

Total Synthesis of (*S*)-(-)-Curvularin: A Ring-Closing-Metathesis-Based Construction of the Macrocyclic Framework

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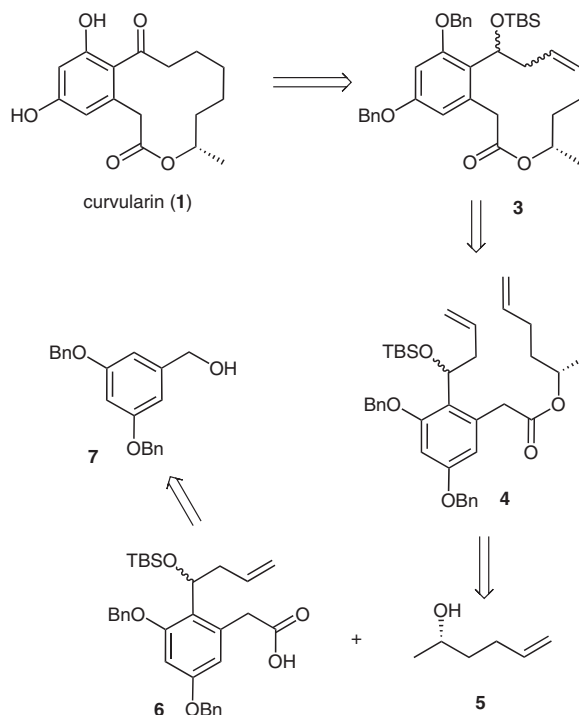
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Abstract: A convergent, flexible, and efficient approach to the synthesis of curvularin is described. Key step is the high-yielding macrocyclic ring formation by ring-closing metathesis (RCM) using the Grubbs second-generation catalyst.

Key words: curvularin, cytotoxic, ring-closing metathesis, Pinnick oxidation, Wittig reaction

Curvularin belongs to a unique class of 12-membered fused resorcinylic macrolides. Curvularin (**1**)^{1,2} and related compounds have been reported as metabolites produced by members of the *Curvularia*,^{3,4} *Alternaria*,^{5,6} and *Penicillium*^{7–9} genera. 11,12-Dehydrocurvularin (**2**) co-occurs with curvularin in *Drechslera australiensis*.¹⁰ These macrolides possess interesting biological properties, including phytotoxicity,⁵ cytotoxicity toward sea urchin embryogenesis,⁸ inhibition of cell division,⁸ inhibition of expression of human inducible nitric oxide synthase,¹¹ and growth-promoting activity in farm animals.¹² The interesting 12-membered fused lactone family and their potent biological activities attracted our attention for the total synthesis.

Despite the great number of reports¹³ regarding the synthesis of curvularin, no effort was made for the construction of macrocyclic framework using ring-closing metathesis. During the final step of our synthesis, Kunz et al.^{13j} for the first time reported the ring-closing metathesis approach for the synthesis of curvularin and its homologues. The retrosynthetic strategy was envisaged utilizing a highly convergent coupling sequence for two intermediates **5** and **6**. The first coupling would entail esterification of an appropriately substituted aromatic acid **6** with an optically pure secondary alcohol **5** containing the



Scheme 1 Retrosynthetic analysis of curvularin (**1**)

single stereogenic center present in curvularin. In the concluding coupling, ring-closing metathesis of a diene **4** would lead to the desired 12-membered macrolide **3**. The substituted aromatic acid could be achieved from readily available alcohol **7**¹⁴ through a tactical combination of functional group manipulation, that is, one-carbon Wittig homologation, sequential hydroboration–oxidation, allylation by Grignard reaction and Pinnick oxidation (Scheme 1).

Synthesis of densely functionalized, substituted aromatic acid began with commercially available 3,5-benzyloxybenzyl alcohol **7**,¹⁵ which was oxidized to aldehyde using PDC, followed by one-carbon Wittig olefination with incipient methylenetriphenyl phosphorane generated in situ from $\text{MeP}^+\text{Ph}_3\text{I}^-$ and *n*-BuLi in THF to provide olefin **8**. The styrene derivative **8** was subjected to the sequential hydroboration–oxidation,¹⁶ protection of the hydroxy group as its PMB ether to provide **9** (Scheme 2). Formylation¹⁷ was effected with POCl_3 , DMF to give aldehyde **10**,¹⁸ which on Grignard reaction with allylmagnesium bromide gave the hydroxyl compound **11**.¹⁹ The alcohol was protected as its TBS ether using TBSCl²⁰ to

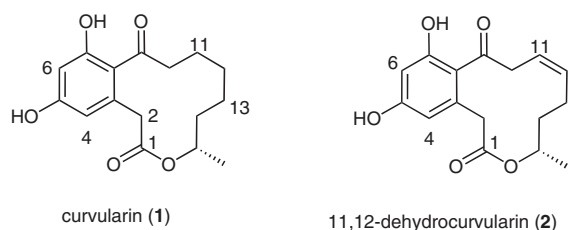
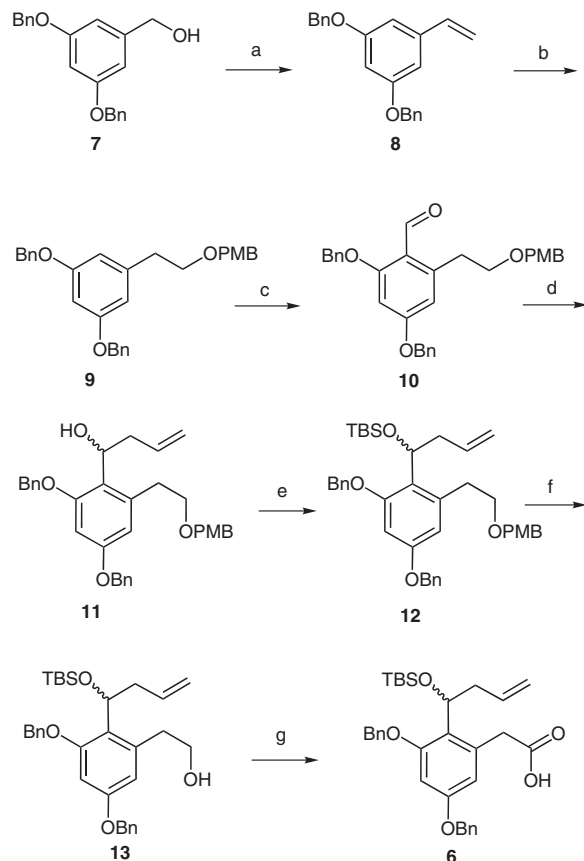
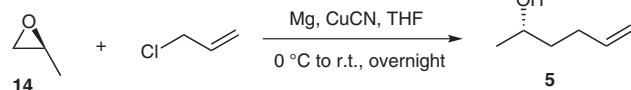


Figure 1



Scheme 2 Reagents and conditions: a) (i) PDC, MS 4 Å, CH₂Cl₂, 0 °C to r.t., 1.5 h; (ii) P⁺Ph₃MeI[−], *n*-BuLi, THF, 0 °C, overnight, 91% (in two steps); b) (i) BH₃·DMS, THF, 0 °C, 3 h, 89%; (ii) PMB-Cl, NaH, DMF, 0 °C, 3 h, 81%; c) DMF, POCl₃, 0–75 °C, 2 h, 82%; d) allyl bromide, Mg, Et₂O, r.t., 3 h, 88%; e) TBSCl, imidazole, DMF, 0 °C, 4 h, 98%; f) DDQ, CH₂Cl₂, buffer (pH = 7), r.t., 1.5 h, 93%; g) (i) IBX, DMSO, THF, r.t., 1 h, 97%, (ii) NaClO₂, NaH₂PO₄, *t*-BuOH, THF, H₂O, 2-methyl-2-butene, 2 h, 94%.



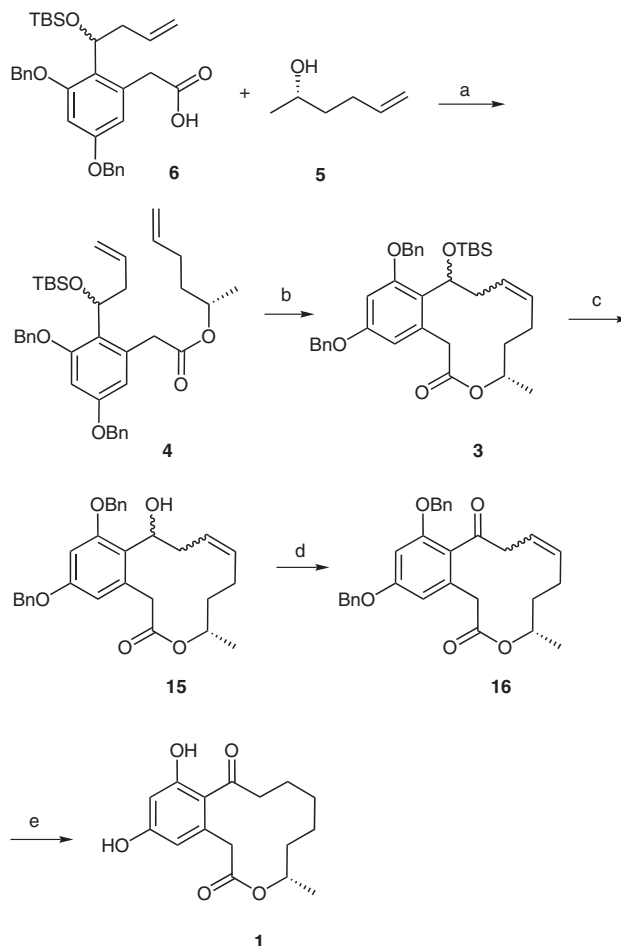
Scheme 3 Synthesis of fragment 5

obtain **12**. Cleavage of PMB ether by treating with DDQ²¹ followed by IBX oxidation of alcohol **13** afforded the corresponding aldehyde which on Pinnick²² oxidation gave the required acid **6**.²³

With the aromatic segment readily in hand, our next attention was to obtain the desired alcohol **5** with commercially available (*S*)-(-)-methyloxirane (**14**) as delineated in Scheme 3.

The epoxide **14** was subjected to CuCN coordinated regioselective nucleophilic opening²⁴ with allylmagnesium chloride to provide the alcohol **5** in 87% yield, the structure of which was adequately substantiated by spectral studies.²⁵

With two subunits in hand, we proceeded to couple both intermediates **5** and **6** as described in Scheme 4. Accord-



Scheme 4 Reagents and conditions: a) DCC, DMAP, CH₂Cl₂, 0 °C to r.t., 2 h, 91%; b) Grubbs' second-generation catalyst, toluene, 70 °C, overnight, 83%; c) TBAF, THF, r.t., 48 h, 97%; d) IBX, DMSO, THF, r.t., 1.5 h, 98%; e) 10% Pd/C, H₂, EtOAc, r.t., 1.5 h,

ingly, the esterification was accomplished using DCC,²⁶ DMAP in CH₂Cl₂ to furnish **4**.²⁷ After having synthesized the diene derivative **4**, the stage was finally set for the construction of macrocyclic framework. Accordingly, on exposure of **4** to the Grubbs second-generation catalyst²⁸ in toluene at 70 °C led to the formation of an *E/Z* mixture of unsaturated macrocycle **3** in 83% yield. The newly generated double bond of **3** was of no consequence, as it would finally be hydrogenated in the later stage of synthetic sequence. All the spectral and elemental analysis secured the assigned structure. Macrolactone **3** was converted into hydroxyl lactone **15** by treating with TBAF (1 M) in THF at ambient temperature.

The IBX oxidation of the secondary hydroxyl functionality in **15** led to the ketolactone **16**.²⁹ Finally, global deprotection and concomitant reduction of the olefin with catalytic hydrogenation^{13b} using 10% Pd/C afforded curvularin (**1**) in 90% yield. The spectroscopic (¹H NMR, ¹³C NMR)³⁰ and analytical data were in all respects identical to those reported in the literature.^{8,13j}

In conclusion, a convergent synthesis of (*S*)-curvularin has been achieved via ring-closing metathesis reaction as

the key step starting from readily available starting materials. This protocol is promising for gram-quantity synthesis of the above molecule and its analogues.

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- (18) **Analytical and Spectral Data of 10**
IR (neat): 3032, 2933, 1673, 1598, 1572, 1513, 1436, 1384, 1320, 1247, 1152, 1090, 1031, 820, 754, 698 cm^{-1} . ^1H NMR (200 MHz, CDCl_3): δ = 3.21 (t, J = 6.6 Hz, 2 H), 3.59 (t, J = 6.6 Hz, 2 H), 3.67 (s, 3 H), 4.35 (s, 2 H), 4.95 (s, 2 H), 4.99 (s, 2 H), 6.40 (d, J = 2.2 Hz, 1 H), 6.42 (d, J = 2.2 Hz, 1 H), 6.73 (d, J = 8.6 Hz, 2 H), 7.12 (d, J = 8.7 Hz, 2 H), 7.22–7.32 (m, 10 H), 10.45 (s, 1 H) ppm. ^{13}C NMR (50 MHz, CDCl_3): δ = 34.9, 55.2, 70.1, 70.2, 70.6, 72.3, 98.4, 110.1, 113.6, 117.4, 127.3, 127.6, 128.2, 128.3, 128.7, 129.2, 130.7, 135.9, 135.9, 145.3, 159.0, 163.4, 164.4, 190.3 ppm. Anal. Calcd for $\text{C}_{31}\text{H}_{30}\text{O}_5$: C, 77.16; H, 6.27. Found: C, 77.46; H, 6.21.
- (19) **Analytical and Spectral Data of 11**
IR (neat): 3400, 2930, 1600, 1512, 1454, 1383, 1302, 1247, 1151, 1084, 1030, 822, 738, 698 cm^{-1} . ^1H NMR (200 MHz, CDCl_3): δ = 2.42–2.55 (m, 1 H), 2.64–2.79 (m, 1 H), 2.84–3.03 (m, 2 H), 3.51–3.67 (m, 2 H), 3.77 (s, 3 H), 4.39 (d, J = 11.5 Hz, 1 H), 4.45 (d, J = 11.6 Hz, 1 H), 4.85–4.95 (m, 2 H), 4.97 (s, 2 H), 5.01–5.02 (m, 1 H), 5.05 (s, 2 H), 5.66–5.87 (m, 1 H), 6.42 (d, J = 2.4 Hz, 1 H), 6.51 (d, J = 2.4 Hz, 1 H), 6.83 (d, J = 8.5 Hz, 2 H), 7.20 (d, J = 8.5 Hz, 2 H), 7.33–7.41 (m, 10 H) ppm. ^{13}C NMR (50 MHz, CDCl_3): δ = 34.6, 55.2, 70.0, 70.1, 70.4, 72.6, 99.2, 108.2, 113.7, 114.3, 116.0, 118.7, 126.3, 126.9, 127.2, 127.6, 127.8, 128.0, 128.5, 128.6, 129.3, 130.4, 132.0, 134.0, 136.7, 136.8, 138.9, 139.3, 158.1, 158.3, 159.1. Anal. Calcd for $\text{C}_{34}\text{H}_{36}\text{O}_5$: C, 77.84; H, 6.92. Found: C, 77.69; H, 6.82.
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- (23) **Analytical and Spectral Data of 6**
IR (neat): 3065, 3031, 2927, 2855, 1730, 1602, 1454, 1385, 1254, 1105, 836, 756, 616 cm^{-1} . ^1H NMR (200 MHz, CDCl_3): δ = -0.11 (s, 3 H), 0.05 (s, 3 H), 0.88 (s, 9 H), 2.40–2.54 (m, 1 H), 2.61–2.75 (m, 1 H), 3.96 (d, J = 16.7 Hz, 1 H), 4.45 (d, J = 16.7 Hz, 1 H), 4.97–5.08 (m, 6 H), 5.55–5.58 (m, 1 H), 5.68–5.89 (m, 1 H), 6.52–6.60 (m, 2 H), 7.30–7.48 (m, 10 H) ppm. ^{13}C NMR (50 MHz, CDCl_3): δ = -3.7, 17.9, 25.6, 34.8, 40.2, 70.0, 70.2, 77.5, 99.1, 104.2, 114.3, 114.7, 119.3, 126.9, 127.1, 127.5, 127.7, 127.8, 127.9, 128.0, 128.1, 128.2, 128.4, 128.5, 128.6, 131.8, 132.4, 136.2, 136.3, 136.6, 136.9, 141.7, 154.9, 159.8, 170.4 ppm. Anal. Calcd for $\text{C}_{32}\text{H}_{40}\text{O}_5\text{Si}$: C, 72.14; H, 7.57. Found: C, 72.31; H, 7.39.
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- (25) **Analytical and Spectral Data of 5**
[α] $_{\text{D}}^{25}$ +9.89 (c 1.1, CHCl_3). IR (neat): 3435, 3019, 1598, 1403, 1215, 1118, 758, 669 cm^{-1} . ^1H NMR (200 MHz, CDCl_3): δ = 1.17 (d, J = 6.2 Hz, 3 H), 1.46–1.58 (m, 2 H), 1.99 (br s, 1 H), 2.06–2.20 (m, 2 H), 3.79 (quin, J = 6.2 Hz, 1 H), 4.90–5.07 (m, 2 H), 5.71–5.91 (m, 1 H) ppm. ^{13}C NMR (50 MHz, CDCl_3): δ = 23.4, 30.1, 38.2, 67.4, 114.7, 138.4 ppm. Anal. Calcd for $\text{C}_6\text{H}_{12}\text{O}$: C, 71.95; H, 12.08. Found: C, 71.89; H, 12.11.
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- (27) **Analytical and Spectral Data of 4**
[α] $_{\text{D}}^{25}$ +7.86 (c 1.5, CHCl_3). IR (neat): 2929, 2371, 2361, 2341, 1730, 1604, 1383, 1257, 1146, 1068, 773, 696, 667 cm^{-1} . ^1H NMR (200 MHz, CDCl_3): δ = -0.16 (s, 3 H), 0.01 (s, 3 H), 0.86 (s, 9 H), 1.22 (d, J = 5.7 Hz, 1.5 H), 1.25 (d, J = 5.7 Hz, 1.5 H), 1.47–1.79 (m, 2 H), 1.99–2.16 (m, 2 H), 2.37–2.50 (m, 1 H), 2.61–2.75 (m, 1 H), 3.70–3.88 (m, 1 H), 4.37–4.50 (m, 1 H), 4.92–5.07 (m, 9 H), 5.50–5.56 (m, 1 H), 5.67–5.91 (m, 2 H), 6.48 (d, J = 2.3 Hz, 1 H), 6.53 (br s, 1 H), 7.31–7.42 (m, 10 H) ppm. ^{13}C NMR (50 MHz, CDCl_3): δ = -5.2, -4.9, 18.2, 19.9, 20.0, 25.9, 29.6, 29.7, 35.1, 69.9, 70.2, 70.5, 70.6, 98.9, 108.5, 108.6, 115.0, 116.4, 124.2, 126.9, 127.2, 127.5, 127.7, 128.0, 128.4, 128.5, 128.6, 135.9, 136.8, 137.0, 137.7, 158.2, 158.3, 170.9, 171.9 ppm. Anal. Calcd for $\text{C}_{38}\text{H}_{50}\text{O}_5\text{Si}$: C, 74.23; H, 8.20. Found: C, 74.01; H, 8.22.

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(29) **Analytical and Spectral Data of 16**

$[\alpha]_{\text{D}}^{25} +45.05$ (*c* 1.0, CHCl_3). IR (neat): 3208, 2928, 1728, 1687, 1655, 1601, 1429, 1311, 1154, 1105, 754 cm^{-1} . ^1H NMR (200 MHz, CDCl_3): δ = 1.09 (d, J = 6.3 Hz, 3 H), 1.47–1.62 (m, 2 H), 1.91–1.99 (m, 1 H), 2.14–2.26 (m, 1 H), 3.05–3.55 (m, 4 H), 4.94–5.02 (m, 5 H), 5.15–5.25 (m, 1 H), 5.38–5.51 (m, 1 H), 6.46 (d, J = 2.1 Hz, 1 H), 6.52 (s, 1 H), 7.22–7.36 (m, 10 H) ppm. ^{13}C NMR (50 MHz, CDCl_3): δ = 20.9, 30.4, 33.7, 38.0, 49.0, 70.2, 70.6, 72.4, 99.6, 109.5, 120.1, 124.3, 127.2, 127.6, 128.0, 128.1, 128.6, 134.1,

136.2, 136.5, 137.2, 156.6, 160.2, 170.8, 204.6 ppm. Anal. Calcd for $\text{C}_{30}\text{H}_{30}\text{O}_5$: C, 76.57; H, 6.43. Found: C, 76.42; H, 6.49.

(30) **Analytical and Spectral Data of 1**

$[\alpha]_{\text{D}}^{25} -31.89$ (*c* 2.0, EtOH); lit¹⁴ $[\alpha]_{\text{D}}^{25} -33.0$ (*c* 2.0, EtOH). IR (Nujol): 3419, 3176, 2933, 2869, 2254, 1719, 1661, 1607, 1589, 1463, 1316, 1264, 1105, 1006, 823, 616. cm^{-1} . ^1H NMR (500 MHz, acetone- d_6): δ = 1.13 (d, J = 6.3 Hz, 3 H), 1.24–1.28 (m, 2 H), 1.40–1.50 (m, 3 H), 1.52–1.57 (m, 1 H), 1.59–1.64 (m, 1 H), 1.73–1.80 (m, 1 H), 2.78 (ddd, J = 2.9, 9.8, 15.5 Hz, 1 H), 3.12 (ddd, J = 2.9, 8.7, 15.5 Hz, 1 H), 3.71 (d, J = 15.7 Hz, 1 H), 3.79 (d, J = 15.6 Hz, 1 H), 4.90–4.96 (m, 1 H), 6.36 (d, J = 2.2 Hz, 1 H), 6.41 (d, J = 2.2 Hz, 1 H), 8.82 (br s, 1 H), 9.19 (br s, 1 H) ppm. ^{13}C NMR (50 MHz, acetone- d_6): δ = 20.5, 23.4, 24.5, 27.4, 32.8, 39.6, 44.0, 72.6, 102.4, 112.2, 121.3, 136.9, 158.2, 160.1, 171.0, 206.7 ppm. Anal. Calcd for $\text{C}_{16}\text{H}_{20}\text{O}_5$: C, 65.74; H, 6.90. Found: C, 65.68; H, 6.91.

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