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Total synthesis of myriocin and mycestericin D employing Rh(II)-catalyzed C–H amination followed by stereoselective alkylation

Narumi Noda^a, Hisanori Nambu^a, Kana Ubukata^a, Tomoya Fujiwara^a, Kiyoshi Tsuge^b, Takayuki Yakura^{a, *}

^a Graduate School of Medicine and Pharmaceutical Sciences, University of Toyama, Sugitani, Toyama 930-0194, Japan
 ^b Department of Chemistry, Graduate School of Science and Engineering, University of Toyama, Gofuku, Toyama 930-8555, Japan

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ABSTRACT

Total synthesis of myriocin and mycestericin D was achieved using the Du Bois Rh(II)catalyzed C–H amination of a sulfamate and subsequent alkylation as a key step. The reaction of a sulfamate with PhI(OAc)₂ and MgO in the presence of Rh₂(OAc)₄ gave oxathiazinane *N*,*O*acetal as the sole product in high yield. Alkylation of *N*,*O*-acetal using vinylmagnesium bromide in the presence of ZnCl₂ proceeded stereoselectively to provide an oxathiazinane bearing a quaternary chiral center in high yield. Myriocin and mycestericin D were synthesized from a common synthetic intermediate. This route includes the first application of the Du Bois procedure for constructing a quaternary chiral center.

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1. Introduction*

Myriocin and mycestericins are complex α . α -disubstituted amino acid natural products structurally related to sphingolipids. Myriocin (1, Figure 1) was isolated from the fermentation broth of the thermophilic fungi Myriococcum albomyces¹ and Mycelia sterilia² in 1972, and from the culture broth of Isalia sinclairii in 1994.3 Notably, myriocin exhibits 10-100 times more potent immunosuppressive activity than cyclosporine A^3 and has been shown to have potent inhibitory activity against serine palmitoyltransferase, which is an essential enzyme in the biosynthesis of sphingolipids.⁴ Mycestericins A-G were isolated from the culture broth of M. sterilia ATCC 20349 and exhibited immunosuppressive activity similar to that of myriocin.⁵ Owing to their challenging structural motif containing a quaternary chiral center and interesting biological activity, myriocin⁶ and the mycestericins⁷ are attractive targets for total synthesis. The first total synthesis of myriocin (1) was achieved from D-fructose using a Strecker reaction to construct a quaternary chiral center, but the desired diastereoisomer was the minor component.^{6a,b} Thereafter, several synthetic chemists reported total and formal syntheses of 1 by employing various methods for stereoselective introduction of the quaternary chiral center, including epoxide opening with external^{6c-g} or internal^{6h} nitrogen nucleophiles, aldol reactions of chiral bislactim ether⁶ⁱ or oxazoline templates, Overman rearrangement of an allylic trichloroacetimidate,

photolytic hydroxymethylation of an α-alkoxycarboxy ketoxime ether,^{6m} enzymatic kinetic resolution of racemic methyl ester,⁶ⁿ and deconjugative alkylation of a dehydroamino acid.⁶⁰ On the contrary, there is only one example of the total synthesis of mycestericin D (2, Figure 1), although 2 has the same stereochemistry at C2 and C3, and an identical long side chain. Node and co-workers reported the asymmetric total synthesis of 2 by employing a stereoselective hydroxymethylation of an oxazoline derivative for the construction of a quaternary chiral center.^{7d,e} Thus, there are no examples of the synthesis of **1** and **2** using the same synthetic strategy. In our laboratory, we have been exploring the synthesis of sphingosine natural products using Rh(II)-catalyzed reactions as key steps.⁸ As part of this program, we herein report a novel strategy for the concise synthesis of myriocin $(1)^9$ and mycestericin D (2) by applying Du Bois' Rh(II)-catalyzed C-H amination-alkylation procedure¹⁰ for construction of a quaternary chiral center.



Fig. 1. Structure of myriocin (1) and mycestericin D (2).

^{*} Corresponding author. Tel.: +81-76-434-7555; fax: +81-76-434-5053; e-mail: yakura@pha.u-toyama.ac.jp (T. Yakura).



2. Results and discussion

Our retrosynthetic strategy for myriocin (1) and mycestericin D (2) is outlined in Scheme 1. Introduction of the long side chain at a later stage in the synthesis was proposed because it would provide high flexibility for analogue synthesis. The conversion would involve cross metathesis of amino alkenes 3 and 4, which contain all asymmetric carbons of 1 and 2, respectively, with their desired configurations, and the known alkene 5.¹¹ It was anticipated that 3 and 4 would be accessible by allylation from aldehyde or alcohol 6 as a key intermediate. Alcohol 6 can be obtained from oxathiazinane 7 by a ring-opening reaction. The key steps of the stereoselective construction of the quaternary chiral center of 7 would be accomplished by applying Du Bois' Rh(II)-catalyzed C-H amination reaction¹² of sulfamate 9, followed by stereoselective alkylation.¹⁰ Although Du Bois' pioneering work has been applied to the synthesis of propargylic amine derivatives^{10a} and (+)-saxitoxin,^{10b,c} there are no reported examples of its use for the construction of quaternary chiral centers. Sulfamate ester 9 would be prepared from commercially available diethyl L-tartrate (10).



Scheme 1. Retrosynthetic analysis of myriocin (1) and mycestericin D (2).

The starting diethyl L-tartrate (10) was converted into the known mono-*tert*-butyldiphenylsilyl (TBDPS) ether 11 according to the literature¹³ (Scheme 2). Reaction of alcohol 11 with chlorosulfonyl isocyanate, formic acid, and pyridine in dichloromethane provided sulfamate ester 12 in 87% yield.^{12b,14} Rhodium(II)-catalyzed C–H amination of 12 using Du Bois' conditions¹² proceeded stereospecifically to yield oxathiazinane *N*,*O*-acetal 13. Thus, treatment of 12 with 4 mol% of dirhodium(II) tetraacetate, 1.1 equivalents of phenyliodine(III) diacetate, and 2.3 equivalents of magnesium oxide in dichloromethane at room temperature for 1 h gave the corresponding C–H amination product 13 in 84% yield.



Scheme 2. Rh(II)-catalyzed C-H amination of sulfamate ester 12.

To construct the quaternary chiral centers in 1 and 2, we investigated alkylation of oxathiazinane N,O-acetal 13. Since ethynylation gave better results than vinylation in Du Bois' original report on the construction of tertiary centers,¹⁰ we first examined ethynylation of 13 using the Du Bois' conditions^{10a} (Table 1). When N,O-acetal 13 was reacted with trimethylsilylethynylzinc chloride generated from 2.1 equivalents of trimethylsilylacetylene, butyllithium (2.0 equiv), and zinc chloride (2.1 equiv) in the presence of boron trifluoride diethyl etherate (BF₃·OEt₂, 3.0 equiv) in tetrahydrofuran (THF) at 50 °C for 2 h, ethynylation product 14 was obtained in only 42% yield as the sole product (entry 1). The use of twice the amounts of all reagents increased the yield of 14 to 62% (entry 2). We then screened the amount of the Lewis acid BF₃·OEt₂. Reaction of 13 with 4.0 equivalents of in situ generated zinc acetylide and 4.2 equivalents of BF₃·OEt₂ afforded 14 in 71% yield (entry 3). When the amount of BF₃·OEt₂ was decreased to 3.0 and 2.1 equivalents, these product yields slightly decreased to 69% and 65%, respectively (entries 4 and 5). Reaction of 13 with a large amount of zinc acetylide (10.0 equiv) in the presence of BF₃·OEt₂ (4.2 equiv) furnished 14 in 81% yield (entry 6). These results demonstrate that Du Bois' C-H amination-alkylation procedure was effective for the stereoselective construction of the quaternary chiral center.

Table 1

Ethynylation of sulfamate ester 13 with zinc acetylide and BF3·OEt2



Entry	Zinc Acetylide ^a (equiv)	BF3·OEt2 (equiv)	Time (h)	Yield (%)
1	2.0	3.0	2	42
2	4.0	6.0	3.5	62
3	4.0	4.2	1	71
4	4.0	3.0	5	69
5	4.0	2.1	5	65
6	10.0	4.2	2	81

^a Zinc acetylide was prepared *in situ* with trimethylsilylacetylene $(1.05 \times X \text{ equiv})$, butyl lithium (X equiv) and zinc chloride $(1.05 \times X \text{ equiv})$.

Next, we examined the vinylation of **13**, which was accomplished according to Du Bois' original conditions^{10c} (Scheme 3). Thus, reaction of **13** with 4.2 equivalents of vinylmagnesium bromide in the presence of zinc chloride (2.2 equiv) in THF at room temperature for 6 h produced **15** as the sole product in 86% yield. The stereochemistry of **15** was determined to be (4R,5R) via single-crystal X-ray analysis of the

p-bromobenzoyl derivative **16**.¹⁵ The stereoselectivity of the C–H amination–alkylation reaction was observed in a manner similar to those reported by Du Bois.¹⁰ Thus, the alkylation would proceed through intermediate A,^{10c} which may be intramolecularly coupled to nucleophiles. Considering the yields of **14** and **15** and the necessary further conversions into a common synthetic intermediate for **1** and **2**, vinyl compound **15** was selected.



Scheme 3. Vinylation of sulfamate ester 13 and determination of the stereochemistry of 15.

With the stereoselective construction of the quaternary chiral center in 1 and 2 accomplished, we next examined opening of the oxathiazinane ring of 15 after protection of OH and NH groups (Table 2). Acetylation of the hydroxy group in 15 followed by tert-butoxycarbonylation of the amino group gave protected oxathiazinane 17 in high yield. We initially investigated the opening of 17 using Du Bois' conditions.^{10c,12b} When 17 was warmed in acetonitrile-water (11:2) at 75 °C, decomposition was observed (Table 2, entry 1). Use of 0.1 M phosphate buffer instead of water at 50 °C led to ring opening and an interesting 1,2-acetyl group shift to give secondary alcohol 18 as the major product in 31% yield, and the initially expected primary alcohol 18' in 7% yield (entry 2). After screening of organic solvents such as dichloromethane, acetone, and THF, we found that THF was the most effective solvent (entries 3–5). Warming of 17 in a THF-buffer solution (11:2) at 50 °C for 24 h was found to afford 18 and 18' in 73% and 23% yields, respectively. Increasing the ratio of buffer (THF/buffer = 5:1 and 3:1) resulted in a decrease in the combined product yield of 18 and 18' (entries 6 and 7). Longer reaction times led to low yields of the desired products due to decomposition; however, treatment of purified 18' under the same reaction conditions gave 18 in 80% yield.

Ring opening of oxathiazinane 17



^a The reaction was conducted at 75 °C.

^b Buffer: 0.1 M phosphate buffer.

We focused on the synthesis of alcohol **21** and aldehyde **22** as key intermediates for **1** and **2** (Scheme 4). Protection of the secondary OH group in **18** with chloroacetyl chloride and pyridine yielded chloroacetate **19**. Reductive ozonolysis of **19** resulted in conversion of the vinyl group to a hydroxymethyl substituent and chloroacetyl deprotection to form diol **20** in 82% yield. After protection of the diol as an acetonide, methanolysis of the acetoxy group provided alcohol **21** in 92% yield. Swern oxidation of **21** afforded aldehyde **22** in 96% yield.



Scheme 4. Synthesis of key intermediates 21 and 22.

With the key intermediate 22 in hand, we investigated the formation of the third stereocenter using chelation-controlled allylation for synthesis of myriocin (1, Table 3). Reaction of 22 with allylmagnesium bromide in THF afforded a 1:3.5 mixture of desired 23 and undesired 23' in 36% combined yield (entry 1). Switching the solvent to diethyl ether greatly improved the product yield and diastereoselectivity, providing a 1.6:1 mixture of 23 and 23' in 69% combined yield (entry 2). We next examined the use of magnesium iodide as a chelating agent. As expected, the reaction of 22 with magnesium iodide increased diastereoselectivity from 1.6:1 to 4.2:1 (entry 3). The allylation

would proceed through chelated complex **B**. To further enhance the product yield and diastereoselectivity, we directed our efforts to allylation using stannane^{7a,b} and indium reagents.¹⁶ The reaction of **22** with allyltributylstannane and magnesium bromide in dichloromethane afforded **23** as the major product (**23**:**23**' = 2.8:1) in 42% yield (entry 4). Gratifyingly, the treatment of **22** with allylbromide and indium metal in the presence of tetrabutylammonium iodide in *N*,*N*-dimethylformamide gave **23** as the major product (**23**:**23**' = 5.1:1) in 80% yield (entry 5). Furthermore, the addition of magnesium iodide under the conditions of entry 4 improved the product yield and diastereoselectivity (80% yield, **23**:**23**' = 6.6:1, entry 6). The two diastereomers were then readily separated by column chromatography.¹⁷

Table 3

Allylation of aldehyde **22** with allylmagnesium bromide, allyltributylstannane, and allylindium reagents



The stage was now set for completion of the synthesis of myriocin (1, Scheme 5). After protection of the secondary hydroxy substituent in 23 as an acetoxy group, introduction of the long alkyl chain using Grubbs' cross metathesis chemistry was examined. According to the procedure reported by Shibasaki, Kumagai, and co-workers,^{7h} alkenes 24 was treated with 4 equivalents of the known alkene 5^{11} in the presence of a catalytic amount of Grubbs' 1st generation catalyst in dichloromethane at reflux for 6 h. Unfortunately, the reaction did not occur at all and the starting 24 was recovered in 96% yield. To our delight, using more reactive Grubbs' 2nd generation catalyst in refluxing dichloromethane gave the good result and coupling product 25, which has all carbons of the myriocin skeleton, was obtained in 96% yield after 5.5 h. Deprotection of TBDPS protecting group in 25 and subsequent oxidation of the resulting hydroxyl group with 2,2,6,6-tetramethylpiperidine 1oxyl (TEMPO), PhI(OAc)₂ and MgO afforded aldehyde 26. Oxidation of 26 to the corresponding carboxylic acid and global deprotection by sequential alkaline and acidic hydrolyses produced crude myriocin (1). Since it was difficult to directly obtain 1 in high purity, the crude product was acetylated with acetic anhydride in pyridine to afford the known γ -lactone 27: $[\alpha]_{D}^{18}$ +55.6 (c 1.0, CHCl₃) {lit.^{6h} $[\alpha]_{D}^{23}$ +52.5 (c 0.85, CHCl₃)}. Finally, saponification of 27 followed by neutralization with Amberlite[®] IRC-86 furnished pure **1**. The synthetic material **1** exhibited spectroscopic data (¹H and ¹³C NMR, and IR) consistent with those reported for natural product **1** and had its melting point (165–168 °C) and an optical rotation { $[\alpha]_D^{23}$ +5.6 (*c* 0.30, DMSO)} in good agreement with literature values {mp 164–168 °C, ${}^3[\alpha]_D^{20}$ +6.1 (*c* 0.26, DMSO)^{6h}}.



Scheme 5. Synthesis of myriocin (1).

Next, the synthesis of mycestericin D (2) from the key intermediate alcohol 21 was investigated (Scheme 6). Initially, we examined a direct coupling of 21 with the allyl component. After conversion of 21 to halides 28a-c by treatment with thionyl chloride (R = Cl: 28a), *N*-bromosuccinimide (NBS, R = Br: 28b), and *N*-iodosuccinimide (NIS, R = I: 28c), their reaction with allyltributylstannane in the presence of azobisisobutyronitrile (AIBN) in THF gave decomposition products. Furthermore, we examined the copper-catalyzed cross-coupling reaction of alkyl iodide with allylmagnesium bromide.¹⁸ However, the allylation did not proceed at all.



Scheme 6. Attempts to introduce an allyl group to alcohol 21.

As cross-coupling reactions between alkyl halides and allylmetal reagents were difficult, we examined introduction of the C3 unit using a Wittig reaction with a key intermediate aldehyde **22** (Scheme 7). Wittig reaction of **22** using a stable ylide reagent followed by hydrogenation provided ester **29** in 92% yield. After reduction of the methyl ester to an aldehyde with diisobutylaluminum hydride (DIBAL-H), synthesis of

alkene 30 was achieved by conversion of the resulting aldehyde with an unstable vlide Wittig reagent. As we obtained the amino alkene component for mycestericin D (2), we finally investigated the completion of the total synthesis of 2. The cross metathesis for the long side chain extension was conducted. The reaction between 30 and 4 equivalents of the alkene 5^{11} in the presence of Grubbs' 2nd generation catalyst in dichloromethane at reflux proceeded smoothly to afford coupling product 31 in 96% yield. Desilvlation of the primary silvl ether in 31, Swern oxidation of the resulting hydroxyl group, oxidation of the resulting aldehyde to the corresponding carboxylic acid, and global deprotection by acidic hydrolysis furnished pure mycestericin D (2). The synthetic compound 2 was spectroscopically (¹H and ¹³C NMR, and IR) identical to natural **2**, and its melting point (164–166 °C) and optical rotation $\{[\alpha]_D^{23} - 7.48 \ (c \ 0.39, \text{ MeOH})\}$ were in good agreement with literature values for the natural product {mp 162- $167 \,^{\circ}\text{C},^{5a} [\alpha]_{\text{D}} - 7.5 (c \, 0.16, \text{MeOH})^{5a}$



Scheme 7. Synthesis of mycestericin D (2).

3. Conclusion

In summary, the total synthesis of myriocin and mycestericin D was achieved using Rh(II)-catalyzed C–H amination of a sulfamate followed by stereoselective alkylation as a key step. This synthesis is the first example of the construction of a quaternary chiral center using sequential Rh(II)-catalyzed C–H amination/alkylation reactions. As the quaternary chiral center can be easily and stereoselectively constructed and the long side chain can be introduced at a later stage in the synthesis, the present method would provide high flexibility for analogue synthesis. Thus, the synthesis of myriocin analogues, including sphingofungins, is currently in progress.

4. Experimental section

4.1. General

Melting points were determined using a Yanaco micro melting point apparatus and are uncorrected. Optical rotations were determined using a JASCO P-2100 polarimeter. Infrared (IR) spectra were recorded on a JASCO FT/IR-460 Plus spectrophotometer and absorbance bands are reported in wavenumber (cm⁻¹). ¹H NMR spectra were recorded on JEOL

JNM-ECX400P (400 MHz) spectrometer. Chemical shifts are reported relative to internal standard (tetramethylsilane at $\delta_H 0.00$ or CDCl₃ at $\delta_{\rm H}$ 7.26). Data are presented as follows: chemical shift (δ , ppm), multiplicity (s = singlet, d = doublet, t = triplet, m = multiplet, br = broad), coupling constant and integration. NMR spectra were recorded on JEOL JNM-ECX400P (100 MHz) spectrometer. The following internal reference was used (CDCl₃ at δ 77.0). All ¹³C NMR spectra were determined with complete proton decoupling. High-resolution mass spectra (HRMS) were determined with JEOL JMS-GCmate II and JEOL JMS-AX505HAD instruments. Column chromatography was performed on Silica Gel 60 PF254 (Nacalai Tesque) and Kanto silica gel 60 N (63-210 mesh) under pressure. Analytical thin layer chromatography (TLC) was carried out on Merck Kieselgel 60 F₂₅₄ plates. Visualization was accomplished with UV light and phosphomolybdic acid stain solution followed by heating.

All reagents such as diethyl L-tartrate (10), chlorosulfonyl isocyanate and trimethylsilylacetylene are commercially available and were purchased from suppliers such as Sigma-Aldrich Co.; Wako Pure Chemical Industries, Ltd.; Tokyo Chemical Industry Co., Ltd.; Nacalai Tesque, INC. Dehydrated CH₂Cl₂, THF, MeOH, benzene, DMF, DMSO and pyridine were purchased from Wako Pure Chemical Industries, Ltd. (4S,5S)-[5-(*tert*-butyldiphenylsilyloxy)methyl-2,2-dimethyl-1,3-dioxolan-4-yl]methanol (11)¹³ and petadec-1-en-9-one (5)¹¹ were prepared according to literature procedures.

4.2. Rh(II)-catalyzed C-H amination followed by alkylation

4.2.1 (4S,5S)-5-(tert-Butyldiphenylsilyloxy)methyl-2,2-dimethyl-4-sulfamoyloxymethyl-1,3-dioxolane (12). Formic acid (1.3 mL, 34.5 mmol) was added dropwise to neat chlorosulfonyl isocyanate (3.0 mL, 34.5 mmol) at 0 °C with rapid stirring. The resulting viscous suspension was stirred at 0 °C for 5 min. CH₂Cl₂ (20 mL) was added to the mixture and the solution was stirred at 0 °C for 20 minutes and then at room temperature overnight. The reaction mixture was cooled to 0 °C, and a solution of the known alcohol 11^{13} (7.0 g, 17.5 mmol) and pyridine (3.5 mL, 43.8 mmol) in CH₂Cl₂ (6 mL) was added via cannula. The reaction mixture was warmed to room temperature and stirred for 1 h. The reaction was quenched with EtOAc and H_2O . The organic phase was collected and the aqueous layer was extracted with EtOAc (2 x 30 mL). The combined organic extracts were washed with brine (20 mL), dried over anhydrous MgSO₄, and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide **12** (7.29 g, 87%) as a colorless oil: $[\alpha]_D^{22}$ – 9.3 (c 1.00, CHCl₃); IR (film, cm⁻¹) v 3366, 3279, 1559, 1428, 1374, 1219, 1187; ¹H NMR (400 MHz, CDCl₃) δ 7.68–7.64 (m. 4H), 7.47-7.38 (m, 6H), 4.92 (br s, 2H), 4.39 (m, 1H), 4.32-4.26 (m, 2H), 3.93 (ddd, J = 7.0, 5.4, 4.4 Hz, 1H), 3.85 (dd, J = 10.6, 4.4 Hz, 1H), 3.76 (dd, J = 10.6, 6.0 Hz, 1H), 1.43 (s, 3H), 1.40 (s, 3H), 1.07 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 135.50, 135.49, 132.72, 132.70, 129.88, 129.85, 127.79, 127.77, 110.2, 77.2, 76.6, 70.5, 63.7, 26.9, 26.8, 26.7, 19.1; HRMS (FAB) m/z calcd for C₂₃H₃₄NO₆SSi (M+H)⁺ 480.1876, found 480.1889.

4.2.2. (4S,5S)-4-(*tert-Butyldiphenylsilyloxy*)*methyl*-(2,2-*dimethyl-*1,3-*dioxolo*)[4,5-*d*]*tetrahydro*-1,2,3-*oxathiazine*-2,2-*dioxide* (13). Rh₂(OAc)₄ (270 mg, 0.611 mmol), PhI(OAc)₂ (5.39 g, 16.7 mmol) and MgO (1.41 g, 35.0 mmol) were added to a solution of sulfamate 12 (7.29 g, 15.2 mmol) in CH₂Cl₂ (96 mL). After stirring at room temperature for 1 h, the reaction mixture was filtered through a pad of Celite. The filter cake was rinsed with CH₂Cl₂, and the combined filtrates were concentrated in vacuo. The residue was purified by column chromatography (silica gel,

10% EtOAc in hexane) to provide **13** (6.11 g, 84%) as a white solid: mp 86–88 °C; $[\alpha]_D^{21}$ +12.9 (*c* 1.00, CHCl₃); IR (KBr, cm⁻¹) v 3281, 1590, 1429, 1422, 1199, 1186; ¹H NMR (400 MHz, CDCl₃) δ 7.65 (d, *J* = 6.4 Hz, 4H), 7.47–7.39 (m, 6H), 4.62 (dd, *J* = 13.2, 1.2 Hz, 1H), 4.38 (s, 1H), 4.37 (dd, *J* = 13.2, 2.0 Hz, 1H), 4.29 (br s, 1H), 4.06 (d, *J* = 12.0 Hz, 1H), 3.90 (d, *J* = 12.0 Hz, 1H), 1.59 (s, 3H), 1.45 (s, 3H), 1.08 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 135.6, 132.21, 132.18, 130.2, 130.1, 128.0, 127.9, 110.8, 94.5, 69.0, 68.8, 63.5, 27.8, 26.9, 26.1, 19.2; HRMS (EI) *m*/*z* calcd for C₂₃H₃₁NO₆SSi (M)⁺ 477.1641, found 477.1627.

4.2.3. (4R,5R)-4-(tert-Butyldiphenylsilyloxy)methyl-5-hydroxy-4-[2-(trimethylsilyl)ethynyl]tetrahydro-1,2,3-oxathiazin-2,2-

dioxide (14). n-butyllithium solution (1.05 mL, 1.6 M in hexane, 1.68 mmol) was added dropwise to a solution of trimethylsilylacetylene (0.25 mL, 1.76 mmol) in THF (0.55 mL) at -78 °C under nitrogen. After stirring for 20 minute, ZnCl₂ in THF (1.76 mL, 1.76 mmol) was added dropwise to the reaction mixture, and the resulting mixture was warmed to room temperature and stirred for 30 minutes. A solution of oxathiazinane N,O-acetal 13 (200 mg, 0.42 mmol) in THF (1.0 mL) was added to the reaction mixture via cannula at 0 °C. After addition of BF3·OEt2 (0.22 mL, 1.76 mmol), the reaction mixture was warmed to 50 °C and stirred for 2 h. The reaction was quenched with saturated NaHCO3 and the whole mixture was extracted with EtOAc (3 x 10 mL). The combined organic layers were washed with water (10 mL) and brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 14 (155 mg, 71%) as a white solid: mp 100–101 °C; $[\alpha]_D^{18}$ –28.9 (*c* 1.00, CHCl₃); IR (KBr, cm⁻¹) v 3493, 3202, 2175; ¹H NMR (400 MHz, CDCl₃) δ 7.66– 7.62 (m, 4H), 7.50–7.42 (m, 6H), 5.11 (s, 1H), 4.58 (t, J = 11.2 Hz, 1H), 4.37 (dd, J = 11.2, 4.4 Hz, 1H), 4.16 (ddd, J = 11.2, 10.6, 4.4 Hz, 1H), 3.95 (d, J = 10.2 Hz, 1H), 3.71 (d, J = 10.2 Hz, 1H), 1.63 (d, J = 10.6 Hz, 1H), 1.09 (s, 9H), 0.18 (s, 9H); ${}^{13}C$ NMR (100 MHz, CDCl₃) δ 135.9, 135.8, 132.04, 132.02, 130.81, 130.80, 128.6, 128.5, 97.0, 77.2, 70.0, 65.0, 62.9, 62.2, 27.3, 19.7, 0.0; HRMS (FAB) m/z calcd for $C_{25}H_{36}NO_5SSi_2$ (M+H)⁺ 518.1853, found 518.1881.

4.2.4. (4R,5R)-4-(tert-Butyldiphenylsilyloxy)methyl-5-hydroxy-4vinyltetrahydro-1,2,3-oxathiazin-2,2-dioxide (15). ZnCl₂ (0.92 mL, 1.0 M in THF, 0.92 mmol) was added dropwise to a solution of vinylmagnesium bromide (1.76 mL, 1.0 M in THF, 1.76 mmol) at -78 °C under nitrogen, and the resulting mixture was warmed to room temperature and stirred for 0.5 h. The reaction mixture was then cooled to 0 °C, and a solution of 13 (200 mg, 0.42 mmol) in THF (1.0 mL) was added dropwise via cannula. After completion of the addition, the reaction mixture was allowed to warm to room temperature and stirred for 6 h, after which time it was quenched with saturated NH₄Cl and the whole mixture was extracted with EtOAc (3 x 10 mL). The combined organic layers were washed with water (10 mL) and brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 15 (163 mg, 86%) as a white solid: mp 105–106 °C; $[\alpha]_D^{18}$ –17.2 (*c* 1.00, CHCl₃); IR (KBr, cm⁻¹) v 3570, 3516, 3251, 1472, 1449, 1429, 1390, 1348, 1190, 1170, 1079, 986, 912; ¹H NMR (400 MHz, CDCl₃) δ 7.68–7.60 (m, 4H), 7.49–7.41 (m, 6H), 6.07 (dd, J = 17.8, 11.6 Hz, 1H), 5.50 (d, J = 17.8 Hz, 1H), 5.44 (d, J =11.6 Hz, 1H), 5.07 (s, 1H), 4.40 (dd, J = 11.2, 9.6 Hz, 1H), 4.24 (dd, J = 11.2, 4.0 Hz, 1H), 4.15 (m, 1H), 3.80 (d, J = 10.8 Hz,1H), 3.56 (d, J = 10.8 Hz, 1H), 1.11 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 135.6, 135.6, 132.1, 131.9, 131.1, 130.5, 130.4, 128.2, 128.1, 120.6, 70.1, 65.5, 65.4, 62.7, 27.0, 19.2; HRMS (EI) *m*/*z* calcd for C₂₂H₂₉NO₅SSi (M)⁺ 447.1536, found 447.1525.

4.2.5. (4R,5R)-4-(tert-Butyldiphenylsilyloxy)methyl-5-(4-bromobenzoyloxy)-4-vinyltetrahydro-1,2,3-oxathiazin-2,2-

dioxide (16). p-Bromobenzoyl chloride (29 mg, 0.134 mmol), N,N-dimethyl-4-aminopyridine (DMAP, 1.6 mg, 0.0134 mmol) and pyridine (0.4 mL) were added to a solution of the alcohol 15 (30 mg, 0.067 mmol) in CH₂Cl₂ (0.5 mL) at 0 °C. After stirring for overnight at room temperature, the reaction was quenched with saturated NH₄Cl and the whole mixture was extracted with EtOAc (3 x 10 mL). The combined organic layers were washed with water (10 mL) and brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 16 (27 mg, 64%) as a white solid: mp 175-176 °C (EtOAc/hexane); $[\alpha]_{D}^{-26}$ -2.88 (c 0.5, CHCl₃); IR (KBr, cm⁻¹) v 3238, 1725, 1591, 1426, 1353, 1263, 1201, 1012, 986; ¹H NMR (400 MHz, CDCl₃) δ 7.79 (d, J = 8.8 Hz, 2H), 7.61 (d, J = 8.4 Hz, 4H), 7.52 (d, J = 8.4 Hz, 2H), 7.46-7.38 (m, 3H), 7.29 (m, 1H), 7.72 (t, J = 7.2 Hz, 2H), 6.12 (dd, J = 17.6, 11.6 Hz, 1H), 5.64 (dd, J = 6.0, 3.8 Hz, 1H), 5.53 (d, J = 17.6 Hz, 1H), 5.45 (d, J = 11.6 Hz, 1H), 5.10 (s, 1H), 4.50 (dd, J = 6.0, 3.8 Hz, 1H), 3.79 (d, J = 10.6 Hz, 1H), 3.69 (d, J = 10.6 Hz, 1H), 1.10 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 164.4, 135.51, 135.50, 132.6, 132.1, 131.8, 131.5, 131.2, 130.3, 130.1, 128.1, 127.8, 119.4, 68.0, 65.7, 65.1, 64.6, 26.9, 19.2; HRMS (FAB) m/z calcd for C₂₉H₃₂BrNO₆SSi (M+H)⁺ 630.0981, found 630.0934.

4.3. Synthesis of a common intermediate aldehyde 22

4.3.1. (4R,5R)-5-Acetoxy-4-(tert-butyldiphenylsilyloxy)methyl-3-(1,1-dimethylethoxycarbonyl)-4-vinyltetrahydro-1,2,3-

oxathiazin-2,2-dioxide (17). Ac₂O (7.6 mL) was added to a solution of alcohol 15 (1.12 g, 2.50 mmol) in pyridine (7.6 mL) at room temperature. After stirring for 1 h, the solvent was removed in vacuo. The resulting residue was solved in EtOAc (50 mL) and washed with water (20 mL) and brine (20 mL), and dried over anhydrous MgSO₄. Filtration and evaporation in vacuo furnished the crude product (1.20 g), which was used in the next step without further purification.

Boc₂O (1.1 mL, 4.88 mmol), Et₃N (1.2 mL, 8.54 mmol) and DMAP (60 mg, 0.49 mmol) were added to a solution of the crude product in THF (23 mL) at room temperature. After stirring for 1 h, the reaction was quenched with saturated NH₄Cl and the whole mixture was extracted with EtOAc (3 x 20 mL). The combined organic layers were washed with water (20 mL) and brine (20 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 17 (1.26 g, 84% for 2 steps) as a white solid: mp 123–124 °C; $[\alpha]_{D}^{18}$ +41.4 (c 1.00, CHCl₃); IR (KBr, cm⁻¹) v 1762, 1739, 1389, 1370, 1227, 1181, 978, 935; ¹H NMR (400 MHz, CDCl₃) δ 7.63–7.58 (m, 4H), 7.44–7.34 (m, 6H), 5.75 (dd, J = 17.2, 10.6 Hz, 1H), 5.71 (dd, J = 8.0, 6.8 Hz, 1H), 5.59 (d, J = 17.2 Hz, 1H), 5.43 (d, J = 10.6 Hz 1H), 4.95 (dd, J = 10.8, 6.8 Hz, 1H), 4.54 (d, J =10.8 Hz, 1H), 3.98 (dd, J = 10.8, 8.0 Hz, 1H), 3.72 (d, J = 10.8Hz, 1H), 1.64 (s, 3H), 1.53 (s, 9H), 1.11 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 168.9, 149.5, 136.0, 135.7, 132.9, 132.4, 130.3, 130.0, 129.8, 127.8, 127.6, 119.3, 85.2, 71.9, 69.9, 65.5, 60.5, 27.8, 26.8, 20.1, 19.2; HRMS (EI) m/z calcd for C₂₉H₃₉NO₈SSi $(M)^+$ 589.2166, found 589.2143.

4.3.2. (2R,3R)-1-Acetoxy-3-(tert-butyldiphenylsilyloxy)methyl-3-(1,1-dimethylethoxycarbonyl)amino-4-penten-2-ol (18) and (2R,3R)-2-Acetoxy-3-(tert-butyldiphenylsilyloxy)methyl-3-(1,1dimethylethoxycarbonyl)amino-4-penten-1-ol (18). 0.1 phosphate buffer (8.0 mL) was added to a solution of 17 (1.50 g. 2.5 mmol) in THF (39 mL). After stirring at 50 °C for 24 h, the reaction was quenched with 5% NaHCO₃ and the whole mixture was extracted with EtOAc (3 x 20 mL). The combined organic layers were washed with water (20 mL) and brine (20 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 18 (982 mg, 73%) as a colorless oil and 18' (313 mg, 23%) as a colorless oil. 18: $[\alpha]_D^{16}$ – 31.6 (c 1.00, CHCl₃); IR (film, cm⁻¹) v 3410, 1740, 1731, 1697, 999, 920; ¹H NMR (400 MHz, CDCl₃) δ 7.62 (t, *J* = 6.0 Hz, 4H), 7.45–7.37 (m, 6H), 5.87 (dd, J = 17.2, 11.0 Hz, 1H), 5.32 (s, 1H), 5.30 (d, J = 11.0 Hz, 1H), 5.18 (d, J = 17.2 Hz, 1H), 4.80 (br s, 1H), 4.03 (dd, J =11.8, 7.8 Hz, 1H), 4.27 (dd, J = 11.8, 2.8 Hz, 1H), 3.86 (t, J = 7.8 Hz, 1H), 3.77 (d, J = 10.0 Hz, 1H) 3.70 (d, J = 10.0 Hz, 1H), 2.02 (s, 3H), 1.45 (s, 9H), 1.07 (s, 9H); ^{13}C NMR (100 MHz, CDCl₃) δ 170.9, 156.3, 136.7, 135.5, 135.4, 132.3, 132.1, 129.9, 127.8, 127.7, 80.3, 72.5, 66.2, 65.9, 63.4, 60.2, 28.1, 26.7, 20.9, 20.8, 19.1; HRMS (EI) m/z calcd for $C_{29}H_{41}NO_6SSi (M)^+$ 527.2703, found 527.2705. **18'**: $[\alpha]_D^{26}$ +22.1 (c 0.5, CHCl₃); IR (film, cm⁻¹) v 3425, 3361, 1739, 1724, 1495, 1042, 925; ¹H NMR (400 MHz, CDCl₃) δ 7.64–7.60 (m, 4H), 7.45–7.37 (m, 6H), 6.04 (dd, J = 17.7, 11.0 Hz, 1H), 5.39 (s, 1H), 5.40 (d, *J* = 11.0 Hz, 1H), 5.180 (t, *J* = 3.2 Hz, 1H), 5.177 (d, *J* = 17.7 Hz, 1H), 3.87 (m, 1H), 3.80 (d, J = 10.2 Hz, 1H), 3.78 (m, 1H), 3.62 (d, J = 10.2 Hz, 1H), 1.96 (s, 3H), 1.44 (s, 9H), 1.09 (s, 9H); 13 C NMR (100 MHz, CDCl₃) δ 170.3, 155.0, 135.7, 135.6, 132.6, 132.5, 129.94, 129.90, 127.9, 127.8, 116.0, 80.0, 77.2, 74.7, 65.8, 61.9, 61.5, 28.3, 26.8, 21.0, 19.2; HRMS (FAB) m/z calcd for C₂₉H₄₁NO₆SSi (M+H)⁺ 527.2781, found 527.2748.

(2R,3R)-3-(tert-Butyldiphenylsilyloxy)methyl-2-(2-4.3.3. chloro)acetoxy-3-(1,1-dimethylethoxycarbonyl)amino-4-pentenyl acetate (19). A solution of chloroacetyl chloride (122 mg, 1.08 mmol) in CH₂Cl₂ (1.0 mL) and pyridine (0.77 mL) were added to a solution of alcohol 18 (284 mg, 0.538 mmol) in CH₂Cl₂ (1.6 mL) at 0 °C. After stirring at room temperature for 1 h, the reaction was quenched with 5% NaHCO₃ and the whole mixture was extracted with EtOAc (3 x 10 mL). The combined organic layers were washed with water (10 mL) and brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 19 (239 mg, 73%) as a colorless oil: $[\alpha]_{D}^{25}$ –1.0 (c 1.00, CHCl₃); IR (film, cm⁻¹) v 3430, 1772, 1747, 1729, 1699, 1008, 956; ¹H NMR (400 MHz, CDCl₃) δ 7.62 (d, *J* = 6.4 Hz, 4H), 7.44–7.36 (m, 6H), 5.93 (dd, *J* = 17.8, 11.2 Hz, 1H), 5.75 (dd, J = 8.2, 2.2 Hz, 1H), 5.30 (d, J = 11.2 Hz, 1H), 5.25 (d, J = 17.8 Hz, 1H), 5.06 (s, 1H), 4.53 (dd, J = 12.4, 2.2 Hz, 1H), 4.11 (dd, *J* = 12.4, 8.2 Hz, 1H), 3.97 (d, *J* = 10.0 Hz, 1H), 3.87 (d, J = 14.4 Hz, 1H), 3.84 (d, J = 10.0 Hz, 1H), 3.80 (d, J = 14.4 Hz, 1H), 2.01 (s, 3H), 1.43 (s, 9H), 1.08 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 170.6 165.9, 154.3, 135.6, 134.9, 132.5, 129.79, 129.76, 127.7, 127.6, 116.5, 79.8, 76.7, 73.4, 63.8, 63.0, 61.6, 40.4, 28.2, 26.7, 20.6, 19.1; HRMS (FAB) m/z calcd for C₃₁H₄₃NO₇SSi (M+H)⁺ 604.2497, found 604.2521.

4.3.4. (2R,3R)-4-Acetoxy-2-(tert-butyldiphenylsilyloxy)methyl-2-(1,1-dimethylethoxycarbonyl)aminobutane-1,3-diol (20). Ozone was bubbling to a solution of **19** (1.13 g, 1.86 mmol) in CH₂Cl₂ (25 mL) and MeOH (25 mL) at -78 °C. After stirring for 15 min, NaBH₄ (105 mg, 2.79 mmol) was added to the reaction mixture at -78 °C. After stirring at room temperature for 1 h, the reaction was quenched with acetone, and the solvent was removed in vacuo. The resulting residue was solved with EtOAc (40 mL), and washed with water (15 mL) and brine (15 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide **20** (817 mg, 82%) as a white solid: mp 100–101 °C; $[\alpha]_D^{21}$ –21.9 (*c* 1.00, CHCl₃); IR (KBr, cm⁻¹) v 3450, 3320, 1719, 1677; ¹H NMR (400 MHz, CDCl₃) δ 7.64 (d, *J* = 7.2 Hz, 4H), 7.48–7.39 (m, 6H), 5.30 (s, 1H), 4.88 (d, *J* = 8.2 Hz, 1H), 4.20 (dd, *J* = 12.0, 2.8 Hz, 1H), 4.07 (dd, *J* = 12.0, 8.2 Hz, 1H), 3.99 (dd, *J* = 11.6, 4.8 Hz, 1H), 3.65 (d, *J* = 9.8 Hz, 1H), 3.80 (m, 1H), 3.74 (t, *J* = 11.6 Hz, 1H), 3.65 (d, *J* = 9.8 Hz, 1H), 2.94 (m, 1H), 2.01 (s, 3H), 1.44 (s, 9H), 1.09 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 171.0, 157.0, 135.53, 135.51, 132.3, 132.2, 130.1, 128.0, 127.9, 80.9, 71.1, 66.1, 63.4, 61.6, 61.5, 28.2, 26.8, 20.9, 19.3; HRMS (EI) *m/z* calcd for C₂₈H₄₁NO₇SSi (M)⁺ 531.2652, found 531.2666.

4.3.5. (4R,5R)-4-Acetoxymethyl-5-(tertbutyldiphenylsilyloxy)methyl-2,2-dimethyl-5-(1,1-

dimethylethoxycarbonyl)amino-1,3-dioxane (32). 2,2-Dimethoxypropane (0.82 mL, 6.7 mmol) and TsOH·H₂O (13 mg, 0.067 mmol) were added to a solution of diol 20 (358 mg, 0.67 mmol) in benzene (20 mL) at room temperature. After stirring for 1 h, the reaction was quenched with 5% NaHCO₃ and the whole mixture was extracted with EtOAc (3 x 10 mL). The combined organic layers were washed with water (10 mL) and brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 32 (359 mg, 94%) as a colorless oil: $[\alpha]_D^{18}$ +10.6 (*c* 1.00, CHCl₃); IR (film, cm⁻¹) v 3430, 1745, 1723; ¹H NMR (400 MHz, CDCl₃) δ 7.63 (d, J = 7.2 Hz, 4H), 7.45–7.37 (m, 6H), 5.12 (s, 1H), 4.46 (dd, J = 7.2, 2.4 Hz, 1H), 4.30 (dd, J = 12.4, 2.4 Hz, 1H), 4.21 (br)s, 1H), 4.06 (dd, J = 12.4, 7.2 Hz, 1H), 4.05 (d, J = 10.4 Hz, 1H), 3.95 (m, 1H), 3.89 (d, J = 10.4 Hz, 1H), 1.48 (s, 3H), 1.37 (s, 3H)9H), 1.08 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 170.6, 155.0, 135.6, 135.5, 132.62, 132.60, 130.0, 129.9, 127.8, 99.2, 79.4, 70.9, 64.0, 63.5, 63.2, 54.9, 28.8, 28.2, 26.8, 20.8, 19.2, 19.1; HRMS (EI) m/z calcd for $C_{31}H_{45}NO_7SSi$ (M)⁺ 571.2965, found 571.2923.

4.3.6. (*4R*,5*R*)-5-(*tert-Butyldiphenylsilyloxy*)*methyl*-2,2-*dimethyl*-5-(*1*,*1*-*dimethylethoxycarbonyl*)*amino*-4-*hydroxymethyl*-1,3-

dioxane (21). K₂CO₃ (179 mg, 1.3 mmol) was added to a solution of 32 (620 mg, 1.08 mmol) in MeOH (7.7 mL) at room temperature. After stirring for 1 h, the reaction was quenched with water and the whole mixture was extracted with EtOAc (3 x 15 mL). The combined organic layers were washed with brine (15 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 21 (565 mg, 98%) as a colorless oil: $[\alpha]_D^{18}$ +2.5 (*c* 1.00, CHCl₃); IR (film, cm⁻¹) v 3430, 3373, 1712, 1506; ¹H NMR (400 MHz, CDCl₃) δ 7.65–7.62 (m. 4H), 7.45–7.36 (m, 6H), 5.54 (s, 1H), 4.21 (d, J = 10.0 Hz, 1H), 4.12 (d, J = 6.8 Hz, 1H), 4.01 (br s, 2H), 3.83 (d, J = 11.2 Hz, 1H), 3.70-3.63 (m, 2H), 2.67 (br s, 1H), 1.44 (s, 3H), 1.43 (s, 3H), 1.39 (s, 9H), 1.07 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 155.5, 135.61, 135.57, 132.8, 129.9, 129.8, 127.8, 99.1, 79.4, 72.0, 64.3, 63.0, 61.6, 55.8, 28.4, 28.3, 26.9, 19.6, 19.3; HRMS (EI) m/z calcd for C₂₉H₄₃NO₆SSi (M)⁺ 529.2860, found 529.2805.

4.3.7. (4R,5R)-5-(*tert-Butyldiphenylsilyloxy*)*methyl*-2,2-*dimethyl*-5-(1,1-*dimethylethoxycarbonyl*)*amino*-4-*formyl*-1,3-*dioxane* (22). A solution of DMSO (0.15 mL, 2.13 mmol) in CH₂Cl₂ (1.5 mL) was added dropwise to a solution of oxalyl chloride (0.12 mL, 1.42 mmol) in CH₂Cl₂ (1.5 mL) at -78 °C and stirred for 5 min. A solution of alcohol **21** (376 mg, 0.71 mmol) in CH₂Cl₂ (4.4 mL) was added dropwise to the reaction mixture and stirred for 20 min. Et₃N (0.50 mL, 3.55 mmol) was added dropwise to the reaction mixture and stirred at -78 °C for 2.5 h, and the reaction mixture was allowed to warm to room temperature and stirred for 30 min. The reaction was quenched with saturated NH₄Cl and the whole mixture was extracted with CH₂Cl₂ (3 x 10 mL). The combined organic layers were washed with water (10 mL) and brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 22 (361 mg, 96%) as a colorless oil: $[\alpha]_{D}^{19}$ –6.3 (*c* 1.00, CHCl₃); IR (film, cm⁻¹) v 3422, 1733, 1714, 1502; ¹H NMR (400 MHz, CDCl₃) § 9.52 (s, 1H), 7.68–7.63 (m, 4H), 7.47–7.38 (m, 6H), 5.12 (s, 1H), 4.56 (s, 1H), 4.20 (d, J = 11.0 Hz, 1H), 4.09 (d, J = 12.0 Hz, 1H), 4.04 (d, J = 10.4 Hz, 1H), 3.84 (d, J = 11.0 Hz, 1H), 1.52 (s, 3H), 1.44 (s, 3H), 1.36 (s, 9H), 1.09 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 171.1, 154.9, 135.6, 135.5, 132.5, 132.4, 130.02, 129.97, 127.9, 99.7, 76.4, 63.9, 62.9, 60.3, 56.1, 28.3, 28.2, 26.8, 21.0, 19.2; HRMS (EI) m/z calcd for $C_{29}H_{41}NO_6SSi(M)^+$ 527.2703, found 527.2692.

4.4. Total synthesis of myriocin (1)

4.4.1. [1R,1(4R,5R)]-1-[5-(tert-Butyldiphenylsilyloxy)methyl-2,2dimethyl-5-(1,1-dimethylethoxycarbonyl)amino-1,3-dioxane-4yl]-3-butene-1-ol (23) [1S,1(4R,5R)]-1-[5-(tert-Butyldiphenylsilyloxy)methyl-2,2-dimethyl-5-(1,1-

dimethylethoxycarbonyl)amino-1,3-dioxane-4-yl]-3-butene-1-ol (23'). MgI₂ (37 mg, 0.13 mmol) was added to a solution of aldehyde 22 (35 mg, 0.066 mmol) in DMF (1.0 mL) at room temperature and stirred for 30 min. Indium (11 mg, 0.099 mmol), a solution of allyl bromide (16 mg, 0.13 mmol) in DMF (1.0 mL), and tetrabutylammonium iodide (TBAI, 24 mg, 0.066 mmol) were added to the reaction mixture. After stirring overnight, the reaction was quenched with saturated NH₄Cl and the whole mixture was extracted with Et₂O (3 x 10 mL). The combined organic layers were washed with water (10 mL) and brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 23 (27 mg, 73%) and 23' (4 mg, 11%) as a colorless oil. 23: $[\alpha]_D$ +17.5 (c 1.00, CHCl₃); IR (film, cm⁻¹) v 3553, 3377, 3073, 1710, 1511; ¹H NMR (400 MHz, CDCl₃) δ 7.64–7.61 (m, 4H), 7.43– 7.36 (m, 6H), 6.66 (s, 1H), 5.71 (m, 1H), 5.09 (d, J = 17.9 Hz, 1H), 5.08 (d, J = 10.0 Hz, 1H), 4.25 (d, J = 10.8 Hz, 1H), 4.06 (ddd, *J* = 9.9, 8.0, 6.0 Hz, 1H), 3.99 (s, 2H), 3.97 (d, *J* = 9.8 Hz, 1H), 3.89 (d, J = 10.8 Hz, 1H), 2.37 (m, 1H), 2.34 (d, J = 9.8 Hz, 1H), 2.19 (m, 1H), 1.44 (s, 3H), 1.42 (s, 3H), 1.37 (s, 9H), 1.07 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 155.2, 135.6, 135.5, 134.1, 132.8, 132.7, 129.9, 129.8, 127.8, 127.7, 118.0, 99.5, 78.9, 70.9, 68.9, 63.9, 63.4, 57.1, 40.1, 28.4, 27.0, 26.9, 20.8, 19.2; HRMS (FAB) m/z calcd for $C_{32}H_{47}NO_6SSi$ (M+H)⁺ 570.3251, found 570.3216. **23'**: $[\alpha]_D^{23}$ –2.50 (*c* 0.5, CHCl₃); IR (film, cm⁻ ¹) \vee 3432, 3073, 1695, 1506; ¹H NMR (400 MHz, CDCl₃) δ7.67–7.64 (m, 4H), 7.45–7.34 (m, 6H), 5.90 (m, 1H), 5.30 (s, 1H), 5.07 (d, J = 17.6 Hz, 1H), 5.06 (d, J = 10.0 Hz, 1H), 4.69 (br s, 1H), 4.53 (d, J = 10.6 Hz, 1H), 3.90 (s, 2H), 3.79 (d, J = 10.6 Hz, 1H), 3.65-3.57 (m, 2H), 2.48-2.44 (m, 2H), 2.15 (ddd, J =13.6, 12.3, 10.2 Hz, 1H), 1.42 (s, 3H), 1.40 (s, 3H), 1.36 (s, 9H), 1.05 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 156.5, 135.6, 135.1, 132.9, 132.8, 129.8, 129.7, 127.7, 127.6, 117.0, 98.8, 79.9, 74.6, 70.0, 66.8, 63.0, 56.7, 37.7, 28.34, 28.29, 26.8, 19.3; HRMS (FAB) m/z calcd for C₃₂H₄₈NO₆SSi (M+H)⁺ 570.3251, found 570.3267.

4.4.2. [1R,1(4R,5R)]-1-[5-(tert-Butyldiphenylsilyloxy)methyl-2,2dimethyl-5-(1,1-dimethylethoxycarbonyl)amino-1,3-dioxane-4-

yl]-3-butenyl acetate (24). Ac₂O (4 mL) was added to a solution of 23 (408 mg, 0.71 mmol) in pyridine (4 mL) at room temperature. After stirring overnight, the solvent was removed in vacuo. The resulting residue was solved with EtOAc (20 mL), and washed water (10 mL) and brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 24 (417 mg, 96%) as a colorless oil: $[\alpha]_{D}^{21}$ +50.0 (c 0.50, CHCl₃); IR (film, cm⁻¹) v 3441, 3074, 3051, 1748, 1722, 1496; ¹H NMR (400 MHz, CDCl₃) δ 7.67–7.61 (m, 4H), 7,47-7.36 (m, 6H), 5.64 (br s, 1H), 5.48 (br s, 1H), 5.39 (br s, 1H), 5.11 (d, J = 17.6 Hz, 1H), 5.06 (d, J = 10.0 Hz, 1H), 4.41 (s, 1H), 4.30 (d, J = 10.8 Hz, 1H), 4.16 (m, 1H), 4.00 (d, J = 10.8 Hz, 1H), 3.84 (d, J = 10.8 Hz, 1H), 2.39 (t, J = 6.4 Hz, 2H), 2.08(s, 3H), 1.50 (s, 3H), 1.46 (s, 3H), 1.34 (s, 9H), 1.08 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 170.0, 154.7, 135.6, 135.5, 133.2, 132.7, 129.9, 127.82, 127.81, 118.5, 99.4, 78.9, 70.2, 70.0, 63.5, 62.9, 55.6, 44.3, 36.1, 29.2, 28.3, 27.0, 21.0, 19.2, 18.8; HRMS (EI) m/z calcd for C₃₄H₄₉NO₇SSi (M)⁺ 611.3278, found 611.3238.

4.4.3. [1R,1(4R,5R),3E]-1-Acetoxy-1-[5-(tertbutyldiphenylsilyloxy)methyl-2,2-dimethyl-5-(1,1-

dimethylethoxycarbonyl)amino-1,3-dioxane-4-yl]heptadec-3-en-11-one (25). Pentadec-1-en-9-one (5,11 88 mg, 0.42 mmol) and second generation Grubbs catalyst (4.5 mg, 0.0053 mmol) were added to a solution of alkene 24 (64 mg, 0.11 mmol) in CH₂Cl₂ (3.3 mL). After stirring at reflux for 5.5 h, the solvent was removed in vacuo. The resulting residue was purified by column chromatography (silica gel, 5% EtOAc in hexane) to provide **25** (81 mg, 96%) as a colorless oil: $[\alpha]_D^{13}$ +33.5 (*c* 1.00, CHCl₃); IR (film, cm^{-1}) v 3450, 3073, 1745, 1718, 1499; ¹H NMR (400 MHz, CDCl₃) δ 7.67–7.62 (m, 4H), 7.45–7.36 (m, 6H), 5.49 (br s, 2H), 5.37 (br s, 1H), 5.26 (br s, 1H), 4.40 (s, 1H), 4.28 (d, J = 10.0 Hz, 1H), 4.17 (m, 1H), 4.00 (d, J = 11.2 Hz, 1H), 3.83 (d, J = 10.0 Hz, 1H), 2.38 (t, J = 7.6 Hz, 4H), 2.33 (m, 1H), 2.07 (br s, 3H), 1.93 (d, J = 5.6 Hz, 2H), 1.55 (dd, J = 14.4, 7.2 Hz, 2H), 1.55 (m, 1H), 1.50 (s, 3H), 1.46 (s, 3H), 1.33 (s, 9H), 1.20 (br s, 11H), 1.08 (s, 9H), 0.88 (t, J = 7.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) & 211.5, 169.9, 154.6, 150.0, 135.5, 134.7, 132.7, 129.9, 127.8, 124.3, 124.2, 99.4, 78.8, 70.5, 70.1, 63.4, 62.9, 55.7, 49.3, 42.8, 34.9, 32.5, 31.6, 29.2, 29.1, 28.9, 26.9, 23.82, 23.76, 23.6, 22.5, 21.1, 19.2, 18.8, 14.0; HRMS (FAB) m/z calcd for $C_{47}H_{74}NO_7SSi (M+H)^+ 808.5184$, found 808.5166.

4.4.4. [1R,1(4R,5S),3E]-1-Acetoxy-1-[2,2-dimethyl-5-(1,1dimethylethoxycarbonyl)amino-5-hydroxymethyl-1,3-dioxane-4yl]heptadec-3-en-11-one (33). Tetrabutylammonium fluoride (TBAF, 0.14 mL, 1 M in THF, 0.14 mmol) and a solution of acetic acid (8 mg, 0.14 mmol) in THF (0.2 mL) were added to a solution of 25 (37 mg, 0.046 mmol) in THF (0.24 mL) at 0 °C. After stirring at 50 °C for 24 h, the reaction was quenched with water and the whole mixture was extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with water (5 mL) and brine (5 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide **33** (22 mg, 84%) as a colorless oil: $[\alpha]_D^{13}$ +56.6 (*c* 1.00, CHCl₃); IR (film, cm⁻¹) v 3438, 1744, 1711, 1701; ¹H NMR (400 MHz, CDCl₃) δ 5.61 (s, 1H), 5.50 (dt, *J* = 15.2, 6.8 Hz, 1H), 5.26 (dt, J = 15.2, 6.8 Hz, 1H), 5.02 (t, J = 6.8 Hz, 1H), 4.75 (d, J =9.6 Hz, 1H), 4.08 (d, J = 12.4 Hz, 1H), 4.00 (d, J = 12.0 Hz, 1H), 3.75 (d, J =12.4 Hz, 1H), 3.75 (d, J =12.0 Hz, 1H), 3.45 (t, J = 10.8 Hz, 1H), 2.39 (t, J = 7.2 Hz, 4H), 2.30–2.26 (m, 2H), 2.10 (s, 3H), 2.01-1.95 (m, 2H), 1.57-1.54 (m, 3H), 1.52 (s, 3H), 1.43 (s, 12H), 1.27 (br s, 12H), 0.88 (t, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) & 211.5, 169.9, 156.4, 135.2, 123.9, 99.6,

80.0, 72.6, 69.9, 64.7, 64.6, 55.7, 42.8, 42.6, 34.5, 32.4, 31.5, 29.1, 28.95, 28.95, 28.7, 28.3, 23.8, 23.5, 22.4, 21.0, 18.3, 14.0; HRMS (EI) m/z calcd for $C_{31}H_{55}NO_8$ (M)⁺ 569.3928, found 569.3979.

4.4.5. [1R,1(4R,5S),3E]-1-Acetoxy-1-[2,2-dimethyl-5-(1,1-dimethylethoxycarbonyl)amino-5-formyl-1,3-dioxane-4-

yl]heptadec-3-en-11-one (26). TEMPO (3.8 mg, 0.024 mmol), PhI(OAc)₂ (51 mg, 0.16 mmol) and MgO (13 mg, 0.32 mmol) were added to a solution of alcohol 33 (45 mg, 0.079 mmol) in CH₂Cl₂ (1.8 mL). After stirring at room temperature overnight, the reaction mixture was filtered through a pad of Celite. The filter cake was rinsed with EtOAc and the combined filtrate was concentrated in vacuo. The resulting residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide **26** (36 mg, 81%) as a colorless oil: $[\alpha]_D^{-25}$ +46.7 (*c* 1.00, CHCl₃); IR (film, cm⁻¹) v 3456, 1746, 1723, 1717; ¹H NMR (400 MHz, CDCl₃) δ 9.67 (s, 1H), 5.65 (s, 1H), 5.47 (dt, J = 15.2, 6.8Hz, 1H), 5.20 (dt, J = 15.2, 6.8 Hz, 1H), 5.02 (td, J = 7.2, 2.0 Hz, 1H), 4.15 (d, J = 12.4 Hz, 1H), 4.01 (d, J = 12.4 Hz, 1H), 3.95 (d, *J* = 2.0 Hz, 1H), 2.39 (t, *J* = 7.2 Hz, 4H), 2.34–2.26 (m, 2H), 2.13 (s, 3H), 2.00-1.93 (m, 2H), 1.57-1.52 (m, 3H), 1.50 (s, 3H), 1.46 (s, 12 H), 1.32–1.26 (m, 12 H), 0.88 (t, J = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 211.5, 200.8, 170.0, 155.3, 135.2, 123.5, 99.9, 80.7, 72.1, 71.0, 63.4, 63.0, 42.7, 42.6, 34.2, 32.2, 31.5, 29.02, 28.98, 28.8, 28.6, 28.2, 23.8, 23.5, 22.4, 20.1, 18.2, 14.0; HRMS (FAB) m/z calcd for $C_{31}H_{54}NO_8 (M+H)^+$ 568.3849, found 568.3842.

4.4.6. (2S,3R,4R)-2-Acetamido-3-acetoxy-2-acetoxymethyl-4-[(E)-10-oxohexadec-2-en-1-yl]-4-butanolide (27).^{6i,k,l,o} A solution of NaClO₂ (29 mg, 0.32 mmol) and NaH₂PO₄·H₂O (22 mg, 0.16 mmol) in water (0.45 mL) was added to a solution of **26** (45 mg, 0.0793 mmol) and 2-methyl-2-butene (0.21 ml) in *t*-BuOH (1.4 mL). After stirring at room temperature for 3 h, the reaction was quenched with H₂O and the whole mixture was extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo to provide the crude carboxylic acid (45 mg), which was used in the next step without further purification.

10% NaOH (0.63 mL) was added to a solution of the crude carboxylic acid in MeOH (0.63 mL). After stirring at 70 °C overnight, the reaction mixture was acidified to pH 3 with 10 % HCl, and extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (5 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo to provide the crude alcohol (50 mg), which was used in the next step without further purification.

10% HCl (0.63 mL) was added to a solution of the crude alcohol in MeOH (0.63 mL). After stirring at 80°C overnight, the reaction mixture was alkalified with K_2CO_3 . Insoluble materials were filtered off, and the filtrate was concentrated in vacuo to provide the crude product (90 mg), which was used in the next step without further purification.

Ac₂O (0.63 mL, 6.7 mmol) was added to a solution of the crude product in pyridine (0.63 mL). After stirring at room temperature overnight, the solvent was removed in vacuo. The resulting residue was solved with EtOAc (10 mL), and washed with water (5 mL) and brine (5 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 40% EtOAc in hexane) to provide **27** (20 mg, 51% for 4 steps) as a colorless oil: $[\alpha]_D^{18}$ +55.6 (*c* 1.00, CHCl₃) {lit.^{6h} $[\alpha]_D^{23}$ +52.5 (*c* 0.85, CHCl₃)}; ¹H NMR (400 MHz, CDCl₃) δ 6.30 (s, 1H), 5.79 (d, *J* = 4.0 Hz, 1H), 5.56 (dt, *J* = 15.2, 6.8 Hz, 1H), 5.39 (dt, *J* = 15.2, 6.4 Hz, 1H),

4.72 (dt, J = 8.4, 4.8 Hz, 1H), 4.52 (s 2H), 2.38 (t, J = 7.2 Hz, 4H), 2.46–2.36 (m, 2H), 2.10 (s, 3H), 2.05 (s, 3H), 2.03 (s, 3H), 2.07–1.98 (m, 2H), 1.59–1.52 (m, 4H), 1.37–1.27 (m, 12H), 0.88 (t, J = 7.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 211.7, 172.4, 170.2, 169.4, 168.9, 135.0, 123.1, 81.6, 71.9, 62.62, 62.57, 42.8, 42.7, 32.4, 32.1, 31.6, 29.0, 28.93, 28.90, 28.86, 23.80, 23.76, 22.7, 22.5, 20.6, 20.3, 14.0.

4.4.7. Myriocin (1). 10% NaOH (1.3 mL) was added to 27 (20 mg, 0.039 mmol) in MeOH (1.3 mL) and stirring at 80 °C for 2 h. The reaction mixture was cooled to 0 °C, and then neutralized with Amberlite[®] IRC-86 (H⁺ type). The insoluble material was removed by filtration and the filtrate was concentrated in vacuo. The resulting residue was purified by column chromatography (1:1:10 to 1:3:10 (gradient) H₂O/MeOH/CHCl₃ (lower phase)) to afford myriocin (1, 14 mg, 88%) as white crystals: mp 165–168 °C (lit.³ mp 164–168 °C); $[\alpha]_D^{23}$ +5.6 (*c* 0.30, DMSO) {lit.^{6h} $[\alpha]_D^{20}$ +6.1 (*c* 0.26, DMSO)}; IR (KBr, cm⁻¹) v 3278, 3203, 1714, 1635, 1465, 1387, 1104, 970; ¹H NMR (400 MHz, CD₃OD) δ 5.54 (dt, J = 16.4, 6.4 Hz, 1H), 5.40 (dt, J = 14.8, 7.2 Hz, 1H), 4.00 (d, J = 11.2 Hz, 1H), 3.87 (d, J = 11.6 Hz, 1H), 3.83 (t, J = 5.6 Hz, 1H), 3.81 (s, 1H), 2.43 (t, J = 7.2 Hz, 4H), 2.27 (t, J = 6.8 Hz, 2 H), 2.00 (dd, J = 14.0, 7.2 Hz, 2H), 1.55 (t, J = 6.8 Hz, 4H), 1.29 (br s, 12H), 0.90 (t, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CD₃OD) δ 214.4, 173.5, 134.7, 126.9, 73.7, 71.3, 70.4, 65.1, 43.54, 43.53, 38.7, 33.8, 32.9, 30.5, 30.23, 30.16, 30.1, 30.0, 24.9, 23.6, 14.4.

4.5. Total synthesis of mycestericin D (2)

4.5.1. *Methyl* [1(4R,5R),2E]-3-[5-(tertbutyldiphenylsilyloxy)methyl-2,2-dimethyl-5-(1,1-

dimethylethoxycarbonyl)amino-1,3-dioxane-4-yl]-2-butenoate (34). Ph₃PCH=CO₂Me (257 mg, 0.77 mmol) was added to a solution of aldehyde 22 (369 mg, 0.70 mmol) in benzene (6.7 mL) at room temperature. After stirring 1 h, the solvent was removed in vacuo. The resulting residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 34 (392 mg, 96%) as a colorless oil: $[\alpha]_D^{22} + 17.8$ (*c* 1.00, CHCl₃); IR (film, cm⁻¹) v 3431, 3073, 1723, 1716, 1495; ¹H NMR (400 MHz, CDCl₃) δ 7.66-7.63 (m, 4H), 7.47-7.38 (m, 6H), 6.93 (dd, J = 15.8, 2.5 Hz, 1H), 6.17 (dd, J = 15.8, 2.5 Hz, 1H), 5.00 (s, 1H), 4.73 (s, 1H), 4.16–4.09 (m, 3H), 3.88 (d, J = 11.0 Hz, 1H), 3.74 (s, 3H), 1.48 (s, 3H), 1.45 (s, 3H), 1.34 (s, 9H), 1.10 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 166.3, 154.9, 142.4, 135.6, 135.5, 132.5, 129.97, 129.93, 127.9, 122.7, 99.1, 79.5, 77.2, 70.7, 63.5, 62.9, 55.8, 51.5, 28.6, 28.2, 26.9, 19.23, 19.21; HRMS (FAB) m/z calcd for C₃₂H₄₆NO₇Si (M+H)⁺ 584.3044, found 584.3066.

4.5.2. Methyl [1(4R,5R)]-3-[5-(tert-butyldiphenylsilyloxy)methyl-2,2-dimethyl-5-(1,1-dimethylethoxycarbonyl)amino-1,3-dioxane-4-yl]butanoate (29). Pd/C (13 mg) was added to a solution of alkene 34 (130 mg, 0.223 mmol) in MeOH (3.9 mL). After stirring at room temperature for 4.5 h under hydrogen, the reaction mixture was filtered through a pad of Celite. The filter cake was rinsed with EtOAc and the combined filtrate was concentrated in vacuo. The resulting residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide **29** (125 mg, 96%) as a colorless oil: $[\alpha]_D^{25}$ +17.6 (*c* 0.5, CHCl₃); IR (film, cm⁻¹) v 3427, 1739, 1715, 1495; ¹H NMR (400 MHz, CDCl₃) δ 7.65–7.61 (m, 4H), 7.45–7.36 (m, 6H), 4.97 (s, 1H), 4.21 (br s, 1H), 4.20 (d, J = 10.0 Hz, 1H), 4.06 (d, J = 11.2 Hz, 1H), 4.01 (br s, 1H), 3.81 (d, *J* = 11.2 Hz, 1H), 3.65 (s, 1H), 2.43 (ddd, J = 15.0, 8.4, 7.1 Hz, 1H), 2.23 (td, J = 15.0, 7.1 Hz, 1H), 1.82 (td, J = 15.0, 7.1 Hz, 1H), 1.74-1.64 (m, 1H), 1.43 (s, 3H), 1.39 (s, 3H), 1.37 (s, 9H), 1.08 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) & 173.6, 155.3, 135.60, 135.58, 132.81, 132.76, 129.9, 129.83, 127.78, 99.0, 79.1, 77.2, 71.5, 63.9, 63.2, 55.4, 51.4, 30.6, 28.9, 28.3, 26.9, 24.1, 19.2, 19.0; HRMS (FAB) m/z calcd for C₃₂H₄₈NO₇SSi (M+H)⁺ 585.3200, found 585.3206.

4.5.3. [1(4R,5R)]-1-[5-(tert-Butyldiphenylsilyloxy)methyl-2,2dimethyl-5-(1,1-dimethylethoxycarbonyl)amino-1,3-dioxane-4-

yl]-3-butene (**30**). DIBAL-H (0.28 ml, 0.28 mmol) was added dropwise to a solution of ester **29** (83 mg, 0.14 mmol) in CH₂Cl₂ (4.0 mL) at -78 °C under nitrogen. After stirring for 1 h, the reaction was quenched with saturated potassium sodium tartrate and the whole mixture was extracted with EtOAc (3 x 10 mL). The combined organic layers were washed with water (10 mL) and brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was filtered through a pad of Celite to provide crude aldehyde (81 mg), which was used in the next step without further purification.

Ph₃PCH₃Br (550 mg, 1.54 mmol) was added slowly to a suspension of NaH (60% in mineral oil, 56 mg, 1.4 mmol) in THF (2 mL) at 0 °C. After stirring for 3 h under nitrogen at room temperature, crude aldehyde (81 mg) in THF (3.3 mL) was added dropwise to the reaction mixture at 0 °C, and the resulting mixture was warmed to room temperature and stirred for 2 h. The reaction mixture was quenched with saturated NH₄Cl and the whole mixture was extracted with EtOAc (3 x 10 mL). The combined organic layers were washed with water (10 mL) and brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 30 (53 mg, 70% for 2 steps) as a colorless oil: $[\alpha]_{D}^{15}$ +22.9 (c 0.5, CHCl₃); IR (film, cm⁻¹) v 3427, 3073, 1722, 1495, 998, 913; ¹H NMR (400 MHz, CDCl₃) δ7.64-7.62 (m, 4H), 7.45-7.36 (m, 6H), 5.76 (m, 1H), 5.01 (d, J = 18.8 Hz, 1H), 4.96 (d, J = 10.0Hz, 1H), 4.24 (d, J = 10.0 Hz, 1H), 4.08 (d, J = 11.4 Hz, 1H), 4.00 (d, J = 11.4 Hz, 1H), 4.00 (d, J = 10.0 Hz, 1H), 3.81 (d, J = 10.0 Hz, 1H), 2.24 (m, 1H), 2.02 (m, 1H), 1.66-1.59 (m, 2H), 1.45 (s, 3H), 1.42 (s, 3H), 1.36 (s, 9H), 1.07 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 155.3, 138.1, 135.62, 135.60, 132.9, 132.8, 129.9, 129.8, 127.8, 115.1, 99.0, 79.0, 71.5, 64.0, 63.4, 55.5, 29.9, 29.0, 28.3, 27.7, 26.9, 19.28, 19.24; HRMS (FAB) m/z calcd for C₃₂H₄₈NO₅SSi (M+H)⁺ 554.3302, found 554.3307.

4.5.4. [1(4R,5R),3E]-1-[5-(tert-Butyldiphenylsilyloxy)methyl-2,2dimethyl-5-(1,1-dimethylethoxycarbonyl)amino-1,3-dioxane-4-

yl]heptadec-3-en-11-one (31). Pentadec-1-en-9-one (5,¹¹ 61 mg, 0.29 mmol) and second generation Grubbs catalyst (3 mg, 0.0036 mmol) were added to a solution of alkene 30 (40 mg, 0.072 mmol) in CH₂Cl₂ (2.3 mL). After stirring at reflux for 1 h, the solvent was removed in vacuo. The resulting residue was purified by column chromatography (silica gel, 5% EtOAc in hexane) to provide **31** (52 mg, 96%) as a colorless oil: $[\alpha]_D^{22}$ +22.6 (c 0.5, CHCl₃); IR (film, cm⁻¹) v 3425, 3072, 1721, 1717, 1495; ¹H NMR (400 MHz, CDCl₃) δ7.64–7.62 (m, 4H), 7.45–7.36 (m, 6H), 5.44-5.31 (m, 2H), 4.98 (s, 1H), 4.23 (d, J = 9.0 Hz, 1H), 4.17 (br s, 1H), 4.08 (dd, J = 11.6, 4.8 Hz, 1H), 4.01 (br s, 1H), 3.80 (d, J = 9.0 Hz, 1H), 2.38 (t, J = 7.2 Hz, 4H), 2.16 (m, 1H), 2.08-1.94 (m, 3H), 1.62-1.54 (m, 6H), 1.44 (s, 3H), 1.41 (s, 3H), 1.36 (s, 9H), 1.27 (br s, 12H), 1.07 (s, 9H), 0.88 (t, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 211.6, 181.3, 155.3, 135.62, 135.59, 132.9, 131.2, 130.4, 129.4, 127.8, 98.9, 78.9, 71.5, 64.0, 63.4, 55.6, 42.80, 42.77, 32.6, 31.6, 29.5, 29.3, 29.1, 29.01, 28.96, 28.9, 28.7, 28.3, 28.2, 26.9, 23.8, 23.7, 23.2, 22.5, 19.3, 19.2, 14.0; HRMS (FAB) m/z calcd for C₄₅H₇₂NO₆SSi (M+H)⁺ 750.5129, found 750.5103.

4.5.5. [1(4R,5S),3E]-1-[2,2-Dimethyl-5-(1,1dimethylethoxycarbonyl)amino-5-hydroxymethyl-1,3-dioxane-4yl]heptadec-3-en-11-one (35). TBAF (0.13 mL, 1 M in THF,

0.13 mmol) were added to a solution of **31** (47 mg, 0.063 mmol) in THF (0.58 mL) at 0 °C. After stirring at room temperature for 3 h, the reaction was guenched with water and the whole mixture was extracted with EtOAc (3 x 10 mL). The combined organic layers were washed with water (10 mL) and brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 20% EtOAc in hexane) to provide 35 (28 mg, 88%) as a colorless oil: $[\alpha]_{D}^{20}$ +25.9 (*c* 0.5, CHCl₃); IR (film, cm⁻¹) v 3424, 1712, 1697; ¹H NMR (400 MHz, CDCl₃) δ 5.50–5.25 (m, 2H), 4.78 (d, J = 12 Hz, 1H), 3.99 (d, J = 12.4 Hz, 1H), 3.93 (dd, J = 12.2, 2.4 Hz, 1H), 3.76 (d, J = 12.4 Hz, 1H), 3.68 (dd, J = 12.2, 2.4 Hz, 1H), 3.42 (t, J = 12.2 Hz, 1H), 2.38 (t, J = 7.2 Hz, 4H), 2.17-1.89 (m, 5H), 1.61-1.52 (m, 5H), 1.45 (s, 9H), 1.43 (s, 3H), 1.41 (s, 3H), 1.34–1.27 (m, 12H), 0.88 (t, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 211.6, 157.2, 131.6, 128.9, 98.9, 80.3, 73.6, 65.0, 64.8, 55.6, 42.8, 42.7, 32.5, 31.6, 29.4, 29.1, 28.94, 28.91, 28.4, 28.3, 28.0, 23.82, 23.79, 22.5, 18.6, 14.0; HRMS (FAB) m/z calcd for C₂₉H₅₄NO₆ (M+H)⁺ 512.3951, found 512.3946.

4.5.6. [1(4R,5S),3E]-1-[2,2-Dimethyl-5-(1,1dimethylethoxycarbonyl)amino-5-formyl-1,3-dioxane-4yl]heptadec-3-en-11-one (36). A solution of DMSO (26 mg, 0.328 mmol) in CH₂Cl₂ (0.3 mL) was added dropwise to a solution of oxalyl chloride (28 mg, 0.219 mmol) in CH₂Cl₂ (0.4 mL) at -78 °C under nitrogen and stirred for 5 min. A solution of alcohol 35 (28 mg, 0.0547 mmol) in CH₂Cl₂ (0.5 mL) was added dropwise to the reaction mixture and stirred for 20 min. A solution of Et₃N (56 mg, 0.547 mmol) in CH₂Cl₂ (0.3 mL) was added dropwise to the reaction mixture at -78 °C. The reaction mixture was warmed to -40 °C and stirred for 1 h. The reaction was quenched with saturated NH₄Cl and the whole mixture was extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with water (5 mL) and brine (5 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo, and the residue was purified by column chromatography (silica gel, 10% EtOAc in hexane) to provide 36 (26 mg, 93%) as a colorless oil: $[\alpha]_{D}^{21}$ +20.4 (c 0.5, CHCl₃); IR (film, cm⁻¹) v 3437, 3362, 1727, 1713, 1700; ¹H NMR (400 MHz, CDCl₃) δ 9.65 (s, 1H), 5.41 (s, 1H), 5.38 (m, 1H), 5.28 (m, 1H), 4.15 (d, J = 11.6 Hz, 1H), 3.97 (dd, J = 10.4, 2.0 Hz, 1H), 3.95 (d, J = 11.6 Hz, 1H), 2.38 (t, J = 8.0 Hz, 4H), 2.25-2.05 (m, 2H), 2.00-1.89 (m, 2H), 1.65 (m, 1H), 1.61-1.52 (m, 5H), 1.44 (s, 3H), 1.43 (s, 3H), 1.47 (s, 9H), 1.31–1.27 (m, 12H), 0.88 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) & 211.6, 200.3, 156.1, 131.8, 128.5, 99.3, 80.8, 72.8, 72.6, 63.3, 63.0, 42.79, 42.75, 32.4, 32.3, 31.6, 29.7, 29.3, 29.2, 29.1, 28.9, 28.2, 28.0, 23.8, 22.5, 18.5, 14.0; HRMS (FAB) m/z calcd for C₂₉H₅₂NO₆ (M+H)⁺ 510.3795, found 510.3799.

4.5.7. Mycestericin D (2). A solution of NaClO₂ (18 mg, 0.204 mmol) and NaH₂PO₄·H₂O (14 mg, 0.102 mmol) in water (0.28 mL) was added to a solution of **36** (26 mg, 0.051 mmol) and 2-methyl-2-butene (0.14 ml) in *t*-BuOH (0.9 mL). After stirring at room temperature for 3 h, the reaction was quenched with H₂O and the whole mixture was extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (10 mL), and dried over anhydrous MgSO₄. Filtrate was concentrated in vacuo to provide the crude carboxylic acid (45 mg), which was used in the next step without further purification.

10% HCl (1 mL) was added to a solution of the crude carboxylic acid in MeOH (1 mL). After stirring at 80 °C overnight, the reaction mixture was alkalified to pH 7 with K_2CO_3 . Insoluble materials were filtered off, and the filtrate was concentrated in vacuo. The resulting residue was purified by column chromatography (1:1:10 to 1:3:10 (gradient) H₂O/MeOH/CHCl₃

(lower phase)) to afford mycestericin D (2, 18 mg, 90% for 2 steps) as a white solid: mp 164–166 °C (lit.^{5a} mp 162–167 °C); $[\alpha]_D^{25}$ –7.48 (*c* 0.30, MeOH) {lit.^{5a} [α]_D –7.5 (*c* 0.16, MeOH)}; IR (KBr, cm⁻¹) v 3348, 3087, 1711, 1618, 1466, 968; ¹H NMR (400 MHz, CD₃OD) δ 5.52–5.34 (m, 2H), 3.99 (d, *J* = 11.0 Hz, 1H), 3.84–3.80 (m, 1H), 3.81 (d, *J* = 11.0 Hz, 1H), 2.43 (t, *J* = 7.2 Hz, 4H), 2.24 (m, 1H), 2.07–1.94 (m, 2H), 1.56–1.49 (m, 6H), 1.34–1.26 (m, 12H), 0.89 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CD₃OD) δ 214.4, 132.0, 130.6, 70.9, 64.70, 64.69, 43.49, 43.48, 33.6, 32.8, 32.6, 30.6, 30.3, 30.2, 30.1, 30.0, 24.9, 23.6, 14.4.

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Supplementary Material

Supplementary data associated with this article can be found, in the online version, at http://....

References and notes

- (a) Kluepfel, D.; Bagli, J.; Baker, H.; Charest, M.-P.; Kudelski, A.; Sehgal, S. N.; Vézina, C. J. Antibiot. 1972, 25, 109–115; (b) Bagli, J. F.; Kluepfel, D.; St-Jacques, M. J. Org. Chem. 1973, 38, 1253–1260.
- Aragozzini, F.; Manachini, P. L.; Craveri, R.; Rindone, B.; Scolastico, C. Tetrahedron 1972, 28, 5493–5498.
- Fujita, T.; Inoue, K.; Yamamoto, S.; Ikumoto, T.; Sasaki, S.; Toyama, R.; Chiba, K.; Hoshino, Y.; Okumoto, T. J. Antibiot. 1994, 47, 208–215.
- (a) Miyake, Y.; Kozutsumi, Y.; Nakamura, S.; Fujita, T.; Kawasaki, T. Biochem. Biophys. Res. Commun. 1995, 211, 396–403; (b) Fujita, T.; Hirose, R.; Yoneta, M.; Sasaki, S.; Inoue, K.; Kiuchi, M.; Hirase, S.; Chiba, K.; Sakamoto, H.; Arita, M. J. Med. Chem. 1996, 39, 4451–4459; (c) Chen, J. K.; Lane, W. S.; Schreiber, S. L. Chem. Biol. 1999, 6, 221–235; (d) Wadsworth, J. M.; Clarke, D. J.; McMahon, S. A.; Lowther, J. P.; Beattie, A. E.; Langridge-Smith, P. R. R.; Broughton, H. B.; Dunn, T. M.; Naismith, J. H.; Campopiano, D. J. J. Am. Chem. Soc. 2013, 135, 14276–14285.
- (a) Sasaki, S.; Hashimoto, R.; Kiuchi, M.; Inoue, K.; Ikumoto, T.; Hirose, R.; Chiba, K.; Hoshino, Y.; Okumoto, T.; Fujita, T. J. Antibiot. 1994, 47, 420–433; (b) Fujita, T.; Hamamichi, N.; Kiuchi, M.; Matsuzaki, T.; Kitao, Y.; Inoue, K.; Hirose, R.; Yoneta, M.; Sasaki, S.; Chiba, K. J. Antibiot. 1996, 49, 846–853.
- For total syntheses of myriocin, see: (a) Banfi, L.; Beretta, M. G.; Colombo, L.; Gennari, C.; Scolastico, C. J. Chem. Soc., Chem. Commun. 1982, 488-490; (b) Banfi, L.; Beretta, M. G.; Colombo, L.; Gennari, C.; Scolastico, C. J. Chem. Soc., Perkin Trans. 1 1983, 1613-1619; (c) Rao, A. V. R.; Gurjar, M. K.; Devi, T. R.; Kumar, K. R. Tetrahedron Lett. 1993, 34, 1653-1656; (d) Deloisy, S.; Thang, T. T.; Olesker, A.; Lukacs, G. Tetrahedron Lett. 1994, 35, 4783-4786; (e) Deloisy, S.; Thang, T. T.; Olesker, A.; Lukacs, G. Bull. Chim. Soc. Fr. 1996, 133, 581-585; (f) Yoshikawa, M.; Yokokawa, Y.; Okuno, Y.; Murakami, N. Chem. Pharm. Bull. 1994, 42, 994-996; (g) Yoshikawa, M.; Yokokawa, Y.; Okuno, Y.; Murakami, N. Tetrahedron 1995, 51, 6209-6228; (h) Hatakeyama, S.; Yoshida, M.; Esumi, T.; Iwabuchi, Y.; Irie, H.; Kawamoto, T.; Yamada, H.; Nishizawa, M. Tetrahedron Lett. 1997, 38, 7887-7890; (i) Sano, S.; Kobayashi, Y.; Kondo, T.; Takebayashi, M.; Maruyama, S.; Fujita, T.; Nagao, Y. Tetrahedron Lett. 1995, 36, 2097-2100; (j) Lee, K.-Y.; Oh, C.-Y.; Kim, Y.-H.; Joo, J.-E.; Ham, W.-H. Tetrahedron Lett. 2002, 43, 9361-9396; (k) Oishi, T.; Ando, K.; Chida, N. Chem. Commun. 2001, 1932-1933; (l) Oishi, T.; Ando, K.; Inomiya, K.; Sato, H.; Iida, M.; Chida, N. Bull. Chem. Soc. Jpn. 2002, 75, 1927-1947; (m) Torrente, S.; Alonso, R. Org. Lett. 2001, 3, 1985-1987; (n) Inai, M.; Goto, T.; Furuta, T.; Wakimoto, T.; Kan, T. Tetrahedron: Asymmetry 2008, 19, 2771-2773; (o) Jones, M. C.; Marsden, S. P. Org. Lett. 2008, 10, 4125-4128.
- For total synthesis of mycestericin A, see: (a) Sato, H.; Sato, K.; Iida, M.; Yamanaka, H.; Oishi, T.; Chida, N. *Tetrahedron Lett.* 2008, 49, 1943–1947; (b) Yamanaka, H.; Sato, K.; Sato, H.; Iida, M.; Oishi, T.; Chida, N. *Tetrahedron* 2009, 65, 9188–9201; For total synthesis of mycestericin C, see: (c) Sakamoto, S.; Kazumi, N.; Kobayashi, Y.;

- Tsukano, C.; Takemoto, Y. Org. Lett. 2014, 16, 4758-4761; For total synthesis of mycestericin D, see: (d) Shibata, K.; Shingu, K.; Vassile, V. P.; Nishide, K.; Fujita, T.; Node, M.; Kajimoto, T.; Wong, C.-H. Tetrahedron Lett. 1996, 37, 2791-2794; (e) Nishide, K.; Shibata, K.; Fujita, T.; Kajimoto, T.; Wong, C.-H.; Node, M. Heterocycles, 2000, 52, 1191-1201; For total synthesis of mycestericins E and G, see: (f) Fujita, T.; Hamamichi, N.; Matsuzaki, T.; Kitao, Y.; Kiuchi, M.; Node, M.; Hirose, R. Tetrahedron Lett. 1995, 36, 8599-8602; For total synthesis of mycestericin E, see: (g) Iwabuchi, Y.; Furukawa, M.; Esumi, T.; Hatakeyama, S. Chem. Commun. 2001, 2030-2031; For total syntheses of mycestericins F and G, see: (h) Berhal, F.; Takechi, S.; Kumagai, N.; Shibasaki, M. Chem. Eur. J. 2011, 17, 1915-1921; (i) Martinková, M.; Gonda, J.; Uhríková, A.; Raschmanová, J. Š.; Vilková, M.; Oroszová, B. Tetrahedron: Asymmetry 2013, 24, 121-133; For total synthesis of mycestericin G, see: (j) Fairhurst, N. W. G.; Mahon, M. F.; Munday, R. H.; Carbery, D. R. Org. Lett., 2012, 14, 756-759.
- (a) Yakura, T.; Yoshimoto, Y.; Ishida, C.; Mabuchi, S. Synlett 2006, 930–932;
 (b) Yakura, T.; Yoshimoto, Y.; Ishida, C.; Mabuchi, S. Tetrahedron 2007, 63, 4429–4438;
 (c) Yakura, T.; Sato, S.; Yoshimoto, Y. Chem. Pharm. Bull. 2007, 55, 1284–1286.
- 9. For a preliminary communication on the total synthesis of myriocin, see: Nambu, H.; Noda, N.; Niu, W.; Fujiwara, T.; Yakura, T. *Asian J. Org. Chem.* **2015**, *4*, 1246–1249.
- (a) Fleming, J. J.; Fiori, K. W.; Du Bois, J. J. Am. Chem. Soc. 2003, 125, 2028–2029; (b) Fleming, J. J.; Du Bois, J. J. Am. Chem. Soc. 2006, 128, 3926–3927; (c) Fleming, J. J.; McReynolds, M. D.; Du Bois, J. J. Am. Chem. Soc. 2007, 129, 9964–9975.
- 11. Hayes, C. J.; Bradley, D. M.; Thomson, N. M. J. Org. Chem. 2006, 71, 2661–2665.
- 12. (a) Espino, C. G.; Du Bois, J. Angew. Chem. 2001, 113, 618-620; Angew. Chem. Int. Ed. 2001, 40, 598-600; (b) Espino, C. G.; Wehn, P. M.; Chow, J.; Du Bois, J. J. Am. Chem. Soc. 2001, 123, 6935-6936; (c) Wehn, P. M.; Lee, J.; Du Bois, J. Org. Lett. 2003, 5, 4823-4826; (d) Espino, C. G.; Fiori, K. W.; Kim, M.; Du Bois, J. J. Am. Chem. Soc. 2004, 126, 15378-15379; (e) Fiori, K. W.; Fleming, J. J.; Du Bois, J. Angew. Chem. 2004, 116, 4449-4452; Angew. Chem. Int. Ed. 2004, 43, 4349-4352; (f) Kim, M.; Mulcahy, J. V.; Espino, C. G.; Du Bois, J. Org. Lett. 2006, 8, 1073-1076; (g) Conrad, R. M.; Du Bois, J. Org. Lett. 2007, 9, 5465-5468; (h) Zalatan, D. N.; Du Bois, J. J. Am. Chem. Soc. 2008, 130, 9220-9221; (i) Olson, D. E.; Du Bois, J. J. Am. Chem. Soc. 2008, 130, 11248-11249; (j) Fiori, K. W.; Espino, C. G.; Brodsky, B. H.; Du Bois, J. Tetrahedron 2009, 3042-3051; (k) Wehn, P. M.; Du Bois, J. Angew. Chem. 2009, 121, 3860–3863; Angew. Chem. Int. Ed. 2009, 48, 3802-3805; (1) Du Bois, J. Org. Process Res. Dev. 2011, 15, 758-762; (m) Olson, D. E.; Roberts, D. A.; Du Bois, J. Org. Lett. 2012, 14, 6174-6177; (n) Roizen, J. L.; Zalatan, D. N.; Du Bois, J. Angew. Chem. 2013, 125, 11553-11556; Angew. Chem. Int. Ed. 2013, 52, 11343-11346; (o) Bess, E. N.; DeLuca, R. J.; Tindall, D. J.; Oderinde, M. S.; Roizen, J. L.; Du Bois, J.; Sigman, M. S. J. Am. Chem. Soc. 2014, 136 5783-5789
- (a) Uchida, K.; Kato, K.; Akita, H. Synthesis 1999, 1678–1686; (b) Yang, J.-H.; Liu, J.; Hsung, R. P. Org. Lett. 2008, 10, 2525–2528; (c) David, O.; Blot, J.; Bellec, C.; Fargeau-Bellassoued, M.-C.; Haviari, G.; Célérier, J.-P.; Lhommet, G.; Gramain, J.-C.; Gardette, D. Bioconjugate Chem. 2008, 19, 1855–1863.
- Thornton, A. R.; Martin, V. I.; Blakey, S. B. J. Am. Chem. Soc. 2009, 131, 2434–2435.
- 15. CCDC 1517011 (16) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre (CCDC) via www.ccdc.cam.ac.uk.
- (a) Araki, S.; Ito, H.; Butsugan, Y. J. Org. Chem. 1988, 53, 1831–1833;
 (b) Kim, E.; Gordon, D. M.; Schmid, W.; Whitesides, G. M. J. Org. Chem. 1993, 58, 5500–5507; (c) Paquette, L. A.; Mitzel, T. M. J. Am. Chem. Soc. 1996, 118, 1931–1937; (d) Chan, T. H.; Yang, Y. J. Am. Chem. Soc. 1999, 121, 3228–3229.
- 17. The stereochemistry of the newly formed chiral center in 23 was determined by its conversion to myriocin.
- Derguini-Boumechal, F.; Lorne, R.; Linstrumelle, G. *Tetrahedron Lett.* 1977, 1181–1184.