

Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry

journal homepage: www.elsevier.com/locate/bmc

Synthesis of highly deuterium-labeled (R)-K-13675, PPAR α agonist, for use as an internal standard for low-level quantification of drugs in plasma

Yukiyoshi Yamazaki, Shin-ichiro Ogawa, Kimiyuki Shibuya *

Tokyo New Drug Research Laboratories, Pharmaceutical Division, Kowa Co., Ltd, 2-17-43, Noguchicho, Higashimurayama, Tokyo 189-0022, Japan

ARTICLE INFO

Article history: Received 4 December 2008 Revised 20 January 2009 Accepted 21 January 2009 Available online 13 February 2009

Keywords: Deuterium-labeled Pt/C catalyzed deuteration Precise quantification Internal standard

1. Introduction

Peroxisome proliferator-activated receptor α (PPAR α) is a member of one of the nuclear receptor superfamilies, activation of which leads to a decrease in triglyceride levels and an increase in HDL-cholesterol levels in humans.¹ During the course of our drug discovery program, we identified (R)-2-[3-[[benzoxazol-2-yl]3-(4methoxyphenoxy)propyl]amino]methyl]phenoxy]butanoic acid ((R)-K-13675 (1), Fig. 1) as a highly potent and selective PPAR α agonist, and its in vivo efficacy was shown to be more than 100 times higher than that of fenofibrate.²

As we might anticipate a clinical dose as low as one hundredth that of fenofibrate, it was necessary to develop and validate a method for quantifying trace amounts (less than ng/mL) of 1 in human plasma for pharmacokinetic studies. Deuterium-labeled compounds are well known to be useful as internal standards for such applications.³ However, for precise low-level quantifications, the purities of deuterated compounds must be very high. Otherwise residual non-deuterated compounds that can still be detected with high sensitivity, might interfere. Therefore the efficient synthesis of such compounds from appropriate reagents is a significant challenge in medicinal chemistry. Although a number of deuteration methods have been reported,⁴ most reactions require harsh reaction conditions or special equipment, are limited by substrate availability, and result in unsatisfactory deuterium incorporation. Despite these obstacles we succeeded in the synthesis of highly pure deuterium-labeled (R)-K-13675 as an excellent internal stan-

ABSTRACT

Two highly deuterium-labeled compounds, (R)-K-13675- d_{11} and (R)-K-13675- d_7 , were prepared for use as internal standards for low-level quantification of plasma drugs by LC/MS/MS. We successfully demonstrated their utility in pharmacokinetic studies for sensitive and precise drug quantification.

© 2009 Elsevier Ltd. All rights reserved.



Figure 1. Structure of (*R*)-K-13675.

dard for sensitive and precise quantification in pharmacokinetic studies. Herein we report details of synthetic procedures and preliminary pharmacokinetic studies.

2. Results and discussion

Our synthetic strategy towards deuterated (R)-K-13675 was based on the construction of two major precursors, 2-aminophe $nol-d_4(2)$ and 4-methoxyphenol- $d_7(3)$ (Scheme 1). Envisaging that post-synthetic deuteration would be feasible in this synthetic scheme. We planned to synthesize 2 by hydrogen-deuterium (H-D) exchange displacement of 2-aminophenol (5) in deuterium oxide (D_2O) ⁵ while **3** was to be derived from a synthetic combination of the commercially available 1,4-hydroquinone- d_6 (9) and methanol- d_4 .

First, we focused on the H-D exchange reaction of Sajiki et al.,⁶ because it is applicable to a variety of aromatic compounds and its implementation is straightforward. Thus we conducted the H-D exchange reaction of 2-aminophenol (5) using 5% Pt/C (Aldrich, Bach#03505ME) in D₂O (99.9 atom % D; Aldrich) under a hydrogen

Corresponding author. Tel.: +81 42 391 6211; fax: +81 42 395 0312. E-mail address: k-sibuya@kowa.co.jp (K. Shibuya).

^{0968-0896/\$ -} see front matter © 2009 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmc.2009.01.049



Scheme 1. Synthetic strategy of (R)-K-13675- d_{11} .

atmosphere at 100 °C for 24 h. The deuterium content was determined on the basis of acetyl groups by ¹H NMR after diacetylation of 2-aminophenol- d_4 (**2**) (Scheme 2, Table 1). Surprisingly, we could achieve a deuterium content of only 86% D, significantly lower than the values reported in the literature (89% D at 80 °C, 97% D at 180 °C).^{6d} After these unsatisfying results, we decided on a different approach.

We then devised a method for exploiting the kinetic isotope effect,⁷ in which almost full deuterium incorporation can be achieved by multiple cycles of Pt/C-catalyzed deuteration. The ratio of deuterium content was increased markedly in each cycle and reached 97% D by the third cycle. A slight increase was observed from the fourth cycle (fourth cycle, 97% D; fifth cycle, 98% D). As detailed in Table 1, this efficient method let to easily reproducible results even on a 10-g-scale (third cycle, 94% D; fifth cycle, 98% D). In summary, we have evidenced that it is possible to obtain highly deuterated **2** by repeating the Pt/C-catalyzed reaction under a hydrogen atmosphere, without requiring either extremely high temperatures or specialized equipment.



Scheme 2. Synthesis of 2-aminophenol- $d_4(2)$ and the diacetylated 2-aminophenol- $d_4 6$.

Table	1
	-

Doutorium	incorneration	of 2 amino	mbanal d4 (2) -1	ftor II D	ovehange	roaction
Deuterium	IIICOLDOLATIOII	01 Z-dillill	DHEHOI-04 (Z di	$\Pi = D$	excitative	reaction

Reaction cycle	Deuterium content ^b (% D)
1	86
2	91
3	97
4	97
5	98

^a The first reaction involved treatment of **5** (1.0 g) with 5% Pt/C (200 mg) in D₂O (50 mL, 99.9 atom% D) at 100 °C under a hydrogen atmosphere for 24 h. From the second cycle, the reactions were performed with the same ratio of reagents under the same conditions.

^b Deuterium content (% D) of **6** was determined on the basis of the acetyl groups by ¹H NMR after diacetylation of **2**.

Next, we turned our attention to construction of the two key parts of (R)-K-13675- d_{11} (**4**) (Scheme 3).⁸ Treatment of **2** with potassium xanthogenate in ethanol afforded the cyclized 2-mercaptobenzoxazole- d_4 (7), subsequent chlorination of which with thionyl chloride furnished 2-chlorobenzoxazole- d_4 (8) in moderate yield. For the other key part, after several attempts⁹ we achieved the synthesis of 4-methoxyphenol- d_7 (3) by application of Bellas' method.¹⁰ Exposure of **9** to benzoquinone- d_4 and sulfuric acid in methanol- d_4 at room temperature proceeded smoothly to provide the desired product 3 in 95% yield. Thus obtained, 3 was alkylated by Michael reaction with acrylonitrile in the presence of Triton B at 80 °C to give the nitrile 10 (64%), followed by reduction with borane-THF at 80 °C to provide the amine **11** (73%). Reductive alkylation of **11** with 3-hydroxybenzaldehyde (**12**) and sodium borohydride yielded the secondary amine **13** (90%). Interestingly, the ratio of deuterium content was unaffected in a series of reductive reaction conditions. The seven-deuterium-labeled amine 13 was reacted with 8 to afford 14 in 83% yield. Etherification of the labeled phenol 14 with *n*-butyl (S)-2-trifluoromethanesulfonyloxybutanoate in the presence of K₂CO₃ in acetonitrile at room temperature gave the phenyl ether 15 in 97% yield. Finally, hydrolysis of **15** in aqueous NaOH provided (*R*)-K-13675-*d*₁₁ (**4**) in 72% yield.

Analogously (*R*)-K-13675- d_7 (**18**) was prepared from the nondeuterated 2-chlorobenzoxazole following the same synthetic operation (Scheme 4).

The deuterium-labeled compounds **4** and **18** were then analyzed by monitoring with LC/MS/MS.¹¹ The multiple reaction monitoring (MRM) ion chromatograms of (*R*)-K-13675- d_7 (**18**) and (*R*)-K-13675- d_{11} (**4**) are shown in Figure 2a and b. When both compounds were measured after injection of 5 ng, **18** was observed to contain a small amount (0.19%) of non-deuterated (*R*)-K-13675, and **4** showed a trace amount (0.02%) of (*R*)-K-13675, as summarized in Table 2.

For quantification of (*R*)-K-13675 (**1**) in rat plasma using isotope dilution mass spectrometry, a calibration curve for **1** was constructed by spiking 100 µL of rat plasma with 0.25 ng of (*R*)-K-13675- d_{11} (**4**) and 0.02, 0.04, 0.1, 0.2, 0.4, 1, 2, or 4 ng of **1**, respectively (Fig. 3 and Table 3).¹² The calibration curve showed excellent linearity (accuracy: 97.4–102%) and the correlation coefficient was 0.9999. Interestingly, compounds labeled with more deuterium were shown to be superior to those with less deuterium for validating low-level quantifications of plasma drugs by LC/MS/MS. Indeed, we carried out the quantifications of drugs in plasma using (*R*)-K-13675- d_{11} (**4**) as an internal standard administered to rats and dogs at a dose of 0.1 mg/kg. The results indicated that we have successfully established a method to determine low-level concentrations (0.1 ng/mL) of drugs in plasma in each species. Details of these pharmacokinetic studies will be reported elsewhere in due time.

3. Conclusions

We have developed a useful method to obtain 2-aminophenol d_4 with excellent deuterium incorporation by repetitive cycles of H–D exchange, and have succeeded in the synthesis of highly deuterated (*R*)-K-13675- d_{11} and (*R*)-K-13675- d_7 . Furthermore, we demonstrated that (*R*)-K-13675- d_{11} served as an excellent internal standard for sensitive and precise quantification in pharmacokinetic studies.

4. Experimental

4.1. General

Commercially available reagents and solvents were used without further purification. TLC analyses were carried out on silica gel 60 F₂₅₄ plates (Merck). ¹H NMR and ¹³C NMR spectra were re-



Scheme 3. Reagents and conditions: (a) potassium xanthogenate, EtOH, 55%; (b) SOCl₂, DMF, 49%; (c) benzoquinone-*d*₄, H₂SO₄, methanol-*d*₄, 95%; (d) acrylonitrile, Triton B, 64%; (e) BH₃–THF, THF, 73%; (f) **11**, NaBH₄, MeOH, 90%; (g) **8**, ^{*i*}Pr₂NEt, DMF, 83%; (h) *n*-butyl 2-(*S*)-trifluoromethanesulfonyloxybutanoate, K₂CO₃, MeCN, 97%; (i) 4 M aq NaOH, EtOH, 72%.



Scheme 4. Reagents and conditions: (j) 2-chlorobenzoxazole, ⁱPr₂NEt, DMF, 87%; (k) benzyl (*S*)-2-trifluoromethanesulfonyloxybutanoate, K₂CO₃, MeCN, 100%; (l) 4 M aq NaOH, EtOH, 85%.

corded on a JEOL JNM-LA 400 MHz and JEOL GSX-270. Tetramethylsilane was used as an internal standard. Chemical shifts (δ) are given in parts per million (ppm), coupling constants *J* values are given in Hertz (Hz) and are reported to the nearest 0.1 Hz. Starred (^{*}) values in the ¹³C NMR are for small peaks. Infrared (IR) spectra were recorded on a Thermo Nicolet 370 FT-IR (ATR) spectrometer. Mass spectra were obtained on a JEOL MS-BU20 mass spectrometer. Elemental analyses (C, H, N) were performed by Yanaco MT-5. Melting points were determined in open glass capillaries on a Buchi B-545 melting point apparatus.

4.2. Synthesis of 2-aminophenol- d_4 (2) (large scale)

T o a suspension of 2-aminophenol (**5**) (10.0 g, 91.6 mmol) in D₂O (70 mL, 99.9 atom % D; Aldrich) was added 5% Pt/C (2.0 g, Aldrich, Bach; #03505ME) under an argon atmosphere. The reaction mixture was stirred at 100 °C under a hydrogen atmosphere for 24 h and diluted with EtOAc. The mixture was filtered through a pad of Celite. The organic layer was washed with brine, dried over Na₂SO₄ and concentrated in vacuo to give **2** as a brown solid (8.77 g). This obtained product was used in the next reaction without purification. The second reaction was performed with the same ratio of reagents under the same conditions. Finally, the desired product **2** was obtained after the fifth reaction cycle (5.57 g, 54%): ¹³C NMR (68 MHz, DMSO-*d*₆) δ 113.71^{*}, 114.20^{*}, 118.63^{*}, 119.25^{*}, 136.41, 143.90; IR (solid sample): 3376, 3351, 3328, 3305, 1578, 1438, 1300 cm⁻¹; HRMS (EI): *m/z* [M⁺] calcd for C₆H₃D₄NO: 113.0775; found: 113.0773.

4.3. Synthesis of the diacetylated 2-aminophenol- d_4 6. Determination of deuterium content (% D)

To a solution of 2-aminophenol- d_4 (**2**) (295 mg, 2.61 mmol) and triethylamine (527 mg, 5.21 mmol) in EtOAc (10 mL) was added acetyl chloride (409 mg, 5.21 mmol) at 0 °C. The reaction mixture was stirred at room temperature for 1 h and quenched with water. The organic layer was washed with water and brine, dried over



Figure 2. MRM ion chromatograms of the deuterated and non-deuterated peaks.

Table 2Peak area and ratio in MRM ion chromatograms.¹¹

Compounds	Peak are	a ^a	Ratio ^b (%)
	Deuterated	Non-deuterated	
4 (<i>d</i> ₁₁)	71,200,000 ± 1,100,000	16,900 ± 800	0.02 ± 0.00
18 (<i>d</i> ₇)	88,800,000 ± 7,000,000	167,000 ± 7000	0.19 ± 0.01

^a Values are means \pm S.D. (n = 10).

^b Ratio of non-deuterated compound (%) = (peak area of non-deuterated compound/ deuterated compound) \times 100.



Figure 3. Calibration curve for (*R*)-K-13675 (**1**) in rat plasma.

 Na_2SO_4 and concentrated in vacuo. The residue was purified by preparative TLC (*n*-hexane/EtOAc = 1:2) to give **6** as a yellow solid

Table 3	
Accuracy of (R)-K-13675 (1) for calibration curve samples in rat plasma	

	1	2	3	4	5	6	7	8
Concentration (ng/mL)	0.2	0.4	1	2	4	10	20	40
Accuracy ^a (%)	99.8	99.6	102	100	99.7	101	101	97.

^a Accuracy (%) = (calculated concentration/nominal concentration) \times 100.12.

(131 mg, 26%): ¹H NMR (400 MHz, CD₃OD) δ 2.13 (s, 3H), 2.30 (s, 3H), 7.12 (s, 0.02H), 7.18 (s, 0.02H), 7.21 (s, 0.02H), 7.74 (s, 0.02H); Deuterium content on aromatic ring = 98% D; Anal. Calcd for C₁₀H₇D₄NO₃: C, 60.90; H, 3.58; D, 4.08; N, 7.10. Found: C, 60.92; H, 3.64; D, 4.04; N, 7.12; IR (solid sample) 3334, 1736, 1686, 1588, 1509, 1439, 1413 cm⁻¹; HRMS (EI): *m/z* [M⁺] calcd for C₁₀H₇D₄NO₃: 197.09858; found: 197.09783; mp 123–124 °C.

4.4. Synthesis of 2-mercaptobenzoxazole-d₄ (7)

To a solution of 2-aminophenol- d_4 (2) (5.15 g, 45.5 mmol) in ethanol (100 mL) was added potassium xanthogenate (8.03 g, 50.1 mmol) at room temperature. The reaction mixture was stirred at 100 °C for 5 h and cooled to room temperature. Solvent was evaporated under reduced pressure. The residue was dissolved in water. The aqueous solution was acidified with 4 M HCl and extracted with EtOAc. The organic layer was washed with water and brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was recrystallized from EtOAc/*n*-hexane to give **7** as a redbrown crystal (2.94 g, 42%). The filtrate was concentrated in vacuo. The residue was purified by silica gel column chromatography (*n*hexane/EtOAc = 4:1) to give **7** as a red-brown crystal (933 mg, 13%. Total yield; 3.87 g, 55%): ¹H NMR (400 MHz, DMSO- d_6) δ 7.03 (s, 0.07H), 7.25 (s, 0.07H), 7.30 (s, 0.07H), 7.51 (s, 0.07H), 13.86 (br s, 1H); ¹³C NMR (100 MHz, DMSO- d_6) δ 109.88^{*}, 110.33^{*}, 123.56^{*}, 124.92^{*}, 131.08, 148.07, 180.12; Anal. Calcd for C₇HD₄NOS: C, 54.17; H, 0.65; D, 5.19; N, 9.02. Found: C, 54.33; H, 0.68; D, 5.29; N, 8.91; IR (solid sample): 3341, 1599, 1476, 1368, 1327, 1112, 932 cm⁻¹; MS (EI) *m/z* 155 [M⁺]; mp 186–189 °C.

4.5. Synthesis of 2-chlorobenzoxazole-d₄ (8)

To a solution of 2-mercaptobenzoxazole- d_4 (**7**) (2.00 g, 12.9 mmol) in thionyl chloride (4 mL) was added *N*,*N*-dimethyl-formamide (940 mg, 12.9 mmol) dropwise over 5 min at 0 °C under an argon atmosphere. The reaction mixture was stirred at same temperature for 2 h and concentrated in vacuo. The residue was dissolved in EtOAc. Water was added. The organic layer was washed with water and brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by silica gel column chromatog-raphy (*n*-hexane/EtOAc = 10:1) to give **8** as a yellow oil (999 mg, 49%): ¹³C NMR (68 MHz, CDCl₃) δ 110.11^{*}, 119.39^{*}, 124.07^{*}, 125.11^{*}, 143.73, 150.78, 151.39; Anal. Calcd for C₇D₄ClNO: C, 53.35; D, 5.11; N, 8.89. Found: C, 53.21; D, 5.39; N, 8.77; IR (neat): 1783, 1584, 1523, 1436, 1370, 1208, 1113 cm⁻¹; MS (EI) *m/z* 157 [M⁺], 159 [M⁺+2].

4.6. Synthesis of 4-methoxyphenol-d₇ (3)

To a solution of 1,4-hydroquinone- d_6 (**9**) (98 atom% D, 4.48 g, 38.6 mmol) and benzoquinone- d_4 (98 atom% D, 325 mg, 2.90 mmol) in methanol- d_4 (99.8 atom % D, 25 mL) was added sulfuric acid (4.80 g) over 5 min at room temperature. The reaction mixture was stirred for 14 h, poured into ice-water and neutralized with 4 M aqueous NaOH. The aqueous solution was extracted with Et₂O. The organic layer was washed with brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by silica gel column chromatography (*n*-hexane/EtOAc = 5:1) to give **3** as a pale yellow solid (4.83 g, 95%): ¹H NMR (400 MHz, CDCl₃) δ 3.72–3.76 (m, 0.06H), 4.87 (s, 1H), 6.76 (s, 0.10H), 6.79 (s, 0.10H); ¹³C NMR (100 MHz, CDCl₃) δ 55.06^{*}, 114.59^{*}, 115.80^{*}, 149.41, 153.48; IR (solid sample): 3381, 2521, 2262, 2069, 1419, 1141, 1112 cm⁻¹; HRMS (EI): *m/z* [M⁺] calcd for C₇HD₇O₂: 131.09564; found: 131.09642; mp 56–57 °C.

4.7. Synthesis of 3-(4-methoxyphenoxy-*d*₇)propionitrile (10)

To acrylonitrile (3.50 g, 65.3 mmol) were added 4-methoxyphenol- d_7 (**3**) (4.28 g, 32.7 mmol) and Triton B (0.3 mL) at room temperature. The reaction mixture was stirred at 80 °C under an argon atmosphere for 22 h and diluted with EtOAc and water. The organic layer was washed with 1 M aqueous NaOH, water and brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was recrystallized from EtOAc/*n*-heptane to give **10** as colorless needles (3.87 g, 64%): ¹H NMR (400 MHz, CDCl₃) δ 2.79 (t, *J* = 6.4 Hz, 2H), 3.72–3.75 (m, 0.01H), 4.13 (t, *J* = 6.4 Hz, 2H), 6.84 (s, 0.03H), 6.86 (s, 0.03H); deuterium content = 99% D; ¹³C NMR (100 MHz, CDCl₃) δ 18.63, 54.87^{*}, 63.55, 114.38^{*}, 115.66^{*}, 117.35, 151.66, 154.55; Anal. Calcd for C₁₀H₄D₇NO₂: C, 65.19; H, 2.19; D, 7.65; N, 7.60. Found: C, 65.09; H, 2.22; D, 7.54; N, 7.54; IR (solid sample): 2930, 2886, 2255, 2219, 2069, 1446, 1420 cm⁻¹; MS (EI) *m/z* 184 [M⁺]; mp 65–66 °C.

4.8. Synthesis of 3-(4-methoxyphenoxy-d₇)propylamine (11)

To a solution of **10** (3.82 g, 20.7 mol) in THF (20 mL) was added BH_3 -THF (1.17 mol/L, 21.3 mL, 24.9 mmol) dropwise at room temperature under an argon atmosphere. The reaction mixture was stirred at 80 °C for 3 h and cooled to room temperature. Methanol

(50 mL) was added slowly to the mixture at 0 °C. The mixture was stirred at 80 °C for 0.5 h, acidified with 4 M HCl until pH 1 at 0 °C and stirred at 80 °C for 14 h. Solvent was evaporated under reduced pressure. The residue was dissolved in water. The aqueous solution was washed with Et₂O. The aqueous layer was alkalinized with 4 M aqueous NaOH and extracted with toluene. The organic layer was washed with water and brine, dried over Na₂SO₄ and concentrated in vacuo to give **11** as a colorless solid (2.87 g, 73%): ¹H NMR (400 MHz, CDCl₃) δ 1.33 (br s, 2H), 1.90 (quintet, J = 6.1 Hz, 2H), 2.90 (t, J = 6.1 Hz, 2H), 3.60-3.64 (m, 0.01H), 4.00 (t, J = 6.1 Hz, 2H), 6.82 (s, 0.03H), 6.83 (s, 0.03H); ¹³C NMR (100 MHz, CD₃OD) δ 33.48, 39.82, 55.24^{*}, 67.64, 115.25^{*}, 116.08^{*}, 154.37, 155.20; IR (solid sample): 2937, 2872, 2216, 2070, 1600, 1426, 1378 cm⁻¹; HRMS (EI): m/z [M⁺] calcd for C₁₀H₈D₇NO₂: 188.15349; found: 188.15294.

4.9. Synthesis of 3-[[[3-(4-methoxyphenoxy-*d*₇)propyl] amino]methyl]phenol (13)

To a solution of **11** (1.88 g, 10.0 mmol) in MeOH (25 mL) was added 3-hydroxybenzaldehyde (1.22 g, 10 mmol) at room temperature. The reaction mixture was stirred at 60 °C for 3 h. A solution of NaBH₄ (567 mg, 15.0 mmol) in water (10 mL) was added slowly to this mixture at 0 °C. The mixture was stirred at room temperature for 14 h and filtered off. The isolated solid was rinsed with water and dried in vacuo to give **13** as a colorless crystal (2.66 g, 90%): ¹H NMR (400 MHz, DMSO- d_6) δ 1.82 (quintet, J = 6.8 Hz, 2H), 2.06 (br s, 1H), 2.60 (t, J = 6.8 Hz, 2H), 3.60 (s, 2H), 3.94 (t, J = 6.8 Hz, 2H), 6.59 (dd, J = 7.8, 2.2 Hz, 1H), 6.72 (d, J = 7.8 Hz, 1H), 6.74 (s, 1H), 7.07 (t, J = 7.8 Hz, 1H), 9.24 (s, 1H); ¹³C NMR $(100 \text{ MHz}, \text{DMSO-}d_6) \delta 29.33, 45.50, 53.08, 54.43^*, 66.37, 113.38,$ 114.72, 114.15^{*}, 114.89^{*}, 118.44, 128.92, 142.51, 152.57, 153.10, 157.26; Anal. Calcd for C₁₇H₁₄D₇NO₃: C, 69.36; H, 4.79; D, 4.79; N, 4.76. Found: C, 69.34; H, 4.90; D, 4.76; N, 4.75; IR (solid sample): 3275, 1580, 1480, 1431, 1390, 1156, 1109 cm⁻¹; MS (EI) *m/z* 294 [M⁺]; mp 142–143 °C.

4.10. Synthesis of 3-[[benzoxazol-2-yl-d₄[3-(4-methoxy phenoxy-d₇)propyl]amino]methyl]phenol (14)

To a solution of 13 (1.87 g, 6.34 mmol) in DMF (15 mL) and N,N-diisopropylethylamine (983 mg, 7.61 mmol) was added 8 (999 mg, 6.34 mmol) at room temperature. The reaction mixture was stirred at 80 °C for 15 h and diluted with EtOAc and water. The organic layer was washed with water and brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by silica gel column chromatography (*n*-hexane/EtOAc = 2:1) to give 14 as an amorphous brown solid (2.19 g, 83%): ¹H NMR (400 MHz, CDCl₃) δ 2.03 (quintet, J = 6.8 Hz, 2H), 3.45 (t, J = 6.8 Hz, 2H), 3.89 (t, J = 6.8 Hz, 2H), 4.65 (s, 2H), 6.70–6.73 (m, 2H), 6.75 (s, 1H), 7.14 (t, J = 7.8 Hz, 1H), 8.75 (s, 1H); ¹³C NMR (68 MHz, CDCl₃) δ 27.62, 45.01, 52.07, 54.89, 65.32, 108.84, 113.12, 114.25[°], 114.81[°], 115.08[°], 115.33, 118.77, 120.22^{*}, 123.53, 129.79, 137.60, 141.40, 148.07, 152.61, 153.77, 157.82, 162.28; Anal. Calcd for $C_{24}H_{13}D_{11}N_2O_4$: C, 69.37; H, 3.15; D, 5.33; N, 6.74. Found: C, 69.47; H, 3.24; D, 5.32; N, 6.75; IR (solid sample): 2941, 1633, 1567, 1424, 1393, 1153, 1111 cm⁻¹; HRMS (EI): m/z [M⁺] calcd for C₂₄H₁₃D₁₁N₂O₄: 415.24150; found: 415.24342.

4.11. Synthesis of *n*-butyl (*R*)-2-[3-[[benzoxazol-2-yl-*d*₄ [3-(4-methoxyphenoxy-*d*₇)propyl]amino]methyl]phenoxy] butanoate (15)

To a solution of **14** (2.16 g, 5.20 mmol) and K_2CO_3 (1.08 g, 7.79 mmol) in MeCN (30 mL) was added a solution of *n*-butyl

(S)-2-trifluoromethanesulfonyloxybutanoate (1.97 mg. 6.76 mmol) in MeCN (10 mL) under an argon atmosphere. The reaction mixture was stirred at room temperature for 19 h and filtered off. The filtrate was concentrated in vacuo. The residue was dissolved in EtOAc. Water was added. The organic layer was washed with water and brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by silica gel column chromatography (n-hexane/EtOAc = 5:1) to give **15** as a brown oil (2.82 g, 97%): ¹H NMR (400 MHz, CDCl₃) δ 0.85 (t, J = 7.3 Hz, 3H), 1.05 (t, J = 7.1 Hz, 3H), 1.22–1.28 (m, 2H), 1.49–1.55 (m, 2H), 1.96 (quintet, J = 6.7 Hz, 2H), 2.14 (quintet, J = 6.8 Hz, 2H), 3.70 (t, J = 6.7 Hz, 2H), 3.96 (t, J = 6.7 Hz, 2H), 4.00-4.14 (m, 2H), 4.53 (t, J = 6.5 Hz, 1H), 4.72 (d, J = 15.6 Hz, 1H), 4.77 (d, J = 15.6 Hz, 1H), 6.76 (dd, J = 7.8, 2.2 Hz,1H), 6.86 (s, 1H), 6.89 (d, J = 7.8 Hz, 1H), 7.21 (t, J = 7.8 Hz, 1H); ¹³C NMR (68 MHz, $CDCl_3$ δ 9.67, 13.59, 18.92, 26.17, 27.51, 30.48, 45.11, 52.06, $55.19^{*},\ 64.96,\ 65.50,\ 77.60,\ 108.64^{*},\ 113.73,\ 113.86^{*},\ 114.66,\\ 115.32^{*},\ 115.78^{*},\ 120.15^{*},\ 120.68,\ 123.44^{*},\ 129.82,\ 138.67,$ 143.41, 148.56, 152.69, 153.76, 158.31, 162.15, 171.67; Anal. Calcd for C₃₂H₂₇D₁₁N₂O₆: C, 68.91; H, 4.88; D, 3.97; N, 5.02. Found: C, 68.80; H, 4.97; D, 3.94; N, 4.94; IR (neat): 2960, 2067, 1751, 1633, 1563, 1425, 1154 cm⁻¹; HRMS (EI): *m/z* [M⁺] calcd for $C_{32}H_{27}D_{11}N_2O_6$: 557.34087; found: 557.34196; $[\alpha]_D^{20}$ +19.1 (c 0.89, CHCl₃).

4.12. Synthesis of (*R*)-2-[3-[[benzoxazol-2-yl-*d*₄ [3-(4-methoxyphenoxy-*d*₇)propyl]amino]methyl]phenoxy] butanoic acid; (*R*)-K-13675-*d*₁₁ (4)

To a solution of 15 (2.70 g, 4.84 mmol) in EtOH (20 mL) was added 4 M aqueous NaOH (2.4 mL) at 0 °C. The reaction mixture was stirred at room temperature for 1 h, diluted with water, acidified with 4 M HCl and extracted with EtOAc. The organic layer was washed with water and brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by silica gel column chromatography (CHCl₃/MeOH = 10:1) and recrystallized from EtOAc/n-heptane to give (*R*)-K-13675 d_{11} as a colorless crystal (1.74 g, 72%): ¹H NMR (400 MHz, DMSO- d_6) δ 0.96 (t, J = 7.1 Hz, 3H), 1.79-1.89 (m, 2H), 2.06 (quintet, J = 6.8 Hz, 2H), 3.65 (t, J = 6.8 Hz, 2H), 3.94 (t, J = 6.8 Hz, 2H), 4.60 (t, J = 6.4 Hz, 1H), 4.72 (s, 2H), 6.78 (dd, *I* = 7.8, 2.4 Hz, 1H), 6.86 (s, 1H), 6.89 (d, *I* = 7.8 Hz, 1H), 7.25 (t, I = 7.8 Hz, 1H), 13.1 (br s, 1H); ¹³C NMR (100 MHz, DMSO- d_6) δ 9.53, 25.42, 27.12, 45.29, 51.27, 65.47, 76.43, 113.30, 114.13, 119.77, 129.76, 139.02, 143.19, 148.43, 152.31, 153.25, 158.04, 162.24, 172.46; Anal. Calcd for C₂₈H₁₉D₁₁N₂O₆: C, 67.04; H, 3.82; D, 4.42; N, 5.58. Found: C, 66.91; H, 3.86; D, 4.34; N, 5.50; IR (solid sample): 2959, 2939, 2887, 1716, 1628, 1426, 1377 cm⁻¹; MS (FAB) m/z 502 [M⁺+1]; mp 95–96 °C; $[\alpha]_{D}^{24}$ +17.1 (*c* 0.67, MeOH).

4.13. Synthesis of 3-[[benzoxazol-2-yl]3-(4-methoxyphenoxy*d*₇)propyl]amino]methyl]phenol (16)

To a solution of **13** (6.22 g, 21.1 mmol) in DMF (60 mL) and *N*,*N*diisopropylethylamine (2.73 g, 21.1 mmol) was added 2-chlorobenzoxazole (3.24 g, 21.1 mmol) at room temperature. The reaction mixture was stirred at 80 °C for 14 h and diluted with EtOAc and water. The organic layer was washed with water and brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by silica gel column chromatography (*n*-hexane/ EtOAc = 2:1) to give a yellow oil. The oil was recrystallized from *tert*-butyl methyl ether to give **16** as a colorless crystal (7.56 g, 87%): ¹H NMR (400 MHz, DMSO-*d*₆) δ 2.06 (quintet, *J* = 6.8 Hz, 2H), 3.65 (t, *J* = 6.8 Hz, 2H), 3.94 (t, *J* = 6.8 Hz, 2H), 4.70 (s, 2H), 6.70 (dd, *J* = 7.8, 1.7 Hz, 1H), 6.69-6.76 (m, 2H), 7.00 (td, *J* = 7.8, 1.2 Hz, 1H), 7.15 (t, *J* = 7.8 Hz, 2H), 7.30 (d, *J* = 7.8 Hz, 1H), 7.36 (d, *J* = 7.8 Hz, 1H), 9.43 (s, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 27.13, 45.22, 51.31, 54.45^{*}, 65.50, 108.81, 113.92, 114.16^{*}, 114.42, 115.04^{*}, 115.59, 117.90, 120.13, 123.88, 129.64, 138.73, 143.31, 148.46, 152.30, 153.27, 157.63, 162.26; Anal. Calcd for $C_{24}H_{17}D_7N_2O_4$: C, 70.05; H, 4.16; D, 3.43; N, 6.81. Found: C, 70.05; H, 4.27; D, 3.42; N, 6.77; IR (solid sample): 2937, 1638, 1600, 1582, 1459, 1424, 1394 cm⁻¹; MS (EI) *m/z* 411 [M⁺]; mp 100–101 °C.

4.14. Synthesis of benzyl (*R*)-2-[3-[[benzoxazol-2-yl-*d*₄ [3-(4-methoxyphenoxy-*d*₇)propyl]amino]methyl]phenoxy] butanoate (17)

To a solution of **16** (4.59 g, 11.2 mmol) and K_2CO_3 (2.31 g, 16.7 mmol) in MeCN (60 mL) was added a solution of benzyl (S)-2-trifluoromethanesulfonyloxybutanoate (4.00 g, 12.3 mmol) under an argon atmosphere. The reaction mixture was stirred at room temperature for 11 h and filtered off. The filtrate was concentrated in vacuo. The residue was dissolved in EtOAc. Water was added. The organic layer was washed with water and brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by silica gel column chromatography (n-hexane/EtOAc = 5:1) to give 17 as a colorless oil (6.97 g, 100%): ¹H NMR (400 MHz, CDCl₃) δ 1.02 (t, J = 7.2 Hz, 3H), 1.96 (quintet, J = 7.1 Hz, 2H), 2.13 (quintet, *J* = 6.5 Hz, 2H), 3.68 (t, *J* = 6.5 Hz, 2H), 3.95 (t, *J* = 6.5 Hz, 2H), 4.57 (t, J = 6.7 Hz, 1H), 4.69 (d, J = 15.9 Hz, 1H), 4.74 (d, J = 15.9 Hz, 1H)1H), 5.06 (d, J = 12.2 Hz, 1H), 5.15 (d, J = 12.2 Hz, 1H), 6.74 (dd, J = 7.8, 2.4 Hz, 1H), 6.84 (s, 1H), 6.89 (d, J = 7.8 Hz, 1H), 7.00 (d, J = 7.8 Hz, 1H), 7.14-7.30 (m, 8H), 7.37 (d, J = 7.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 9.52, 26.03, 27.27, 45.12, 52.00, 54.48, 65.47, 66.64, 77.48, 108.69, 113.83, 114.19[°], 114.61, 115.23[°], 116.10, 120.32, 120.66, 123.88, 128.15, 128.27, 128.45, 129.78, 135.30, 138.65, 143.43, 148.85, 152.63, 153.72, 158.17, 162.56, 171.30; Anal. Calcd for $C_{35}H_{29}D_7N_2O_6$: C, 71.53; H, 4.97; D, 2.40; N, 4.77. Found: C, 71.42; H, 5.11; D, 2.40; N, 4.75; IR (neat): 2938, 1752, 1638, 1459, 1578, 1425, 1153 cm⁻¹; HRMS (EI): *m/z* $[M^+]$ calcd for C₃₅H₂₉D₇N₂O₆: 587.30052; found: 587.30184; $[\alpha]_{p}^{20}$ +16.4 (c 1.00, CHCl₃).

4.15. Synthesis of (*R*)-2-[3-[[benzoxazol-2-yl [3-(4-methoxy phenoxy-*d*₇)propyl]amino]methyl]phenoxy]butanoic acid; (*R*)-K-13675-*d*₇ (18)

To a solution of 17 (6.30 g, 10.7 mmol) in EtOH (60 mL) was added 4 M aqueous NaOH (5.4 mL) at 0 °C. The reaction mixture was stirred at room temperature for 2 h and diluted with water. The aqueous solution was washed with Et₂O, acidified with *c*HCl at 0 °C and extracted with EtOAc. The organic layer was washed with water and brine, dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by silica gel column chromatography (CHCl₃/MeOH = 10:1) and recrystallized from EtOAc-nheptane to give (*R*)-K-13675- d_7 as colorless needles (4.55 g, 85%): ¹H NMR (400 MHz, DMSO- d_6) δ 0.98 (t, J = 7.0 Hz, 3H), 1.83-1.92 (m, 2H), 2.07 (quintet, J = 6.5 Hz, 2H), 3.67 (t, J = 6.5 Hz, 2H), 3.95 (t, J = 6.5 Hz, 2H), 4.64 (t, J = 6.3 Hz, 1H), 4.74 (s, 2H), 6.80 (dd, J = 7.8, 2.0 Hz, 1H), 6.90 (s, 1H), 6.91 (d, J = 7.8 Hz, 1H), 7.00 (t, J = 7.8 Hz, 1H), 7.15 (t, J = 7.8 Hz, 1H), 7.26 (t, J = 7.8 Hz, 1H), 7.31 (d, J = 7.8 Hz, 1H), 7.36 (d, J = 7.8 Hz, 1H), 13.1 (br s, 1H); ¹³C-NMR (100 MHz, DMSO- d_6) δ 9.46, 25.41, 27.14, 45.29, 51.30, 54.45, 65.49, 76.41, 108.85, 113.36, 114.17, 114.18, 115.05, 115.67, 119.85, 120.18, 123.89, 129.74, 139.01, 143.29, 148.51, 152.34, 153.29, 158.04, 162.25, 172.39; Anal. Calcd for C₂₈H₂₃D₇N₂O₆: C, 67.59; H, 4.66; D, 2.83; N, 5.63. Found: C, 67.66; H, 4.77; D, 2.82; N, 5.54; IR (solid sample): 2967, 2935, 1725, 1635, 1584, 1426, 1382 cm⁻¹; MS (FAB) m/z 498 [M⁺+1]; mp 91–93 °C; $[\alpha]_{D}^{24}$ +16.5 (c 0.62, MeOH).

Acknowledgments

We are grateful to Professor H. Sajiki of Gifu Pharmaceutical University for helpful advice. We also wish to thank Dr. David Ricketts (Discovery Business Development) and Dr. Karsten Marx (Polyphor) for their help in preparing the manuscript.

References and notes

- (a) Staels, B.; Auwerx, J. Curr. Pharm. Des. 1997, 3, 1–14; (b) Staels, B.; Dallongeville, J.; Auwerx, J.; Schoonjans, K.; Leitersdorf, E.; Fruchart, J. C. Circulation 1998, 89, 2088–2093; (c) Vakkilainen, J.; Steiner, G.; Ansquer, J. C.; Aubin, F.; Rattier, S.; Foucher, C.; Hamsten, A.; Taskinen, M. R. Circulation 2003, 107, 1733–1737. on behalf of the DAIS Group; (d) Robins, S. J.; Bloomfield, H. E. Curr. Opin. Lipidol. 2006, 17, 431–439; (e) Fruchart, J. C.; Duriez, P.; Staels, B. Curr. Opin. Lipidol. 1999, 10, 245–258; (f) Staels, B.; Koenig, W.; Habib, A.; Merval, R.; Lebret, M. Y.; Torra, I. P.; Delerive, P.; Fadel, A.; Chinetti, G.; Fruchart, J. C.; Najib, J.; Maclouf, J.; Tedgui, A. Nature 1998, 393, 790–793; (g) Marx, N.; Sukhova, G. K.; Collins, T.; Libby, P.; Plutzky, J. Circulation 1999, 99, 3125–3131; (h) Willson, T. M.; Brown, P. J.; Sternbach, D. D.; Henke, B. R. J. Med. Chem. 2000, 43, 527–550; (i) Staels, B. Cell Metab. 2005, 2, 77–78; (j) Vosper, H.; Patel, L.; Graham, T. L.; Khoudoli, G. A.; Hill, A.; Macphee, C. H.; Pinto, I.; Smith, S. A.; Sukklova, E. K.; E; Wolf, C. R.; Palmer, C. N. A. J. Biol. Chem. 2001, 276(.), 44258– 44265.
- Yamazaki, Y.; Abe, K.; Toma, T.; Nishikawa, M.; Ozawa, H.; Okuda, A.; Araki, T.; Oda, S.; Inoue, K.; Shibuya, K.; Staels, B.; Fruchart, J. C. *Bioorg. Med. Chem. Lett.* 2007, 17, 4689–4693.
- (a) Tarui, N.; Ikeura, Y.; Natsugari, H.; Nakahama, K. J. Label. Comp. Radiopharm.
 2001, 44, 865–870; (b) Walsky, R. L.; Obach, R. S. Drug Metab. Dispos. 2004, 32, 647–660; (c) Grasso, P.; Benfenate, E.; Terreni, M.; Pregnolato, M.; Natangelo, M.; Pagani, G. J. Chromatogr., A 1998, 822, 91–99; (d) Wille, S. M. R; Hee, P. V.; Neels, H. M.; Van Peteghem, C. H.; Lambert, W. E. J. Chromatogr., A 2007, 1176, 236–245; (e) Bullen, W. W.; Miller, R. A.; Hayes, R. N. J. Am. Soc. Mass Spectrom.
 1999, 10, 55–66; (f) Edwards, S. R.; Smith, M. T. J. Chromatogr., B 2007, 848, 264–270.
- (a) Junk, T.; Catallo, W. J. Chem. Soc. Rev. 1997, 26, 401–406; (b) Garnett, J. L.; Hodges, R. J. J. Am. Chem. Soc. 1967, 89, 4546–4547; (c) Ofosu-Asante, K.; Stock, L. M. J. Org. Chem. 1986, 51, 5452–5454; (d) Fodor-Csorba, K.; Galli, G.; Holly, S.; Gacs-Baitz, E. Tetrahedron Lett. 2002, 43, 3789–3792; (e) Heys, R. J. Chem. Soc. Chem. Commun. 1992, 680–681; (f) Lenges, C. P.; White, P. S.; Brookhart, M. J. Am. Chem. Soc. 1999, 121, 4385–4396; (g) Klei, S. R.; Golden, J. T.; Tilley, T. D.; Bergman, R. G. J. Am. Chem. Soc. 2002, 124, 2092–2093; (h) Weil, T. A. J. Org. Chem. 1974, 39, 48–50; (i) Maeda, M.; Ogawa, O.; Kawazoe, Y. Chem. Pharm. Bull. 1977, 25, 3329–3333; (j) Jones, J. R.; Lockley, W. J. S.; Lu, S. Y.; Thompson, S. P. Tetrahedron Lett. 2001, 42, 331–332; (k) Junk, T.; Catallo, W. J. Tetrahedron Lett. 1996, 37, 3445–3448; (l) Yamamoto, M.; Yokota, Y.; Oshima, K.; Matsubara, S. Chem. Commun. 2004, 1714–1715; (m) Matsubara, S.; Yokota, Y.; Oshima, K. Chem. Lett. 2001, 33, 294–295; (n) Hardacre, C.; Holbrey, J. D.; McMath, S. E. J. Chem. Commun. 2013, 367–368; (o) Vaidyanathan, S.; Surber, B. W. Tetrahedron Lett. 2005, 46, 5195–5197; (p) Atzrodt, J.; Derdau, V.; Fey, T.; Zimmermann, J. Angew. Chem., Int. Ed. 2007, 46, 7744–7765.
- 5. Vaidyanathan et al. reported a simple and efficient method for post-synthesis of 4-aminophenol-d₄ by microwave irradiation in a CEM microwave oven at 140 W and 175 °C for 20 min in the presence of 35% deuterium chloride in

 D_2O .⁴⁰ We attempted to apply this method for preparation of 2-aminophenold₄ (2) under the same reaction conditions except using a Biotage microwave oven. Unfortunately, satisfactory results could not be obtained due to the mechanistic differences in the microwave oven. The maximum pressure of the Biotage microwave oven was set as 22 bar. When the deuteration reaction was carried out at 140 W and 175 °C, the reaction stopped within 2 min because the pressure exceeded 22 bar.

- (a) Sajiki, H.; Hattori, K.; Aoki, F.; Yasunaga, K.; Hirota, K. Synlett 2002, 1149–1151; (b) Sajiki, H.; Aoki, F.; Esaki, H.; Maegawa, T.; Hirota, K. Org. Lett. 2004, 6, 1485–1487; (c) Sajiki, H.; Esaki, H.; Aoki, F.; Maegawa, T.; Hirota, K. Synlett 2005, 1385–1388; (d) Sajiki, H.; Ito, N.; Esaki, H.; Maegawa, T.; Maegawa, T.; Hirota, K. Tetrahedron Lett. 2005, 46, 6995–6998; (e) Maegawa, T.; Akashi, A.; Esaki, H.; Aoki, F.; Sajiki, H.; Hirota, K. Synlett 2005, 845–847; (f) Ito, N.; Watahiki, T.; Maesawa, T.; Maegawa, T.; Sajiki, H. dv. Synth. Catal. 2006, 348, 1025–1028; (g) Ito, N.; Watahiki, T.; Maesawa, T.; Sajiki, H. dv. Synth. Catal. 2006, 348, 1025–1028; (g) Ito, N.; Watahiki, T.; Maesawa, T.; Sajiki, H. Synthesis 2008, 9, 1467–1478; (h) Esaki, H.; Aoki, F.; Umemura, M.; Kato, M.; Maegawa, T.; Monguchi, Y.; Sajiki, H. Chem. Eur. J. 2007, 13, 4052–4063; (i) Kurita, T.; Hattori, K.; Seki, S.; Mizumoto, T.; Aoki, F.; Vamada, Y.; Ikawa, K.; Maegawa, T.; Fujiwara, Y.; Inagaki, Y.; Esaki, H.; Monguchi, Y.; Sajiki, H. Angew. Chem., Int. Ed. 2008, 47, 5394–5397.
- (a) March, J. Advanced Organic Chemistry, 4th ed.; Wiley-VCH: Weinheim, 1992;
 (b) Bell, R. P. Chem. Soc. Rev. 1974, 3, 513–544.
- Refer to synthesis of (*R*)-K-13675: (a) Yamazaki, Y.; Araki, T.; Koura, M.; Shibuya, K. *Synthesis* **2008**, *7*, 1017–1022; (b) Yamazaki, Y.; Araki, T.; Koura, M.; Shibuya, K. *Tetrahedron* **2008**, *64*, 8155–8158.
- 9. We initially attempted the post-synthetic deuterium reaction of 4methoxyphenol to give 4-methoxyphenol- d_7 (**3**) using 5% Pt/C in D₂O under a hydrogen atmosphere at 100 °C for 24 h. However, the H–D exchange displacement of the phenol occurred only at the aromatic ring and afforded 4methoxyphenol- d_4 (64%, 66% D). When we conducted the methylation of 1,4hydroquinone- d_6 (**9**) with iodomethane- d_3 and K₂CO₃ in acetonitrile, **9** was easily transformed into benzoquinone- d_4 via oxidation.
- 10. Bellas, M.; Cahill, R.; Hayes, L. G.B. Patent 1557237.
- 11. Liquid chromatography: Autosampler; 3133 HTS autosampler Z (Shiseido), degasser/binary pump/column compartment; Agilent 1100 Series (Agilent Technologies), Column; Symmetry C18 2.1 × 50 mm id, 5 μ m (Waters). Mass spectrometer: API4000 triple-quadrupole mass spectrometer (Applied Biosystems) was equipped with an electrospray source, operating in the positive ion mode. Quantification was performed in multiple reaction monitoring (MRM) mode. The MS/MS was set to monitor at a transition of m/z 491/107 for (R)-K-13675. m/z 498/107 for (R)-K-13675- d_{11} , respectively. MRM data were acquired and integrated using Analyst software (Ver.1.4.1, Applied Biosystems).
- 12. Methanol solution containing different amounts of **1** (0.02, 0.04, 0.1, 0.2, 0.4, 1, 2, and 4 ng/tube) was added to 250 μ L of 2% propylene glycol in ethanol. After evaporation under nitrogen at 40 °C, rat plasma (100 μ L) was added to the dried residue, and then 100 μ L of aqueous solution of **4** (2.5 ng/mL) and 900 μ L of 0.1% aqueous acetic acid were added to the sample. Each sample was vortexed, extracted with *tert*-butylmethyl ether (5 mL) with shaking for 10 min and centrifuged at 3000 rpm for 10 min. The organic layer (3 mL) was transferred into a glass tube and 250 μ L of 2% propylene glycol in ethanol was added to the solution. After evaporation under nitrogen at 40 °C, the dried residue was dissolved in 150 μ L of 50% aqueous methanol. The samples thus obtained (20 μ L) were analyzed using LC/MS/MS and monitored at *m/z* 491 and 502.¹¹