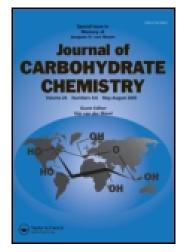
This article was downloaded by: [North Dakota State University]

On: 19 October 2014, At: 12:29

Publisher: Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information:

http://www.tandfonline.com/loi/lcar20

Easy Chemo-Enzymatic Synthesis of Human Milk Trisaccharides from a Common Selectively Protected Lactose Building Block

Barbara La Ferla , Luigi Lay , Laura Poletti , Giovanni Russo & Luigi Panza

- ^a Università degli Studi di Milano, Dipartimento di Chimica Organica e Industriale, Centro di Studio sulle Sostanze Organiche Naturali del CNR, via G. Venezian, 21 20133 Milano, (Italy)
- ^b Università degli Studi di Milano, Dipartimento di Chimica Organica e Industriale, Centro di Studio sulle Sostanze Organiche Naturali del CNR, via G. Venezian, 21 20133 Milano, (Italy)
- ^c Università degli Studi di Milano, Dipartimento di Chimica Organica e Industriale, Centro di Studio sulle Sostanze Organiche Naturali del CNR, via G. Venezian, 21 20133 Milano, (Italy)
- ^d Università degli Studi di Milano, Dipartimento di Chimica Organica e Industriale, Centro di Studio sulle Sostanze Organiche Naturali del CNR, via G. Venezian, 21 20133 Milano, (Italy)
- Università degli Studi del Piemonte Orientale, Facoltà di Farmacia, Dipartimento di Scienze Mediche, Viale Ferrucci, 33 - 28100 Novara, (Italy)

Published online: 27 Feb 2008.

To cite this article: Barbara La Ferla , Luigi Lay , Laura Poletti , Giovanni Russo & Luigi Panza (2000) Easy Chemo-Enzymatic Synthesis of Human Milk Trisaccharides from a Common Selectively Protected Lactose Building Block, Journal of Carbohydrate Chemistry, 19:3, 331-343, DOI: 10.1080/07328300008544082

To link to this article: http://dx.doi.org/10.1080/07328300008544082

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms & Conditions of access and use can be found at http://www.tandfonline.com/page/terms-and-conditions

EASY CHEMO-ENZYMATIC SYNTHESIS OF HUMAN MILK TRISACCHARIDES FROM A COMMON SELECTIVELY PROTECTED LACTOSE BUILDING BLOCK

Barbara La Ferla, Luigi Lay, Laura Poletti, Giovanni Russo, and Luigi Panzaba

*Università degli Studi di Milano, Dipartimento di Chimica Organica e Industriale, Centro di Studio sulle Sostanze Organiche Naturali del CNR, via G. Venezian, 21 20133 Milano (Italy)

bUniversità degli Studi del Piemonte Orientale, Facoltà di Farmacia, Dipartimento di Scienze Mediche, Viale Ferrucci, 33 – 28100 Novara (Italy)

Received September 8, 1999 - Final Form February 3, 2000

ABSTRACT

Three glycosyllactosides, contained in the neutral fraction of human milk oligosaccharides, were synthesised in a simple and straightforward manner through a sequence based on a chemo-enzymatic approach. Lipase catalysed regioselective 6'-O-acylation of benzyl β-lactoside, followed by the introduction of an isopropylidene group and acetylation afforded, depending on the reaction conditions, compounds 4a and 4b, which allow selective access to positions 3, 3' and 6'. Glycosylation with proper donors gave trisaccharides 6, 9 and 12.

INTRODUCTION

In recent years there has been a growing interest in the use of lipases and proteases both for hydrolysis and transesterification reactions as a tool for protection and deprotection of carbohydrates. In many cases the enzymatic method showed noteworthy

regioselectivity, often offering much simpler experimental conditions with respect to the corresponding regioselective chemical acylation (if the latter is feasible at all).

Despite the large number of examples of such reactions on different mono- and disaccharides published so far,²⁻⁶ the enzymatically protected sugars have rarely been utilised for the synthesis of more complex derivatives.

We recently gained interest in human milk oligosaccharides, when it has been demonstrated that these compounds, most of which are unique to the human species, possess antiadhesive properties and inhibitory activity towards many pathogens, representing a protection against infections for breast-fed infants during the lactation period. It could then be of great importance to have access to such compounds to be used as additives in artificial milk for infants or aged people.

However, it is still unclear which of the above oligosaccharides exert this function. It will therefore be useful to have a flexible method for an easy access to different oligosaccharidic structures from common building blocks. We recently published a study on regioselective enzymatic acylation of common, commercially available disaccharides such as cellobiose, maltose and lactose. As lactose is the most abundant component and a fundamental constituent of almost all the milk oligosaccharidic structures characterised so far, we focussed our attention on benzyl lactoside as a substrate for enzymatic acylation, showing that it is possible to selectively and sequentially acylate the hydroxy groups in position 6' and 2' with two different esters.

Exploiting some of the results previously described, we planned a chemoenzymatic approach aimed at obtaining versatile building blocks for the preparation of some human milk oligosaccharidic structures. The philosophy of such an approach consists in exploring a way to generate a family of oligosaccharides for biological testing in a reasonable amount of time instead of optimisation of a synthetic scheme for the preparation of specific target molecules. Using an appropriate pattern of protecting groups, introduced both with chemical and enzymatic methods, it is possible in principle to have selective access to each hydroxy group of lactose. As an example of such a strategy, we wish now to describe the synthesis of three lactose derived trisaccharides contained in the neutral fraction of human milk oligosaccharides. Antiadhesion studies on these compounds, after their complete deprotection, will be reported in due course.

RESULTS AND DISCUSSION

As mentioned above, we reported the highly regioselective 6'-O-acylation of benzyl β -lactoside 1 using lipase from Candida antarctica. The reaction was carried out in tert-amyl alcohol with trifluoroethyl chloroacetate as acylating agent and afforded benzyl 6'-O-chloroacetyl- β -lactoside 2 in 81% yield. The choice of tert-amyl alcohol was determined by the necessity to solubilize the substrate, but the high boiling point of this solvent makes the work-up of the reaction and the purification of the product a laborious task, especially in light of a possible large scale synthesis. These difficulties and the observation of the activity of these lipases in other solvents, suggested that we perform the reaction under different conditions. Using THF as solvent we obtained the desired selectively acylated benzyl 6'-O-chloroacetyl- β -lactoside 2 in comparable yield (78%) and with a much easier procedure for the recovery of the product (Scheme 1).

Scheme 1

Positions 3',4' of compound 2 were then protected by the introduction of an isopropylidene group; subsequent acetylation of compound 3 afforded the triacetylated compound 4a and the tetraacetylated compound 4b, depending on the reaction conditions. As a matter of fact, when acetyl chloride and 2,6-di-tert-butyl-4-methyl-

pyridine were used, an easily separable mixture of compounds 4a (45%) and 4b (39%) was obtained. In contrast, carrying out the reaction with acetyl chloride in the presence of sym-collidine gave the fully acetylated product, and compound 4b was recovered in 90% yield. Compounds 4a and 4b give selective access to positions 3, 3' and 6' (Scheme 1).

Scheme 2

Protected 3-O-fucosyl lactose 6 (Scheme 2) was obtained in excellent yield (99%) by fucosylation of compound 4a with donor 5, ¹⁰ using TMSOTf (0.01 eq) in CH₂Cl₂ at 0 °C. The newly synthesized glycosidic linkage was confirmed as having an α configuration by ¹H NMR spectroscopy; the signal corresponding to H-1" appeared as a doublet at 5.31 ppm ($J_{1,2} = 4.0$ Hz), as expected for a 1,2-cis linkage.

Chemoselective removal of the 6'-O-chloroacetyl group from compound 4b (DABCO, 87% yield) gave compound 7 which was then glycosylated with galactosyl trichloroacetimidate 8^{11} to afford the protected 6'-O-galactosyl lactose 9 (Scheme 2, 71% yield). The ¹H NMR spectrum of trisaccharide 9 showed a doublet at 4.64 ppm ($J_{1,2} = 7.9$ Hz), confirming the β configuration of the newly formed glycosidic linkage. Finally, hydrolysis of the isopropylidene group on compound 4b, followed by regioselective 4'-

N.,

O-acetylation, led to acceptor 10 (Scheme 3). Glycosylation of 10 with donor 8 gave unexpectedly poor yields of the trisaccharide, in spite of many attempts with different Lewis acids (TMSOTF or BF₃·OEt₂, from -20 to 0 °C). Using the more reactive donor 11,¹² trisaccharide 12 was obtained. However, the presence of an inseparable by-product prevented compound 12 from being isolated in pure enough form for full characterization. Removal of the chloroacetyl group from 12 afforded trisaccharide 13 (29% overall yield from compound 10). The structure of trisaccharide 13 was ascertained by ¹H NMR spectroscopy; H-1" showed a doublet at 4.48 ppm ($J_{1,2} = 7.8$ Hz), as expected for a 1,2-trans (β) linkage.

Scheme 3

The lipase catalyzed regioselective acylation will be further investigated on disaccharides others than lactose, in order to obtain building blocks suitable for the synthesis of higher human milk oligosaccharides.

EXPERIMENTAL

General methods. Melting points are uncorrected. Optical rotations were measured with a Perkin Elmer 241 digital polarimeter. NMR spectra were recorded on Varian XL200 (200 MHz for ¹H and 50.29 MHz for ¹³C) and Bruker AC300 (300 MHz for ¹H and 75.44 MHz for ¹³C) spectrometers. Chemical shifts are expressed in parts per million downfield from TMS. In ¹³C NMR spectra description, the signals corresponding to aromatic carbons are omitted. Reactions were followed on TLC using silica gel 60F₂₅₄

(E. Merck); flash column chromatography was performed on silica gel 60 (0.040-0.063 mm, E. Merck). Lipase from *Candida antarctica* was purchased from Boehringer Mannheim (Chirazyme® L-2, c.-f. C2 lyo).

Benzyl (6-O-chloroacetyl-β-D-galactopyranosyl)-(1->4)-β-D-glucopyranoside (2). Benzyl β-D-lactoside 1¹³ (1.57 g, 3.63 mmol) was suspended in dry THF (100 mL). Vinyl chloroacetate (2 mL, 19.75 mmol) and lipase from Candida antarctica (1 g) were added, the suspension was stirred mechanically for 48 h at rt and monitored by TLC (EtOAc/MeOH/H2O 8:1.5:0.5 v/v). The enzyme was filtered, and the solvent was removed under reduced pressure; purification by flash chromatography (eluent ethyl acetate/methanol 10:1.5 v/v) afforded compound 2 as a white solid (1.44 g, 78% yield): mp 71-75 °C; [α]_D -10.1° (c 1.1, MeOH); ¹H NMR (300 MHz, CD₃OD) δ 3.33 (m, 1H, H-5'), 3.43 (m, 1H, H-5), 3.46-3.59 (m, 4H, H-2, H-3, H-4, H-2'), 3.85 (dd, 1H, $J_{6a,5} =$ 4.2 Hz, $J_{6a.6b} = 12.5$ Hz, H-6a), 3.86-3.94 (m, 2H, H-3', H-4'), 3.93 (dd, 1H, $J_{5.6b} = 2.3$ Hz, H-6b), 4.24 (d, 1H, J = 15.5 Hz, CHCl), 4.30 (d, 1H, CHCl), 4.34 (dd, 1H, $J_{5.6} = 4.3$ Hz, $J_{6'a,6'b} = 11.4$ Hz, H-6'a), 4.37 (d, 1H, $J_{1',2'} = 8.1$ Hz, H-1'), 4.39 (d, 1H, $J_{1,2} = 8.0$ Hz, H-1), 4.41 (dd, 1H, $J_{5,6'b} = 8.5$ Hz, H-6'b), 4.67 (d, 1H, J = 11.8 Hz, CHPh), 4.91 (d, 1H, CHPh), 7.26-7.48 (m, 5H, H_{Ar}); ¹³C NMR (50.29 MHz, CD₃OD) δ 41.85 (t, CH₂Cl), 62.15, 66.24, 71.90 (3t, C-6, C-6', CH₂Ph), 70.10, 72.24, 74.10, 74.61, 74.85, 76.39, 76.39, 81.95 (8d, C-2, C-3, C-4, C-5, C-2', C-3', C-4', C-5'), 103.2, 105.3 (2d, C-1, C-1'), 169.2 (s, CO).

Anal. Calcd for $C_{21}H_{29}O_{12}Cl$ (508.91): C, 49.56; H, 5.74; Cl, 6.97. Found: C, 49.50; H, 5.77; Cl, 6.92.

Benzyl (6-O-chloroacetyl-3,4-O-isopropylidene-β-D-galactopyranosyl)-(1 \rightarrow 4)-β-D-glucopyranoside (3). Compound 2 (620 mg, 1.21 mmol) was dissolved in dry CH₃CN (7 mL) under an inert atmosphere, then dimethoxypropane (451 μL, 3 equiv) and a catalytic amount of CSA was added. The reaction was monitored by TLC (eluent ethyl acetate/methanol 10:1.5 v/v); after 1 h the solution was neutralized with NaHCO₃ and the solvent was removed under reduced pressure. Chromatographic purification (eluent ethyl acetate) afforded compound 3 as a white foam (584 mg, 88% yield); $[\alpha]_D$ +7.7° (c 1.0, MeOH); ¹H NMR (300 MHz, CD₃OD) δ 1.34, 1.50 (2s, 6H, C(CH₃)₂), 3.32 (m, 1H, H-5'), 3.43 (m, 1H, H-5), 3.43-3.57 (m, 4H, H-2, H-3, H-4, H-2'), 3.82 (dd, 1H, J_{6a,5} = 4.1 Hz, J_{6a,6b} = 12.0 Hz, H-6a), 3.91 (dd, 1H, J_{5,6b} = 2.1 Hz, H-6b), 4.08 (bt, 1H, J_{2',3'} = J_{3',4'}

= 6.1 Hz, H-3'), 4.21-4.25 (m, 2H, H-4', H-6'a), 4.29 (s, 2H, CH₂Cl), 4.37 (d, 1H, $J_{1',2'}$ = 8.1 Hz, H-1'), 4.39 (d, 1H, $J_{1,2}$ = 8.0 Hz, H-1), 4.41 (bd, 1H, $J_{6'a,6'b}$ = 6.3 Hz, H-6'b), 4.66 (d, 1H, J = 11.8 Hz, CHPh), 4.91 (d, 1H, CHPh), 7.25-7.46 (m, 5H, H_{Ar}); ¹³C NMR (75.44 MHz, CD₃OD) δ 26.80, 28.65 (2q, CH₃C), 41.97 (t, CH₂Cl), 62.26, 66.13, 72.16 (3t, C-6, C-6', CH₂Ph), 72.48, 74.43, 75.05, 75.18, 76.56, 76.56, 81.18, 82.35 (8d, C-2, C-3, C-4, C-5, C-2', C-3', C-4', C-5'), 103.42, 104.57 (2d, C-1, C-1'), 111.72 (s, C(CH₃)₂), 169.4 (s, CO).

Anal. Calcd for $C_{24}H_{33}O_{12}Cl$ (548.97): C, 52.51; H, 6.06; Cl, 6.46. Found: C, 52.49; H, 6.00; Cl, 6.41.

Benzyl (2-O-acetyl-6-O-chloroacetyl-3,4-O-isopropylidene-β-D-galactopyranosyl)-(1->4)-2,6-di-O-acetyl-β-D-glucopyranoside (4a). Compound 3 (272 mg, 0.49 mmol) was dissolved in dry CH2Cl2 (5 mL) under an inert atmosphere, then 2,6-di-tertbutyl-4-methyl pyridine (1.63 g, 7.92 mmol) and acetyl chloride (352 µL, 494 mmol) were added in two portions over 24 h. After 48 h the reaction was quenched with a saturated solution of NaHCO₃ (5 mL) and the organic layer was washed with HCl 5% and then with water. It was dried over Na₂SO₄ and the solvent removed under reduced pressure; purification of the crude by flash chromatography (eluent petroleum ether/ethyl acetate = 1:1 v/v) afforded compound 4a as an amorphous white solid (152 mg, 45% yield) and compound 4b also as an amorphous white solid (138 mg, 39% yield). 4a: [α]_D -2.0° (c 1.5, CHCl₃); ¹H NMR (200 MHz, CDCl₃) δ 1.30, 1.56 (2s, 6H, C(CH₃)₂), 2.12 (s, 3H, OAc), 2.14 (s, 6H, 2 OAc), 3.43-3.62 (m, 2H, H-4, H-5), 3.69 (bt, 1H, $J_{23} = J_{3.4} =$ 8.6 Hz, H-3), 4.08 (dd, 1H, $J_{5,6a} = 4.5$ Hz, $J_{6a,6b} = 11.9$ Hz, H-6a), 4.16 (s, 2H, CH_2Cl), 4.11-4.26 (m, 3H, H-3', H-4', H-5'), 4.36 (dd, 1H, $J_{5.6b} = 1.9$ Hz, H-6b), 4.39 (d, 1H, $J_{1'2'}$ = 8.2 Hz, H-1'), 4.46 (bd, 1H, $J_{6'a,6'b}$ = 12.3 Hz, H-6'a), 4.47 (d, 1H, $J_{1,2}$ = 8.0 Hz, H-1), 4.50 (bd, 1H, H-6'b), 4.61 (d, 1H, 12.2 Hz, CHPh), 4.88 (d, 1H, CHPh), 4.95 (dd, 1H, $J_{2',3'} = 9.2 \text{ Hz}$, H-2'), 4.99 (bt, 1H, H-2), 7.25-7.45 (m, 5H, H_{Ar}); ¹³C NMR (75.44 MHz, CDCl₃) δ 21.39 (3q, CH₃CO), 26.84, 28.05 (2q, CH₃C), 41.19 (t, CH₂Cl), 63.24, 64.99, 71.21 (3t, C-6, C-6', CH₂Ph), 71.72, 72.46, 73.08, 73.08, 73.50, 73.87, 77.84, 83.24 (8d, C-2, C-3, C-4, C-5, C-2', C-3', C-4', C-5'), 99.86, 102.0 (2d, C-1, C-1'), 111.9 (s, $C(CH_3)_2$, 167.9, 170.3, 170.3, 171.2 (4s, CO).

Anal. Calcd for $C_{30}H_{39}O_{15}Cl$ (675.08): C, 53.38; H, 5.82; Cl, 5.25. Found: C, 53.45; H, 5.80; Cl, 5.15.

For characterization of 4b, see below.

(2-O-acetyl-6-O-chloroacetyl-3,4-O-isopropylidene-β-D-galactopyranosyl)-(1->4)-2,3,6-tri-O-acetyl-β-D-glucopyranoside (4b). Compound 3 (540 mg, 0.98 mmol) was dissolved in dry CH₂Cl₂ (10 mL) under inert atmosphere, then sym-collidine (1.1 mL, 7.84 mmol) and acetyl chloride (490 µL, 6.86) were added. The reaction was monitored by TLC (eluent petroleum ether/ethyl acetate = 5:7 v/v). After 10 h symcollidine (2.2 mL, 15.68 mmol) and acetyl chloride (980 µL, 13.72 mmol) were added and the solution was left stirring for 48 h. The reaction was quenched with a satured solution of NaHCO3 (5 mL) and the organic layer was washed with 5% HCl, then with water, dried over Na₂SO₄ and concentrated under reduced pressure. Purification of the crude by flash chromatography (eluent petroleum ether/ethyl acetate = 6:4, then 1:1 v/v) afforded compound 4b as a glassy solid (630 mg, 90% yield); [α]_D -8.4° (c 1.1, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 1.31, 1.53 (2s, 6H, C(CH₃)₂), 2.00, 2.07, 2.08, 2.12 (4s, 12H, OAc), 3.59 (ddd, 1H, $J_{5.6b} = 2.0$ Hz, $J_{5.6a} = 5.1$ Hz, $J_{5.4} = 9.5$ Hz, H-5), 3.82 (bt, 1H, H-4), 3.78-3.95 (m, 4H, H-3', H-4', H-5', H-6'a), 4.10-4.16 (m, 4H, H-6a, H-6'b, CH_2Cl), 4.39 (d, 1H, $J_{1'2'} = 7.7$ Hz, H-1'), 4.49 (dd, 1H, $J_{6a.6b} = 12.2$ Hz, H-6b), 4.51 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1), 4.58 (d, 1H, J = 12.3 Hz, CHPh), 4.83 (m, 1H, H-2'), 4.84 (d, 1H, CHPh), 4.97 (bt, 1H, H-2), 5.12 (t, 1H, $J_{2,3} = 9.1$ Hz, H-3), 7.22-7.38 (m, 5H, H_{Ar}); ^{13}C NMR (75.44 MHz, CDCl₃) δ 20.62, 20.80 (4q, CH₃CO), 26.04, 27.24 (2q, CH₃C), 40.61 (t, CH₂Cl), 62.23, 64.50, 70.70 (3t, C-6, C-6', CH₂Ph), 70.61, 71.71, 72.41, 72.65, 72.82, 72.82, 75.77, 76.77 (8d, C-2, C-3, C-4, C-5, C-2', C-3', C-4', C-5'), 99.11, 100.2 (2d, C-1, C-1'), 110.9 (s, C(CH₃)₂), 167.1, 169.2, 169.5, 169.9, 170.4 (5s, CO).

Anal. Calcd for $C_{32}H_{41}O_{16}Cl$ (717.12): C, 53.60; H, 5.76; Cl, 4.94. Found: C, 53.63; H, 5.74; Cl, 4.90.

Benzyl (2-O-acetyl-6-O-chloroacetyl-3,4-O-isopropylidene-β-D-galactopyranosyl)-(1->4)-[(3,4-di-O-acetyl-2-O-benzyl-α-L-fucopyranosyl)-(1->3)]-2,6-di-O-acetyl-β-D-glucopyranoside (6). Compound 4a (100 mg, 0.148 mmol) was dissolved under an inert atmosphere in dry CH₂Cl₂ (2 mL). At 0 °C TMSOTf (16 μL of a 0.1M solution in dry CH₂Cl₂, 0.0016 mmol) was added. Compound 5 dissolved in dry CH₂Cl₂ (1 mL) was added dropwise to the solution. After 30 min the reaction was neutralized with NaHCO₃ and chromatographic purification (cluent petroleum ether/ethyl acetate = 1:1 v/v) of the crude afforded pure compound 6 as a white foam (148 mg, 99% yield): [α]_D - 50.1° (c 1.2, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 1.26 (d, 3H, J_{5'',6''} = 6.6 Hz, H-6''),

1.31, 1.50 (2s, 6H, C(CH₃)₂), 1.91, 1.96, 2.07, 2.10, 2.13 (5s, 15H, OAc), 3.47 (m, 1H, H-5), 3.82-3.96 (m, 2H, H-2", H-4"), 3.89 (t, 1H, $J_{3,4} = J_{4,5} = 8.5$ Hz, H-4), 3.98 (t, 1H, $J_{2,3} = 8.5$ Hz, H-3), 3.89-4.08 (m, 1H, H-5"), 4.09-4.17 (m, 4H, H-6a, H-3", CH₂Cl), 4.28 (d, 1H, $J_{1',2'} = 8.6$ Hz, H-1"), 4.42 (d, 1H, $J_{1,2} = 7.5$ Hz, H-1), 4.46 (d, 1H, J = 11.5 Hz, CHPh), 4.53 (d, 1H, J = 12.5 Hz, CHPh), 4.62 (d, 1H, CHPh), 4.51-4.64 (m, 2H, H-6b, H-6"a), 4.72 (dd, 1H, $J_{5',6'b} = 8.0$ Hz, $J_{6'a,6'b} = 11.9$ Hz, H-6"b), 4.83 (d, 1H, CHPh), 4.81-4.93 (m, 2H, H-2", H-5"), 5.15 (bt, 1H, H-2), 5.24 (bs, 1H, H-4"), 5.27 (m, 1H, H-3"), 5.31 (d, 1H, $J_{1'',2''} = 4.0$ Hz, H-1"), 7.18-7.40 (m, 10H, $J_{1,1'',2''} = 4.0$ Hz, H-1"), 7.18-7.40 (m, 10H, $J_{1,1'',2''} = 4.0$ Hz, H-2"), 20.63, 20.77, 20.97 (3q, 5C, CH₃CO), 26.09, 27.56 (2q, CH₃C), 40.90 (t, CH₂Cl), 61.63, 64.01, 70.13, 72.61 (4t, C-6, C-6", CH₂Ph, CH₂Ph), 64.25, 70.00, 70.65, 71.85, 72.73, 73.23, 73.23, 73.23, 73.35, 74.00, 74.32, 74.51, 77.18 (12 d, C-2, C-3, C-4, C-5, C-2", C-3", C-4", C-5", C-2", C-3", C-4", C-5"), 96.34, 98.96, 100.2 (3d, C-1, C-1", C-1"), 110.9 (s, C(CH₃)₂), 167.7, 168.8, 169.1, 169.6, 170.5, 170.5 (6s, CO).

Anal. Calcd for $C_{47}H_{59}O_{21}Cl$ (995.43): C, 56.71; H, 5.97; Cl, 3.56. Found: C, 56.68; H, 6.01; Cl, 3.55.

Benzyl (2-O-acetyl-3,4-O-isopropylidene- β -D-galactopyranosyl)-(1->4)-2,3,6tri-O-acetyl-β-D-glucopyranoside (7). Compound 4b (88 mg, 0.12 mmol) was dissolved in toluene/ethanol = 1:1 v/v (8 mL), then DABCO (100 mg, 8 equiv) was added, and the solution was stirred at 60 °C. After 30 min the reaction mixture was concentrated and chromatographic purification (eluent petroleum ether/ethyl acetate = 4:6 v/v) afforded compound 7 as a white foam (68 mg, 88% yield): $[\alpha]_D$ -9.9° (c 1.1, CHCl₃); ¹H NMR (200 MHz, CDCl₃) δ 1.30, 1.52 (2s, 6H, C(CH₃)₂), 1.99, 2.06, 2.08, 2.11 (4s, 12H, OAc), 2.50 (bs, OH), 3.58 (ddd, 1H, $J_{5,6b} = 1.9$ Hz, $J_{5,6a} = 5.1$ Hz, $J_{5,4} = 9.4$ Hz, H-5), 3.82 (t, 1H, $J_{3,4} = 9.4$ Hz, H-4), 3.76-3.95 (m, 3H, H-5', H-6'a, H-6'b), 4.00-4.29 (m, 3H, H-3', H-4', H-6a), 4.38 (d, 1H, $J_{1',2'} = 7.7$ Hz, H-1'), 4.49 (dd, 1H, $J_{6a,6b} = 12.2$ Hz, H-6b), 4.50 (d, 1H, $J_{1,2} = 7.6$ Hz, H-1), 4.52 (d, 1H, J = 12.3 Hz, CHPh), 4.85 (m, 1H, H-2'), 4.85 (d, 1H, CHPh), 4.97 (dd, 1H, $J_{2,3} = 9.4$ Hz, H-2), 5.12 (t, 1H, H-3), 7.26-7.48 (m, 5H, H_{Ar}); ¹³C NMR (50.29 MHz, CDCl₃) δ 20.58, 20.78 (4q, CH₃CO), 26.12, 27.33 (2q, CH₃C), 62.10, 62.27, 70.65 (3t, C-6, C-6', CH₂Ph), 71.58, 72.68, 72.97, 72.97, 73.60, 73.60, 75.67, 77.06 (8d, C-2, C-3, C-4, C-5, C-2', C-3', C-4', C-5'), 99.02, 100.3 (2d, C-1, C-1'), 110.8 (s, $=C(CH_3)_2$), 169.2, 169.6, 170.3, 170.4 (4s, CO).

Anal. Calcd for C₃₀H₄₀O₁₅ (640.64): C, 56.25; H, 6.29. Found: C, 56.28; H, 6.32.

Benzyl (2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 6)-(2-O-acetyl-3,4-Oisopropylidene- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -2,3,6-tri-O-acetyl- β -D-glucopyranoside (9). Compound 7 (50 mg, 0.078 mmol.) and compound 8 (77 mg, 0.156 mmol) were dissolved in dry CH2Cl2 (3 mL) under an inert atmosphere. TBDMSOTf (39 µL of a 0.1 M solution in dry CH₂Cl₂, 0.0039 mmol.) was then added. After 1 h the reaction was quenched with TEA and the solvent removed under reduced pressure. Chromatographic purification (eluent petroleum ether/ethyl acetate = 6:4, then 1:1 v/v) afforded compound 9 as a white foam (53 mg, 70% yield): $[\alpha]_D$ -13.3° (c 1.5, CHCl₃); ¹H NMR (300 MHz, CDCl₃) & 1.29, 1.52 (2s, 6H, C(CH₃)₂), 1.92, 1.98, 2.02, 2.04, 2.07, 2.11, 2.15, 2.17 (8s, 24H, OAc), 3.57 (ddd, 1H, $J_{5.6b} = 1.4$ Hz, $J_{5.6a} = 4.7$ Hz, $J_{5.4} = 9.5$ Hz, H-5), 3.78 (t, 1H, $J_{3,4} = 9.5 \text{ Hz}$, H-4), 3.77-3.83 (m, 2H, H-6'a, H-6'b), 3.94 (bt, 1H, $J_{5'',6''} = 6.6 \text{ Hz}$, H-5''), 4.00-4.22 (m, 6H, H-6a, H-3', H-4', H-5', H-6''a, H-6''b), 4.36 (d, 1H, $J_{1',2'} = 7.1$ Hz, H-1'), 4.44 (dd, 1H, $J_{6a,6b}$ = 11.9 Hz, H-6b), 4.51 (d, 1H, $J_{1,2}$ = 7.9 Hz, H-1), 4.59 (d, 1H, J= 12.3 Hz, CHPh), 4.64 (d, 1H, $J_{1",2"} = 7.9$ Hz, H-1"), 4.81 (m, 1H, H-2"), 4.84 (d, 1H, CHPh), 4.95 (bd, 1H, $J_{2,3} = 9.5$ Hz, H-2), 5.07 (dd, 1H, $J_{3'',4''} = 3.4$ Hz, $J_{2'',3''} = 10.4$ Hz, H-3"), 5.13 (t, 1H, H-3), 5.21 (dd, 1H, H-2"), 5.40 (d, 1H, H-4"), 7.23-7.35 (m, 5H, H_{Af}); ¹³C NMR (50.29 MHz, CDCl₃) δ 20.70 (1q, 8C, CH₃CO), 26.00, 27.21 (2q, CH₃C), 61.10, 62.22, 68.11, 70.70 (4t, C-6, C-6', C-6'', CH₂Ph), 67.02, 68.82, 70.71, 71.45, 72.61, 72.61, 72.61, 72.61, 72.93, 73.13, 75.32, 76.60 (12d, C-2, C-3, C-4, C-5, C-2', C-3', C-4', C-5', C-2", C-3", C-4", C-5"), 99.13, 100.3, 100.8 (3d, C-1, C-1', C-1"). 110.7 (s, C(CH₃)₂), 169.1, 169.4, 169.4, 169.6, 169.9, 170.1, 170.2, 170.5 (8s, CO).

Anal. Calcd for C₄₄H₅₈O₂₄ (970.93): C, 54.43; H, 6.02. Found: C, 54.45; H, 5.99.

Benzyl (2,4-di-O-acetyl-6-O-chloroacetyl-β-D-galactopyranosyl)-(1->4)-2,3,6-tri-O-acetyl-β-D-glucopyranoside (10). Compound 4b (100 mg, 0.14 mmol) was dissolved in CH₂Cl₂ (3 mL) and 50% aq CF₃COOH was added. The reaction was left stirring at 50 °C and after 2 h was neutralized with a saturated solution of NaHCO₃. The organic layer was washed with water then dried over Na₂SO₄, and the solvent was removed under reduced pressure. The crude product was dissolved in dry CH₃CN under an inert atmosphere and (CH₃O)₃CCH₃ (48 μL, 0.42 mmol.) and a catalytic amount of CSA was added. After 10 min AcOH 80% was added, and the reaction was left stirring for 30 min. The reaction was diluted with CH₂Cl₂ and neutralized with a saturated solution of NaHCO₃, the organic layer was washed with water then dried over Na₂SO₄,

and the solvent was removed under reduced pressure. Chromatographic purification (eluent petroleum ether/ethyl acetate = 3:7 v/v) afforded 68 mg (68% yield) of compound 8 as a glassy solid: $[\alpha]_D$ -20.6° (c 1.1, CHCl₃); 1H NMR (200 MHz, CDCl₃) δ 2.03, 2.05, 2.11, 2.14, 2.17 (5s, 15H, OAc), 2.74 (bs, OH), 3.60 (ddd, 1H, $J_{5,6b}$ = 1.9 Hz, $J_{5,6a}$ = 5.1 Hz, $J_{5,4}$ = 9.9 Hz, H-5), 3.78 (dd, 1H, $J_{3',4'}$ = 3.5 Hz, $J_{2',3'}$ = 10.0 Hz, H-3'), 3.81 (bt, 1H, H-4), 3.85 (bt, 1H, $J_{5',6'}$ = 6.4 Hz, H-5'), 4.08 (s, 2H, CH₂Cl), 4.10 (dd, 1H, $J_{6a,6b}$ = 12.6 Hz, H-6a), 4.14-4.20 (m, 2H, H-6'a, H-6'b), 4.43 (d, 1H, $J_{1',2'}$ = 7.8 Hz, H-1'), 4.52 (dd, 1H, H-6b), 4.53 (d, 1H, $J_{1,2}$ = 7.8 Hz, H-1), 4.59 (d, 1H, $J_{2,3}$ = 9.2 Hz, H-2), 5.14 (t, 1H, $J_{3,2}$ = $J_{3,4}$ = 9.2 Hz, H-3), 5.27 (d, 1H, H-4'), 7.24-7.37 (m, 5H, J_{Ar}); 13 C NMR (50.29 MHz, CDCl₃) δ 20.74 (5q, CH₃CO), 40.44 (t, CH₂Cl), 62.10, 62.97, 70.71 (3t, C-6, C-6', CH₂Ph), 69.09, 71.22, 71.22, 71.51, 72.61, 72.61, 72.77, 76.02 (8d, C-2, C-3, C-4, C-5, C-2', C-3', C-4', C-5'), 93.99, 100.5 (2d, C-1, C-1'), 166.9, 169.5, 169.9, 170.5, 170.6, 170.9 (6s, CO).

Anal. Calcd for $C_{31}H_{39}O_{17}Cl$ (719.09): C, 51.78; H, 5.47; Cl, 4.93. Found: C, 51.80; H, 5.45; Cl, 4.98.

Benzyl (2,3,4-tri-O-acetyl-6-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-(2,4-di-Oacetyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -2,3,6-tri-O-acetyl- β -D-glucopyranoside (13).Compound 10 (190 mg, 0.26 mmol) and compound 11 (226 mg, 0.42 mmol) were dissolved in dry CH2Cl2 (2 mL) under an inert atmosphere. Then TMSOTf (39 µL of a 0.1 M solution in dry CH₂Cl₂, 0.05 equiv) was added. After 1 h the reaction was neutralized with NaHCO3 and the solvent removed under reduced pressure. The crude was partially purified by flash chromatography (eluent petroleum ether/ethyl acetate = 4:6 v/v). The fractions containing the product were dissolved in EtOH/toluene = 1:1 (4 mL) and a catalytic amount of DABCO was added. After 6 h the solvent was removed and chromatographic purification (eluent petroleum ether/ethyl acetate 1:1, then 4:6 v/v) afforded compound 13 as a glassy solid (77 mg, 29% yield): [α]_D -15.4° (c 1.9, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 1.94, 1.99, 1.99, 2.01, 2.07, 2.08, 2.12, 2.16 (8s, 24H, OAc), 2.82 (bt, OH), 3.33-3.66 (m, 5H, H-4, H-5, H-5', H-6'a, H-6'b), 3.74-3.81 (m, 2H, H-3', H-5''), 4.08 (dd, 1H, $J_{5.6a} = 5.0$ Hz, $J_{6a,6b} = 11.8$ Hz, H-6a), 4.11 (dd, 1H, $J_{5'',6a''} =$ 7.1 Hz, $J_{6''a,6''b} = 14.1$ Hz, H-6''a), 4.36 (d, 1H, $J_{1',2'} = 8.1$ Hz, H-1'), 4.42 (d, 1H, J = 12.0Hz, CHPh), 4.42-4.54 (m, 2H, H-6b, H-6''b), 4.48 (d, 1H, $J_{1",2"} = 7.8$ Hz, H-1''), 4.49

(d, 1H, $J_{1,2} = 7.5$ Hz, H-1), 4.52 (d, 1H, CHPh), 4.57 (d, 1H, J = 12.3 Hz, CHPh), 4.83 (d, 1H, CHPh), 4.91 (dd, 1H, $J_{3'',4''} = 3.7$ Hz, $J_{2'',3''} = 10.4$ Hz, H-3''), 4.93 (dd, 1H, $J_{2,3} = 10.0$ Hz, H-2), 5.04 (dd, 1H, H-2''), 5.10 (t, 1H, $J_{3,4} = 9.2$ Hz, H-3), 5.11 (t, 1H, $J_{2',3'} = 8.1$ Hz, H-2'), 5.24 (d, 1H, $J_{3',4'} = 3.3$ Hz, H-4'), 5.40 (d, 1H, H-4''), 7.22-7.39 (m, 10H, H_{Ar}); ¹³C NMR (75.44 MHz, CDCl₃) δ 20.46, 20.56, 20.69, 20.78 (4q, 8C, CH₃CO), 60.18, 62.16, 67.47, 70.70, 73.59 (5t, C-6, C-6', C-6'', CH₂Ph, CH₂Ph), 67.35, 68.80, 70.01, 70.87, 71.05, 71.74, 72.43, 72.67, 73.36, 74.00, 76.03, 76.12 (12d, C-2, C-3, C-4, C-5, C-2', C-3', C-4', C-5', C-2'', C-3'', C-4'', C-5''), 99.01, 101.0, 101.4 (3d, C-1, C-1', C-1''), 168.6, 169.1, 169.5, 169.8, 170.0, 170.1, 170.4, 172.2 (8s, CO).

Anal. Calcd for C₄₈H₆₀O₂₄ (1020.99): C, 56.47; H, 5.92. Found: C, 56.43; H, 5.90.

ACKNOWLEDGEMENTS

This work was supported by EU-NOFA project (grant FAIR CT973142).

REFERENCES AND NOTES

- 1. H. Waldmann and D. Sebastian, Chem. Rev., 94, 911 (1994).
- 2. M. Therisod and A. M. Klibanov, J. Am Chem. Soc., 108, 5638 (1986).
- 3. R. Khan, L. Gropen, P. A. Konowicz, M. Matulovà and S. Paoletti, *Tetrahedron Lett.*, 34, 7767 (1993).
- 4. D. C. Palmer and F. Terradas, Tetrahedron Lett., 38, 1673 (1994).
- 5. a) L. Panza, S. Brasca, S. Riva and G. Russo, Tetrahedron: Asymmetry, 4, 931 (1993).
 - b) L. Panza, M. Luisetti, E. Crociati and S. Riva, J. Carbohydr. Chem., 12, 125 (1993).
- 6. S. Cai, S. Hakomori and T. Toyokumi, J. Org. Chem., 57, 3432 (1992).
- 7. a) G. V. Coppa, O. Gabrielli, P. Giorgi, C. Catassi, M. P. Montanari, P. E. Varaldo and B. L. Nichols, *The Lancet*, 335, 569 (1990).
 - b) A. Cravioto, A. Tello, H. Villafan, J. Ruiz, S. Del Vedovo and J-R. Neeser, J. Infect. Dis., 6, 163 (1991).
 - c) A. Laegreid, A. B. K. Otnaessi and J. Fuglesan, Pediatr. Res., 20, 416 (1986).
 - d) B. Andersson, O. Porras, L. A. Hanson, T. Lagergard and C. Svanborg-Eden, J. Infect. Dis., 153, 232 (1986).
 - e) D. S. Newburg, L. K. Pickering, R. H. McCluer and T. G. Cleary, J. Infect. Dis., 162, 1075 (1990).
- 8. L. Lay, L. Panza, S. Riva, M. Khitri and S. Tirendi, Carbohydr. Res., 291, 197 (1996).
- 9. a) B. Danieli, M. Luisetti, G. Sampognaro, G. Carrea and S. Riva, J. Mol. Catal., 3, 193 (1997).
 - b) V. V. Mozhaev, C. L. Budde, J. O. Rich, A. Y. Usyatinsky, P. C. Michels, Y. L. Khmelnitsky, D. S. Clark and J. S. Dordick, *Tetrahedron*, 54, 3971 (1998).

- 10. L. Manzoni, L. Lay and R.R. Schmidt, *J. Carbohydr. Chem.*, 17, 739 (1998) and references therein.
- 11. P. Henri, A. Zollo and P. Sinaÿ, Carbohydr. Res., 150, 199 (1986).
- 12. G. Hummel and R. R. Schmidt, Tetrahedron Lett., 38, 1173 (1997).
- 13. K.-H. Jung, M. Hoch and R. R. Schmidt, Liebigs Ann. Chem., 1099 (1989).