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# Direct Experimental Characterization of Glycosyl Cations by Infrared Ion Spectroscopy.

Hidde Elferink,<sup>‡,a</sup> Marion E. Severijnen,<sup>‡,b</sup> Jonathan Martens,<sup>b</sup> Rens A. Mensink<sup>a</sup>, Giel Berden,<sup>b</sup> Jos Oomens,<sup>b</sup> Floris P. J. T. Rutjes,<sup>a</sup> Anouk M. Rijs,<sup>\*,b</sup> and Thomas J. Boltje<sup>\*,a</sup>

<sup>a</sup>Radboud University, Institute for Molecules and Materials, Synthetic Organic Chemistry, Heyendaalseweg 135, 6525 AJ, Nijmegen, The Netherlands.

<sup>b</sup>Radboud University, Institute for Molecules and Materials, FELIX laboratory, Toernooiveld 7c, 6525 ED, Nijmegen, The Netherlands.

Supporting Information Placeholder

**ABSTRACT:** Glycosyl cations are crucial intermediates formed during enzymatic and chemical glycosylation. The intrinsic high reactivity and short life-time of these reaction intermediates makes them very challenging to characterize using spectroscopic techniques. Herein, we report the use of collision induced dissociation tandem mass spectrometry to generate glycosyl cations in the gas phase followed by infrared ion spectroscopy using the FELIX infrared free electron laser. The experimentally observed IR spectra were compared to DFT calculated spectra enabling the detailed structural elucidation of elusive glycosyl oxocarbenium and dioxolenium ions.

The principle challenge in chemical oligosaccharide synthesis is the stereoselective synthesis of glycosidic bonds.<sup>1</sup> Glycosidic bonds connecting monosaccharides can exist as  $\alpha$ - or  $\beta$ -diastereomers. The most common approach to chemically prepare glycosidic bonds is a nucleophilic substitution reaction between a glycosyl donor carrying an anomeric leaving group, and a glycosyl acceptor containing a nucleophilic alcohol. Depending on the nature of the glycosyl donor, acceptor and reaction parameters, the mechanism of a glycosylation is best described as a continuum between S<sub>N1</sub>-like and S<sub>N2</sub>-like reaction pathways (Scheme 1).<sup>2</sup>

Scheme 1: Reactive intermediates in glycosylation reactions and their respective reaction pathways



LG = Leaving group, E = Electrophile, A = Anion, P = Protecting group Stereoselective glycosylation can be achieved by the  $S_N2$ -like displacement of a contact ion pair or covalent intermediate such as glycosyl triflates<sup>3</sup>, sulfonium ions<sup>4,5</sup> and dioxolenium ions<sup>6</sup> (Scheme 1). Reactions via this pathway are well studied as the reaction intermediates can be characterized by low temperature NMR spectroscopy.<sup>3,7,8</sup> Loss of stereoselectivity during the displacement of the contact ion pair intermediates is often attributed to an alternative reaction pathway via

the solvent separated ion pair, also known as the oxocarbenium ion. Hence, to fully understand the mechanistic pathways of glycosylation reactions, it is crucial to elucidate the structure of its intermediates such as glycosyl oxocarbenium ions. However, glycosyl oxocarbenium ions are very challenging to characterize due to their intrinsic high reactivity, short life-times and equilibrium with the corresponding contact ion pair. Very recently, NMR spectroscopy of glycosyl oxocarbenium ions was reported in super acid solution.<sup>9</sup> However, the harsh conditions to generate the oxocarbenium ions are not compatible with all types of protecting groups commonly used in oligosaccharide synthesis. Tandem mass spectrometry (MS/MS) can be employed to generate glycosyl cations in complete isolation and their gas-phase fragmentation has been extensively studied.<sup>10-14</sup> Recently, the use of MS in combination with infrared (IR) ion spectroscopy has emerged as a powerful method to determine the gas-phase structures of molecular ions in MS/MS 15-20 and to probe glycan structure 21-26.

Herein we report the use of collisional dissociation to generate glycosyl cations followed by characterization using infrared (IR) ion spectroscopy. Electrospray ionization (ESI) of four glycosyl donors was employed to form a parent ion which was fragmented using collision induced dissociation (CID) affording the desired glycosyl cations. Crucially, the absence of a counter ion and solvent led to "naked" cations that could be characterized using IR ion spectroscopy. The IR spectra showed specific diagnostic vibrational bands enabling the assignment of oxocarbenium and dioxolenium ions. In addition, a mixture of two dioxolenium ion isomers could be resolved thereby providing insight into the dynamics of these elusive intermediates.

We synthesized mannosyl donors 1-4 as substrates and the commonly used benzyl ethers were replaced by methyl ethers in order to eliminate IR signals resulting from the aromatic system (Scheme 2). Thioglycosides were selected as they are frequently used glycosyl donors and have been used to study the formation and fragmentation of glycosyl cations using CID.<sup>27-29</sup>

Scheme 2: Structures of the mannosyl donors used in this study



Ionization of mannosyl donor 1 was achieved using ESI resulting in the parent ion [M+NH<sub>4</sub>]<sup>+</sup> with a m/z ratio of 458. Subsequent isolation and CID led to the formation of various fragment ions, most notably the mannosyl cation with m/z ratio of 331 (Fig. S1). This fragment ion was isolated in a quadrupole ion trap<sup>30</sup> and subsequently characterized by IR ion spectroscopy using the FELIX laser operating in the 700-1850 cm<sup>-1</sup> region (for details see SI). The mannosyl cation derived from 1, produced an IR spectrum containing a wealth of well-resolved and diagnostic peaks (Fig. 1, black line). To explore the generality of this approach, gluco- and galactoside were also prepared, fragmented and characterized by IR ion spectroscopy producing similar, yet distinct IR spectra (Fig. S2).

To obtain structural information on the formed ions, infrared ion spectroscopy is combined with high level ab initio calculations.<sup>31</sup> Calculated IR spectra from selected low energy structures were compared with the experimental IR spectra for structural assignments.



Figure 1: Comparison of the calculated spectra (filled) of 2-Oacetyl- (1a) or 3-O-acetyl- (1b) participation with the measured IR ion spectrum of mannosyl cation derived from 1 (black line in both panels).

In our spectral assessment, peak positions (cm<sup>-1</sup>) are most important since peak intensities in the calculated spectrum are based on linear IR absorption, whereas the measured spectra rely on multiple-photon absorption.Initial optimization indicated that DFT calculations at the  $B_3LYP/6-31++G(d, d)$ p) level, with relative energies determined using MP<sub>2</sub>/6-311++G(2d, 2p), proved to provide the most representative spectra (Fig. S<sub>3</sub>). It is well established that acetyl protected donors produce dioxolenium ions via neighboring group participation.<sup>32</sup> The experimental IR spectrum was compared with the calculated IR spectra of the lowest energy isomers of four different structural families with m/z=331. Participation of the C-2, C-3, C-4 or C-6 acetyl ester was considered together with the unstabilized oxocarbenium ion (Fig. S<sub>4</sub>). The lowest energy structure resulted from C-2 acetyl participation followed by C-3 participation (+5.8 kJ/mol), whilst C-4/C-6 participation or no participation display significant higher energies (+34.2-40.3 kJ/mol). Based on the energies themselves, the latter three structures are unlikely to be present. This is further supported by comparing the calculated and experimental IR spectra. The calculated C=O<sup>+</sup> stretch of the oxocarbenium ion would appear at 1616  $\text{cm}^{-1}$  (Fig. S4), however, this signal is absent in the measured spectrum. The participation of acetyl esters can be assigned by the calculated O-C=O<sup>+</sup> stretch (~1540 cm<sup>-1</sup>), and the C-C stretch (~1495 cm<sup>-1</sup>) modes of the dioxolenium ion, whereas the C=O stretch vibration of the non-participating acetyl esters are found at 1725-1800 cm<sup>-1</sup>. The C-6 acetyl C=O stretch (1725 cm<sup>-1</sup>) is distinct from the other esters and clearly visible in the measured IR spectrum. Calculations show that this signature is lost upon C-6 acetyl participation thereby excluding it (Fig. S<sub>4</sub>). Dioxolenium ions resulting from either C-2 (1a) or C-3 (**1b**) ester participation both produce low energy structures (Fig. 1). However, when comparing the calculated peaks originating from the O-C=O<sup>+</sup> stretch and C-C stretch vibrations of the dioxolenium ion (1538 and 1496 cm<sup>-1</sup>), it is clear that C-2 participation is in much better agreement with the experimentally observed spectrum (1a, Fig 1A) than C-3 participation (**1b**, Fig 1B). Finally, the assignment that the activation of 1 leads to a C-2-dioxolenium ion is consistent with previous NMR studies.<sup>32</sup> Hence, isobaric glycosyl cation fragments can be distinguished on the basis of their IR spectrum using DFT calculations.

Next, we investigated the characterization of the more challenging oxocarbenium ion using permethylated mannosyl donor 2 to limit the possibilities for neighboring group participation. ESI of 2 followed by CID resulted in the desired fragment ion with m/z = 219 (Fig. S<sub>5</sub>). Subsequent IR ion spectroscopy yielded an IR spectrum rich in diagnostic peaks for structural characterization (Fig. 2, black line). DFT calculations for oxocarbenium ions resulting from 2 yielded two main structural families which differ in energy by ~13 kJ/mol (2a and 2b, Fig. 2, color fill). Both structures are oxocarbenium ions, but differ in the conformation of the pyranose ring,  ${}^{3}E$  (2a) and  ${}^{4}H_{3}$  (2b). Most notably, the calculated C=O<sup>+</sup> stretch, in 2a and 2b, corresponding to the C-1 oxocarbenium ion (1609 cm<sup>-1</sup>) was indeed observed in the measured spectrum at 1585 cm<sup>-1</sup> (Fig. 2A,B). The lowest energy structure, the  ${}^{3}E$  envelope, displayed the best fit with the observed spectrum Previous calculations by Whitfield suggested a two conformer hypothesis with  ${}^{4}H_{3}$  and  ${}^{3}E$  conformations as the lowest energy states.<sup>33</sup> The calculated spectra of the  ${}^{3}E$  (2a)

and  ${}^{4}\text{H}_{3}$  (**2b**) conformers are very similar but the additional peak at 1058 cm<sup>-1</sup> (C-3-C-4 and C-2-*O*-2 stretch) predicted for the  ${}^{4}\text{H}_{3}$  conformer is absent in the measured spectrum (Fig. 2B). In the calculated  ${}^{3}E$  conformation, the same C-3-C-4 and C-2-*O*-2 interactions are calculated at slightly lower wavenumbers (991 cm<sup>-1</sup>) and are in better agreement with the measured spectrum. Together with the fact that the  ${}^{4}\text{H}_{3}$  has a higher energy, this suggests the  ${}^{3}E$  is the preferred conformer. These data indicate that elusive oxocarbenium ions can be generated and characterized at a highly detailed level using diagnostic peaks indicative of pyranose conformation.

The conformational flexibility of glycosyl oxocarbenium ions is an important factor determining the stereochemical outcome of a glycosylation reaction. Much effort has been directed to the creation of oxocarbenium ions that display a well-defined conformation to induce stereoselectivity via the preferential attack on one diastereotopic face.<sup>34</sup> A prime example are oxocarbenium ions derived from mannuronic acid esters 3 and 4 which are expected to favor the  ${}^{3}H_{4}$  conformation due to stabilization of the oxocarbenium ion by the pseudo axial C-6 carboxylate ester.<sup>34-37</sup> <sup>35,38</sup>. Subsequent pseudo axial attack is favorable as it directly leads to the βproduct in the chair conformation. In addition, it is also known that the  $\alpha$ -triflate intermediate contact ion pair is formed in reactions with 3, leading to the  $\beta$ -product via S<sub>N</sub>2like displacement.<sup>39</sup> Hence, to elucidate the solvent separated ion pair side of this continuum we characterized cations resulting from the fragmentation of 3 and 4.



Ionization and fragmentation resulted in the corresponding mannuronyl cations (Fig. S7). Well resolved IR spectra were obtained displaying distinct diagnostic peaks for permethylated mannuronic acid **3** (Fig. 3A, black line). DFT-calculations indicated that participation of the C-6 ester provided the lowest energy structure **3a** (Fig. 3A, color fill).



Figure 2: Comparison of the calculated spectra (filled) of the  ${}^{3}E$  (2a) or  ${}^{4}H_{3}$  (2b) oxocarbenium ion conformer with the measured IR ion spectrum of the m/z=219 CID fragment of 2 (black line in both panels). Energies are relative to the o.o kJ/mol structure as reported in Fig. S6.

Figure 3: Comparison of calculated spectra (filled) with the measured IR ion spectra (black line) of mannuronic acids 3a, 4a, 4b (A-C) and a mixture of isomers 4a and 4b (D).

The measured spectrum is in good agreement with the calculated spectrum of this structure. The peak observed at 1637 cm<sup>-1</sup> corresponds well with the calculated peak at 1644 cm<sup>-1</sup> describing the diagnostic O-C=O<sup>+</sup> stretch vibration. The CH bend modes of the methyl ester located at ~1409-1487 cm<sup>-1</sup> in the stabilized oxocarbenium ion also have clear overlap (**3a**, Fig. 3A). Furthermore, the various CH bending modes belonging to the methoxy and ring hydrogens were identified in the calculated spectrum and showed clear overlap with peaks in the measured spectrum. Isomers in which the C-6 ester does not participate were also considered, however as the characteristic calculated C=O stretch vibration at ~1800 cm<sup>-1</sup> is not observed in the experimental IR spectrum, we can discard these structures (Fig. S8). Hence, these data strongly suggest C-6 ester participation upon activation of **3**.

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14 Finally, mannuronic acid ester donor 4 containing a C-4 ester 15 was investigated. In this case, ester participation from both 16 diastereotopic faces of the oxocarbenium ion should be pos-17 sible. The IR spectra of the isolated ions of 4 were obtained 18 and yield a well resolved spectrum (Fig. 3B-D, black line). 19 Initial DFT calculations pointed towards structure 4a resulting from C-6 ester participation as observed with 3 (Fig. 3B). 20 All major calculated peaks such as CH bending, C-6  $O-C=O^+$ 21 stretch and 4-Ac C=O stretch could be assigned and showed 22 excellent agreement between the experimental- and theoret-23 ical IR spectrum (Fig. 3B). However, distinct peaks in the 24 measured spectrum observed at 1557 cm<sup>-1</sup> and 1267 cm<sup>-1</sup> were 25 not accounted for by the calculation. However, the comput-26 ed spectrum of isomer 4b with C-4 acetyl participation shows 27 clear activity at these vibrational frequencies (Fig. 3C). The 28 most characteristic peak calculated at 1554 cm<sup>-1</sup> (4-Ac, O-29  $C=O^+$  stretch) is highly diagnostic for C-4 acetyl participation whereas the peak at 1261 cm<sup>-1</sup> (C-6-OMe stretch) belongs to 30 the non-participating methyl ester. Mixing of the two calcu-31 lated spectra belonging to both participation modes (4a and 32 **4b**) led to an excellent fit of the observed spectrum (Fig. 3D). 33 This result highlights the possibility to characterize a mix-34 ture of isomers using IR ion spectroscopy thereby providing 35 crucial insight in the dynamics of glycosyl cations. 36

In conclusion, the use of mass spectrometry coupled with IR 37 ion spectroscopy and quantum chemical calculations is a 38 mild and versatile method to generate and structurally char-39 acterize elusive glycosyl cations in the gas phase. Whilst we 40 acknowledge that gas phase experiments are unlikely to be 41 fully representative of glycosyl cation structure in solution, 42 the absence of counter ions and solvent allows for the de-43 tailed study of the intrinsic structure and dynamics of these 44 intermediates. Furthermore, the IR spectra displayed a 45 wealth of diagnostic peaks that could be used to elucidate 46 conformational preferences as well as mixtures of isomeric 47 dioxolenium ions. We therefore believe that this MS-IR based methodology will be valuable to advance our funda-48 mental understanding S<sub>N1</sub> pathways in glycosylation reac-49 tions and likely also other reactions proceeding through ion-50 ic intermediates. 51

#### ASSOCIATED CONTENT

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#### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website. The supporting info contains experimental procedure and spectra for the synthesis of 1-4 as

well as additional ESI and CID spectra and DFT calculations in PDF format.

#### **AUTHOR INFORMATION**

#### **Corresponding Author**

\* Dr. A.M. Rijs, <u>a.rijs@science.ru.nl</u> and Dr. T.J. Boltje, <u>t.boltje@science.ru.nl</u>.

#### **Author Contributions**

‡HE and MS contributed equally.

#### Notes

The authors declare no competing financial interests.

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#### REFERENCES

- (1) Boltje, T. J.; Buskas, T.; Boons, G.-J. Nat. Chem. 2009, 1, 611.
- (2) Horenstein, N. A. In *Adv. Phys. Org. Chem.*; Richard, J. P., Ed.; Elsevier: New York, 2006; Vol. 41, p 275.
- (3) Crich, D. Acc. Chem. Res. 2010, 43, 1144.
- (4) Fang, T.; Gu, Y.; Huang, W.; Boons, G.-J. J. Am. Chem. Soc. 2016, 138, 3002.
- (5) Mensink, R. A.; Boltje, T. J. Chem. Eur. J. 2017, 23, 17637.
- (6) Paulsen, H.; Herold, C.-P. *Chem. Ber.* **1970**, *103*, 2450.
- (7) Frihed, T. G.; Bols, M.; Pedersen, C. M. Chem. Rev. 2015, 115, 4963.
- (8) Crich, D.; Dai, Z.; Gastaldi, S. J. Org. Chem. 1999, 64, 5224.
- Martin, A.; Arda, A.; Désiré, J.; Martin-Mingot, A.; Probst, N.; Sinaÿ, P.; Jiménez-Barbero, J.; Thibaudeau, S.; Blériot, Y. Nat. Chem. 2015, 8, 186.
- Khanal, N.; Masellis, C.; Kamrath, M. Z.; Clemmer, D. E.; Rizzo, T. R. Anal. Chem. 2017, 89, 7601.
- Molina, E. R.; Eizaguirre, A.; Haldys, V.; Urban, D.; Doisneau, G.; Bourdreux, Y.; Beau, J.-M.; Salpin, J.-Y.; Spezia, R. ChemPhysChem 2017, 18, 2812.
- (12) Rabus, J. M.; Abutokaikah, M. T.; Ross, R. T.; Bythell, B. J. *Phys. Chem. Chem. Phys.* **2017**, *19*, 25643.
- (13) Chen, J.-L.; Nguan, H. S.; Hsu, P.-J.; Tsai, S.-T.; Liew, C. Y.;
  Kuo, J.-L.; Hu, W.-P.; Ni, C.-K. *Phys. Chem. Chem. Phys.* 2017, 19, 15454.
- Bythell, B. J.; Abutokaikah, M. T.; Wagoner, A. R.; Guan,
  S.; Rabus, J. M. J. Am. Soc. Mass. Spectrom. 2017, 28, 688.
- (15) Martens, J.; Grzetic, J.; Berden, G.; Oomens, J. Nat. Commun. 2016, 7, 11754.
- (16) Patrick, A. L.; Stedwell, C. N.; Polfer, N. C. Anal. Chem. 2014, 86, 5547.
- (17) Seo, J.; Warnke, S.; Pagel, K.; Bowers, M. T.; von Helden, G. Nat. Chem. 2017, 9, 1263.
- Zhu, Y.; Roy, H. A.; Cunningham, N. A.; Strobehn, S. F.;
  Gao, J.; Munshi, M. U.; Berden, G.; Oomens, J.; Rodgers,
  M. T. Phys. Chem. Chem. Phys. 2017, 19, 17637.
- Zhu, Y.; Hamlow, L. A.; He, C. C.; Lee, J. K.; Gao, J.; Berden, G.; Oomens, J.; Rodgers, M. T. J. Phys. Chem. B, 2017, 121, 4048.
- (20) Zhu, Y.; Yang, Z.; Rodgers, M. T. J. Am. Soc. Mass. Spectrom. 2017, 28, 2602.
- (21) Gray, C. J.; Schindler, B.; Migas, L. G.; Pičmanová, M.; Allouche, A. R.; Green, A. P.; Mandal, S.; Motawia, M. S.;

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5	(23)	Martens, J.; Berden, G.; van Outersterp, R. E.; Kluijtmans,	(33)	Nuka
6		L. A. J.; Engelke, U. F.; van Karnebeek, C. D. M.; Wevers,		Whit
7		R. A.; Oomens, J. Sci. Rep. 2017, 7, 3363.	(34)	Walv
/	(24)	Mucha, E.; Florez, A. I. G.; Marianski, M.; Thomas, D. A.;		Lodd
8		Hoffmann, W.; Struwe, W. B.; Hahm, H. S.; Gewinner, S.;		Mare
9		Schollkopf, W.; Seeberger, P. H.; von Helden, G.; Pagel, K.	(35)	Codé
10		Angew. ChemInt. Edit. 2017, 56, 11248.		G.; O
11	(25)	Contreras, C. S.; Polfer, N. C.; Oomens, J.; Steill, J. D.;	(	Chem
12		Bendiak, B.; Eyler, J. R. Int. J. Mass Spectrom. 2012, 330,	(36)	Zhu,
13	(26)	205. Coginara E. L. Carcabal, R. Vadan, T. D. Simong, I. D.	(27)	50C. 2
14	(20)	Davis B C. Natura 2011 (60 76	(37)	Abe,
14	(27)	Kancharla P. K · Navuluri C · Crich D. Angew Chem Int	(28)	Avala
15	(27)	Ed. 2012. 51, 11105.	(50)	Woer
16	(28)	Denekamp, C.; Sandlers, Y. J. Mass Spectrom. 2005, 40,	(39)	Walv
17	1055.			S.; Co
18	(29)	Denekamp, C.; Sandlers, Y. J. Mass Spectrom. 2005, 40,		2009
19	765.			
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- ens, J.; Berden, G.; Gebhardt, C. R.; Oomens, J. Rev. nstrum. 2016, 87, 103108.
- A. M.; Oomens, J. In Gas-Phase IR Spectroscopy and ture of Biological Molecules; Rijs, A. M., Oomens, J., Springer International Publishing: Cham, 2015, p 1.
- n, A. S. Can. J. Chem. 1963, 41, 399.
- nda, T.; Bérces, A.; Wang, L.; Zgierski, M. Z.; field, D. M. Carbohydr. Res. 2005, 340, 841.
- oort, M. T. C.; Dinkelaar, J.; van den Bos, L. J.; er, G.; Overkleeft, H. S.; Codée, J. D. C.; van der el, G. A. Carbohydr. Res. **2010**, 345, 1252.
- e, J. D. C.; Walvoort, M. T. C.; de Jong, A.-R.; Lodder, Dverkleeft, H. S.; van der Marel, G. A. J. Carbohydr. 1. **2011**, 30, 438.
- X.; Kawatkar, S.; Rao, Y.; Boons, G.-J. J. Am. Chem. **2006**, *1*28, 11948.
- H.; Shuto, S.; Matsuda, A. J. Am. Chem. Soc. 2001, 123,
- a, L.; Lucero, C. G.; Romero, J. A. C.; Tabacco, S. A.; rpel, K. A. J. Am. Chem. Soc. 2003, 125, 15521.
- voort, M. T. C.; Lodder, G.; Mazurek, J.; Overkleeft, H. odée, J. D. C.; van der Marel, G. A. J. Am. Chem. Soc. 131, 12080.

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