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Monoterpenoids Dithiophosphates. Synthesis and Biological Activity

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Abstract—O,O-Dialkyldithiophosphoric acids adds at the double bond of the racemic camphene and (+)limonene in the presence of Lewis acids in accordance with the Markownikoff rule with the formation of Sterpenyl esters of dithiophosphoric acids. The reaction with camphene is accompanied by the rearrangement of camphane structure to that of bornane. Addition of dithiophosphoric acid to (+)-limonene proceeds with the participation of the exocyclic double bond. Toxic and genotoxic properties of the monoterpenoid dithiophosphates were studied.

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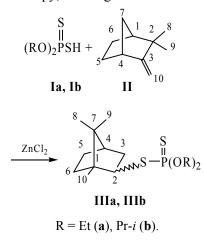
A fundamental problem in the chemistry of natural compounds is the creation of new types of phosphorusmodified terpenoid derivatives. These compounds are promising low-molecular bioregulators playing an important role in the production by living organisms of carbohydrate biopolymers of (lipo)-polysaccharide series, glycoproteins and peptidoglycans. Among the natural substances terpenoids were found exhibiting antiulcer, wound healing, hypertensive, antithrombotic, antitumor, antihypertensive, adrenergic, antiulcerogenic, hyperprotective and antihypercholesterolemic activity, as well as participating in the normalization of immune function, in the restoring liver function, and are solvents of gallstones [1, 2]. Meanwhile, at the present time both veterinary and medicine need new synthetic drugs of bioregulatory type. On the basis of terpenoids, including their phosphate derivatives, practically useful materials can be obtained for forestry, petrochemical, pharmaceutical, and perfume industries [3–5]. According to the early patent data [4, 5], the reaction of O,O-dialkyldithiophosphoric acids with pinene and dipentene [racemic mixture of (+)and (-)-limonene] at 100-200°C led to formation of the mixtures of phosphorus-containing products that were suggested for application as additives to

lubricants. The structure of the product mixtures was not established. It is presumable that the primary products formed in these reactions decompose under the rigid conditions of the process through isomerization and fragmentation characteristic of labile terpene molecules. In order to obtain stable adducts and elucidate their structure and biological activity we carried out the reaction of mono- and bicyclic monoterpenoids with dithiophosphoric acids.

It is known that in the addition reactions of organophosphorus compounds at the double bond of unsaturated compounds, the regiochemistry of the phosphorus fragment addition is determined by the structure of unsaturated compound, the nature of the catalyst, and the reaction conditions [6–9]. At the homolytic addition of dialkylphosphites to olefins in the presence of free radical initiators or at UV irradiation, the adducts are formed contrary to the Markownikoff rule. In the reaction with acidic phosphites of unsaturated compounds containing a double bond activated by electron-donating substituents the electrophilic mode of the Pudovik reaction is realized. The electrophilic addition is carried out with hydrophosphites with pronounced proton–donor properties

(e.g., cyclic hydrophosphites) [8]. It is expectable that for the implementation of catalyzed electrophilic addition catalysts with strong electron-acceptor properties (e.g., Lewis acid) or with a high protondonor action (e.g., strong mineral acid) should be used.

It is known that thiols and dithiophosphoric acid containing the SH group depending on their structure and the catalyst can be added to limonene both along and contrary to the Markownikoff rule, as well as bisadducts can form [10-12]. In our work, the reaction of O.O-dialkyldithiophosphoric acids Ia and Ib with monoterpenes was carried out in the presence of Lewis acids. We found that addition of O.O-dialkyldithiophosphoric acids Ia and Ib to the double bond of the racemic camphene (II) proceeds in the presence of catalytic amounts of anhydrous zinc chloride at 50-60°C over 2-3 h with the formation of S-2-(1-methyl-7dimethylbicyclo[2.2.1]heptyl)-O.O-dialkyldithiophosphates (IIIa, IIIb). Upon completion of the reaction the catalyst was removed by washing the reaction mixture with water. According to the data of vibration and NMR spectroscopy, washing did not lead to hydrolysis.



It follows from the data of ³¹P NMR spectroscopy that the degree of conversion of dithiophosphoric acids **Ia** and **Ib** is 100%. In accordance with ¹H NMR spectra, addition of acids **Ia** and **Ib** to the double bond of camphene (**II**) proceeds according to Markownikoff rule and is accompanied by the Wagner–Meerwein skeletal rearrangement of camphane structure to that of bornane, leading to the formation of a mixture of *exo*and *endo*-isomers. In the ³¹P NMR spectra of products **III** there are the signals at δ_P 94.0 (**IIIa**) and 92.3 (**IIIb**) ppm characteristic of the esters of dithiophosphoric acids [13]. Formation of the bornane structure is indicated also by the presence in the ¹H

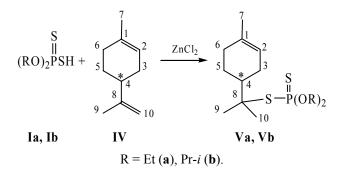
NMR spectra of products IIIa and IIIb of three strong singlets of methyl protons. In the ¹H NMR spectrum the singlets at δ 0.84, 0.93 and 1.00 ppm correspond to the diisopropyl protons of IIIb homolog. Doublet of triplets at δ 3.37 ppm with the spin-spin coupling constants ${}^{3}J_{\text{HH}}$ 7.3 Hz and ${}^{3}J_{\text{PH}}$ 19.9 Hz belongs to methine proton at the sulfur atom $C^{3}H_{A}H_{B}C^{2}HSP$. These data indicate that the two protons at C^3 atom in the compound IIIb are magnetically equivalent. In contrast, in the ¹H NMR spectrum the methine proton in the CH_AH_BCHSC fragment of the exo-isomer of the product obtained by the mercaptans addition to camphene gives rise to a doublet of doublets, whereas the endo-adduct gives a multiplet due to the additional splitting on the protons of the ring [14]. In the case of retention of the camphene structure, in the ¹H NMR spectrum the signal of methine proton in geminal position to the dithiophosphoryl fragment would be absent. Note that a similar reaction of camphene with methylmercaptoacetate in the presence of zinc chloride also results in a product of addition along the Markownikoff rule with the bornane structure, ethyl 1methyl(6-dimethylbicyclo[2.2.1]heptyl)-2-mercaptoacetate; the respective methine proton at the C^2 atom of $C^{3}H_{A}H_{B}C^{2}HSC$ fragment gives in the ¹H NMR spectrum a signal shifted upfield (δ 2.63 ppm) [15]. This result is consistent with the presence in the adducts IIIa and IIIb of dithiophosphoryl group which is more electronegative compared to the alkylthio group in alkylthiobornanes. In the ¹H NMR spectrum of adducts IIIa and IIIb were not detected the signals of vinyl protons at δ 4.5 and 4.7 ppm of the parent camphene II, which indicates the reaction completeness. In the ¹³C NMR spectra of compound IIIb obtained in the proton decoupling mode there is a singlet signal of carbon fragment PSC²H at $\delta_{\rm C}$ 53.8 ppm, which without the proton decoupling appears as a doublet (${}^{1}J_{\text{HC}}$ 151.1 Hz). In contrast, at the retention of the camphene structure, the carbon atom to which is added the thiophosphoryl fragment (the PSC fragment, which does not contain protons) would appear as a singlet at recording ¹³C NMR spectra in both modes.

The IR spectra of the products **IIIa** and **IIIb** do not contain the absorption bands at $v = 2500-2400 \text{ cm}^{-1}$, characteristic of stretching vibrations of the SH bond in the acids **Ia** and **Ib**. The C=C absorption band of terpene **II** in the region of $v = 1600-1650 \text{ cm}^{-1}$ in the IR spectra of compounds **IIIa** and **IIIb** was not detected also. The absorption bands of the stretching vibrations of the P=S and P–S bonds in dithiophosphates **IIIa** and **IIIb** are located at v = 659-655and 547-527 cm⁻¹, respectively. The electron impact mass spectra of the products **IIIa** and **IIIb** include the mass peaks m/z = 322.7 and 350.2, respectively, of their molecular ions $[M]^+$ (calculated M 322.2 and 350.2, respectively).

The adducts IIIa and IIIb are of low thermal stability and could not be distilled even in a high vacuum. Their purification was carried out by column chromatography. Compounds IIIa and IIIb, thus, are the primary products of reactions, which under the rigid conditions of vacuum distillation decompose. In this regard, it is presumable that compounds previously described as products of addition of O,O-dialkyldithiophosphoric acids to pinene and dipentene at 100-200°C [4, 5] are actually the products of various secondary reactions. Inasmuch as this reaction proceeds in accordance with the Markownikoff rule, as well as in the case of thiols, mercaptoacetic acid or methyl mercaptoacetate [12, 14, 15] the dithiophosphoric acids possess the ability to the electrophilic addition to electron-rich alkenes. It is known that dithiophosphoric acids show a dual reactivity in the addition reactions: the presence of labile protons and nucleophilic dithiophosphoryl fragment gives them the opportunity of entering into both nucleophilic and electrophilic addition reactions [9].

In order to expand the synthetic potential of unsaturated terpenes by involving them in the tiophosphorylation reactions and obtaining potential biologically active products, is interesting to involve into the reaction with dithiophosphoric acids Ia and Ib some unsaturated monocyclic terpenes. Among them of special interest is (+)-limonene, one of the most stable terpenes, which contains an exocyclic and an endocyclic double bonds differing in the reactivity. Therefore we could expect that the electrophilic addition might occur in various directions. We found that addition of O,O-dialkyldithiophosphoric acids Ia and Ib to (+)-limonene (IV) in the presence of zinc chloride proceeded at room temperature over 1-2 h at the exocyclic double bond, affording O,O-dialkyl-S-8-[(+)-1-methyl-4-isopropylcyclohex-1-enyl]dithiophosphates Va and Vb. At heating the initial reagents at 60°C for 3 h the formation of compounds of the phosphorylation at the endocyclic double bond was not observed.

The compounds **Va** and **Vb** are not stable at high temperatures in the distillation process and therefore were isolated by column chromatography. The signals



in ³¹P NMR spectra of adducts Va and Vb are shifted upfield $[\delta_P 90.4 (Va) and 87.1 (Vb) ppm]$ compared to products IIIa and IIIb obtained in the reactions with camphene (δ_P 92.3 – 94.0 ppm). In the ¹H NMR spectrum of adduct Vb there are three singlet signals of protons of three methyl groups at δ 1.53, 1.59 and 1.66 ppm, and the signal of endocyclic vinyl proton remains at δ 5.37 ppm. The signals of two exocyclic vinyl protons disappear in ¹H NMR spectra of the products Va and Vb completely. Note that in the ¹H NMR spectrum the exocyclic vinyl protons of (+)limonene (IV) give a singlet signal at d 4.71 ppm. The IR spectrum of products Va and Vb shows a weak absorption band of the C=C bond stretching vibrations at v = 1643 cm⁻¹. The mass peak m/z = 323.2 in the mass spectrum of chemical ionization (CI) of dithiophosphate Va is consistent with its molecular ion $[M+H]^+$. The mass spectrum of electron impact of the isopropyl homolog Vb contains the mass peak m/z =350.2 corresponding to its molecular ion $[M]^+$.

It was established that the electrophilic addition of dithiophosphoric acids Ia and Ib to unsaturated terpenes II and IV is accelerated in the presence of other Lewis acids (NiCl₂, CuCl, CuCl₂, FeCl₃) at room temperature. The less active catalysts are BF₃. Et₂O, and AlCl₃. From a number of Lewis acids we selected zinc chloride as a "soft" Lewis acid, which unlike most catalysts with higher acceptor strength, for example, BF₃·Et₂O, commonly does not induce a secondary transformation of the terpene molecule. Note also that the use of protic acids (HCl, H₂SO₄, HClO₄) as catalysts in the reactions of thiols with unsaturated monoterpenes leads to the isomerization of terpene molecules with the formation of a complex mixture of products [16]. Presumably the high hygroscopicity of zinc chloride may contribute to the proceeding of the reactions of dithiophosphoric acids with monoterpenes promoting formation in the reaction medium of HCl, which may also act as catalyst in the reactions of dithiophosphoric acids with the unsaturated monoterpenes. Thus, we first found the phenomenon of the catalysis with the Lewis acids in a series of derivatives of the tetracoordinated phosphorus thioacids by the example of reactions of electrophilic addition of dithiophosphoric acids at the double bond of unsaturated monoterpenoids.

The studied reaction showed a route to monoterpenoid dithiophosphates with potential biological activity. This article presents the principal results of studies of toxic and genotoxic properties of compounds IIIa, IIIb, Va and Vb. The methods used in these studies were published in [17-19]. Full description of the results will be published in a special biological journal. The study of toxic and genotoxic properties of dithiophosphates IIIa, IIIb, Va and Vb was carried out on Salmonella typhimurium TA 100 and Escherichia coli PQ37 as the test bacteria. We found that dithiophosphates IIIa. IIIb. Va and Vb are weak toxicants for the test bacteria. Compound IIIa exhibits the characteristics of a weak inhibitor of alkaline phosphatase activity in the Escherichia coli PQ 37 cells, initiates the highest SOS-response and shows mutagenic activity in Ames test. Dithiophosphate IIIa is a direct mutagen and true genotoxicant, since this compound is practically non-toxic to the test bacteria, but causes the induction of gene mutations. Remaining compounds, IIIb, Va and Vb do not show mutagenic properties.

EXPERIMENTAL

IR spectra were recorded on a Bruker Vector 22 and a Tensor 27 IR Fourier Spectrometers (400-4000 cm⁻¹) from liquid films between KBr plates. Chemical shifts of ³¹P nuclei of phosphorus compounds were measured on a Bruker CXP-100 spectrometer with an operating frequency of 36.47 MHz with external 85% H₃PO₃. Positive value of the chemical shift δ_P corresponds to the downfield shift. The ¹H NMR spectra were registered on a Bruker Avance-400 with operating frequency 400 MHz and a Bruker Avance-600 (600 MHz) spectrometers in CDCl₃ solutions. The ¹³C NMR spectra were recorded on a Bruker Avance-600 spectrometer (100.6 MHz) in CDCl₃ solutions. The electron impact and chemical ionization mass spectra were recorded on a Finnigan MAT-212 and a TRACE MS Finnigan MAT mass spectrometers.

S-2-(1-Methyl-7-dimethylbicyclo[2.2.1]heptyl)-O,O'-diethyldithiophosphate (IIIa). To a mixture of

3.9 g of acid Ia and 2.9 g of camphene II at ~20°C in a stream of dry argon was added at stirring in portions 0.1 g (3.4 wt %) of ZnCl₂. The mixture was heated for 3 h at 50-60°C. After cooling, the mixture was diluted with 10 ml of Et₂O and washed with three 10-ml portions of water. The organic layer was separated and dried over anhydrous CaCl₂ for ~12 h. After separating the drying agent, the filtrate was evaporated in a vacuum of 0.5 mm Hg at 40°C for 1 h and then in a vacuum of 0.06 mm Hg at 40°C for 1 h. 5.7 g (84%) of dithiophosphate IIIa was obtained, which was purified column chromatography (silica gel, eluent bv petroleum ether boiling at 70–100°C), R_f 0.27 (petroleum ether), n_D^{20} 1.5325. IR spectrum, v, cm⁻¹: 2983 s, 2956 s, 2880 s [vas.s(CH3), vas.s(CH2)]; 1465 m $[\delta_{as}(CH_3)]; 1390 \text{ s} [\delta_s(CH_3)]; 1016 \text{ v.s.br}, 958 \text{ s} [v]$ (POC]; 826 m, 796 m [v_{as,s}(PO₂)], 659 s [v(P=S)], 527 m [v(PS)]. ¹H NMR spectrum, δ , ppm, (J, Hz): 0.85 s, 0.93 s and 1.00 s (9H, CH₃-ring), 1.37 m [6H, (CH₃CH₂O)₂P, ³J_{HH} 7.1], 1.24 m, 1.74 m and 2.01 m (CH₂-ring and CH-ring.) 3.31 d.t (1H, CH_AH_BCHSP, ${}^{3}J_{\rm HH}$ 7.0, ${}^{3}J_{\rm PH}$ 19.0) 4.12 m {2H, [(CH₃CH₂O)₂]₂P, ${}^{3}J_{\rm HH}$ 7.1}. Found, %: C 52.27, H 8.46, P 9.43; S 19.48. C₁₄H₂₇O₂PS₂. Calculated, %: C 52.14, H 8.46, P 9.61; S 19.85.

S-2-(1-Methyl-7-dimethylbicyclo[2.2.1]heptyl)-**O,O'-diisopropyldithiophosphate** (IIIb) was prepared similarly from 3.6 g of acid Ib and 2.3 g of camphene II using 0.23 g (3.5 wt %) of ZnCl₂, yield 4.4 g (75%), purified by column chromatography (silica gel, eluent petroleum ether boiling at 70–100°C/Et₂O 1:1), R_f 0.60 (petroleum ether/Et₂O 1:1), n_D^{20} 1.5025. IR spectrum, v, cm⁻¹: 2979 s, 2956 s, 2879 s $[v_{as,s}(CH_3), v_{as,s}(CH_2)];$ 1454 m [$\delta_{as}(CH_3)$]; 1386 m, 1374 m [$\delta_{s}(CH_3)_2C$ gem] o.s 993, 970 o.s [v(POC]; 888 m, 777 s [v_{as s}(PO₂)], 655 s [v(P=S)]; 547 m [v(PS)]. ¹H NMR spectrum, δ , ppm, (J, Hz): 0.84 s, 0.93 s and 1.00 s [9H, CH₃-ring), 1.34 d and 1.36 d [12H, $(CH_3)_2$ CHOP, ${}^3J_{HH}$ 5.7], 1.41 m, 1.72 m and 2.02 m (CH₂-ring. and CH-ring), 3.37 d.t (1H, CH_AH_BCHSP, ${}^{3}J_{HH}$ 7.3, ${}^{3}J_{PH}$ 19.9), 4.80 m {2H, [(CH₃)₂CHO]₂P, ³J_{HH} 5.7}. Found, %: C 54.42, H 8.96, P 9.32; S 18.18. C₁₆H₃₁O₂PS₂. Calculated, %: C 54.82, H 8.94, P 8.84; S 18.26.

0,0'-Diethyl-S-8-[(+)-1-methyl-4-isopropylcyclohex-1-enyl]dithiophosphate (Va) was obtained similarly from 8.0 g of acid Ia and 5.6 g of (+)-limonene (IV) using 0.24 g (3 wt %) of ZnCl₂, yield 7.0 g (50%), purified by column chromatography (silica gel, eluent CCl₄), R_f 0.28 (CCl₄), n_D^{20} 1.5320. IR spectrum, v, cm⁻¹: 2962 s, 2926 s, 2667 m [v_{as,s}(CH₃), v_{as,s}(CH₂)]; 1642 w [v(C=C)], 1443 m [δ_{as} (CH₃)]; 1384 m, 1373 m [δ_{s} (CH₃)₂C gem.] 1017 v.s.br, 957 m [v(POC]; 798 m [$v_{as,s}$ (PO₂)]; 656 m [v(P=S)]; 533 m [v(PS)]. ¹H NMR spectrum, δ , ppm, (*J*, Hz): δ_{1} 1.35 t [3H, (CH₃CH₂O)₂P, ³*J*_{HH} 7.1] and δ_{2} 1.36 t [3H, (CH₃CH₂O)₂P, ³*J*_{HH} 6.8], 1.46 s and 1.53 s [6H, (CH₃)₂CS], 1.63 s (3H, CH₃-ring.) 1.71–2.15 m (7H, CH₂-ring and CH-ring); d₁ 4.13 d.q [4H, (CH₃CH₂O)₂P, ³*J*_{HH} 7.1, ³*J*_{PH} 9.7] and d₂ 4.23 d.q [4H, (CH₃CH₂O)₂P, ³*J*_{HH} 6.8, ³*J*_{PH} 7.0]; 5.34 m (H, C=CH-ring). Found, %: C 51.78, H 8.13, P 9.33; S 19.99. C₁₄H₂₇O₂PS₂. Calculated, %: C 52.14, H 8.46, P 9.61; S 19.85.

O,O-Diisopropyl-S-8-[(+)-1-methyl-4-isopropylcyclohex-1-enyl|dithiophosphate (Vb) was obtained similarly from 8.0 g of acid Ib and 5.1 g of (+)limonene (IV) using 0.056 g (0.7 wt %) of ZnCl₂, yield 1.11 g (85%), purified by column chromatography (silica gel, eluent CCl₄), $R_f 0.24$ (CCl₄), $n_D^{20} 1.5079$. IR spectrum, v, cm⁻¹: 2976 s, 2928 s, 2836 s [v_{as,s}(CH₃), v_{as.s}(CH₂)]; 1643 w [v(C=C)], 1450 m [δ_{as}(CH₃)]; 1384 m, 1373 m [δ_s (CH₃)₂C gem] v.s.br 968 [v(POC]; 887 m, 775 m $[v(PO_2)]$; 650 m [v(P=S)]; 548 m [v(PS)]. ¹H NMR spectrum, δ , ppm, (J, Hz): 1.35 d and 1.37 d $[12H, (CH_3)_2 CHOP, {}^3J_{HH} 6.0]; 1.53 \text{ s and } 1.59 \text{ s } [6H,$ (CH₃)₂CS]; 1.66 s [3H, CH₃-ring]; 1.54–2.18 m (7H, CH₂-ring and CH-ring); 4.91 m {2H, [(CH₃)₂CHO]₂P, ${}^{3}J_{\text{HH}}$ 6.0}; 5.37 m (1H, C=CH-ring). Found, %: C 54.58; H 8.38; P 9.25; S 18.17. C₁₆H₃₁O₂PS₂. Calculated, % : C 54.82; H 8.94; P 8.84; S 18.26.

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