, M.; Collins, E.; Xu, Z.; Skotnicki, J.; Levin, J. I.; Lovering, F. E. *Bioorg. Med. Chem. Lett.* **2005**, 15, 4105.
- Freeman-Cook, K. D.; Reiter, L. A.; Noe, M. C.; Antipas, A. S.; Danley, D. E.; Datta, K.; Downs, J. T.; Eisenbeis, S.; Eskra, J. D.; Garmene, D. J.; Greer, E. M.; Griffiths, R. J.; Guzman, R.; Hardink, J. R.; Janat, F.; Jones, C. S.; Martinelli, G. J.; Mitchell, P. G.; Laird, E. R.; Liras, J. L.; Lopresti-Morrow, L. L.; Pandit, J.; Reilly, U. D.; Robertson, D.; Vaughn-Bowser, M. L.; Wolf-Gouviea, L. A.; Yocum, S. A. *Bioorg. Med. Chem. Lett.* **2007**, 17, 6529.
- Becker, D. P.; Barta, T. E.; Bedell, L.; DeCrescenzo, G.; Freskos, J.; Getman, D. P.; Hockerman, S. L.; Li, M.; Mehta, P.; Mischke, B.; Munie, G. E.; Swearingen, C.; Villamil, C. I. *Bioorg. Med. Chem. Lett.* **2001**, 11, 2719.
- Becker, D. P.; DeCrescenzo, G.; Freskos, J.; Getman, D. P.; Hockerman, S. L.; Li, M.; Mehta, P.; Munie, G. E.; Swearingen, C. *Bioorg. Med. Chem. Lett.* **2001**, 11, 2723.
- Barta, T. E.; Becker, D. P.; Bedell, L. J.; De Crescenzo, G. A.; McDonald, J. J.; Munie, G. E.; Rao, S.; Shieh, H.; Stegeman, R.; Stevens, A. M.; Villamil, C. I. *Bioorg. Med. Chem. Lett.* **2000**, 10, 2815.
- Barta, T. E.; Becker, D. P.; Bedell, L. J.; De Crescenzo, G. A.; McDonald, J. J.; Mehta, P.; Munie, G. E.; Villamil, C. I. *Bioorg. Med. Chem. Lett.* **2001**, 11, 2481.
- Becker, D. P.; Villamil, C. I.; Barta, T. E.; Bedell, L. J.; Boehm, T. L.; DeCrescenzo, G. A.; Freskos, J. N.; Getman, D. P.; Hockerman, S.; Heintz, R.; Howard, S. C.; Li, M. H.; McDonald, J. J.; Carron, C. P.; Funkes-Shippy, C. L.; Mehta, P. P.; Munie, G. E.; Swearingen, C. A. *J. Med. Chem.* **2005**, 48, 6713.
- Aranapakam, V.; Davis, J. M.; Grosu, G. T.; Baker, J.; Ellingboe, J.; Zask, A.; Levin, J. I.; Sandanayaka, V. P.; Du, M.; Skotnicki, J. S.; DiJoseph, J. F.; Sung, A.; Sharr, M. A.; Killar, L. M.; Walter, T.; Jin, G.; Cowling, R.; Tillett, J.; Zhao, W.; McDevitt, J.; Xu, Z. B. *J. Med. Chem.* **2003**, 46, 2376.
- Aranapakam, V.; Grosu, G. T.; Davis, J. M.; Hu, B.; Ellingboe, J.; Baker, J. L.; Skotnicki, J. S.; Zask, A.; DiJoseph, J. F.; Sung, A.; Sharr, M. A.; Killar, L. M.; Walter, T.; Jin, G.; Cowling, R. *J. Med. Chem.* **2003**, 46, 2361.
- Condon, J. S.; Joseph-McCarthy, D.; Levin, J. I.; Lombart, H.; Lovering, F. E.; Sun, L.; Wang, W.; Xu, W.; Zhang, Y. *Bioorg. Med. Chem. Lett.* **2007**, 17, 34.
- Park, K.; Aplasca, A.; Du, M. T.; Sun, L.; Zhu, Y.; Zhang, Y.; Levin, J. I. *Bioorg. Med. Chem. Lett.* **2006**, 16, 3927.
- Zhang, Y.; Fan, X.; Xiang, B.; Chakravarty, D.; Scannevin, R.; Burke, S.; Karnachi, P.; Rhodes, K.; Jackson, P. *Bioorg. Med. Chem. Lett.* **2006**, 16, 3096.
- Floyd, C. D.; Lewis, C. N.; Patel, S. R.; Whittaker, M. *Tetrahedron Lett.* **1996**, 37, 8045.
- Cignarella, G.; Villa, S.; Barlocco, D. *Farmaco* **1993**, 48, 1439.
- Lipinski, C. A.; Lombardo, F.; Dominy, B. W.; Feeney, P. J. *Adv. Drug Deliv. Rev.* **1997**, 23, 3.
- Kolodziej, S. A.; Hockerman, S. L.; Boehm, T. L.; Carroll, J. N.; DeCrescenzo, G. A.; McDonald, J. J.; Mischke, D. A.; Munie, G. E.; Fletcher, T. R.; Rico, J. G.; Stehle, N. W.; Swearingen, C.; Becker, D. P. *Bioorg. Med. Chem. Lett.* **2010**, 20, 3557.