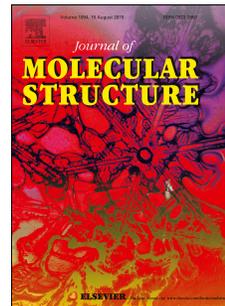


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Sulfonated starch nanoparticles: An effective, heterogeneous and bio-based catalyst for synthesis of 14-aryl-14-*H*-dibenzo[a,j]xanthenes

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Highlights

- Preparation of sulfonated starch nanoparticles (SO₃H-SNPs) as catalyst for organic synthesis.
- Introducing an efficient new catalyst for the synthesis of xanthenes.
- Excellent yields of products and short reaction times.

1 Sulfonated starch nanoparticles: An effective, heterogeneous and bio-based
2 catalyst for synthesis of 14-aryl-14-*H*-dibenzo[a,j]xanthenes

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6 Abstract

7 In recent years, biodegradable polymer based nanoparticles have attracted wide attention for
8 the synthesis of high-performance and green catalytic species. Polymeric nanoparticles used
9 for catalytic processes must be biocompatible and biodegradable. The objective of this study
10 is to fabricate starch nanoparticles from native starch and preparation of sulfonated starch
11 nanoparticles (HO₃S-SNPs) as acidic nanocatalyst in the synthesis of 14-aryl-14-*H*-
12 dibenzo[a,j]xanthenes under solvent free conditions. This procedure has a lot of advantages
13 such as very easy reaction conditions, low-cost production and natural catalyst and absence of
14 any tedious workup or purification. The corresponding products have been obtained in
15 excellent yields, high purity and short reaction times.

16 Keywords: Sulfonated starch, Starch nanoparticles, Heterogeneous catalysis, 14-Aryl-14-*H*-
17 dibenzo[a,j]xanthenes, Solvent-free conditions.

18 1. Introduction

19 Homogeneous inorganic acids such as H₂SO₄ and HF are essential and efficient catalysts for
20 the production of industrial important chemicals. However, the use of these homogeneous
21 catalysts requires costly and inefficient processes for separation, recycling, and treatment of
22 the spent acids. For example, the neutralization of the H₂SO₄ produces sulfate wastes, and

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23 thus, the liquid acids do not fulfill the general requirement of a catalyst as a reusable material
24 in chemical processes. The use of recyclable strong solid acids instead of unrecyclable
25 homogeneous catalysts is a major area of research for the development of cleaner and more
26 efficient processes [1-3].

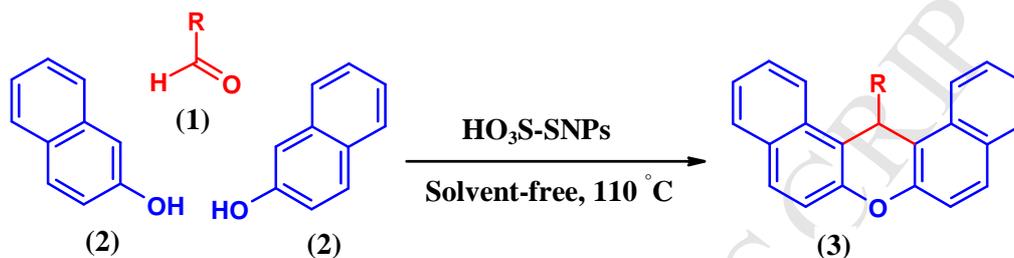
27 Recently, the utilization of polymer bound catalysts is well recognized because of their ease
28 of workup, simple and clean separation of products and catalysts from the reaction mixture. In
29 this context, cellulose and starch are good candidates as natural, renewable, biocompatible,
30 and biodegradable polymers [4-6]. As an ideal support material for catalysts, starch has
31 unique properties including availability, safety, non-toxicity, and inexpensive and
32 compatibility with many other materials. Owing to these properties, starch has been used as a
33 bio-based support for a wide range of catalytic species. This catalyst gained much attention
34 due to the low-cost production, easy preparation process, high effectiveness, greener and
35 suitable for chemical transformations [4-8]. The polar hydroxyl groups in starch provide
36 active sites for numerous attractive chemical modifications [3]. One such modification is
37 sulfonation which leads to the formation of starch-SO₃H. Starch-SO₃H has been utilized as
38 solid acid catalyst in organic reactions [4, 5]. Therefore, the modification of starch is rapidly
39 developed along with its growing applications. Starch-based nanoparticles have received a
40 great deal of attention because of their nano-size, large surface area, biodegradable, and good
41 biocompatibility. These characteristics favour the employment of starch nanoparticles in a
42 wide range of various industrial applications especially food products, biomedical fields
43 especially drug carriers, composites filler, water treatment agent, fat replacer and catalytic
44 processes [9-13]. Moreover, starch nanoparticles have raised much attention, as sustainable
45 development policies tend to expand with the decreasing reserve of fossil fuel and the
46 growing concern for the environment [10]. These bio-based nanoparticles have received a

47 tremendous level of attention as a promising natural material in waste water treatment due to
48 their unique biological and adsorptive properties [11, 12].

49 In recent times, xanthenes and benzoxanthenes have attracted attention of medicinal chemists
50 as well as organic chemists due to their wide range of biological and pharmacological
51 activities such as antibacterial and anti-inflammatory activities [14], photodynamic therapy
52 and as antagonism for the paralyzing action of zoxazolamine [15]. Moreover, these valuable
53 compounds have found wide usage such as fluorescent dyes [16], in laser technologies [17]
54 and as pH-sensitive fluorescent materials for visualization of biomolecules [18]. Due to the
55 high medicinal and biological importance of xanthenes, several methods have been reported
56 for the synthesis of these heterocycles, which include trapping of benzyne by phenols,
57 cyclodehydrations, condensation between aromatic 2-hydroxyl aldehydes and 2-tetralone and
58 intramolecular phenyl carbonyl coupling reactions of benzaldehyde and acetophenone
59 derivatives. Also, 14-aryl-14*H*-di benzo[*a,j*]xanthenes and related products have been
60 prepared by the reaction of 2-naphthol with formamide, 2-naphthol-1-methanol and carbon
61 monoxide [19]. On the other hand, a number of methods have been developed for the
62 synthesis of 14-*H*-dibenzo[*a,j*]xanthene by condensation of 2-naphthol and aldehydes in the
63 presence of strontium sulfamic acid [20], Amberlyst-15 [21], $K_5CoW_{12}O_{40} \cdot 3H_2O$ [22], $FeCl_3$
64 [23], $WO_3/ZrOb$ [24], $PS/AlCl_3$ [25], Alum [26], $In(OTf)_3$ [27]. Therefore, a significant
65 attention has been focused on designing environmentally benign catalysts and synthetic
66 strategies for the preparation of xanthenes derivatives. The green processes in modern
67 chemical synthesis involve the use of biodegradable, cost-effective and efficient catalysts,
68 non-toxic system such as water, supercritical fluids and the reactions under solvent-free
69 conditions. Advantages of solvent-free reactions include cost savings, energy consumption,
70 atom economy, or improved atom utilization reduced decreased reaction times, and a

71 considerable reduction in reactor size and simplicity in processing, which are beneficial to the
72 industry as well as to the environment [28].

73 In the present work, we report a simple and efficient procedure for one-pot synthesis of 14-
74 aryl-14-*H*-dibenzo[*a,j*]xanthenes using sulfonated starch nanoparticles (HO₃S-SNPs) as an
75 effective, neutral and heterogeneous acidic catalyst under solvent-free conditions (Scheme 1).



77 Scheme 1. Synthesis of 14-aryl-14*H*-dibenzo[*a,i*]xanthenes using HO₃S-SNPs as catalyst.

78 2. Experimental

79 Chemical substances were purchased from Merck or Aldrich. Melting points were determined
80 on an Electrothermal 9100 apparatus using open-glass capillary and are uncorrected. FT-IR
81 spectra were recorded as KBr pellets on a Shimadzu FT-IR-8400S spectrometer. ¹H NMR
82 (400 MHz) spectra were obtained using a Bruker DRX-400 AVANCE spectrometer.
83 Analytical thin-layer chromatography (TLC) was accomplished on 0.2 mm precoated plates of
84 silica gel 60 F-254 (Merck) or neutral alumina oxide gel 60F 254 (Merck). Morphological
85 characteristics of chitosan derivatives were observed by a scanning electron microscope
86 (SEM, LEO 1544VP) operating at an accelerating voltage of 20 kV under high vacuum.
87 Nanostructures were characterized using a Holland Philips Xpert X-ray diffraction (XRD)
88 diffractometer (CuK_α radiation, λ = 0.154056 nm), at a scanning speed of 2°/min from 10° to
89 100° (2θ).

90 2.1. Synthesis of starch nanoparticles

91 0.35 g of maleic acid was firstly dissolved in 100 mL of distilled water, then 1 g of starch
92 was added to the this solution the resulted mixture was stirred for 3 h. Then, hydrochloric acid

93 and sodium hydroxide solution (0.1 M) was dropped slowly into the mixture solution until the
94 pH value was titrated to 4.0 with vigorous mechanical stirring and under continuous Ar
95 atmosphere bubbling for 8 h. Afterwards, 0.054 g of potassium persulfate was added to the
96 mixture and stirred for 2 h at 70 °C under Ar atmosphere. The mixture was cooled in ice-bath
97 and the formed nanoparticles were dried by dry ice.

98 2.2. Synthesis of sulfonated starch nanoparticles:

99 HO₃S-SNPs were prepared according to reported procedure in the literature [4]. A 500 mL
100 suction flask was equipped with a pressure equalizing dropping funnel containing
101 chlorosulfonic acid and gas inlet tube for conducting HCl gas over adsorbing the solution
102 water. A flask was charged with starch nanoparticles (500 g) and dispersed in dry CHCl₃ (20
103 mL). Subsequently, chlorosulfonic acid (1.00 g, 9 mmol) was added drop-wise manner to a
104 cooled (ice-bath) solution of starch over a 2 h period, which HCl gas evolved from the
105 reaction vessel immediately. After the addition was completed the mixture was filtered and
106 washed with methanol (30 mL) and dried at room temperature to obtain HO₃S-SNPs (5.06 g)
107 as cream powder. The number of H⁺ sites of starch-SO₃H was determined by acid–base
108 titration and found to be 1.50 meq/g of modified starch at 25 °C. The sulfur content of the
109 samples was determined 1.45 mmol/g by conventional elemental analysis.

110 2.3. General method for the synthesis of benzoxanthenes

111 In a glass tube a mixture of aldehyde (1.0 mmol), 2-naphthol (2.0 mmol) and HO₃S-SNPs
112 (0.08 g) were stirred at 110 °C in an oil bath for the appropriate time, as indicated by TLC for
113 a complete reaction. Ethyl acetate (10 mL) was added and the reaction mixture filtered to
114 separate the catalyst. Pure products were afforded by evaporation of the solvent, followed by
115 recrystallization from ethanol. All the products were characterized by comparison of IR and
116 NMR spectral data with the values of authentic samples.

117 14-(Phenyl)14*H*-dibenzo[*a,j*]xanthenes (3a): pale yellow solid; IR (KBr) ν_{\max} (cm^{-1}): 3071
118 (aromatic C–H), 2923 (aliphatic C–H), 1624 (C=C), 1249 (C–O); ^1H NMR (CDCl_3 , 400 MHz)
119 δ (ppm): 6.5 (s, 1H), 6.98–7.01 (t, 1H $J=7.2$ Hz), 7.13 (s, 1H), 7.15–7.17 (d, 1H $J=8$ Hz), 7.39
120 (s, 1H), 7.41–7.43 (d, 1H, $J=7.2$ Hz), 7.48–7.60 (m, 6H), 7.79–7.84 (m, 4H), 8.39– 8.41 (2H, d,
121 $J=8$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ (ppm): 148.8, 145.0, 131.5, 131.1, 128.8, 128.5,
122 128.2 126.8, 126.5, 126.4, 124.2, 122.7, 118.0, 117.4, 38.1.

123 14-(3-Nitrophenyl)14*H*-dibenzo[*a,j*]xanthenes (3b): yellow solid; IR (KBr) ν_{\max} (cm^{-1}): 3074
124 (aromatic C–H), 2923 (aliphatic C–H), 1625 (C=C), 1559 (NO_2), 1346 (NO_2), 1249 (C–O);
125 ^1H NMR (CDCl_3 , 400 MHz) δ (ppm): $\delta=$ 6.61 (1H, s), 7.29–7.31 (1H, d, $J=7.2$ Hz), 7.44–7.46
126 (2H, d, $J=7.2$ Hz), 7.51–7.53 (2H, d, $J=8.4$ Hz), 7.62 (2H, t, $J=8.4$ Hz), 7.83–7.85 (6H, m),
127 8.30–8.32 (2H, d, $J=7.6$ Hz), 8.42 (1H, s); ^{13}C NMR (CDCl_3 , 100 MHz) δ (ppm): 37.3, 116.3,
128 116.8, 117.4, 121.1, 122.3, 125.4, 126.9, 128.2, 129.1, 134.7, 136.3, 137.1, 138.2, 146.5,
129 147.9, 158.3.

130 14-(4-Nitrophenyl)14*H*-dibenzo[*a,j*]xanthenes (3c): pale yellow solid; IR (KBr) ν_{\max} (cm^{-1}):
131 3041 (aromatic C–H), 1622 (C=C), 1583, 1548 (NO_2), 1348 (NO_2), 1242 (C–O), 800–850; ^1H
132 NMR (CDCl_3 , 400 MHz) δ (ppm): 6.61 (1H, s), 7.43–7.45 (2H, t, $J=8.0$ Hz), 7.51–7.53 (2H,
133 d, $J=8.1$ Hz), 7.59–7.61(2H, t, $J=8.0$ Hz), 7.68–7.70 (2H, d, $J=8.1$ Hz), 7.84–7.87 (4H, m),
134 8.00–8.02 (2H, d, $J=8.0$ Hz), 8.28–8-30 (2H, d, $J=8.0$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ
135 (ppm): 36.9, 115.9, 116.2, 117.1, 120.2, 122.5, 124.8, 126.1, 128.1, 129.7, 132.6, 134.1,
136 135.9, 148.3, 159.1.

137 14-(2-Flouoro)14*H*-dibenzo[*a,j*]xanthenes (3d): cream solid; IR (KBr) ν_{\max} (cm^{-1}): 3090
138 (aromatic C–H), 2923 (aliphatic C–H), 1623 (C–C), 1458 (C=C), 1243 (C–O), 1100–1250; ^1H
139 NMR (CDCl_3 , 400 MHz) δ (ppm): 6.79 (1H, s), 6.81–6.83 (1H, d, $J=6.8$ Hz), 6.96–7.04 (2H,
140 m), 7.20–7.24 (3H,m), 7.41–7.62 (4H, m), 7.79–7.84 (4H, m), 8.40–8.42 (2H, d, $J=8.8$ Hz);

141 ^{13}C NMR (CDCl_3 , 100 MHz) δ (ppm): 30.2, 115.1, 115.2, 116.7, 117.9, 122.3, 124.4, 125.1,
142 127.1, 128.2, 128.6, 129.1, 130.9, 131.1, 131.5, 132.4.

143 14-(2-Chlorophenyl)14*H*-dibenzo[*a,j*]xanthenes (3e): white solid; IR (KBr) ν_{max} (cm^{-1}): 3056
144 (aromatic C–H), 2924–2992 (aliphatic C–H), 1247 (C–O); ^1H NMR (CDCl_3 , 400 MHz) δ
145 (ppm): 6.80 (1H, s), 6.89–6.96 (2H, m), 7.25–7.26 (1H, d, $J=8.4$ Hz), 7.38–7.46 (3H, m),
146 7.49–7.51 (2H, d, $J=8.8$ Hz), 7.61–7.65 (2H, t, $J=7.6$ Hz), 7.79–7.84 (4H, m), 8.76–8.76 (2H,
147 d, $J=8.4$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ (ppm): 36.4, 117.9, 123.3, 124.6, 125.5, 127.8,
148 128.6, 129.4, 129.5, 131, 132.5, 132.6, 134.5, 141.3, 147.5, 149.8, 152.

149 14-(4-Chlorophenyl)14*H*-dibenzo[*a,j*]xanthenes (3f): yellow solid; IR (KBr) ν_{max} (cm^{-1}):
150 3050 (aromatic C–H), 2922 (aliphatic C–H), 1625 (C–C), 1244 (C–O); ^1H NMR (CDCl_3 , 400
151 MHz) δ (ppm): 6.48 (1H, s), 7.10–7.12 (2H, d, $J=7.2$ Hz), 7.42–7.50 (6H, m), 7.58–7.61 (2H,
152 t, $J=7.2$ Hz), 7.80–7.86 (4H, m), 8.32–8.34 (2H, d, $J=8$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ
153 (ppm): 37.39, 116.74, 118.02, 122.42, 124.38, 126.93, 128.65, 128.92, 129.10, 129.50,
154 131.05, 131.25, 132.08, 143.47, 148.68.

155 14-(2,4-Dichlorophenyl)14*H*-dibenzo[*a,j*]xanthenes (3g): pale yellow solid; IR (KBr) ν_{max}
156 (cm^{-1}): 3061 (aromatic C–H), 2923 (aliphatic C–H), 1624 (C–C), 1247 (C–O); ^1H NMR
157 (CDCl_3 , 400 MHz) δ (ppm): 6.75 (1H, s), 6.89–6.91 (1H, t, $J=6.4$ Hz), 7.27–7.31 (2H, m),
158 7.42–7.49 (4H, m), 7.60–7.64 (2H, t, $J=8.4$ Hz), 7.79–7.84 (4H, t), 8.64–8.66 (2H, d, $J=8$ Hz);
159 ^{13}C NMR (CDCl_3 , 100 MHz) δ (ppm): 36.6, 117.3, 117.5, 118.4, 123.3, 126.7, 127.5, 128.4,
160 130.8, 133.5, 135, 141.5, 150.9.

161 14-(4-Methyl)14*H*-dibenzo[*a,j*]xanthenes (3h): pale yellow solid, IR (KBr) ν_{max} (cm^{-1}): 3061
162 (aromatic C–H), 2923 (aliphatic C–H), 1624 (C–C), 1247 (C–O); ^1H NMR (CDCl_3 , 400 MHz)
163 δ (ppm): 2.13 (3H, s), 6.46 (1H, s), 6.95–6.97 (2H, d, $J=7.6$ Hz), 7.39–7.43 (4H, m), 7.47–7.50
164 (2H, d, $J=8.8$ Hz), 7.56–7.60 (2H, t, $J=8$ Hz), 7.78–7.84 (4H, m), 8.39–8.41 (2H, d, $J=8.4$ Hz);

165 ^{13}C NMR (CDCl_3 , 100 MHz) δ (ppm): 20.95, 37.67, 117.48, 118.04, 122.76, 124.25, 126.79,
166 128.15, 128.81, 128.82, 129.22, 131.11, 131.49, 135.93, 142.18, 148.

167 14-(2-Methoxy)14*H*-dibenzo[*a,j*]xanthenes (3i): cream solid, IR (KBr) ν_{max} (cm^{-1}): 3051
168 aromatic C–H), 2921 (aliphatic C–H), 1622 (C=C), 1247 (C–O); ^1H NMR (CDCl_3 , 400 MHz)
169 δ (ppm): 3.62 (s, 3H, OCH_3), 6.46 (s, 1H, ArH), 6.67–6.69 (2H, d, $J=8.4$ Hz), 7.40–7.44 (4H,
170 m), 7.47–7.49 (2H, d, $J=8.8$ Hz), 7.57–7.60 (2H, t, $J=7.6$ Hz), 7.78–7.84 (4H, m), 8.38–8.40
171 (2H, d, $J=8.4$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ (ppm): 44.11, 55.17, 114.78, 118.59,
172 119.21, 122.54, 125.23, 127.77, 129.65, 129.72, 130.01, 132.16, 132.76, 137.45, 149.04, 158.

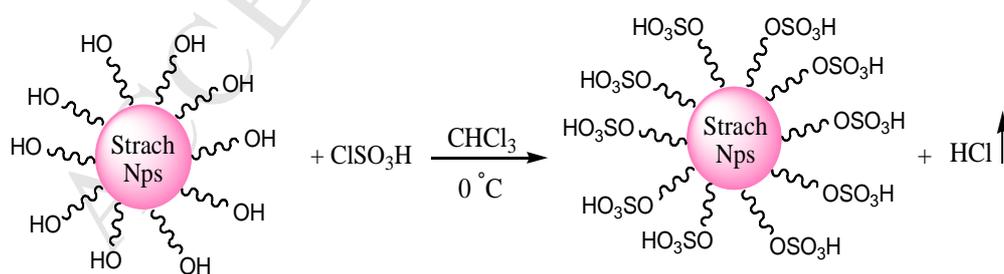
173 14-(4-Methoxy)14*H*-dibenzo[*a,j*]xanthenes (3j): pink solid; IR (KBr) ν_{max} (cm^{-1}): 3050
174 (aromatic C–H), 2922 (aliphatic C–H), 1593 (C=C), 1247 (C–O); ^1H NMR (CDCl_3 , 400 MHz)
175 δ (ppm): 3.62 (3H, s), 6.45 (1H, s), 6.67–6.69 (2H, d, $J=8.8$ Hz), 7.40–7.49 (6H, m), 7.57–7.60
176 (2H, t, $J=7.2$ Hz), 7.78–7.80 (2H, d, $J=8.8$ Hz), 7.82–7.84 (2H, d, $J=8.8$ Hz), 8.38–8.40 (2H, d,
177 $J=8.4$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ (ppm): 37.11, 55.05, 113.85, 117.54, 118.02,
178 122.70, 124.23, 126.77, 128.74, 128.82, 129.17, 131.08, 131.43, 137.39, 148.68, 157.21.

179 14-(3-Hydroxyphenyl)14*H*-dibenzo[*a,j*]xanthenes (3k): Pale Pink solid; IR (KBr) ν_{max} (cm^{-1}):
180 3070 (OH), 1595 (C=C), 1246 (C–O); ^1H NMR (CDCl_3 , 400 MHz) δ (ppm): 3.45 (s, 3H), 6.45
181 (s, 1H), 6.33 (d, 1H, $J=8.2$ Hz), 7.08 (t, 2H, $J=8.4$ Hz), 7.21 (d, 1H, $J=7.5$ Hz), 7.33 (t, 2H,
182 $J=7.2$ Hz), 7.52 (d, 2H, $J=9.0$ Hz), 7.58 (t, 2H, $J=7.6$ Hz), 7.72–7.80 (m, 4H), 8.40 (d, 2H,
183 $J=8.7$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ (ppm): 37.22, 116.05, 118.02, 118.75, 123.66,
184 125.02, 126.45, 130.30, 130.56, 131.03, 131.76, 132.11, 138.54, 149.45, 154.43.

185 14-(4-Hydroxyphenyl)14*H*-dibenzo[*a,j*]xanthenes (3l): Pink solid; IR (KBr) ν_{max} (cm^{-1}): 3405
186 (OH), 1590 (C=C), 1245 (C–O); ^1H NMR (CDCl_3 , 400 MHz) δ (ppm): 5.01 (s, 1H, OH), 6.42
187 (s, 1H, CH), 6.55–8.36 (m, 16H, Ar); ^{13}C NMR (CDCl_3 , 100 MHz) δ (ppm): 37.45, 115.85,
188 117.85, 118.50, 123.20, 124.62, 127.23, 129.30, 129.56, 130.00, 131.58, 131.61, 138.02,
189 149.18, 154.32.

190 3. Results and discussion

191 Sulfonic acid-based solid catalysts have used in organic reactions due to their unique
 192 properties, such as high reactivity, availability, selectivity, eco-friendly reaction conditions,
 193 and ability to promote a wide range of chemical reactions. Since the sulfonation of D-glucose
 194 was first reported as a carbohydrate acid catalyst by Hara et al., [29] much efforts have been
 195 devoted to developing various types of solid acid catalysts from bio-based polymers, such as
 196 sucrose, starch, cellulose and bagasse [30-33] This class of biocompatible and biodegradable
 197 catalysts have shown much better performance than commercially available solid acid
 198 catalysts, such as Nafion NR50, zeolites and Amerlyst-15 [34]. Furthermore, nanoscale
 199 polymers have been attracting more research interest in catalytic systems because of their
 200 unique physical and chemical properties, such as nano size, high surface-to-volume ratio and
 201 stiffness and improved dispersibility. The polymeric nanoparticles offer a number of distinct
 202 advantages over bulk polymers. They have been used in different types of chemical reactions
 203 as a suitable support for different catalytic species [35]. In the first of research, starch was
 204 modified to starch nanoparticles by a nanoprecipitation process. Next, HO₃S-SNPs were
 205 prepared by the reaction of chlorosulfonic acid with starch nanoparticles and the by-product,
 206 HCl gas, was easily removed from the reaction vessel (Scheme 2).



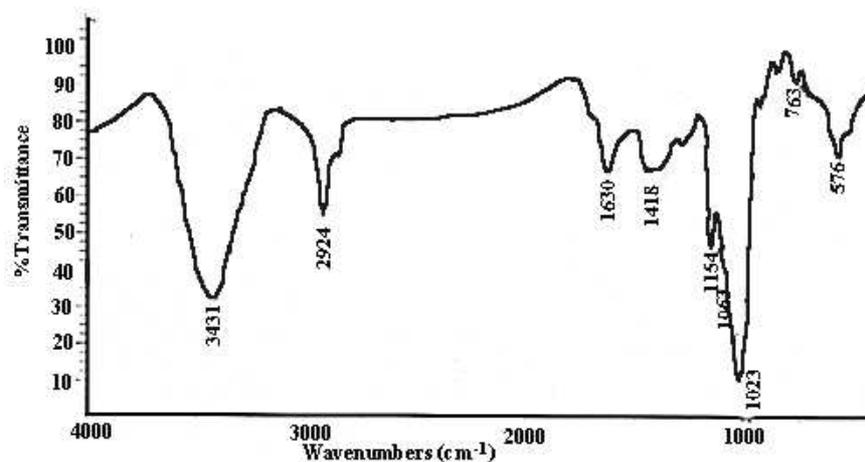
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208

Scheme 2: Preparation steps for fabricating HO₃S-SNPs.

209 The morphology and structure of starch nanoparticles were investigated by FT-IR, XRD,
 210 TEM and SEM. The FT-IR spectrum of starch nanoparticles are shown in Fig. 1. The O-H
 211 stretching and the C-H stretching vibration gave strong signals at 3431 and 2924 cm⁻¹,

212 respectively. Meanwhile, the band at 1630 cm^{-1} was ascribed to O-H bending vibration.
213 Besides, several discernible absorbencies at 1154 and 1023 cm^{-1} were assigned to the C-O
214 bond stretching vibrations of glucose units [36].

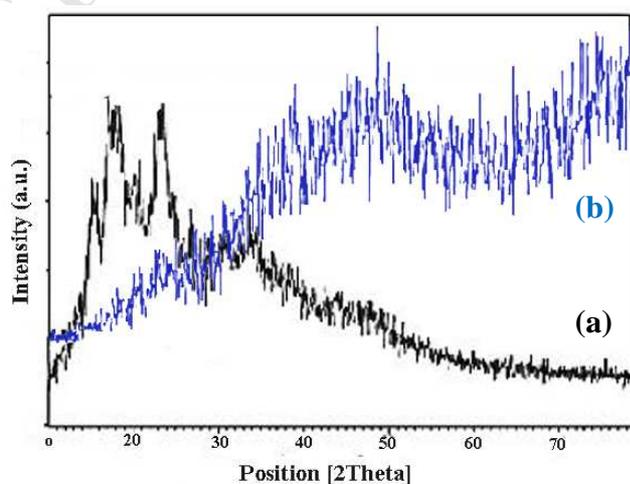


215

216

Fig. 1: FT-IR of starch nanoparticles.

217 Fig. 2 (a) and (b) display XRD result of native starch and starch nanoparticles, respectively.
218 The sharp diffraction peaks at 5.8° , 17.4° , 22.3° , and 24.2° were presented in spectrogram of
219 unmodified starch, which indicated that starch exhibited a typical B-type X-ray pattern. The
220 spectrogram of starch nanoparticles showed a broad peak, suggesting that it was the
221 amorphous nature. Fu et al. also reported the similar XRD results for starch nanoparticles
222 [36].

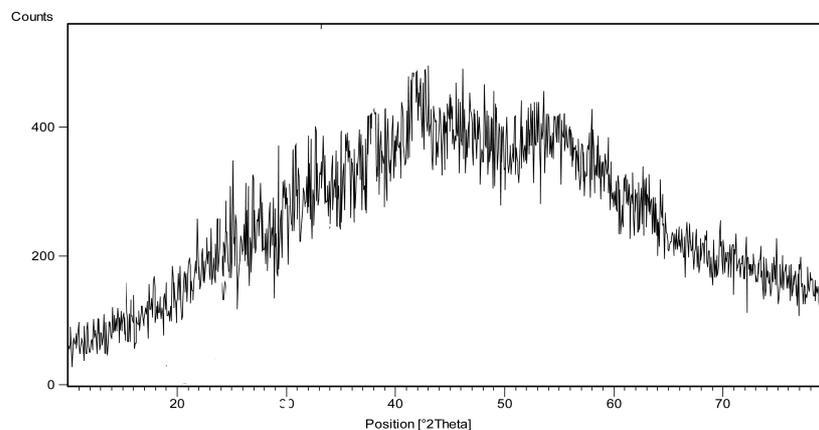


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Fig. 2. XRD patterns of (a) starch and (b) starch nanoparticles.

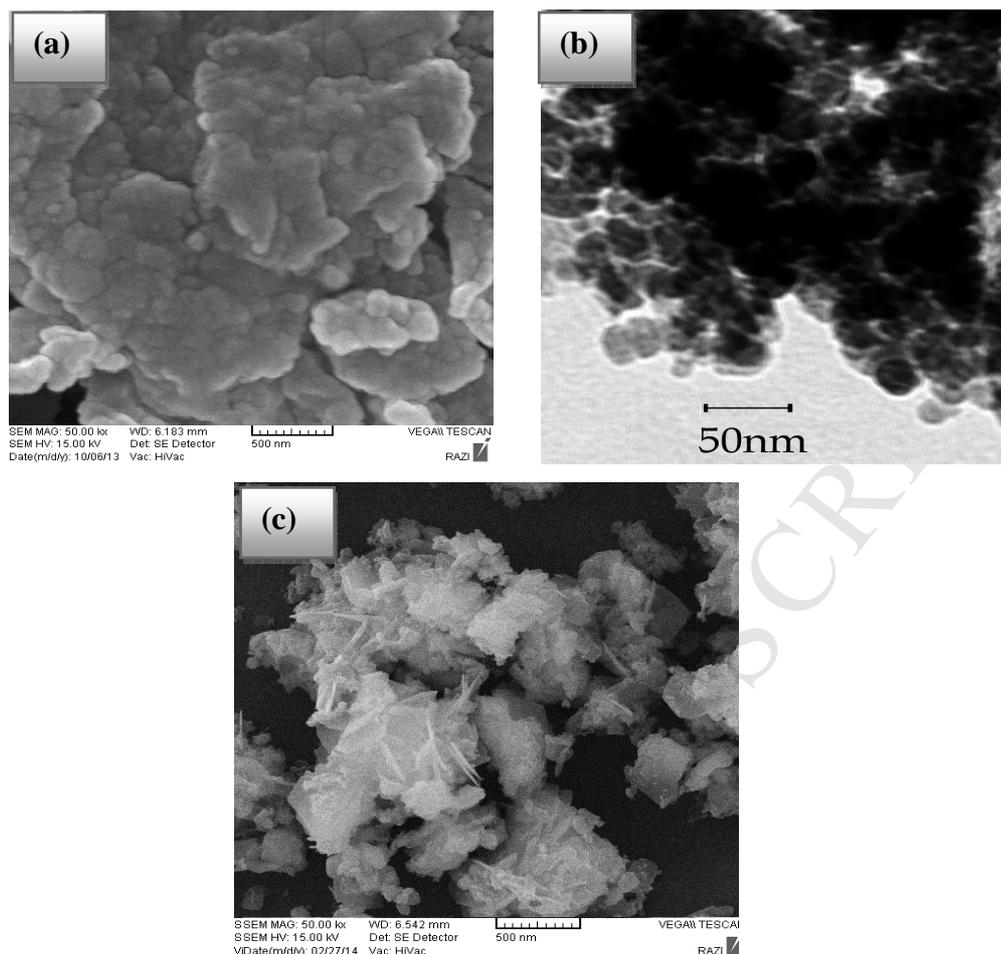
225 Fig. 3 presents the XRD-diffraction patterns of the HO₃S-SNPs. The same broad peak was
226 observed in XRD pattern of HO₃S-SNPs, indicating retention of the amorphous nature of the
227 starch nanoparticles during the sulfonation process.



228
229

Fig. 3. XRD pattern of HO₃S-SNPs.

230 Fig. 4 (a) and (b) showed SEM and TEM micrographs of starch nanoparticles, respectively.
231 SEM in Fig. 4(a) shows the surface morphology of the starch nanoparticles. TEM image in
232 Fig. 4(b) indicates that the particles are nearly spherical and they have a range diameter of 50-
233 80 nm. SEM image for HO₃S-SNPs in Fig. 4(c), shows significant difference between starch
234 nanoparticles and sulfonated starch. After sulfonation of the starch nanoparticles, the
235 aggregates were foliated with rougher surfaces.



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237

238 Fig. 4. (a) SEM and (b) TEM image of starch nanoparticles and (c) SEM of HO₃S-SNPs.

239 To investigate the feasibility of the synthetic methodology for the synthesis of
 240 benzoxanthenes, initial experiments were performed by using 2-naphthol and benzaldehyde as
 241 reactants at 110 °C under solvent-free conditions. Initially, the Starch-SO₃H was studied for its
 242 activity in the model reaction. The results showed that 0.08 g of catalyst was optimal for this
 243 reaction (Table 1, Entry 4). In a blank experiment no desired product was observed under
 244 similar reaction conditions in the absence of acidic catalyst (Table 1, Entry 1). In the next
 245 stage for getting the best conditions, we carried out a model study with the same reagent
 246 system at 90°C, 100°C, 110°C and 120°C temperatures, (Table 1, Entries, 4 and 8-11).
 247 Increasing the temperature enhances the reaction rate substantially with respective 70%, 85%,
 248 92% and 80% yield. According to the results, we obtained the best results at 110 °C, and
 249 therefore all the reactions were performed at this temperature for the synthesis of 14-aryl-14-

250 *H*-dibenzo[*a,j*]xanthenes. The reaction time was still long and needs to be improved and for
 251 this reason, HO₃S-SNPs were used in this organic reaction. It was observed that SO₃H-SNP is
 252 better than the Starch-SO₃H tested in this study in terms of reaction yield (92%) and reaction
 253 time (45 min) (Table 1, entry 13). Some solvents such as EtOH, MeOH, CH₃CN, H₂O and
 254 CH₂Cl₂, did not show improvement in the yield of product (Table 1, Entries 14-18).

255 Table 1. Results of screening the conditions.

Entry	Catalyst	Catalyst(g)	Solvent	Temp (°C)	Time (min)	Yield (%)
1	-	-	Solvent-free	110	240	-
2	Starch	0.08	Solvent-free	110	240	-
3	Starch NPs	0.08	Solvent-free	110	240	-
4	Starch-SO ₃ H	0.08	Solvent-free	110	105	90
5	Starch-SO ₃ H	0.05	Solvent-free	110	180	75
6	Starch-SO ₃ H	0.01	Solvent-free	110	210	70
7	Starch-SO ₃ H	0.1	Solvent-free	110	105	84
8	Starch-SO ₃ H	0.15	Solvent-free	110	105	81
9	Starch-SO ₃ H	0.08	Solvent-free	120	105	80
10	Starch-SO ₃ H	0.08	Solvent-free	100	180	85
11	Starch-SO ₃ H	0.08	Solvent-free	90	180	70
12	Starch-SO ₃ H	0.08	Solvent-free	r.t.	240	-
13	HO ₃ S-SNPs	0.08	Solvent-free	110	45	92
14	HO ₃ S-SNPs	0.08	EtOH	Reflux	120	65
15	HO ₃ S-SNPs	0.08	MeOH	Reflux	120	58
16	HO ₃ S-SNPs	0.08	CH ₃ CN	Reflux	120	70
17	HO ₃ S-SNPs	0.08	CH ₂ Cl ₂	Reflux	120	35
18	HO ₃ S-SNPs	0.08	H ₂ O	Reflux	120	40

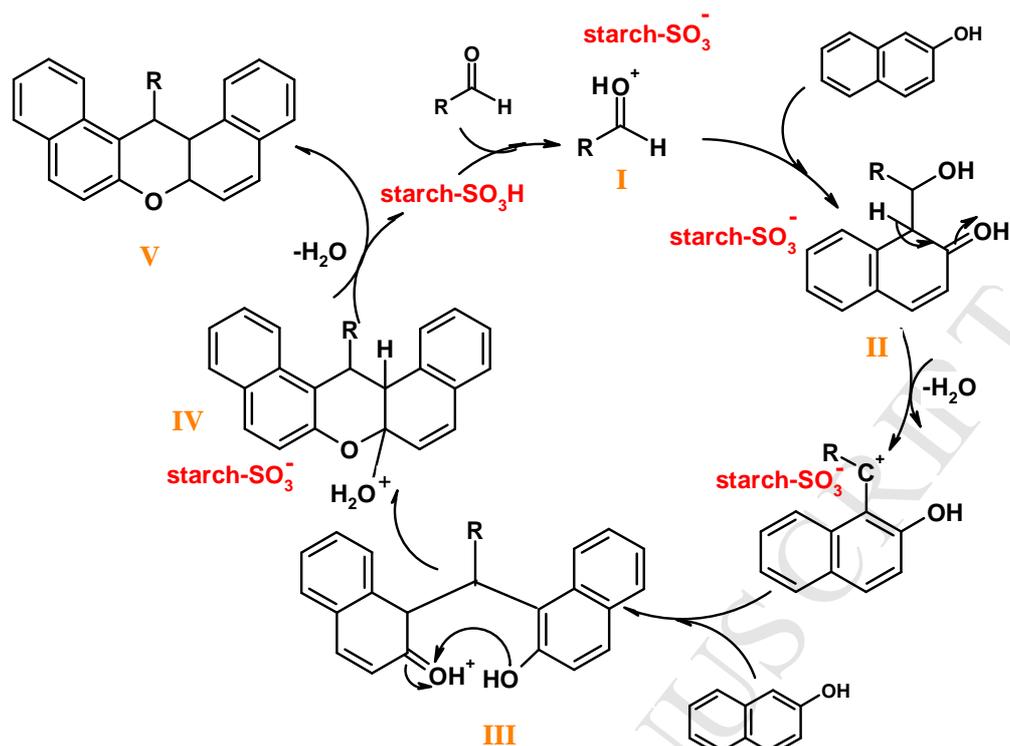
256 Under the optimized reaction conditions, a series of aldehydes with 2-naphthol were explored
 257 in the presence of HO₃S-SNPs. The results were summarized in Table 2. The procedure was
 258 highly effective for the preparation of 14-aryl-14-*H*-dibenzo[*a,j*]xanthenes. As expected, the
 259 aromatic aldehyde substrates containing electron donor groups (such as OMe and OH),
 260 showed weaker results than the cases containing electron withdrawing groups (such as NO₂,
 261 Cl).

262 Table2. Synthesis of 14-aryl-14-*H*-dibenzo[*a,j*]xanthenes using HO₃S-SNPs.

Entry	R	Product	SO ₃ H-SNPs		M.p (°C)		Ref
			Time(min)	Yield(%) ^a	(Found)	(Reported)	
1	C ₆ H ₅	3a	45	92	183-185	181-183	37
2	3-NO ₂ C ₆ H ₄	3b	40	95	208-210	211-212	38
3	4-NO ₂ C ₆ H ₄	3c	30	97	310-312	310-312	38
4	2-FC ₆ H ₄	3d	40	95	209-210	211-212	38
5	2-ClC ₆ H ₄	3e	45	92	213-215	214-215	39
6	4-ClC ₆ H ₄	3f	35	95	287	287-289	37
7	2,4-ClC ₆ H ₄	3g	30	95	252-253	254-255	40
8	4-CH ₃ C ₆ H ₄	3h	70	95	226	227-230	41
9	2-OCH ₃ C ₆ H ₄	3i	80	91	263-265	263-265	41
10	4-OCH ₃ C ₆ H ₄	3j	70	94	203-204	202-204	42
11	3-OHC ₆ H ₄	3k	70	90	241-242	240-241	42
12	4-OHC ₆ H ₄	3l	70	92	139-140	138-140	42

263 ^a Isolated yields

264 The formation of benzoxanthenes from aldehyde derivatives and 2-naphthol in the presence of
265 HO₃S-SNPs as catalyst can be explained by a tentative mechanism is presented in Scheme 3.
266 HO₃S-SNP is supposed as a Lewis acid catalyst to facilitate the activation of the aldehyde.
267 The acidic SO₃H groups distributed on the surface of starch nanoparticles in high
268 concentrations activate the carbonyl group of the aldehyde. Then, one molecule of 2-naphthol
269 condense with an activated aldehyde (I) to provide intermediate (II), which can be regarded as
270 a fast Knoevenagel addition. In the next step, the active methylene of the second molecule of
271 2-naphthol react with intermediate (II) *via* conjugate Michael addition to produce the
272 intermediate (III), which undergoes intramolecular cyclodehydration to furnish the desired 14-
273 aryl-14-*H*-dibenzo[*a,j*]xanthenes (V) [42, 43].



274

275

Scheme.3: Proposed reaction mechanism.

276 To have a better understanding of the performance of our catalytic system, the effectiveness
 277 of the $\text{SO}_3\text{H-SNPs}$ was also compared to that of catalysts reported previously in the synthesis
 278 of 14-aryl-14-*H*-dibenzo[*a,j*]xanthenes from the reaction of 2-naphthol and various aromatic
 279 aldehydes and the results are listed in Table 3. As can be seen in this table, the present catalyst
 280 was found to be the most efficient catalyst among all of the catalysts in the literatures. The
 281 reaction in the presence of $\text{HO}_3\text{S-SNPs}$ was indicated a lot of significant such as; simplicity of
 282 the reaction, excellent yields, low reaction times, naturally safe, biodegradable, economical,
 283 and eco-friendly catalyst. These advantages can be related to the highly activity due to its
 284 nanoporosity and high surface area.

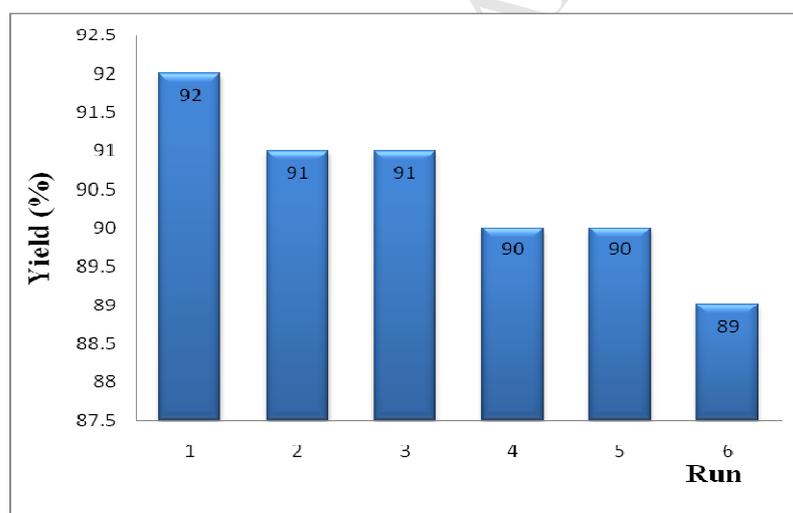
285 Table 3: The synthesis of 3a using different catalysts.

Entry	Catalyst	Time (min)	Yield(%) ^a	Temp (°C)	Ref.
1	$\text{HO}_3\text{S-SNPs}$	45	92	110	This research
2	Dowex-50 W	160	84	100	22
3	TCCA	50	74	110	44

4	Iodine	18	85	90	45
5	H ₅ PW ₁₀ V ₂ O ₄₀	60	92	100	46
6	H ₂ SO ₄ /SiO ₂	60	97	125	47
7	PFPAT	189	96	r.t.	48
8	Cellulose sulfuric acid	120	90	110	49
9	SBA-15-SO ₃ H	24h	91	85	50
10	Sulfamic acid	8h	93	125	20
11	AgBr	90	50	140	41
12	AgI	70	70	140	41
13	CuCl	120	40	140	41
14	NiO	140	45	140	41
15	CaO	160	50	140	41
16	AgI-nano	40	92	140	41
17	SBA-15-CO ₂ H	6h	80	r.t.	43
18	MCM-41-SO ₃ H	60	90	r.t.	43

286 ^a Isolated yields

287 The reusability of the HO₃S-SNPs was tested for the model reaction. The separated catalyst
 288 was used several times and loss of catalytic activity was not observed (Fig. 5).



289

290

Fig. 5. Recoverability of HO₃S-SNPs.

291 Conclusions

292 In summary, the present work was the first report on using HO₃S-SNPs as a novel and very
 293 promising solid acid catalyst for the synthesis of 14-aryl-14-*H*-dibenzo[a,j]xanthenes. The
 294 application of starch-SO₃H as a highly efficient, inexpensive, easy work-up, and natural solid
 295 acid catalyst makes the present methodology economically and eco-friendly acceptable.

296 Moreover, solvent free, low-cost, non-toxicity, high yields of the desired benzoxanthenes and
297 short reaction times are supporting the protocol toward the green chemistry. Further
298 applications of these biopolymer catalysts are currently under investigation in our laboratory.

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301 of Kashan for supporting this work by Grant No. (363022/16).

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471

472 Figure captions

473 Table 1. Results of screening the conditions.

474 Table 2. Synthesis of 14-aryl-14-*H*-dibenzo[*a,j*]xanthenes using SO₃H-SNPs.

475 Table 3: The synthesis of 3a using different catalysts.

476 Scheme 1. Synthesis of 14-aryl-14-*H*-dibenzo[*a,i*]xanthenes using SO₃H-SNPs as catalyst.

477 Scheme 2: Preparation steps for fabricating SO₃H-SNPs.

478 Scheme 3: Proposed reaction mechanism.

479 Fig. 1: FT-IR of starch nanoparticles.

480 Fig. 2. XRD patterns of (a) starch and (b) starch nanoparticles.

481 Fig. 3. XRD pattern of SO₃H-SNPs.

482 Fig. 4. (a) SEM and (b) TEM image of starch nanoparticles and (c) SEM of SO₃H-SNPs.

483 Fig. 5. Recoverability of SO₃H-SNPs.

