



A microwave-assisted stereoselective synthesis of polyandrocarpamines A and B

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ARTICLE INFO

Article history:

Received 20 October 2008

Revised 24 November 2008

Accepted 3 December 2008

Available online 7 December 2008

ABSTRACT

A stereoselective synthesis of the marine natural products, polyandrocarpamines A and B, has been achieved using a high-yielding one-step aldol condensation reaction under microwave conditions. The structures of both synthetic compounds were confirmed following 1D and 2D NMR, UV, IR and MS spectral analysis, and by comparison with literature data. Both synthetic natural products were assigned *Z* geometry about their exocyclic double bond on the basis of ¹³C/¹H long-range coupling constants, which were measured using a gHSQMBC experiment. Polyandrocarpamines A and B were evaluated for their cytotoxicity towards the tumour cell lines, MCF-7 (breast), H460 (lung) and SF268 (central nervous system). Polyandrocarpamine A displayed selective cytotoxicity towards the SF268 cell line with a GI₅₀ value of 65 μM.

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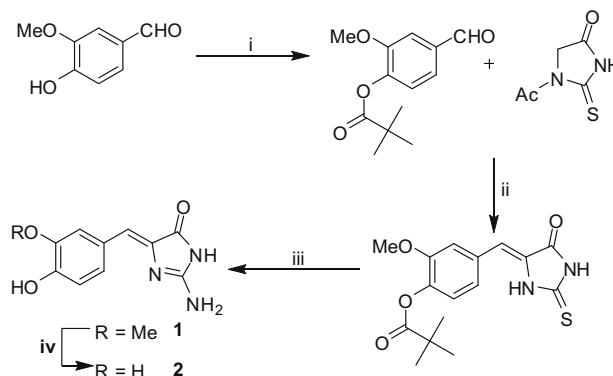
Marine organisms have proven to be a rich source of novel, and structurally diverse metabolites that display a wide variety of biologically significant activities.^{1–3} Examples include the anti-fouling oroidin dimer, mauritamine,⁴ the MEK-1 inhibitor, hymenialdine,⁵ the EGF inhibitor, purealidin K⁶ and the histaminergic antagonist, dispacamide A **3**.⁷ All the marine metabolites listed above belong to the 2-aminoimidazolone class of compounds, and all have been isolated from marine sponges. Other marine organisms, such as ascidians, have also yielded a small number of alkaloids belonging to this structure class, although no bioactivity has been reported.⁸ Polyandrocarpamines A **1** and B **2** are examples of ascidian-derived 2-aminoimidazolones. These simple alkaloids were first isolated and synthesized in 2002,⁹ and although sufficient quantities of these molecules were obtained by total synthesis, no biological activity was reported. Due to our interest in the synthesis of bioactive natural products and their structural analogues,^{10–15} we wished to obtain polyandrocarpamines A **1** and B **2** in sufficient quantities in order to undertake biological evaluations. Herein, we report an alternative synthesis for polyandrocarpamines A and B, along with their cytotoxicity profile against three different tumour cell lines.

The previous synthesis of polyandrocarpamines A **1** and B **2** (Scheme 1),⁹ along with the related marine-derived imidazolones dispacamide A **3**,¹⁶ and leucettamine B **4**,¹⁷ have all been reported using the same methodology for the construction of the 2-aminoimidazolone moiety (Fig. 1).^{9,16,17}

The synthesis of the 2-aminoimidazolone component for **1–4** involved the generation of an alkyl- or aryl-thiohydantoin, followed by the facile conversion of the thiohydantoin to the corresponding

2-aminoimidazolones using *tert*-butyl hydroperoxide as an oxidant in the presence of aqueous ammonia (Scheme 1).^{9,16,17} This method generally requires the protection of reactive groups (e.g., –OH) and long reaction times (up to 72 h), which often result in variable yields.⁹

An alternative synthetic route for the synthesis of 2-aminoimidazolones has been reported using glycoylamine **5**, or a glycoylamine derivative, in the presence of an aldehyde, NaOAc and AcOH.^{17–19} This method uses a readily synthesized reagent (i.e., **5**), requires no protection chemistry and involves a one-step condensation reaction that we thought might be suitable for microwave chemistry. The sponge metabolite, leucettamine B **4**,



Scheme 1. Reagents and conditions for the original synthesis of polyandrocarpamines A and B:⁹ (i) pivaloyl chloride, pyridine, Ar, rt, 7 h; (ii) NaOAc, AcOH, reflux, 2 h; (iii) *tert*-butyl hydroperoxide, aq NH₄OH, MeOH, rt, 72 h; (iv) BBr₃·S(Me)₂, DCE, reflux, 15 min.

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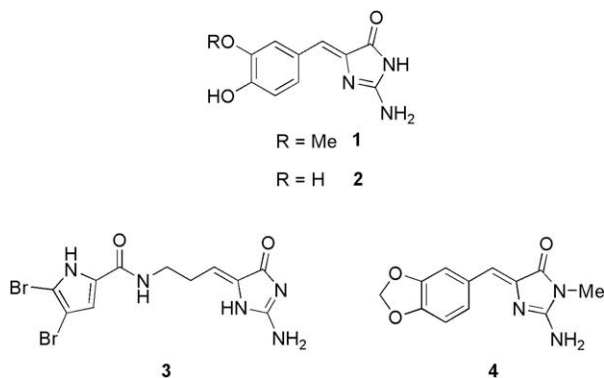


Figure 1. Chemical structures of marine natural products 1–4.

has been successfully synthesized using this glycohydrazide methodology under reflux conditions, although the reaction generated a *Z/E* mixture (9.5:0.5) of **4**.¹⁷

In our laboratory, the HCl salt of glycohydrazide **5** was synthesized using a microwave-modified literature preparation.²⁰ This involved the addition of glycohydrazide to a solution of H₂O/32% aqueous HCl (3:1) followed by heating under microwave conditions at 160 °C for 30 min. Following purification by recrystallisation, the HCl salt of glycohydrazide **5** was condensed with 4-hydroxy-3-methoxybenzaldehyde **6** or 3,4-dihydroxybenzaldehyde **7** in a NaOAc/AcOH slurry under microwave conditions of 160 °C for 30 min (Scheme 2). Purification of each separate reaction mixture using C₁₈ flash chromatography yielded the TFA salts of polyandrocarpamines A²¹ **1** (193 mg, 56%) and B²² **2** (267 mg, 80%) in high yields.

By comparing the overall yields for both **1** (31% vs 56%) and **2** (6% vs 80%) from the two different synthetic routes outlined in Schemes 1 and 2, we showed that the new, faster and simpler methodology also resulted in substantial yield improvements.

The structural confirmation and ¹H/¹³C NMR assignments of compounds **1** and **2** were performed following extensive 1D and 2D NMR spectroscopy (gCOSY, gHSQC, gHMBC and ROESY). The geometry about the exocyclic double bonds of **1** and **2** was determined on the basis of ¹³C/¹H long-range coupling constants, which were measured using a gHSQMBC experiment.²³ Analysis of each gHSQMBC experiment identified that the coupling constant between the exocyclic protons [δ 6.81 (H-6) for **1**; δ 6.79 (H-6) for **2**] and the imidazolone carbonyl [δ 165.2 ppm (C-4) for **1**; 165.7 ppm (C-4) for **2**] was 5.4 Hz for both molecules, which is consistent with the *Z* geometry.^{24,25} Both synthetic **1** and **2** were obtained solely as their *Z* isomers and no isomerisation about the exocyclic double bond was observed during our studies.

Interestingly, several chemical shift values for the TFA salts of **1** and **2** differed compared to the original data reported for the natural products and synthetics.⁹ In particular, the ¹³C NMR data for the 2-aminoimidazolone moiety of **1** and **2** reported in this Letter showed some differences (Δ 5.2–8.7 ppm for C-2; Δ 11.1–13.0 ppm for C-4; Δ 8.4–8.8 ppm for C-5) with those published in the original isolation and synthesis paper of polyandrocarp-

amines A and B.⁹ These differences were postulated to arise since the isolated natural products were purified as their free bases.⁹ In order to confirm this hypothesis, a small amount of polyandrocarpamine A TFA salt was purified on a base-stable C₁₈ HPLC column using aqueous NH₄OH/MeOH. The free base of polyandrocarpamine A was obtained, and NMR assignments were made following analysis of the 1D and 2D NMR spectra, and were shown to be essentially identical to those of the natural product originally reported.⁹

Polyandrocarpamines A and B were tested for their cytotoxic activity against the tumour cell lines, H460 (lung), MCF-7 (breast), and SF268 (central nervous system).^{26,27} Initial dosing at 10 μ M for 72 h for both alkaloids showed no growth inhibition towards H460 and MCF-7; however, moderate cytotoxicity was identified towards the cancer cell line, SF268. Further biological testing revealed that **1** and **2** showed cytotoxicity towards this cell line with GI₅₀ values of 65 and >80 μ M, respectively.

In conclusion, we have successfully synthesized polyandrocarpamines A and B stereoselectively in high-yield using a microwave reactor, and have shown that both natural products have selective cytotoxicity towards the SF268 tumour cell line. With the synthesis of large quantities of **1** and **2**, these molecules will now be added to the Queensland compound library (QCL)²⁸ at the Eskitis Institute, where they will be available for further biological evaluations. This new synthetic route to polyandrocarpamines has substantially reduced the synthesis time, by up to 80 h, and uses less reagents compared to the original procedure.⁹ This method makes this class of marine alkaloids now more amenable to natural product-based combinatorial synthesis.

Acknowledgements

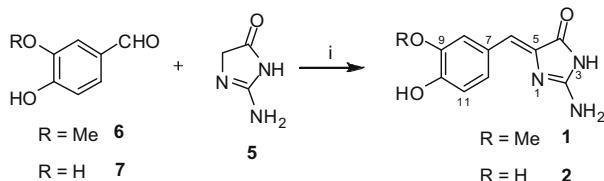
The authors acknowledge financial support of this work by Griffith University and the Eskitis Institute for Cell and Molecular Therapies. We thank Hoan The Vu from Griffith University for acquiring the HRESIMS measurements. One of us (P.S.B.) thanks Griffith University for an Honours scholarship. Alison Slater is acknowledged for technical assistance in performing the cell culture assays.

Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2008.12.010.

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Scheme 2. Reagents and conditions: (i) NaOAc, AcOH, μ w, 160 °C, 20 min.

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21. Yellow amorphous solid; mp 182–186 °C; UV (MeOH) λ_{max} (log ϵ) 216 (3.66), 240 (2.55), 357 (2.76) nm; IR ν_{max} (KBr) 3500–2800, 1712, 1667, 1592, 1519, 1436, 1395, 1291, 1201, 1136, 1055, 1029, 840, 802, 723 cm^{-1} ; ^1H NMR (500 MHz, CD_3OD) δ 3.91 (3H, s, 9-OCH₃), 6.81 (1H, s, H-6), 6.87 (1H, d, J = 8.5 Hz, H-11), 7.10 (1H, dd, J = 8.5, 1.5 Hz, H-12), 7.12 (1H, d, J = 1.5 Hz, H-8); ^{13}C NMR (125 MHz, CD_3OD) δ 56.8 (9-OCH₃), 114.3 (C-8), 116.9 (C-11), 119.7 (C-6), 123.6 (C-5), 124.8 (C-7), 125.8 (C-12), 149.5 (C-9), 150.6 (C-10), 157.6 (C-2), 165.2 (C-4); (+)-LRESIMS m/z (rel. int.) 234 (100) $[\text{M}-\text{CF}_3\text{COO}]^+$; (+)-HRESIMS m/z 234.08709 ($\text{C}_{11}\text{H}_{12}\text{N}_3\text{O}_3$ $[\text{M}-\text{CF}_3\text{COO}]^+$ requires 234.08732).
22. Yellow amorphous solid; mp >250 °C (decomp.); UV (MeOH) λ_{max} (log ϵ) 218 (2.98), 250 (2.73), 359 (3.00) nm; IR ν_{max} (KBr) 3500–2800, 1704, 1655, 1594, 1438, 1396, 1270, 1191, 1135, 841, 797, 722, 669 cm^{-1} ; ^1H NMR (500 MHz, CD_3OD) δ 6.79 (1H, s, H-6), 6.86 (1H, d, J = 8.0 Hz, H-11), 7.01 (1H, d, J = 8.0 Hz, H-12), 7.02 (1H, s, H-8); ^{13}C NMR (125 MHz, CD_3OD) δ 117.0 (C-11), 118.0 (C-8), 119.8 (C-6), 124.0 (C-5), 124.1 (C-12), 125.0 (C-7), 147.1 (C-9), 149.5 (C-10), 157.9 (C-2), 165.7 (C-4); (+)-LRESIMS m/z (rel. int.) 220 (100) $[\text{M}-\text{CF}_3\text{COO}]^+$; (+)-HRESIMS m/z 220.07273 ($\text{C}_{10}\text{H}_{10}\text{N}_3\text{O}_3$ $[\text{M}-\text{CF}_3\text{COO}]^+$ requires 220.07167).
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