

QUATERNARY SUBSTITUTED PDE IV INHIBITORS II : THE SYNTHESIS AND IN VITRO EVALUATION OF A NOVEL SERIES OF γ-LACTAMS

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Abstract: This communication describes the synthesis and in vitro evaluation of a novel potent series of phosphodiesterase type (IV) (PDE IV) inhibitors. Several of the quaternary substituted lactams presented possess low nanomolar IC₅₀'s for PDE IV inhibition. © 1998 Elsevier Science Ltd. All rights reserved.

PDE IV, the major PDE isozyme found within inflammatory cells, regulates levels of the secondary messenger cyclic adenosine monophosphate (cAMP) via hydrolysis to its acyclic analogue 5'-adenosine monophosphate (5'-AMP).¹ The pivotal role that increased cAMP levels play in retarding production of the proinflammatory cytokine tumor necrosis factor (TNF- α)² has pushed the cAMP specific PDE IV to the forefront of molecular targets for the potential treatment of autoimmune diseases. Early inhibitors of PDE IV include the antidepressant rolipram 1 and analogs thereof.³ Of more recent note are the 3,4-disubstituted pyrrolidine derivatives 4 reported by Stafford et al.⁴ and the Celltech inhibitor CDP 840.⁵ The latter has been shown to possess selectivity for the catalytic binding site over the rolipram binding site, a highly desirable property for overcoming the emetic side effect often seen with potent PDE IV inhibitors.⁶ This letter reports the synthesis and PDE IV inhibitory activity of a novel series of quaternary substituted γ -lactams with the general structure 2.



The structural resemblance of 2 to the series of quaternary substituted oxindoles 3, originally designed as conformationally constrained analogs of CDP 840 and reported in a earlier communication,⁷ is clear. From a design aspect, we hoped to increase the PDE IV potency of the latter series by maintaining the novel spirocyclic feature, while accessing compounds which were similar to the pyrrolidine series 4.

The following synthetic scheme was therefore developed to access lactams of type 2 amenable to straightforward SAR studies at R_1 and R_2 . Commercially available 3-hydroxy-4-methoxyphenylacetic acid was methylated giving 5 and alkylated with cyclopentyl bromide to afford the rolipram-like 3-methoxy-4-cyclopentoxy moiety 6. Treatment with base and bromoacetonitrile produced 7, followed by reduction with Raney Nickel to access the desired lactam core structure 8.⁸ BOC protection of the lactam nitrogen proved facile

and subsequent C-alkylation with 4-picolyl chloride gave 10 in modest yield. Ring opened products 11 and 12, presumably formed via cleavage by adventitious NaOH, were also detected in the reaction mixture. BOC removal and reaction with a variety of alkylating/acylating/sulfonylating agents (R_2CI) gave final products 16 to 28. *N*-oxide formation proceeded smoothly with *m*-CPBA giving 29 and 30 respectively. Reduction of the lactam 13 and acylation afforded products 14 and 15, respectively.



Reagents & conditions :- (i) HCl (g), MeOH, rt, 20 h, 95%. (ii) cyclopentylbromide (1.5 equiv), K_2CO_3 (1.5 equiv), DMF, 70 °C, 20 h, 56%. (iii) KHMDS (1 equiv), dimethoxyethane, -78 °C, then BrCH₂CN (1 equiv), 3 h, -78 °C, mono-alkylated product (17%), di-alkylated product (39%). (iv) Ra-Ni (xs), MeOH, H₂, 50 °C, 20 h, 57%. (v) (BOC)₂O (1.2 equiv), Et₃N (1.2 equiv), DMAP (cat), rt, 20 h, 63%. (vi) 4-picolyl chloride (3 equiv), NaH (3 equiv), DMF, rt, 20 h, 48%. (vii) TFA, CH₂Cl₂, rt, 2 h, 90%. (viii) R₂Cl, NaH (1.1 equiv), DMF, 60-90%. (ix) *m*-CPBA (1 equiv), CH₂Cl₂, rt, 4 h, 90%. (x) Red-Al, toluene, 80 °C, 2 h. (xi) BOC₂O, Et₃N, DMAP (cat), CH₂Cl₂, rt, 3 h, 40% for (x) and (xi) combined.

Compounds were evaluated for PDE IV inhibition (IC₅₀, nM) and rolipram displacement (K_i, nM) using the methods reported by Thompson and Schmeichen respectively.⁹ The results are summarized in Table 1. Weak signs of PDE IV activity were observed at an early stage with the evaluation of intermediates 8 (IC₅₀ (PDE IV) 5000 nM, K_i (rolipram binding) > 3745 nM) and 9 (IC₅₀ 700 nM, K_i 337 nM). This immediately highlighted the importance of the BOC group, which provided almost a ten fold increase in PDE IV potency. A similar occurrence was previously observed in the pyrrolidine series 4 by Stafford et al.⁴ Formation of the quarternary

X X N General Structure					
Cmpd #	X,Y	Ri	R ₂	PDEIV IC ₅₀ nM	K _i rolipram bind. nM
8	0	н	н	5000	>3745
9	0	н	CO ₂ (CH ₃) ₃	700	337
10	0	ᡃᡗᢕᢅ᠉	CO ₂ (CH ₃) ₃	6.3	7.5
13	0	J~U_N	н	800	210
15	н,н	ᡃᠵᠧᢅ᠉	CO ₂ (CH ₃) ₃	807	470
16	0	٦ <u>)</u> ۲	CO ₂ CH(CH ₃) ₂	44	113
17	0	r_{n}	CO ₂ CH ₂ CH ₃	137	139
18	0	J CN	CO₂CH₃	293	822
19	0	L.N	∿℃	56	51
20	0	J ()	J T.N	401	443
21	0	57 (°N	SO₂C ₆ H₄p-CH₃	61	217
22	ο	r Co	CON(CH ₃) ₂	319	274
23	0	57(N	COCH ₃	461	367
24	0	5 L N	ب ۲۰۰۰	123	137
25	0	5 L.N	- بم بم	81	80
26	0	م کر تر م	50	48	43
27	0	ς Υ.»		17	4.3
28	0	r L.»	20	293	822
29	0	ᡃᠧ ᡘ ᠄	н	1600	2247
30	0	h LN.O.	CO ₂ (CH ₃) ₃	59	217
31	0	50	н	1000	787
32	ο	50	CO ₂ (CH ₃) ₃	220	2996
33	<u>o</u>	ч~см	CO ₂ (CH ₃) ₃	82	476

centre via alkylation of 9 with picolyl chloride giving 10 produced an excellent 100-fold increase in potency (IC₅₀ 6.3 nM, K_i 7 nM).¹⁰ However, neither of the two ring opened side products 11 and 12 possessed PDE IV activity.

The high potency of the γ -lactam 10, when compared to its analogue in the related oxindole series (For 3 where R₁= 4-CH₂py, R₂ = BOC, IC₅₀ (PDE IV) 700 nM, K_i (rolipram binding) 2800 nM), exemplifies the dramatic improvement in PDE IV activity on removal of the fused phenyl ring.⁷ BOC removal from 10 to give 13 resulted, as expected, in a corresponding drop in activity (IC₅₀ 800 nM, K_i 210 nM). Reduction of lactam 13 to the corresponding pyrrolidine 14 and BOC protection to give 15 showed the existence of an important lactam carbonyl interaction with a >100-fold drop in PDE IV activity (IC₅₀ 800 nM, K_i 470 nM). Interesting decreases in PDE IV activity on going from butyloxy-isopropyloxy-ethoxy-methoxy 10, 16, 17, 18 clearly indicates a preference for larger lipophilic groups in this region of space. A further drop in activity was also observed for the N-acetyl analog 23 (IC₅₀ 461 nM, K_i 367 nM) and the N-dimethyl urea derivative 22 (IC₅₀ 319 nM, K_i 274 nM). The benzyl derivative 19 showed it was possible to deviate from carbamate protection while still maintaining good PDE IV potency (IC₅₀ 56 nM, K_i 51 nM). A preference for such bulky non polar groups in this position could again be seen via the subsequent drop in activity for the analogous and more polar 4-pyridyl derivative 20 (IC₅₀ 401 nM, K_i 403 nM). Good activity was also observed for the sulfonamide 21 (IC₅₀ 61 nM, K_i 217 nM). Based on these observations a variety of lipophilic groups were investigated for their PDE IV potency at R₂. Once again increasing the size and lipophilicity of the substituents 24, 25, 26 from butyl - isopentyl methylcyclohexyl gave gradual increases in PDE IV potency. A similar trend was observed with affinity for the rolipram binding site. The 2-napthyl derivative 27¹¹ possessed excellent PDE IV inhibitory activity (IC₅₀ 17 nM) with the highest recorded rolipram binding affinity in the series (K_i 4.3 nM). Interestingly, the 1-napthyl analog 28 showed a drop in PDE IV activity (IC₅₀ 293 nM) and an even greater reduction in rolipram binding affinity (K_i 822 nM). This perhaps suggests a definite shape to the binding pocket that exists around the lactam nitrogen. Encouraged with the potent inhibitors 10 and 27, a brief SAR study was undertaken around R_1 , maintaining R_2 as t-butyloxycarbonyl. A drop in activity was observed for the N-oxide derivative 30 (IC₅₀ 59 nM, K_i 217 nM). Removal of the pyridyl nitrogen 32 showed a 30-fold decrease in activity (IC₅₀ 220 nM, K_i 2996 nM) when compared to 10. This compound showed highest selectivity (ten fold greater) for the catalytic over the rolipram binding site in the above series. Again BOC removal for the latter two compounds giving 29 (IC₅₀ 1600 nM, K_i 2247 nM) and 31 (IC₅₀ 1000 nM, K_i 787 nM) lead to dramatic decreases in PDE IV activity. Interestingly it was possible to replace the pyridyl functionality with a nitrile group 33 and still maintain good PDE IV activity (IC_{50}) 82 nM, K_i 476 nM). Related nitrile functionality at a similarly positioned quarternary centre has been previously reported by Barnette et al in their series of cyanocyclohexylcarboxylic acid PDE IV inhibitors.¹²

In summary, a novel series of potent spirocyclic γ -lactam PDE IV inhibitors has been discovered with improved PDE IV activity over the spirocyclic oxindole series reported earlier.⁷ A sterically large lipophilic group at R₂ gives optimal PDE IV activity, exemplified by inhibitors 10 and 27. The lactam carbonyl also plays a key role in maintaining PDE IV activity. The pyrrolidine 15 showed a ten fold decrease in PDE IV potency.

Substitution at R_1 is a definite prerequisite for PDE IV activity with 4-picolyl substitution proving most advantageous. Several compounds possessed selectivity for the catalytic over the rolipram binding site.¹³ Most encouraging was the 10-fold selectivity observed for compound **32**. In vivo studies and emesis data will be reported in the near future.

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References and Notes

- (a) For a review see : Palfreyman, M. N.; Souness, J. E. Progress in Medicinal Chemistry, 1996, 33, 1.
 (b) Butcher, R. W.; Sutherland, E. W. J. Biol. Chem. 1962, 237, 1244. (c) Conti, M. and Swinnen, J. V. Cyclic Nucleotide Phosphodiesterases: Structure, Regulation and Drug Action, 1990, 244-266.
- (a) Feldmann, M.; Brennan, F. M.; Elliot, M.; Katsikis, P.; Maini, R. N. Circulatory Shock, 1994, 43, 179. (b) Williams, R. D.; Feldmann, M.; Maini, R. N. Proc. Natl. Acad. Sci. USA 1992, 89, 9783. (c) Tracey, K. J.; Cerami, A. Annu. Rev. Cell. Biol. 9, 317-343. (d) Ochalski, S. J.; Hartman, D. A.; Belfast, M. T.; Walter, T. L.; Glaser, K. B.; Carlson, R. P., Agents Actions, 1993, 39, C52-C54. (e) Everitt, D. E.; Boike, S.; Davies, B.; Zariffa, N.; Audet, P.; Brown, L.; Freed, M.; Ilson, B.; Esser, K.; O'Leary, Bartus, J.; Slack, G.; Smith, G.; Jordasky, D. Clin. Pharmacol. Ther. 1994, 55, 150.
- (a) Schwabe, U.; Miyake, M.; Ohga, Y.; Daly, J. W. Mol. Pharmacol. 1976, 12, 900. (b) Marivet,
 M. C.; Bourguignon, J-J.; Lugnier, C.; Mann, A.; Stoclet, J-C.; Wermuth, C-G. J. Med. Chem. 1989, 32,
 1450. (c) Fenton, G.; Mason, J. S.; Palfreyman, M. N.; Ratcliffe, A. J. W.O. Pat. 9427947, 1994. (d)
 Fenton, G.; Mason, J. S.; Palfreyman, M. N.; Ratcliffe, A. J. W.O. Pat. 9318024, 1993. (e) Bender, P. E.;
 Christensen, S. B.; Esser, K. M.; Forster, C. J.; Ryan, M. D.; Simon, P. L. W.O. Pat. 9200968, 1992.
- Stafford, J. A.; Valvano, N. L.; Feldman, E.; Brawley, E. S.; Cowan, D. J.; Domanico, P. L.; Leesnitzer, M. A.; Rose, D. A.; Stimpson, S. A.; Strickland, A. B.; Unwalla, R.J.; Verghese, M. W. *Bioorg. Med. Chem. Lett.* 1995, 5, 1977.
- (a) Warrellow, G. J.; Alexander, R. P.; Boyd, E. C.; Eaton, M. A.; Head, J. C.; Higgs, G. A. The design of an orally active PDE IV inhibitor (CDP 840) for asthma. 8th RSC-SCI Medicinal Chemistry Symposium, Cambridge, U.K. 1995. (b) Hughes, B. CDP 840 2nd World Congress on Inflammation, Brighton, U.K. 1995.
- (a) Eban, E.; Ruther, E. Pharmacopsychiatry, 1985, 18, 69. (b) Carpenter, D. O.; Briggs, D. B.;
 Knox, A. P. J. Neurophysiol. 1988, 59, 358. (c) Israel, E.; Mathur, P. N.; Tachkin, D.; Drazen, J. M. Chest, 1988, 94, 71S.

- 7. Hulme, C.; Poli, G.; Fu-Chih Huang, F.-C.; Souness, J.; Djuric, S. *Bioorg. Med. Chem. Lett.* 1998, in press.
- (a) Winans, C. F.; Adkins, H. J. Am. Chem. Soc. 1933?, 55, 4167. (b) Altomare, C.; Carotti, A.; Casini, G.; Cellamore, S.; Ferappi, M.; Vitali, C. Arzneim.-Forsch, 1992, 42, 152-155.
- Batches of PDE IV were obtained from guinea-pig macrophages. See Turner, N. C.; Wood, L. J.; Burns, F. M.; Guermy, T.; Souness, J. E. Br. J. Pharmacol. 1993, 108, 876. PDE IV IC₅₀'s were determined in macrophage homogenates via a two step radioisotopic method. See Thompson, W. J.; Teraski, W.; Epstein, P. M.; Strada, S. J. Adv. Cyclic Nucleotide Res., 1979, 10, 69. K_i values were determined using [³H] rolipram in a guinea pig brain membrane binding assay. See Schmeichen, R.; Schneider, H. H.; Watchtel, H. Psychopharmacology, 1990, 102, 17-20.
- 10. Compound 10 : 3-(3-Cyclopentyloxy-4-methoxy-phenyl)-2-oxo-3-pyridin-4-ylmethyl-pyrrolidine-1 carboxylic acid *tert*-butyl ester, mp 148-149 °C, white solid. For C₂₇H₃₄O₅N₂ calcd C 69.51 H 7.34 N 6.0. Found C 69.37 H 7.33 N 5.86. MS (FAB). Exact mass measurement- Theoretical (M+H⁺) 467.2546, measured 467.2560. ¹H (CDCl₃) 1.5 (9H, s, 3x CH₃), 1.55-1.60, 1.75-1.95 (8H, 2xm, cyclopentyl), 2.0-2.1, 2.3-2.4, 3.35-3.45, 3.6-3.7 (4H, 3xm, CH₂CH₂), 3.05-3.25 (2H, s, CH₂py), 3.8 (3H, s, OCH₃), 4.7-4.75 (1H, m, CH), 6.75-6.80, 7.0 (5H, m, s, C₆H₃, 2 from C₃H₄N), 8.35-8.40 (2H, br m, 2 from C₃H₄N).
- Compound 17 : 3-(3-Cyclopentyloxy-4-methoxy-phenyl)-naphthalen-2-ylmethyl-3-pyridin-4-ylmethylpyrrolidin-2-one, an oil. MS (FAB) 507 (MH⁺), 414. Exact mass measurement- Theoretical (M+H⁺) 507.2648, measured 507.2679. ¹H (CDCl₃)1.55-1.65, 1.75-2.0 (8H, 3xm cyclopentyl), 2.05-2.2, 2.25-2.4, 2.9-3.0, 3.05-3.1 (4H, m, CH₂CH₂), 3.15-3.30 (2H, m, CH₂py), 3.7 (3H, s, OCH₃), 4.75-4.80 (1H, m, CH), 4.5-4.7 (2H, m, CH₂napth), 6.75-6.8, 6.85-6.9, 6.9-6.95, 7.1-7.2, 7.4-7.5, 7.6-7.65, 7.65-7.7, 7.75-7.8 (12H, 8xm, C₆H₃, 2 from C₅H₄N, C₁₀H₇), 8.35-8.45 (2H, br m, 2 from C₅H₄N).
- (a) Christensen, S. B. W.O. Pat. 9319749, 1993. (b) Barnette, M. S.; Christensen, S. B.; Essayan, D. M.;
 Esser, K. M.; Grous, M.; Huang, S-K.; Manning, C. D.; Prabhaker, U.; Rush, J.; Torphy, T. J. Am. J.
 Resp. Crit. Care Med. 1994, 149, Suppl. A209.
- For the relationship between the inhibition constant (K_i) and IC₅₀ of an enzymatic reaction see: Cheng,
 Y-C; Prusoff, W. H. *Biochem. Pharmacol.* 1973, 22, 3099.