

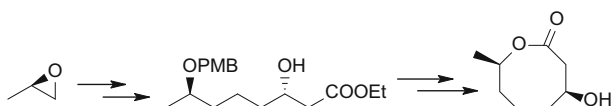
A concise stereoselective total synthesis of (–)-cephalosporolide D

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Received: 30 January 2015 / Accepted: 29 June 2015
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Abstract A concise stereoselective total synthesis of eight-membered lactone (–)-cephalosporolide D has been derived from inexpensive and commercially available starting material (*S*)-propylene epoxide. This concise synthesis utilizes Grignard reaction, Noyori asymmetric reduction, and Yamaguchi macrolactonization as key steps.

Graphical abstract



Keywords (*S*)-Propylene epoxide ·
Noyori asymmetric reduction ·
Yamaguchi macrolactonization · Stereoselective synthesis ·
(–)-Cephalosporolide D

Electronic supplementary material The online version of this article (doi:10.1007/s00706-015-1526-4) contains supplementary material, which is available to authorized users.

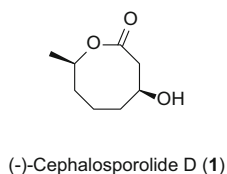
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Introduction

The interesting biological properties of medium-sized ring systems tend to attract the scientists from worldwide toward the synthesis of macrolides [1–4]. The various cephalosporolides represent very important attractive target molecules for the synthesis due to their biological activities which include mild inhibitory activity toward xanthine oxidase and 3 α -hydroxysteroid dehydrogenase and also showed antimicrobial activity against gram-positive cocci, fungi, and structural profiles [5–9]. Among them, the eight-membered ring lactone (–)-cephalosporolide D (**1**) was isolated from fermentation fungus, *Cephalosporium aphidicola* ACC 3490 [5] and shown in Fig. 1. The absolute configuration of **1** was assigned as (4*R*, 8*S*) with chemical, spectroscopic, and X-ray analyses. The first stereoselective total synthesis of (–)-cephalosporolide D (**1**) was done by Shiina et al. [10, 11].

The reported synthetic routes [12, 13] to (–)-cephalosporolide D that mainly involve the low yielding steps, multiple step sequences, and dependence on the chiral pool reagents are some of the disadvantages associated with the earlier methods. We previously reported stereoselective total synthesis of (+)-cephalosporolide D from inexpensive starting material racemic propylene epoxide which includes α -aminoxylation catalyzed by L-proline, followed by in situ reduction using NaBH₄ and Yamaguchi macrolactonization as key steps. This synthesis compromises the long chain reaction with low yields. In continuation of our research work on the synthesis of biologically active natural products, herein we report an efficient straightforward and concise stereoselective total synthesis of (–)-cephalosporolide D starting from commercially available starting material (+)-propylene epoxide with overall high yield.

Fig. 1 Structure of (–)-cephalosporolide D

Results and discussion

Our retrosynthetic analysis of **1** is outlined in Scheme 1. The macrolide **1** could be obtained by Yamaguchi macrolactonization followed by reductive cleavage of benzyl ether of hydroxyl acid **3** which in turn could be synthesized from corresponding ester **4**. The ester **4** could be obtained from (*S*)-propylene epoxide.

The total synthesis of **1** was initiated with (*S*)-propylene epoxide (**5**) which was obtained from (±)-propylene oxide [14–16] and shown in Scheme 2. Regioselective ring opening of epoxide **5** with the Grignard reagent derived from homoallyl bromide at -78°C for 2 h gave secondary alcohol **6** [17], which on subsequent treatment with PMBB and NaH in tetrahydrofuran solvent at 0°C to 25°C for 8 h afforded **7** in 81 % yield. Further extension of the olefin **7** by ozonolysis and subsequent addition reaction with EtOAc on the resulting aldehyde (EtOAc/LiHMDS/THF/ -78°C) afforded a mixture of secondary alcohol product (72 %). In order to increase the diastereoselectivity in favor of the requisite stereocenter (*anti* to the existing one), an oxidation–reduction protocol was undertaken. Hence, secondary alcohol **4** was oxidized with PDC in dry CH_2Cl_2 at reflux temperature for 10 h afforded the corresponding keto compound, which was subjected to Noyori asymmetric hydrogenation reduction [18] with RuCl_2 [(*S*)-binap] in MeOH at 30°C for 36 h afforded alcohol **9** in 81 % (94 % de). The above obtained alcohol **9** on treatment with NaH and benzyl bromide at 0°C gave the Bn ether **10** in 89 % yield. Hydrolysis of **10** in the basic condition (LiOH in THF:MeOH:H₂O-3:1:1) afforded acid

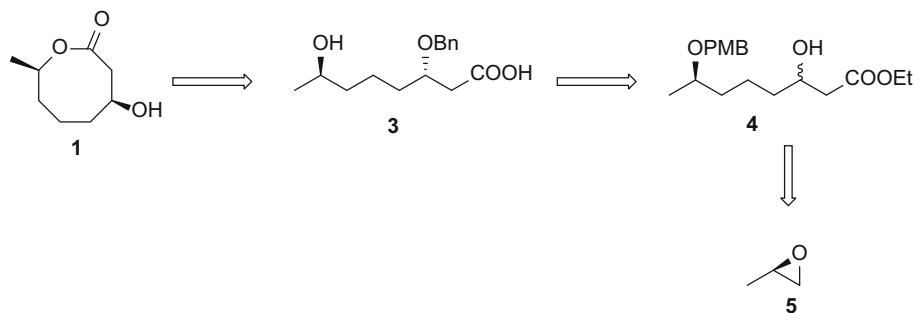
11, which on selective cleavage of PMB ether in the presence of DDQ in aq. CH_2Cl_2 gave the hydroxyacid **3** in 81 % yield [13]. Hydroxyacid **3** was subjected to macrolactonization under Yamaguchi high dilution conditions [19] using 2,4,6-trichlorobenzoyl chloride and Et_3N in dry THF to afford the lactone **12** in 69 % yield [13]. Finally, oxidative deprotection of Bn group in **12** using TiCl_4 in CH_2Cl_2 gave (–)-cephalosporolide D (**1**) in 79 % yield. The ^1H NMR and ^{13}C NMR spectral data and optical rotation value of synthetic **1** were in good accord with those of the natural product [12] (Scheme 2).

Conclusion

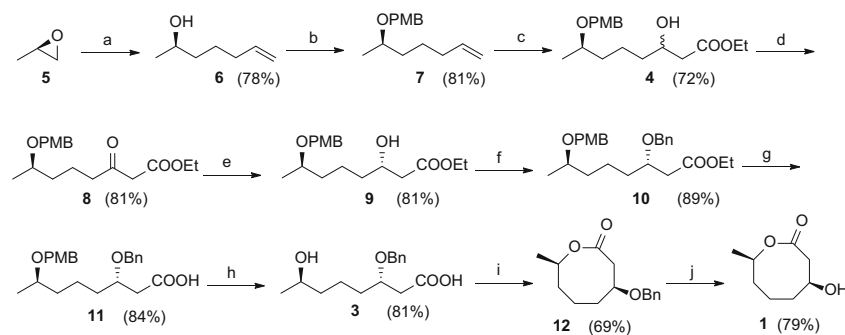
We developed a concise route for the synthesis of (–)-cephalosporolide D (**1**) by a common synthetic strategy. The synthesis of this lactone has been achieved by convergent approach starting from (*S*)-propylene oxide followed by Grignard reaction, Noyori Asymmetric reduction, and Yamaguchi macrolactonization as key steps.

Experimental

All chemicals and reagents were obtained from Aldrich (Sigma-Aldrich, St. Louis, MO, USA), Lancaster (Alfa Aesar, Johnson Matthey Company, Ward Hill, MA, USA) and were used without further purification. Reactions were monitored by TLC, performed on silica gel glass plates containing 60 F-254, and visualization on TLC was achieved by UV light or iodine indicator. ^1H and ^{13}C NMR spectra were recorded on INOVA (400 MHz) or Gemini Varian-VXR-unity (300 MHz) or Bruker UXNMR/XWIN-NMR (300 MHz) instruments. Chemical shifts (δ) are reported in ppm downfield from internal TMS standard. ESI spectra were recorded on Micro mass, Quattro LC using ESI⁺ software with capillary voltage 3.98 kV and

Scheme 1

Scheme 2



(a) Homoallyl bromide, Mg, dry ether, -78°C , 2 h; (b) PMBBBr, NaH, THF, 0°C to 25°C , 8 h; (c) i) O_3 , CH_2Cl_2 , -78°C , 15 min; ii) EtOAc, LiHMDS, THF, -78°C , 1 h; (d) PDC, CH_2Cl_2 , reflux, 10 h; (e) RuCl_2 [(*S*)-binap], MeOH, 30°C , 36 h; (f) benzyl bromide, NaH, THF, 0°C to 25°C , 4 h; (g) LiOH, THF:MeOH:H₂O (3:1:1), 25°C , 4 h; (h) DDQ, CH_2Cl_2 :H₂O (19:1), 25°C , 3 h; (i) 2,4,6-trichlorobenzoyl chloride, Et₃N, THF, DMAP, toluene, 25°C , 12 h; (j) TiCl_4 , CH_2Cl_2 , 0°C to 25°C , 3 h.

ESI mode-positive ion trap detector. Melting points were determined with an Electrothermal melting point apparatus.

(R)-Hept-6-en-2-ol (**6**) [17]

A suspension of 3.8 g Mg (165.51 mmol, 3.0 eq) and 30 cm³ dry ether was treated with 8.4 cm³ homoallyl bromide (82.75 mmol, 1.5 eq) at 25°C and stirred for 30 min. It was cooled to -78°C , and a solution of 4 cm³ **5** (55.17 mmol, 1.0 eq) in 10 cm³ dry ether was added dropwise and the mixture was stirred at 25°C for 2 h. The reaction mixture was quenched with 10 cm³ aq. NH_4Cl solution and extracted with ether (2×50 cm³). Combined extracts were washed with 30 cm³ brine, dried (Na_2SO_4), and concentrated and purified the residue by column chromatography (silica gel, 60–120 mesh, 7 % EtOAc in pet. ether) to afford alcohol **6** (4.9 g, 78 %) as a colorless liquid.

(R)-1-[(Hept-6-en-2-yloxy)methyl]-4-methoxybenzene (**7**, C₁₅H₂₂O₂)

To a cooled (0°C) solution of 4.8 g **6** (42.10 mmol, 1.0 eq) in 30 cm³ dry THF, 4.0 g NaH (168.4 mmol, 4.0 eq) was added, stirred for 30 min, and treated with a solution of 10.0 g PMBBBr (50.52 mmol, 1.2 eq) in 40 cm³ dry THF. After 7.5 h stirring at 25°C , the reaction mixture was quenched with 30 cm³ sat. NH_4Cl solution and extracted with ethyl acetate (2×100 cm³). The organic

layers were washed with water (2×50 cm³), 50 cm³ brine, and dried (Na_2SO_4). Solvent was evaporated under reduced pressure and purified the residue by column chromatography (silica gel, 60–120 mesh, 5 % EtOAc in pet. ether) to furnish **7** (8.01 g, 81 %) as a yellow liquid. $[\alpha]_{\text{D}}^{25} = +43.8$ ($c = 0.91$, CHCl_3); ^1H NMR (500 MHz, CDCl_3): $\delta = 7.18$ (d, 2H, $J = 8.5$ Hz, ArH-PMB), 6.79 (d, 2H, $J = 8.5$ Hz, ArH-PMB), 5.84 (dddd, 1H, $J = 6.6$, 10.2, 13.4, 16.8 Hz, olefinic), 4.98–4.89 (m, 2H, olefinic), 4.46 (d, 1H, $J = 11.3$ Hz, benzylic), 4.34 (d, 1H, $J = 11.3$ Hz, benzylic), 3.78 (s, 3H, $-\text{OCH}_3$), 3.46 (m, 1H, $-\text{OCH}$), 2.01 (m, 2H, $-\text{CH}_2$), 1.61–1.41 (m, 4H, 2 CH₂), 1.14 (d, 3H, $J = 6.0$ Hz, $-\text{CH}_3$) ppm; ^{13}C NMR (100 MHz, CDCl_3): $\delta = 159.0$, 138.9, 129.7, 129.1, 114.4, 113.7, 74.3, 69.9, 55.2, 36.1, 33.8, 24.8, 19.6 ppm; IR (neat): $\bar{\nu} = 3070$, 2932, 2829, 1635, 1466, 1134, 1015 cm^{−1}; ESI-MS: $m/z = 257$ [(M + Na)⁺].

(R)-Ethyl 3-hydroxy-7-(4-methoxybenzyloxy)octanoate (**4**, C₁₈H₂₈O₅)

Ozone was bubbled through a cooled (-78°C) solution of 5.0 g **7** (21.36 mmol, 1.0 eq) in 50 cm³ CH_2Cl_2 until the pale blue color persisted. Excess ozone was removed with 2 cm³ Me₂S and stirred for 30 min at 0°C . The reaction mixture was concentrated under reduced pressure to give aldehyde, which was used for further reaction. To a stirred solution of 10.5 g HMDS (65.24 mmol, 3.5 eq) in 25 cm³

anhydrous THF at 0 °C, 39 cm³ *n*-BuLi (2.0 M in *n*-hexane, 74.56 mmol, 4.0 eq) was added and reaction mixture stirred for 30 min at the same temperature, after that reaction mixture taken to −78 °C and 6.5 g EtOAc (74.56 mmol, 4.0 eq) was added in one pot by syringing. Stirring was continued at −78 °C for 1 h, and then 4.4 g aldehyde (18.64 mmol, 1.0 eq) dissolved in 25 cm³ dry THF was added to above reaction mixture at −78 °C and reaction mixture stirred at the same temperature for 1 h. The reaction mixture was quenched with 50 cm³ sat. aq. NH₄Cl solution at 0 °C, extracted with EtOAc (2 × 100 cm³), washed with 50 cm³ water, 50 cm³ brine, concentrated the organic layer, and purified by column chromatography (silica gel, 60–120 mesh, 12 % EtOAc in pet. ether) to afford **4** (2.17 g) in 72 % yield. ¹H NMR (300 MHz, CDCl₃): δ = 7.21 (d, 2H, *J* = 8.8 Hz, ArH-PMB), 6.81 (d, 2H, *J* = 8.8 Hz, ArH-PMB), 4.44 (d, 1H, *J* = 11.1 Hz, benzylic), 4.38 (d, 1H, *J* = 11.1 Hz, benzylic), 4.14 (q, 2H, *J* = 7.0 Hz, −OCH₂), 3.74 (s, 3H, −OCH₃), 3.66 (m, 1H, −OCH), 3.44 (m, 1H, −OCH), 2.76 (m, 2H, −CH₂), 1.58–1.21 (m, 6H, 3 −CH₂), 1.27 (t, 3H, *J* = 7.0 Hz, −OCH₂CH₃) 1.12 (d, 3H, *J* = 6.1 Hz, −CH₃) ppm; ESI-MS: *m/z* = 325 [(M + H)⁺].

(R)-Ethyl 7-(4-methoxybenzyloxy)-3-oxooctanoate (**8**, C₁₈H₂₆O₅)

To the stirred solution of 2.5 g **4** (7.71 mmol, 1.0 eq) in 25 cm³, anhydrous CH₂Cl₂ MS powder and 8.7 g PDC (23.14 mmol, 3.0 eq) were added, to that (4–5 drops) Ac₂O was added and the whole reaction mixture was put for reflux for 10 h. The reaction mixture was concentrated and purified by column chromatography (silica gel, 60–120 mesh, 10 % EtOAc in pet. ether) to afford **8** (2.01 g, 81 %) as a yellow colored thick syrup. ¹H NMR (300 MHz, CDCl₃): δ = 7.23 (d, 2H, *J* = 8.6 Hz, ArH-PMB), 6.87 (d, 2H, *J* = 8.6 Hz, ArH-PMB), 4.47 (s, 2H, *J* = 10.8 Hz, benzylic), 4.11 (q, 2H, *J* = 6.9 Hz, −OCH₂), 3.67 (s, 3H, −OCH₃), 3.36 (m, 1H, −OCH), 3.31 (s, 2H, −CH₂), 2.19 (t, 2H, *J* = 6.6 Hz, −CH₂), 1.44–1.24 (m, 4H, 2 CH₂), 1.21 (t, 3H, *J* = 6.9 Hz, −OCH₂CH₃) 1.11 (d, 3H, *J* = 6.1 Hz, −CH₃) ppm; ¹³C NMR (100 MHz, CDCl₃): δ = 199.8, 170.8, 158.3, 130.1, 129.3, 113.3, 79.1, 62.2, 56.1, 49.3, 44.6, 36.7, 22.8, 22.3, 14.6 ppm; ESI-MS: *m/z* = 323 [(M + H)⁺].

(3S,7R)-Ethyl 3-hydroxy-7-(4-methoxybenzyloxy)octanoate (**9**, C₁₈H₂₈O₅)

To the stirred solution of 1.9 g **8** (4.96 mmol, 1.0 eq) in 5.0 cm³, anhydrous methanol was treated with catalytic amount of RuCl₂ [(*S*)-binap] [17] at 30 °C for 36 h under hydrogen atmosphere. After completion of the reaction, it was filtered, evaporated, and purified by column chromatography (silica gel, 60–120 mesh, 15 % EtOAc in pet. ether) to give **9** (1.56 g, 81 %) as a pale yellow syrup.

[α]_D = −114.3 (*c* = 1.1, CHCl₃); ¹H NMR (CDCl₃, 300 MHz): δ = 7.20 (d, 2H, *J* = 8.6 Hz, ArH-PMB), 6.79 (d, 2H, *J* = 8.6 Hz, ArH-PMB), 4.47 (d, 1H, *J* = 11.1 Hz, benzylic), 4.36 (d, 1H, *J* = 11.1 Hz, benzylic), 4.14 (q, 2H, *J* = 7.0 Hz, −OCH₂), 3.74 (s, 3H, −OCH₃), 3.63 (m, 1H, −OCH), 3.39 (m, 1H, −OCH), 2.71 (dd, 1H, *J* = 8.0, 15.3 Hz, −CH), 2.50 (dd, 1H, *J* = 5.2, 15.3 Hz, −CH), 1.58–1.29 (m, 6H, 3 CH₂), 1.27 (t, 3H, *J* = 7.1 Hz, −OCH₂CH₃) 1.11 (d, 3H, *J* = 6.1 Hz, −CH₃) ppm; ¹³C NMR (100 MHz, CDCl₃): δ = 171.3, 159.3, 141.2, 130.1, 129.8, 114.3, 75.6, 72.2, 66.8, 59.8, 56.1, 42.4, 37.3, 34.7, 26.0, 23.2, 20.9, 14.7 ppm; IR (neat): $\bar{\nu}$ = 3354, 2928, 2862, 2105, 1613, 1514 cm^{−1}; ESI-MS: *m/z* = 347 [(M + Na)⁺].

(3S,7R)-Ethyl 3-(benzyloxy)-7-(4-methoxybenzyloxy)octanoate (**10**, C₂₅H₃₄O₅)

To a cooled (0 °C) suspension of 0.41 g NaH (17.28 mmol, 4.0 eq 60 % w/w dispersion in paraffin oil) in 15 cm³ dry THF, a solution of 1.4 g **9** (4.32 mmol, 1.0 eq) in 5 cm³ dry THF was added and stirred for 30 min. After that 0.78 cm³ BnBr (6.48 mmol, 1.2 eq) was added dropwise at 0 °C and stirred for 4 h at 25 °C. Saturated aq. NH₄Cl solution (20 cm³) was added dropwise at 0 °C followed by 30 cm³ EtOAc. Organic layers were washed with 20 cm³ water, 20 cm³ brine, dried (Na₂SO₄), and evaporated. The residue was purified by column chromatography (silica gel, 60–120 mesh, 8 % EtOAc in pet. ether) to give **10** (1.59 g, 89 %) as a light yellow syrup. [α]_D = −166.3 (*c* = 1.0, CHCl₃); ¹H NMR (CDCl₃, 300 MHz): δ = 7.27 (m, 5H, ArH-Bn), 7.18 (d, 2H, *J* = 8.3 Hz, ArH-PMB), 6.84 (d, 2H, *J* = 8.3 Hz, ArH-PMB), 4.56 (d, 1H, *J* = 11.4 Hz, benzylic), 4.48 (s, 2H, benzylic), 4.36 (d, 1H, *J* = 11.4 Hz, benzylic), 4.11 (q, 2H, *J* = 7.3 Hz, −OCH₂), 3.69 (s, 3H, −OCH₃), 3.59 (m, 1H, −OCH), 3.39 (m, 1H, −OCH), 2.71 (dd, 1H, *J* = 8.1, 15.1 Hz, −CH), 2.53 (dd, 1H, *J* = 5.1, 15.1 Hz, −CH), 1.66–1.31 (m, 6H, 3 CH₂), 1.27 (t, 3H, *J* = 7.0 Hz, −OCH₂CH₃) 1.14 (d, 3H, *J* = 6.1 Hz, −CH₃) ppm; ¹³C NMR (CDCl₃, 100 MHz): δ = 170.9, 159.6, 146.7, 139.3, 129.8, 129.6, 129.1, 128.5, 128.0, 127.8, 114.0, 75.9, 73.1, 71.3, 69.9, 60.3, 55.4, 40.4, 36.8, 36.0, 23.3, 21.2, 14.9 ppm; IR (neat): $\bar{\nu}$ = 2928, 2862, 2105, 1613, 1514 cm^{−1}; ESI-MS: *m/z* = 437 [(M + Na)⁺].

(3S,7R)-3,7-Bis(4-methoxybenzyloxy)octanoic acid (**11**, C₂₃H₃₀O₅)

To a solution of 1.65 g **10** (3.98 mmol, 1.0 eq) in 10 cm³ THF: MeOH: water (3:1:1), 0.29 g LiOH (18.48 mmol, 4.6 eq) was added and stirred at 25 °C for 4 h. The pH of reaction mixture was adjusted to acidic with 1 N HCl solution and extracted with 30 cm³ ethyl acetate. Organic layers were washed with 15 cm³ water, 15 cm³ brine, dried (Na₂SO₄), evaporated under reduced pressure, and purified the residue by column chromatography (silica gel, 60–120

mesh, 30 % EtOAc in pet. ether) to give **11** (1.29 g, 84 %) as a colorless oil. $[\alpha]_D^{25} = +14.6$ ($c = 0.6$, CHCl_3); ^1H NMR (CDCl_3 , 300 MHz): $\delta = 7.31$ (m, 5H, ArH-Bn), 7.20 (d, 2H, $J = 8.6$ Hz, ArH-PMB), 6.90 (d, 2H, $J = 8.6$ Hz, ArH-PMB), 4.57 (d, 1H, $J = 11.4$ Hz, benzylic), 4.46 (s, 2H, benzylic), 4.37 (d, 1H, $J = 11.4$ Hz, benzylic), 3.71 (s, 3H, $-\text{OCH}_3$), 3.63 (m, 1H, $-\text{OCH}$), 3.37 (m, 1H, $-\text{OCH}$), 2.59 (m, 2H, $-\text{CH}_2$), 1.63–1.41 (m, 4H, 3 CH_2), 1.32–1.20 (m, 2H, $-\text{CH}_2$) 1.16 (d, 3H, $J = 6.0$ Hz, $-\text{CH}_3$) ppm; ^{13}C NMR (75 MHz, CDCl_3): $\delta = 174.8$, 159.8, 138.9, 129.7, 129.4, 128.8, 128.5, 128.0, 114.4, 76.1, 73.8, 72.7, 70.1, 56.3, 41.1, 36.3, 36.1, 22.8, 21.4 ppm; IR (neat): $\bar{\nu} = 3540$, 3031, 2930, 2857, 1710, 1097 cm^{-1} ; ESI-MS: $m/z = 409$ $[(M + \text{Na})^+]$.

(3S,7R)-3-(Benzyloxy)-7-hydroxyoctanoic acid (**3**) [13]

To a solution of 1.2 g **11** (3.10 mmol, 1.0 eq) in 10 cm^3 aq. CH_2Cl_2 (19:1), 1.05 g DDQ (4.66 mmol, 1.5 eq) was added and stirred at 25 °C for 3 h. The reaction mixture was quenched with 5 cm^3 sat. NaHCO_3 solution, filtered, and washed with 10 cm^3 CH_2Cl_2 . The filtrate was washed with 3 cm^3 water, 3 cm^3 brine, dried (Na_2SO_4), and evaporated under reduced pressure. The residue was purified by column chromatography (silica gel, 60–120 mesh, 50 % EtOAc in pet. ether) to furnish **3** (0.66 g, 81 %). $[\alpha]_D^{25} = -11.1$ ($c = 1.1$, CHCl_3).

(4S,8R)-4-(Benzyloxy)-8-methyloxocan-2-one (**12**) [13]

To a stirred solution of 0.42 g **3** (1.57 mmol, 1.0 eq) and 0.66 cm^3 Et_3N (4.71 mmol, 3.0 eq) in 5 cm^3 dry THF, a solution of 0.36 cm^3 2,4,6-trichlorobenzoyl chloride (2.36 mmol, 1.5 eq) in 1 cm^3 dry THF was added. The resulting mixture was stirred for 2 h at 25 °C under nitrogen atmosphere and evaporated to afford the mixed anhydride. It was diluted with 10 cm^3 toluene and filtered quickly through Celite. The filtrate was added dropwise to a stirred solution of 0.38 g DMAP (3.14 mmol, 2.0 eq) in 350 cm^3 toluene at 90 °C over a period of 8 h. After the complete addition, the reaction mixture was stirred at 100 °C for 2 h. It was cooled, washed with 40 cm^3 7 % aq NaHCO_3 , 40 cm^3 2 M aqueous HCl, 40 cm^3 brine, and dried (Na_2SO_4). The organic layer was evaporated and the obtained residue purified by column chromatography (silica gel, 60–120 mesh, 15 % EtOAc in pet. ether) to give **12** (0.27 g, 69 %) as a syrup. $[\alpha]_D^{25} = -69.6$ ($c = 0.9$, CHCl_3).

(–)-Cephalosporolide D (**1**)

To a stirred solution of 0.14 g **12** (0.56 mmol, 1.0 eq) in 2 cm^3 CH_2Cl_2 , 0.11 g TiCl_4 (0.56 mmol, 1.0 eq) in 1 cm^3 CH_2Cl_2 was added at 0 °C and stirred at 25 °C for 2 h. The reaction mixture was treated with 5 cm^3 saturated NaHCO_3 solution and extracted with CHCl_3 (3 \times 10 cm^3). The combined organic layers were washed with 10 cm^3 water, 10 cm^3 brine, dried (Na_2SO_4), and concentrated. The residue was purified by column chromatography (silica gel, 60–120 mesh, 1:3 EtOAc:*n*-hexane) to afford **1** (70 mg, 79 %). $[\alpha]_D^{25} = -38.7$ ($c = 1.6$, CHCl_3); ^1H NMR and ^{13}C NMR spectra were in agreement with those of the natural product [12].

References

1. Celmer WD (1971) Pure Appl Chem 28:413
2. Keller-Schierlein W (1973) Fortschr Chem Org Naturst 30:313
3. Boeckman RK Jr, Fayos J, Clardy J (1974) J Am Chem Soc 96:5954
4. Omura S, Nakagawa A (1975) J Antibiot 28:401
5. Ackland MJ, Hanson JR, Hitchcock PB, Ratcliffe AH (1985) J Chem Soc Perkin Trans 1:843
6. Farooq A, Gordon J, Hanson JR, Takahashi JA (1995) Phytochemistry 38:557
7. Li X, Yao Y, Zheng Y, Sattler I, Lin W (2007) Arch Pharmacol Res 30:812
8. Murthy IS, Sreenivasulu R, Alluraiah G, Raju RR (2014) Lett Org Chem 11:327
9. Oller-Lopez JL, Iranzo M, Mormeneo S, Oliver E, Cuerva JM, Oltra JE (2005) Org Biomol Chem 3:1172
10. Shiina I, Fukuda Y, Ishii T, Fujisawa H, Mukaiyama T (1998) Chem Lett 27:831
11. Shiina I, Fujisawa H, Ishii T, Fukuda Y (2000) Heterocycles 52:1105
12. Sabitha G, Reddy TR, Ramesh M, Srinivas C, Yadav JS (2011) Bull Chem Soc Jpn 84:229
13. Harkala KJ, Eppakayala L, Maringanti TC (2014) Eur J Chem 5:127
14. Schaus SE, Brandes BD, Larrow JF, Tokunaga M, Hansen KB, Gould AE, Furrow ME, Jacobsen EN (2002) J Am Chem Soc 124:1307
15. Tokunaga M, Larrow JF, Kakiuchi F, Jacobsen EN (1997) Science 277:936
16. Louis JT, Nelson WL (1987) J Org Chem 52:1309
17. Dixon DJ, Ley SV, Tate EW (2000) J Chem Soc Perkin Trans 1:2385
18. Noyori R, Ohkuma T, Kitamura M, Takaya H, Sayo N, Kumobayashi H, Akutagawa S (1987) J Am Chem Soc 109:5856
19. Inagawa J, Hirata K, Saeki H, Katsuki T, Yamaguchi M (1979) Bull Chem Soc Jpn 52:1989