OPTICAL-ANALYTICAL STUDIES ON STEROIDS.

I. LIGHT ABSORPTION BETWEEN 230–500 m μ OF α,β -UNSATURATED AND 3,6-DIKETO STEROIDS IN ALKALINE ETHANOL

ANDRÉ S. MEYER

Received April 22, 1955

For the elucidation of structural features of unknown steroids available in minute quantities only, infrared spectrometry has been developed into a most powerful tool. It is possible by its use to determine the types of functional grouping and the number of carbonyls present in the molecule (cf. 1, 2). However, their exact localization is as yet feasible only in a limited number of specific cases. Ultraviolet absorption (cf. 3) and optical rotation measurements (cf. 4) have proved of help in deciding between alternative structural possibilities. The determination of mobilities in paper-chromatographic systems (inter al. 5) can render valuable clues toward the establishment of a more accurate structure and, in combination with the concept of conformational analysis (6), the configuration of certain substituents may be deduced. In addition, oxidative degradations (cf. 7, 8, 9) followed by estimation of the liberated formaldehyde (inter al. 10) and acetaldehyde (11) as well as the Zimmermann chromogen (inter al. 12, 13) can be employed with advantage for the elucidation of side chain structure. The position of a ketone group may be more accurately defined by the ultraviolet spectrum of a derivative such as a phenylhydrazone (inter al. 3, 14, 15). The action of perchloric acid on certain substituted 2,4-dinitrophenylhydrazones should produce a bathochromic shift by extension of the conjugated system (cf. 16). The color formed after reaction of steroids with diphenylamine in acidic medium appears to depend on certain structural arrangements (17). A method has been recently devised for the recognition and estimation of Δ^4 -3-ketosteroids by measuring the formation of intensely fluorescent substances resulting from alkaline treatment (18, 19). On the other hand, absorption spectra in concentrated sulfuric acid (cf. 20, 21), valuable as they are for the characterization of individual compounds, do not lend themselves for extensive interpretation of structural elements (cf. 22, 23).

In the papers presented in this series, an attempt is made to implement the methods enumerated, prompted by the specific need to determine structures of unknown Δ^4 -3-ketosteroids isolated in milligram quantities from biological material. In this communication it is desired to report on the changes encountered in the intense absorption of a number of Δ^4 -3-ketosteroids upon exposure to alkaline ethanol at 23 and 60°. Of particular interest were Δ^4 -3-ketosteroids containing hydroxyl substituents at various positions and related substances with carbonyl functions. Alteration in spectral absorption of the color products has allowed correlation of certain structural features of these compounds. Some applications have been reported previously (24-27).* Paper II of this series pre-

^{*} Incidentally, by this method the presence of a certain impurity was easily detected in a sample, although it had not been recognized in two paper-chromatographic systems.

sents the results of rate studies of formation of the "blue formazan" chromogen which may indicate the site of a substituent (26). Report III summarizes some observations on isolated olefinic centers of α , β -unsaturated ketonic steroids as related to absorption near 200 m μ (28).

METHOD

The source of the *steroids* is acknowledged below. Those compounds which are, throughout the Tables, designated by p were prepared by us following published procedures. Those samples designated by q were commercially available materials purified by direct crystallization, in certain cases preceded by chromatography. The steroids, before weighing, were dried at 0.1 mm. and 80° for 1 hr. The *reagent* was freshly prepared by mixing 94.0 ml. of 95% redistilled ethanol with 6.0 ml. of 10% aqueous tetramethylammonium hydroxide or 1.1 N aqueous potassium hydroxide. The solution thus had a normality of 0.066 with respect to hydroxide and a 90% ethanol content.

A quantity of steroid (between 50–70 μ g.) to make an approx. $7 \times 10^{-5} M$ solution was transferred to a stoppered silica cell of 10 mm. light path. The solvent used for the transfer was evaporated, at a temperature of no more than 35°, under a stream of nitrogen. To the dried sample, 3.0 ml. of alkaline reagent was added and the material readily dissolved. The absorbance, over a reaction blank, was then immediately recorded from 230 to 400 or 500 m μ by use of a Cary spectrophotometer (Model 11 MS). The region of 240 m μ , where Δ^4 -3-ketosteroids absorb intensely, was reached 2–3 minutes after the steroids came in contact with the reagent. Further measurements were carried out at appropriate time intervals. When the reaction was conducted at elevated temperature, the cells were cooled under tap water to room temperature after their exposure. The absorptivity (ϵ) is expressed for concentrations in mole per liter.

In some instances, 0.20 ml. of 1 N aqueous hydrochloric acid was added after completion of the alkaline reaction and the spectrum of the solution, now at a pH between 7 and 3, was again recorded.

RESULTS AND DISCUSSION

A number of α,β -unsaturated steroids investigated were only slightly affected by the alkaline reagent under the described conditions (Table I). The group of Δ^4 -3-ketosteroids in which a moderate decrease in absorbance near 240 m μ was observed, frequently gave a concomittant rise to a second less intense absorption near 380 m μ which became more distinct when the samples were heated to 60°. With 16 α -hydroxyprogesterone (#12), in addition, dehydration to 16-dehydroprogesterone appeared to have occurred, manifested by an increase in absorbance near 240 m μ within 4 hr. All of the six Δ^4 -3-ketosteroids studied, with an unbonded oxygen at C-11, could be classed in the group mentioned (Table II). However, aldosterone (#21) did not show the second band near 380 m μ ; this is probably due to the fact that the 11 β -hydroxyl group is largely tied up in a half acetal linkage with the 18-aldehyde function (cf. 29). Cross, et al. (30) noted a maximum near 375 m μ of unspecified intensity for cortisone acetate, progesterone, and testosterone, after heating these steroids in 1.1 N aqueous tetramethyl-

ANDRÉ S. MEYER

	Compound	After 2-3 min.	After 24 hr.	After additional 1.5 hr. at 60°	Source						
1	Δ^4 -Androstene-										
	3,17-dione	241 (17.2)	241 (17.0)	241 (16.3) 377 (0.5)	q						
2	Δ^4 -Androstene-17 β -				-						
	ol-3-one	241 (17.7)	242 (17.5)		q						
3	3-Keto- Δ^4 -etienic				-						
	acid	242 (17.2)	243 (15.6)	243 (14.4) 377 (0.6)	p						
4	Δ^4 -Pregnen-21-ol-				-						
	3,20-dione	242 (17.0)	242 (16.5)	243 (15.0) 377	q						
5	Δ^4 -Cholesten-3-one.	242 (17.1)	243 (16.4)		d						
6	Δ ^{1, 4} -Androsta-		• •								
	diene-3,17-dione	244 (16.5)	243 (16.6)	243 (16.5)	n						
7	Δ4. 6-Androsta-		, .								
	diene-3,17-dione	284 (25.5)	284 (25.5)		f						
8	7β-Hydroxy-Δ4-										
	cholesten-3-										
	one*	245 (15.6)	286 (25.9)	286 (24.2)	p						
9	12α-Hydroxy-Δ4-				-						
	androstene-3,17-										
	dione	240 (17.0)	241 (16.6)		0						
10	14-Hydroxy-∆4-	. ,									
	androstene-3,17-										
	dione	238 (15.4)	243 (13.7) 377 (0.7)		m						
11	16α-Hydroxy-Δ4-										
	androstene-3,17-										
	dione	241 (17.6)	242 (16.7)		c						
12	16α-Hydroxy-Δ4-										
	pregnene-3,20-										
	dionet	242 (17.7)	242 (21.5) 375 (0.5)	241 (20,5) 375 (1,2)	c						
13	∆ ^s -Cholesten-38-	. ,			1						
	ol-7-one a cetate.	235 (14.0)	239 (12.6) 279 (3.8)	239 (10.0) 279 (9.1)	d						
14	$\Delta^{5,16}$ -Pregnadien-3.				1						
	ol-20-one	238 (9.9)	238 (4.8)		q						
				1							

TABLE I Absorption Maxima in M μ ($\epsilon \times 10^{-3}$) of α,β -unsaturated Ketosteroids in Alkaline Ethanol at 23°C.

* This sample (cf. 32) showed a maximum at 243 m μ (ϵ 15.6 \times 10³) in neutral abs. ethanol; $[\alpha]_{p}^{28}$ +84° ±3° (c, 0.95 in chloroform).

† This sample showed an absorption peak at 241 m μ (ϵ 17.8 \times 10³) in neutral 95 per cent ethanol; the higher extinction coefficient in alkaline ethanol listed was reached before 4 hr. elapsed and afterwards decreased only slightly.

ammonium hydroxide solution at 70° for 35 minutes. Since this band was not present in dehydrocholic acid and some steroidal estrogens, they associated it with the Δ^4 -3-keto moiety. After conclusion of the present investigation, we learned of the successful development of a method for the quantitative estimation of Δ^4 -3-ketosteroids, based on the formation, by 0.3 N potassium *tert*-butoxide in *tert*-butyl alcohol, of an absorption band near 385 m μ in 1 hr. at room temperature (D. Abelson, private communication). The formation of this maximum appears to be greatly facilitated by the anhydrous reagent. Table III illustrates

	Compound		After 2-3min. After 24 Max. Max.		24 hr.			After additional 1.5 hr. at 60°					Source	
					Max.		Max.		Max.		Min.		Max.	
15	Δ^4 -Androstene-												<i>(</i> 2 - 1)	
	3,11,17-trione.	237	(16.7)	238	(14.9)	377	(1.6)	239	(10.0)	308	(0.6)	375	(3.9)	p
16	Δ'-Pregnene-													
	$1/\alpha, 21-0101-$	1												
	(aortisono)	020	(17.0)	020	(13 1)	377	(0 0)	220	(11 1)	308	(0.7)	377	(1.5)	
17	A4-Androsten-	200	(17.0)	200	(10.1)	011	(0.9)	200	(11.1)	000	(0.1)	011	(1.0)	Ŷ
11	118-01-3 17-													
	dione	242	(17.4)	243	(18.0)	377	(0.9)							n
18	Δ^4 -Pregnene-118.		((10.0)		(0.0)							P
	$17\alpha.21$ -triol-													
	3,20-dione	243	(17.5)	244	(15.3)	377		245	(14.0)			377	(0.9)	a
19	Δ^4 -Ándrosten-		. ,	}	. ,									-
	11α -ol-3,17-											Ì		ļ
	dione	242	(17.9)	243	(16.3)	377	(0.8)							
20	Δ^4 -Pregnen-11 α -			l				1		1		ł		
	ol-3,20-dione	242	(17.3)	241	(16.1)	377	(0.7)	243	(13.4)	308	(0.4)	377	(1.8)	e
21	Δ^4 -Pregnene-													ł
	$11\beta, 21$ -diol-													
	18-oxo-3,20-											1		
	dione (aldo-]
	sterone)	237	(17.8)	237	(17.2)	I.	285	237	(17.1)	I.2	285			h

TABLE II

Absorptions in M μ ($\epsilon \times 10^{-3}$) of 11-Oxygenated Δ^4 -3-Ketosteroids in Alkaline Ethanol at 23°C.

I = inflection.

that in Δ^4 -3-ketosteroids in which the angular group at C-10 is actually (19norsteroids) or potentially absent [eliminated by the action of the alkaline medium in 19-hydroxyl derivatives (25)], the second maximum was seen at 367 m μ rather than at 377 m μ as with the 10-methyl series. However, after acidification this second maximum shifted to approximately 310 m μ in the case of both series (e.g. #12, 15, 20, and 22).

Most obvious, as was to be expected, were the changes caused by the alkaline reagent on the spectra of α,β -unsaturated ketosteroids having a hydroxyl substitutent in the vicinity of the unsaturated system. The $\gamma\beta$ -hydroxy- Δ^4 -3-keto structure (#8) reacted in a characteristic manner: The absorption maximum at 245 m μ disappeared (it could no longer be clearly distinguished after 2 hr.), while simultaneously a new maximum at 286 m μ (first visible after 30 min.) developed. Since $\Delta^{4.6}$ -cholestadien-3-one is reported (31) to absorb in the ultraviolet in neutral ethanol maximally at 285 m μ (ϵ 26.0 \times 10³), the absorbance changes in alkaline ethanol of #8 represented the rate of dehydration of this compound. When compound #8 was exposed to 0.03 N hydrochloric acid in 92% ethanol at 23°, the absorption peak at 245 m μ did not change over a period

TABLE III

Absorption Maxima in Mm ($\epsilon \times 10^{-3}$) of 19-Hydroxylated and 19-Nor- Δ^4 -3-ketosteroids in Alkaline Ethanol at 23°C.

Compound		After 2-3 min.		After 24 hr.				After additional 1.5 hr. at 60°				Source
22	19-Nor-∆ ⁴ - androstene-											
	3,17-dione	240	(17.2)	241	(12.8)	367	(0.4)	242	(7.4)	367	(1.2)	d
23	19-Nor-∆⁴-											
	pregnene-											1
	3,20-dione	241	(17.5)	242	(14.2)			242	(10.6)	367	(1.1)	k
24	19-Hydroxy-						Ì					1
	Δ^4 -andros-											
	stene-3,17-											
	dione	243	(16.0)	242	(14.3)	367	(0.5)	241	(9.6)	367	(1.2)	p
25	19-Hydroxy-											
	∆⁴-preg-			{								
	nene-3,20-											
	dione	243	(16.0)	242	(14.4)	367	(0.3)	241	(9.6)	367	(1.2)	a
26	19-Hydroxy-											
	3-keto-∆4-											
	etienic acid	br. 245	(16.2)	244	(14.7)			br. 242	(12.4)	367	(1.1)	a

 $br_{1} = broad.$

TABLE IV

Absorptions in Mm ($\epsilon \times 10^{-3}$) of 2 α -Hydroxylated Δ^4 -3-ketosteroids^a in Alkaline Ethanol at 23°C.

Exposure Time	Δ^4 -Androstene- 2α , 17 β -diol-3-one (27)								
Diposule Time	Max.	Inflection	Min.	Max.					
Neutral	240 (15.8)								
2-3 min.	238 (14.9)								
1.0 hr.	234 (17.6)			354					
4.0 hr.	233 (20.2)	254 (7.9)	290 (1.3)	355 (2.7)					
24 hr.	233 (19.5)	254 (7.9)	290 (1.4)	355 (2.9)					
	Δ ⁴ -Pregnen-2α, ol-3, 20-dione acetate (28)								
	Max.	Inflection	Min.	Max.					
Neutral	240 (16.7)								
2-3 min.	238 (15.0)								
1.0 hr.	234 (19.0)			354					
4.0 hr.	233 (20.4)	254 (7.8)	290 (1.2)	355 (2.6)					
24 hr.	233 (19.8)	254 (7.8)	290 (1.2)	355 (2.7)					
	Δ4-Pregnene-2 <i>a</i> , 17 <i>α</i> -21-triol-3, 20-dione (29)								
	Max.	Inflection	Min.	Max.					
2-3 min.	238 (15.6)								
1.0 hr.	234 (17.1)			354					
4.0 hr.	233 (19.2)	254 (8.7)	290 (1.3)	355 (2.4)					
24 hr.	233 (18.1)	254 (8.7)	290 (1.4)	355 (2.8)					

^a Source of these samples, *l*.

of 24 hr. and it lost only 5% of its intensity after additional heating at 60° for 1.5 hr. This behavior in an acidic medium may well prove to be a means of distinguishing Δ^4 -7 β -hydroxyl from the Δ^4 -7 α -hydroxyl derivatives, since Greenhalgh, *et al.* (32) observed a dehydration when hydrolyzing the 7 α -tetrahydropyranyloxy- Δ^4 -cholesten-3-one in very dilute ethanolic hydrocholoric acid at room temperature. Similarly, the data for Δ^5 -cholesten-3 β -ol-7-one acetate (#13 would suggest that a partial conversion to $\Delta^{3,5}$ -cholestadien-7-one had taken place; this dienone reportedly (33) has a peak at 280 m μ (ϵ 27 \times 10³) in neutral



FIG. 1. SPECTRA OF 58.5 μ g. OF Δ^4 -ANDROSTENE-2 α , 17 β -DIOL-3-ONE (#27) IN 3.0 ML. OF SOLVENT:- - - in 0.066 N tetramethylammonium hydroxide of 90% ethanol content, 2-3 minutes after solution; ______ in the alkaline solution after 4 hour of exposure; ••• same as latter conditions, however acidified with hydrochloric acid to pH 3 (ethanol content 85%).

ethanol. After acidification, the positions of these maxima at 280 m μ remained unchanged.

Three 2α -hydroxylated Δ^4 -3-ketosteroids with various side chains were studied. In all these cases the absorptivity near 240 m μ increased appreciably within the first 4 hr. of reaction; the integrated area of this band, however, remained approximately constant. The appearance of a typical shoulder near 255 m μ and a second maximum near 360 m μ led to an easily recognizable spectrum (Table IV and Fig. 1). The 2β -acetoxy- Δ^4 -3-ketosteroids should show the same pattern, since their mild alkaline hydrolysis afforded the 2α -hydroxyl isomers (34). The profound change in the spectrum after acidification, which can be reverted after renewed addition of alkali, is noteworthy.

The allylic 6α and 6β -hydroxylated Δ^4 -3-ketosteroids were easily distinguished under the described conditions (Tables V and VI). The absorption maximum near 240 m μ of the former compounds (#33-35) decreased to approximately half the intensity and shifted to 260 m μ within 24 hr. at 23° or 2 hr. at 60°. In the same period, a second maximum of somewhat lower intensity than the first one appeared near 380 m μ , the solution turning yellow. The 6β -hydroxylated steroids (#31 and 32) did not produce this shift at room temperature and absorption near 380 m μ was less intense; at 60° the bathochromic shift occurred which, however, was delayed by approximately 1 hr. as compared to the 6α -hydroxylated derivatives. The saturated 3,6-diketone compounds (#36-39), which do not absorb intensely in neutral ethanol, gradually gave rise to the two peaks which

Exposure Time	3β-Hydroxy-∆4	-6-keto-choleste	n acetate (30)	6α-Hydroxy-Δ4-3-keto derivatives of cholestene 6α-acetate (33)				
•	Max.	Min.	Max.	Max.	Min.	Max.		
2-3 min. 4.0 hr. 10 hr. 24 hr.	237 (5.5) 243 (6.0) 243 (6.0) 243 (6.1)	320 315 (0.8)	380 (1.0) 380 (2.2)	241 (16.3) 245 (9.2) 260 (8.7)	315 (1.3) 308 (2.0)	377 (3.4) 377 (6.5)		
<u> </u>	6β-Hydro of an	xy-∆4-3-keto de drosten-17-one	rivatives (31)†	of androsten-17-one (34)‡				
2-3 min. 1.0 hr. 4.0 hr. 10 hr. 24 hr.	236 (14.2) 238 (12.6) 240 (9.3)	308 (0.7)	377 (1.0) 377 (3.5)	241 (16.4) 242 (12.8) 246 (8.7) 258 (9.8) 258 (9.2)	312 310 (1.4) 308 (1.5) 308 (1.7)	377 (2.4) 377 (4.8) 377 (7.7) 377 (7.3)		
<u> </u>	of pregner	ne-17a, 21-diol-20	D-one (32)	of pregr 6α	nene-17α, 21-diol , 21-diacetate (3	-20-one 5)		
2-3 min. 2.0 hr.	237 (13.3)		277 (0.8)	237 (15.7)	222 (0.0)	277 (9.2)		
4.0 hr. 10 hr. 24 hr.	238 (11.6) 240 (8.5)	310 (0.9)	377 (0.8)	240 (9.3) 258 (7.3)	323 (0.9) 310 (1.6)	377 (5.0)		

TABLE V

Absorptions in mµ ($\epsilon \times 10$	*) of 6-Oxygenated Steroids in A	Alkaline Ethanol at 23°C.*
--	----------------------------------	----------------------------

Exposure Time	3, 6- of	Diketo derivativ cholestane (36)	ves §	Δ^{4-3} , 6-Diketo derivatives of cholestene (42)			
	Max.	Max. Min.		Max.	Min.	Max.	
2-3 min.				260 (11.3)	308 (1.9)	377 (9.3)	
4.0 hr.	260 (4.7)		377 (3.3)	260 (10.9)	308 (1.6)	377 (8.7)	
10 hr.	260 (8.2)	308 (1.9)	377 (6.6)				
24 hr.	260 (8.2)	308 (1.2)	377 (6.2)	260 (9.2)	308 (1.3)	377 (7.7)	
	of ar	ndrostan-17-one	(39)	of ar	ndrosten-17-one	(43)¶	
2-3 min.			[259 (10.8)	306 (1.6)	377 (9.1)	
1.0 hr.	258 (3.9)	312	377 (2.7)				
4.0 hr.	258 (7.6)	308 (1.2)	377 (5.7)	259 (10.3)	306 (1.6)	377 (9.0)	
10 hr.	258 (9.6)	308 (1.5)	377 (7.3)				
24 hr.	258 (9.0)	308 (1.7)	377 (6.6)	259 (10.1)	306 (1.6)	377 (8.4)	
	of chol	estane-4a, 5a-di	ol (40)	Δ4-6-Keto-cholestene (44)			
2-3 min.		313 (0.7)	384 (2.2)	242 (7.9)			
2.0 hr.		308 (1.4)	377 (7.8)	242 (7.9)		380 (0.9)	
4.0	-	308 (1.6)	377 (7.8)	244 (7.2)	315 (0.9)	380 (2.8)	
10 hr.	I240	308 (1.7)	377 (7.4)	253 (6.3)	315 (1.1)	380 (3.6)	
24 hr.	I240	308 (1.8)	377 (6.8)	257 (5.5)	310 (1.3)	380 (3.8)	
	of 4-	keto-cholestane	(41)	$\Delta^{2,4}$ -6-Keto-cholestadien (45)			
Neutral	275 (5.2)	298 (2.8)	341 (7.0)			314 (7.4)	
2-3 min.	238 (3.2)	296 (0.6)	357 (4.8)		264 (1.6)	316 (6.9)	
4.0 hr.	233 (5.5)	250 (5.2)			268 (2.8)	316 (6.2)	
	258 (5.3)	296 (1.2)	357 (8.3)				
24 hr.	258 (4.7)	296 (1.1)	357 (7.7)	249 (6.3)	294 (3.3)	313 (3.8)	
		1]	360 (1.6)	405 (1.9)	
+1.5 hr. (60°)				250 (8.8)	340 (1.0)	405 (2.4)	

TABLE V—Continued

* Source of samples: #30, 44 and 45 from g; #31 and 43 from p; #32 from e; #33 and 42 from b; #34, 35 and 39 from a; #36 and 46 from d; #37 and 38 from f; #40, 40a, and 41 from i.—Compounds #31, 39 and 42 were further studied at the same alkalinity in aqueous 60 per cent ethanol and gave similar results, except that the bands near 260 m μ were shifted 2 m μ toward longer wave-lengths and the chromophore formation of compound #39 was slower (the highest extinction values were reached after approximately 24 hours instead of 10 hours).

† The 6β -acetate (from a) showed essentially the same rate.

[‡] The 6α -acetate (from a) showed essentially the same rate, except that the shift toward 241 m μ was attained only after 10 minutes; the absorbance maximum was located at 239.5 m μ after 2-3 minutes.

§ 3,6-Diketocholanic acid (#37) and 3,6-diketoallocholanic acid (#38) gave similar curves, the chromogen developed slower.

¶ The spectra (after 24 hr. exposure to the alkaline medium) were virtually the same when the solutions were diluted with 12 ml. of ethanol in a silica cell of 50 mm. light path.

I = Inflection.

reached their maximum in 10–14 hr. at room temperature. These compounds produced essentially the same absorptivity pattern as the corresponding 6α -hydroxyl derivatives after approximately 12 hr. at room temperature and 2–3 hr. at 60° and the 6β -hydroxy derivatives after 3–4 hr. at 60°. The spectral shift of

ANDRÉ S. MEYER

	Δ^4 -androstene-3-17-dione								
Exposure Time	6 β -Нус	lroxy derivative	e (31)	6α-Hydroxy derivative (34)					
	Max.	Min.	Max.	Max.	Min.	Max.			
30 min. 1 hr. 2 hr. 3 hr. 4 hr.	238 (13.0) 239 (10.6) 240 (9.2) 258 (8.6) 258 (8.9)	320 (1.5) 308 (1.4) 308 (2.3) 208 (2.6) Δ4-	377 (2.2) 377 (3.5) 377 (6.4) 377 (6.3) PREGNENE-17a, 2	243 (8.3) 257 (7.2) 258 (9.7) 258 (9.6) 258 (8.5) 21-DIOL-3, 20-DION	310 (1.0) 308 (1.2) 308 (1.7) 308 (2.3) 308 (2.2)	377 (3.5) 377 (5.4) 377 (7.5) 377 (7.4) 377 (6.5)			
	6 β -Hy	droxy derivative	e (32)	6α-Hydroxy 6, 21-diacetate (35)					
30 min. 1 hr. 2 hr. 3 hr. 4 hr.	$\begin{array}{c} 238 \hspace{0.1cm} (11.9) \\ 238 \hspace{0.1cm} (10.7) \\ 240 \hspace{0.1cm} (8.2) \\ 253 \hspace{0.1cm} (5.6) \\ 258 \hspace{0.1cm} (6.6) \end{array}$	315 313 (1.1) 310 (1.6)	377 (1.4) 377 (2.6) 377 (3.3) 377 (4.1)	240 (8.9) 245 (6.7) 254 (6.8) 258 (7.1) 258 (7.4)	325 320 312 (1.6) 310 (1.7) 310 (2.0)	377 (1.8) 377 (1.9) 377 (4.2) 377 (4.7) 377 (4.7)			

TABLE VI

Absorptions in m μ ($\epsilon \times 10^{-3}$) of 6-Hydroxylated Δ^{4} -3-Ketosteroids in Alkaline Ethanol at 60°C.

 3β -hydroxy- Δ^4 -6-keto-cholestene (#30) occurred at a similar rate to that of the 6β -hydroxy- Δ^4 -3-keto derivatives. The behavior of these two 6-oxygenated substances was not unexpected since isomerization of such structures to the 5α -3, 6-diketo derivatives in alkaline solution at elevated temperature were reported (35, 36, 36a). However, the more rapid shift of the Δ^4 -6 α -hydroxyl compounds, which would indicate that such isomerization would occur more easily (cf. 26), was surprising in view of a report (36) that no isomerization was observed nder alkaline conditions when Δ^4 -pregnene- 6α , 21-diol-3, 20-dione diacetate was hydrolyzed. The varied conditions might well explain the difference in result. Furthermore, the unsaturated Δ^4 -3, 6-diketosteroids (#42 and 43) turned yellow immediately after contact with alkaline ethanol and likewise exhibited absorption near 260 m μ and a second of approximately three-fourth its intensity near 380 m μ (Fig. 2). After acidification, these peaks shifted in all cases to 253 and 315 m μ , respectively.

Exposure of compound #43 to 0.07 N hydrochloric acid in 90% ethanol for 4 hr. at room temperature brought about the appearance of a second maximum at 315 m μ ($\epsilon 2.9 \times 10^3$); the maximum at 252 m μ decreased only slightly to an absorptivity of 10.3 \times 10³. Under the same acidic conditions, the spectrum of compound #34 was not significantly influenced over a period of 24 hr. However isomerization of 6 α - and 6 β -hydroxy- Δ^4 -3-keto compounds to the saturated derivatives was reported to occur when the compounds were refluxed in alcoholic solutions of 0.1–0.6 N hydrochloric acid (41, 36a, see also 36). When microgram quantities of the Δ^4 -3,6-diketosteroids (#42 and 43) were stored for about a month at 4° in a stoppered glass tube in dry state or in neutral ethanolic solution, the appearance of a second maximum near 315 m μ was also observed.



FIG. 2. SPECTRA OF 115.0 μ g. OF Δ^4 -ANDROSTENE-3,6,17-TRIONE (#43) IN 3.0 ML. OF SOL-VENT: --- in 95% ethanol; ______ in 0.066 N potassium hydroxide of 90% ethanol content (after 2-3 minutes); ••• 2 minutes after addition of the potassium hydroxide solution (2.8 ml.) acidified with 1 N hydrochloric acid (0.20 ml.) to pH 6.7 (ethanol content 85%); _____ acidified to pH 2.7; ____ ••• ___ same as latter conditions, steroid however exposed to the alkaline solution at 23° for 24 hours before acidification.

The action of alkali on 3,6-oxygenated steroids was noted previously. Windaus (37) found that 3,6-cholestanedione was not soluble in aqueous alkali. However, it dissolved in ethanolic potassium hydroxide producing a strongly yellow solution from which it was precipitated by the addition of large volumes of water, mostly as the unchanged material. Furthermore, Windaus (38) observed that solutions of aqueous potassium hydroxide had to have a strength of at least 20% in order to extract Δ^4 -cholestene-3, 6-dione from its ethereal solution. The freshly formed, only slightly soluble potassium salt (cf. 39) was easily decomposed to the free compound by addition of water. However, when the dried salt was stored for some days under a vacuum, it had undergone a rearrangement to the salt of a carboxylic acid of unknown nature. Ross (39) achieved total hydrolysis of the two dienol dibenzoates of Δ^4 -cholestene-3,6-dione when the substances were refluxed with an excess of N/10 alcoholic potassium hydroxide for 1 hr. However, in contrast to the 3,6-dibenzoyloxy- $\Delta^{2,4,6}$ -cholestatriene, no Δ^{4} -cholestene-3,6dione was obtained from the experiment with the 3,6-dibenzoyloxy- $\Delta^{3,5,7}$ cholestatriene. Heard and Sobel (40) noted the occurrence of a shift of the main resonance band in the ultraviolet from 251.5 m μ (ϵ 10.6 \times 10³) to 259 m μ (ϵ 10.0 \times 10³) when the spectrum of Δ^4 -cholestene-3,6-dione was examined in the presence of alkali rather than in neutral ethanol; with Δ^4 -cholesten-3-one no such shift was seen. Fieser (41) showed again the formation of various enol anions, all with ultraviolet absorptions in the region of 260 mµ ($\epsilon \sim 5 \times 10^3$), when hexane solutions of cholestane-3,6-dione, Δ^4 -cholestene-3,6-dione, Δ^4 cholesten-6 β -ol-3-one, and Δ^4 -cholesten-3 β -ol-6-one were shaken with 35% potassium hydroxide solution in aqueous methanol. Finally, it might be pertinent to add to this survey an observation of Greenhalgh, et al. (32). These workers found that Δ^5 -cholestene-3,7-dione, which absorbs in ethanolic solution maximally at 320 m μ and is probably present as the $\Delta^{3.5}$ -dien-7-one 3-enolate, produced also a bathochromic shift in its spectrum of approximately 70 m μ when alkali was added to the solution.

It is worthy of note that after exposure to alkaline ethanol a second absorption peak near 380 m μ was common to a number of different 6-ketosteroids. This band invariably shifted to approximately $315 \text{ m}\mu$ after acidification, but reverted to the previous absorptivity pattern after renewed addition of alkali. This typical spectrum was exhibited, as mentioned before, by the Δ^4 -3,6-diketosteroids (#42 and 43) immediately after contact with alkaline ethanol. Interestingly, the saturated 3,6-diketo derivatives also produced the same absorption pattern; however, approximately 10 hr. at room temperature were required until maximal intensity was reached. Again, approximately 10 hr. elapsed until the heteroannular Δ^4 -6-ketocholestene (#44) attained maximal absorptivity (at 380 m μ). On the other hand, $4\alpha(\text{or }\beta)$, 5α -dihydroxy-3, 6-diketocholestane (* 40 and 40a) developed maximal absorption (at 377 m μ) already within 2 hr. (cf. 42). The latter two were the only compounds of the series with a single peak which shifted to 317 mµ after acidification. As exemplified by Δ^4 -androstene-3,6,17-trione (Fig. 2), it can be seen that the intensities of the two bands, especially that near 315 m μ , are highly sensitive to the pH of the solution after acidification. The band near 315 m μ reached maximal intensity when neutrality was approached. 6-Ketocholestanol acetate (#46) did not develop intense ultraviolet absorption in alkaline solution.

Finally, the action of alkali on two further substances is listed in Table V. $\Delta^{2.4}$ -6-Ketocholestadien (#45), after alkaline treatment at 60°, had two maxima, at 250 and 405 m μ , which shifted after acidification to 239 and 327 m μ , respectively. In contrast, 3,4,6-triketocholestane (#41) produced, after alkaline treatment and acidification, a similar spectrum as it originally had exhibited in neutral ethanol (cf. 42), although during exposure to alkaline solution significant changes were observed. This indicates that the reaction products, which are probably enolic in nature, are formed at a relatively slow rate and that their formation is reversible.

Acknowledgment. This study was made possible by the generous contributions of those who furnished the required materials. Thanks are due to Dr. M. Ehrenstein for the compounds designated by (a), to Dr. L. F. Fieser (b), to Dr. J. Fried of Squibb Institute for Medical Research (c), to Dr. M. Gut (d), to Dr. D. H. Peterson of The Upjohn Company (e), to Dr. A. L. Raymond of G. D. Searle and Co. (f), to Dr. H. Reich (g), to Dr. T. Reichstein (h), to Dr. F. E. Romans (i), to Dr. I. V. Sollins of Chemical Specialties Co., Inc. (k), to Dr. F. Sondheimer of Syntex, S.A. (l) to Dr. A. F. St. Andre of Ciba Pharmaceutical Products, Inc. (m), to Dr. F. Ungar (n), and to Dr. A. Wettstein of Ciba Ltd. (o). The helpful assistance of Paul Skogstrom, who recorded a number of the spectra, is gratefully acknowledged. The investigation was supported by a grant of G. D. Searle and Co., Chicago, Illinois.

SUMMARY

The absorption spectra in alkaline medium of a number of Δ^4 -3-ketosteroids and their hydroxylated derivatives have been studied. Measurements were conducted in 0.066 N alkali in 90% ethanol at 23° and/or 60° over varying periods of time. Typical spectral shifts were observed which make it possible to use these data as a microanalytical tool for the characterization of various substituted Δ^4 -3-ketosteroids. In addition, the spectral changes of a number of 6-oxygenated steroids are reported.

SHREWSBURY, MASS.

REFERENCES

- JONES AND HERLING, J. Org. Chem., 19, 1252 (1954); ROSENKRANTZ, in Methods of Biochemical Analysis, Interscience Publishers, New York, 2; 1 (1955).
- (2) COATES, OFFNER, AND SIEGLER, J. Opt. Soc. Amer. 43, 984 (1953); ANDERSON AND WOODALL, Anal. Chem. 25, 1906 (1953).
- (3) DORFMAN, Chem. Revs., 53, 47 (1953).
- (4) BARTON AND KLYNE, Chemistry & Industry, 755 (1948); KLYNE, in Determination of Organic Structures by Physical Methods; BRAUDE AND NACHOD, editors, Academic Press, New York, 1955, p. 73.
- (5) SAVARD, J. Biol. Chem., 202, 457 (1953).
- (6) BARTON, J. Chem. Soc., 1027 (1953).

- (7) APPLEBY AND NORYMBERSKI, Biochem. J., 60, 453, 460 (1955).
- (8) WILSON AND FAIRBANKS, Arch. Biochem. and Biophys., 54, 440 (1955).
- (9) AXELROD, Anal. Chem., 27, 1308 (1955); ROMANOFF AND HUNT, Endocrinology, in press.
- (10) Edwards and Kellie, Biochem. J., 56, 207 (1954).
- (11) Cox, Biochem. J., 52, 339 (1952).
- (12) VESTERGAARD, Acta Endocrinol., 8, 193 (1951).
- (13) ZIMMERMANN AND PONTIUS, Z. physiol. Chem., 297, 157 (1954).
- (14) REICH, SANFILIPPO, AND CRANE, J. Biol. Chem., 198, 713 (1952); J. Org. Chem., 18, 882 (1953).
- (15) SILBER AND PORTER, J. Biol. Chem., 210, 923 (1954).
- (16) MATTOX, MASON, ALBERT, AND CODE, J. Am. Chem. Soc., 75, 4869 (1953).
- (17) CLARK, Nature, 175, 123 (1955).
- (18) BUSH AND SANDBERG, J. Biol. Chem., 205, 783 (1953).
- (19) ABELSON AND BONDY, Arch. Biochem. and Biophys., 57, 208 (1955).
- (20) UMBERGER AND CURTIS, J. Biol. Chem., 178, 275 (1949).
- (21) ZAFFARONI, J. Am. Chem. Soc., 72, 3828 (1950).
- (22) LINFORD AND FLEMING, Can. J. Med. Sciences, 31, 182 (1953).
- (23) BERNSTEIN AND LENHARD, J. Org. Chem., 19, 1269 (1954).
- (24) MEYER, HAYANO, LINDBERG, GUT, AND RODGERS, Acta Endocrinol., 18, 148 (1955).
- (25) MEYER, Experientia, 11, 99 (1955).
- (26) MEYER AND LINDBERG, Anal. Chem., 27, 813 (1955).
- (27) ROSENKRANTZ AND GUT, Science, 120, 3129 (1954).
- (28) MEYER, ROSENKRANTZ, AND PINCUS, Amer. Chem. Soc., 126th Meeting, Abstract p. 53-O, New York 1954.
- (29) SIMPSON, TAIT, WETTSTEIN, NEHER, VON EUW, SCHINDLER, AND REICHSTEIN, Helv. Chim. Acta, 37, 1163 (1954).
- (30) CROSS, EISEN, AND KEDERSHA, Anal. Chem., 24, 1049 (1952).
- (31) WINTERSTEINER AND RUIGH, J. Am. Chem. Soc., 64, 2453 (1942).
- (32) GREENHALGH, HENBEST, AND JONES, J. Chem. Soc. 2375 (1952).
- (33) JACKSON AMD JONES, J. Chem. Soc., 659 (1940).
- (34) SONDHEIMER, ST. KAUFMANN, ROMO, MARTINEZ, AND ROSENKRANZ, J. Am. Chem. Soc., 75, 4712 (1953).
- (35) HEILBRON, JONES, AND SPRING, J. Chem. Soc., 801 (1937).
- (36) HERZIG AND EHRENSTEIN, J. Org. Chem., 16, 1050 (1951).
- (36a) CAMERINO, ALBERTI, VERCELLONE, AND AMMANATI, Gazz. chim. ital., 84, 301 (1954).
- (37) WINDAUS, Ber., 36, 3752 (1903).
- (38) WINDAUS, Ber., 39, 2249 (1906).
- (39) Ross, J. Chem. Soc., 737 (1946).
- (40) HEARD AND SOBEL, J. Biol. Chem., 165, 687 (1946).
- (41) FIESER, J. Am. Chem. Soc., 75, 4377 (1953).
- (42) WINDAUS AND KUHB, Ann., 532, 53 (1937).