ChemComm



View Article Online

COMMUNICATION



Cite this: Chem. Commun., 2014, 50, 10059

Received 3rd June 2014, Accepted 9th July 2014

DOI: 10.1039/c4cc04227a

www.rsc.org/chemcomm

Guest-induced supramolecular chirality in a ditopic azoprobe-cyclodextrin complex in water†

Kentaro Nonaka, Mai Yamaguchi, Masashi Yasui, Shoji Fujiwara, Takeshi Hashimoto and Takashi Hayashita*

We report a novel supramolecular chirality induced by the twisted structural change of two ditopic azoprobes (15C5-Azo-dpa) inside the chiral cavity of γ -cyclodextrin (γ -CyD) due to multi-point recognition of guest ions by 15C5-Azo-dpa molecules in water.

Chirality control by supramolecular assemblies and helical polymers based on chiral templates has received much attention in recent years.¹ Those systems have been widely used in chiroptical devices,² memory devices,³ and chiral catalysis.⁴ Pioneering work on supramolecular chirality was conducted by Yashima's group, who designed chiral recognition systems by using helical polymers.⁵ Recently, Kumar *et al.* reported that naphthalene diimide amphiphiles functionalized with the dipicolylethylenediamine motif self-assembled with tunable chirality upon molecular recognition of various adenosine phosphates.⁶ They showed that competitive guest binding induced the dynamic reversal of the helix assemblies. Such guest-induced chirality control is expected to have potential application in the development of versatile chiral switching and sensing devices.⁷

Here we report a simple system of guest-induced chirality control in the 2:1 inclusion complex of ditopic azoprobes with γ -cyclodextrin (γ -CyD) in water. The ditopic azoprobe is a receptor possessing two different recognition sites that are expected to show excellent recognition function when two guest molecules exist in the system. As shown in Fig. 1, we have designed a ditopic azoprobe (**15C5-Azo-dpa**) that bears benzo-15-crown-5 (B15C5) and dipicolylamine (dpa) as the recognition sites (Fig. 1).

The azochromophore moiety can be used as a photosignal transducer.⁸ Although **15C5-Azo-dpa** is almost insoluble in water, γ -CyD can improve its solubility in water by forming a stable inclusion complex with it. The **15C5-Azo-dpa**- γ -CyD complex was found to show a unique response based on the guest-induced supramolecular chirality in water. An induced circular dichroism



Fig. 1 Structures of 15C5-azo-dpa and γ-CyD

(ICD) response was noted upon the addition of carbonate (CO_3^{2-}) or acetate $(CH_3CO_2^{-})$ ions in the presence of both K⁺ and Zn²⁺ ions. Such supramolecular responses were successfully used to sense guest anions in water.

The synthesis of **15C5-Azo-dpa** proceeded with the azocoupling of 4'-aminobenzo-15-crown-5 with phenol, followed by the introduction of a bromoethylene spacer using the Williamson ether synthesis.⁹ Then, a dpa moiety was introduced under basic conditions with K_2CO_3 , and the obtained product was purified by silica column chromatography. The structure of **15C5-Azo-dpa** was confirmed by ¹H NMR and combustion analyses (Fig. S1–S3, ESI†). Details of the synthesis are available in the ESI (Scheme S1, ESI†).

In our previous work, we showed that B15C5 azoprobes selectively formed a 2:1 inclusion complex with γ -CyD in the presence of K⁺ ions in water.¹⁰ Meanwhile, Hamachi's group and other research groups reported that the Zn²⁺ complexes of dpa units interacted with various phosphate anion derivatives

Department of Materials and Life Sciences, Sophia University, 7-1 Kioi-cho, Chiyoda-ku, Tokyo 102-8554, Japan. E-mail: ta-hayas@sophia.ac.jp

[†] Electronic supplementary information (ESI) available: Measurement conditions, synthesis of azoprobes, and the measurements of NMR spectra. See DOI: 10.1039/c4cc04227a



Fig. 2 ICD and UV-Vis spectra of **15C5-Azo-dpa**- γ -CyD sensors. [**15C5-Azo-dpa**] = 40 μ M, [γ -CyD] = 5 mM, and (a) K⁺, (b) Zn²⁺, (c) K⁺ + Zn²⁺, (d) K⁺ + Zn²⁺ + Tri, in 4% DMSO aq., pH = 11.0 at 25 °C; K⁺ (50 mM K₂CO₃), Zn²⁺ (40 μ M Zn(NO₃)₂), Tri (3.0 mM Na₅P₃O₁₀); (M = mol dm⁻³).

through the unoccupied coordination sites in the Zn²⁺-dpa complexes.¹¹ Thus, it is interesting to examine how such guest species affect the response function of the 15C5-Azo-dpa-γ-CyD complex in water. Fig. 2 shows the ICD spectra and the UV-Vis absorption spectra of 15C5-Azo-dpa (40 µM) in 5.0 mM γ-CyD aqueous solution with K^+ ions (a), with Zn^{2+} ions (b), with K^+ and Zn^{2+} ions (c), and with K⁺, Zn^{2+} , and triphosphate (Tri) ions (d). Interestingly, the split-type Cotton effect was observed only in the presence of both K^+ and Zn^{2+} ions (spectrum c in the top panel of Fig. 2). In addition, the UV-Vis absorbance ratio of 366 nm to 425 nm (A_{366}/A_{425}) exhibited the largest blue shift in the presence of both K⁺ and Zn²⁺ ions. The azobenzene chromophores are known to exhibit strong ICD by forming an inclusion complex with CyDs based on the chiral nature of the CyD cavities.¹² We have shown that this ICD response was affected by the dimer formation of the azoprobes inside γ -CyD to induce a split-type Cotton effect.⁸ In addition, an exciton interaction of the azodimer was found to induce clear changes in the UV-Vis absorption spectra. Thus, the observed split ICD in spectrum c in the top panel of Fig. 2 and the largest blue shift in the UV-Vis spectra indicated the formation of a clockwise twisted dimer of 15C5-Azo-dpa probes inside γ -CyD in the presence of both K⁺ and Zn²⁺ ions. The addition of triphosphate was found to inhibit the twisted structure formation of the 15C5-Azo-dpa- γ -CyD complex in water, probably due to the coordination of the triphosphate anion with the Zn²⁺-dpa complex in 15C5-Azo-dpa.

Fig. 3 shows an absorbance *vs.* wavelength plot and the effects of Zn^{2+} concentration on the UV-Vis absorbance ratio (A_{366}/A_{425}) of the **15C5-Azo-dpa**– γ -CyD complex in the presence of 50 mM K₂CO₃ in water. It is evident that the absorbance ratio gradually increased with the addition of Zn^{2+} and reached a plateau near the equivalence point with **15C5-Azo-dpa** (20 μ M). The finding that an equivalent amount of Zn^{2+} in excess of 20 μ M caused no spectral changes



Fig. 3 The changes in UV-Vis spectra and the absorbance ratio of **15C5-Azo-dpa**- γ -CyD sensor as a function of zinc ion concentration. [**15C5-Azo-dpa**] = 20 μ M, [Zn²⁺] = 0 - 45 μ M, [K₂CO₃] = 50 mM, [γ -CyD] = 5 mM, in 4% aq. DMSO, pH = 11.0, at 25 °C (M = mol dm⁻³).

attests the 1:1 complex formation of Zn^{2+} with the dpa binding site in **15C5-Azo-dpa**. The sigmoid response observed in Fig. 3 indicates the occurrence of a successive binding reaction of Zn^{2+} with the two **15C5-Azo-dpa** molecules inside γ -CyD. The formation of the 2:1 complex of **15C5-Azo-dpa** probes with γ -CyD was confirmed by the analysis of Job's plots (Fig. S4, ESI⁺).

To elucidate the mechanism of the split ICD noted for the **15C5-Azo-dpa**-γ-CyD complex, we examined the salt effect by substituting 100 mM KNO3 and 100 mM KOH for 50 mM K2CO3 in the presence of 40 µM Zn²⁺. Interestingly, no split ICD was observed for the KNO3 and KOH systems, indicating the possibility of CO32bridging between the two Zn²⁺-dpa binding sites (data not shown). The effect of CO_3^{2-} concentration on the ICD spectra of the 15C5-Azodpa-γ-CyD complex is depicted in Fig. 4. It is evident that the addition of CO_3^{2-} ions enhanced the split ICD for the 15C5-Azo-dpa- γ -CyD complex at pH 11.0. The ICD intensity change as a function of CO3²⁻ concentration at a constant K⁺ concentration of 0.10 M was well fitted by a 1:1 binding isotherm, and the apparent 1:1 binding constant of CO_3^{2-} ions was calculated to be 83.5 \pm 6.0 M⁻¹ (Fig. S5, ESI[†]). The triphosphate binding may break this CO_3^{2-} bridging of the two 15C5-Azo-dpa molecules inside γ -CyD to reduce the split ICD. In addition, direct evidence of the relative orientation of the azoprobes and the macrocyclic ring was obtained by NOESY experiments. Cross peaks between H3 protons inside the CyD cavity and protons of the azoprobes were clearly observed (Fig. S8, ESI[†]).

If this CO_3^{2-} bridging were the key mechanism for the induction of the split ICD, a similar split ICD induced by the CH_3CO_2^- bridging would be expected under neutral pH conditions. In fact, we observed the typical split ICD in the presence of 50 mM $\text{CH}_3\text{CO}_2\text{K}$ at pH 7.0 (Fig. 5). However, it should be noted that the split ICD had peak



Fig. 4 ICD spectra of **15C5-Azo-dpa** $-\gamma$ -CyD sensors, [**15C5-Azo-dpa**] = 40 μ M, [Zn²⁺] = 40 μ M, [K₂CO₃] = 5–40 mM, [γ -CyD] = 5 mM, in 4% DMSO aq., pH = 11.0 at 25 °C; (M = mol dm⁻³).



Fig. 5 ICD spectra of **15C5-Azo-dpa**- γ -CyD sensors, [**15C5-Azo-dpa**] = 40 μ M, [Zn²⁺] = 40 μ M, [γ -CyD] = 5 mM, (a) [CH₃CO₂Na] = 50 mM, (b) [CH₃CO₂K] = 50 mM, (c) [CH₃CO₂Cs] = 50 mM, in 4% DMSO aq., pH = 7.0 at 25 °C; (M = mol dm⁻³).

shapes that differed from those of the CO_3^{2-} bridging system, indicating that a somewhat different mode of the twisted structure was formed inside γ -CyD. In contrast, no split ICD was noted in the presence of 50 mM CH₃CO₂Na or 50 mM CH₃CO₂Cs. This indicated that the clockwise twisted structure of the two **15C5-Azo-dpa** molecules inside γ -CyD was induced only in the presence of K⁺, Zn²⁺, and the bridging anions of the two dpa–Zn²⁺ binding sites, such as CO₃²⁻ or CH₃CO₂⁻ ions. By titration analysis of CH₃CO₂⁻ at a constant K⁺ concentration of 0.10 M, the apparent 1:1 binding constant of CH₃CO₂⁻ for the **15C5-Azo-dpa**– γ -CyD complex in the presence of an equivalent amount of Zn²⁺ was determined to be 180 ± 27 M⁻¹ at pH 7.0 (Fig. S6, ESI⁺).

In conclusion, we have shown a novel supramolecular chirality induced by the twisted structural change of two **15C5-Azo-dpa** molecules inside the γ -CyD chiral cavity due to multi-point recognition of guest ions by the ditopic azoprobes in water. In this system, all components of the guest species induce the split ICD response, making the specific detection of each component feasible by maintaining the other two components in the **15C5-Azo-dpa**- γ -CyD complex solution. To the best of our knowledge, this is the first example of a simple ditopic receptor exhibiting multi-recognition function based on supramolecular chirality in water.

This work was supported by Grants-in-Aid for Scientific Research from the Japan Society for the Promotion of Science (24655069 and 26248038). This communication is dedicated to Professor Shinkai Seiji (Kyushu University, Sojo University) on the occasion of his 70th birthday.

Notes and references

- Supramolecular Chirality, In *Top. Curr. Chem.*, ed. M. Crego-Calama and D. N. Reinhovdt, 2006, vol. 26; Amplification of Chirality, In *Top. Curr. Chem.*, ed. K. Soai, 2008, vol. 284; *Chirality at the Nanoscale*, ed. D. B. Amabilino, Wiley-VCH, Weinheim, 2009; R. Raval, *Chem. Soc. Rev.*, 2009, 38, 707; J. Crassovs, *Chem. Soc. Rev.*, 2009, 38, 830.
- 2 K. C.-F. Lrung, C.-P. Chak, C.-M. Lo, W.-Y. Wong, S. Xuan and C. H. K. Cheng, *Chem. – Asian J.*, 2009, 4, 364; R. Klajn, J. F. Stoddart and B. A. Gtrzybowski, *Chem. Soc. Rev.*, 2010, 39, 2203; J. W. Canary, S. Morteznei and J. Liang, *Coord. Chem. Rev.*, 2010, 254, 2249; C. Colvccini, A. Mazzrnti and D. Pasini, *Org. Biomol. Chem.*, 2010, 8, 1807; M. Caricato, C. Colvccini, D. Dondi, D. A. Vander Griend and D. Pasini, *Org. Biomol. Chem.*, 2010, 8, 3272; M. Caricato, A. Olmo, C. Gargivlli, G. Gattvso and D. Pasini, *Tetrahedron*, 2012, 68, 7861; Y. Nakatani, Y. Frosho and E. Yashima, *Org. Biomol. Chem.*, 2013, 11, 1614.
- M. Ziegler, A. V. Davis, D. W. Johnson and K. N. Raymond, Angew. Chem., Int. Ed., 2003, 42, 665; A. Mammana, A. D'Vrso, R. Lauceri and R. Pvrrello, J. Am. Chem. Soc., 2007, 129, 8062; L. Rosaria, A. D'Vrso, A. Mammana and R. Purrello, Chirality, 2008, 20, 411; I. D. Cat, Z. Gvo, S. J. George, E. W. Meijer, A. P. H. J. Schenning and S. D. Feyter, J. Am. Chem. Soc., 2012, 134, 3171; P. A. Korevaar, S. J. George, A. J. Markvoort, M. M. J. Smulders, P. A. J. Hilbers, A. P. H. J. Schenning, T. F. A. de Greef and E. W. Meijer, Nature, 2012, 481, 492.
- 4 M. Weis, C. Waloch, W. Seiche and B. Breif, J. Am. Chem. Soc., 2006, 128, 4188; J. Meevwissen and J. N. H. Reek, Nat. Chem., 2010, 2, 615; P. W. N. M. Van Leevwen, D. Rivillo, M. Ranal and Z. Frexa, J. Am. Chem. Soc., 2011, 133, 18562; M. Hatano and K. Ishihara, Chem. Commun., 2012, 48, 4273.
- E. Yashima, K. Maeda and Y. Okamoto, *Nature*, 1999, 399, 449;
 H. Ito, M. Ikeda, T. Hasegawa, Y. Furusho and E. Yashima, *J. Am. Chem. Soc.*, 2011, 133, 3419;
 H. Yamada, Y. Furusho and E. Yashima, *J. Am. Chem. Soc.*, 2012, 134, 7250;
 H. Yamada, Z. Q. Wu, Y. Furusho and E. Yashima, *J. Am. Chem. Soc.*, 2012, 134, 7250;
 H. Yamada, Z. Q. Wu, Y. Furusho and E. Yashima, *J. Am. Chem. Soc.*, 2012, 134, 7250;
 H. Yamada, Z. Q. Wu, Y. Furusho and E. Yashima, *J. Am. Chem. Soc.*, 2012, 134, 9506;
 Y. Furusho and E. Yashima, *Org. Biomol. Chem.*, 2013, 11, 1614;
 W. Makiguchi, S. Kobayashi, Y. Furusho and E. Yashima, *Angew. Chem., Int. Ed.*, 2013, 52, 5275;
 S. Yashima, *Angew. Chem., Int. Ed.*, 2013, 52, 6849.
- 6 M. Kumar, N. Jonnalagadda and S. J. George, *Chem. Commun.*, 2012, 48, 10948.
- 7 S. Akine, S. Sairenji, T. Taniguchi and T. Nabeshima, *J. Am. Chem. Soc.*, 2013, 135, 12948; J. Crassous, *Chem. Commun.*, 2012, 48, 9684;
 H. Miyake and H. Tsukube, *Chem. Soc. Rev.*, 2012, 41, 6977; R. Lin,
 H. Zhang, S. Li, L. Chen, W. Zhang, T. B. Wen, H. Zhang and H. Xia, *Chem. Eur. J.*, 2011, 17, 2420.
- 8 C. Shimpuku, R. Ozawa, A. Sasaki, F. Sato, T. Hashimoto, A. Yamauchi, I. Suzuki and T. Hayashita, *Chem. Commun.*, 2009, 1709.
- 9 F. Sato, M. Tsukano, K. Sakamoto, W. Umemoto, T. Hashimoto and T. Hayashita, *Bull. Chem. Soc. Jpn.*, 2008, **81**, 1589; F. Sato, K. Sakamoto, W. Umemoto, T. Hashimoto and T. Hayashita, *Chem. Lett.*, 2007, **36**(10), 1243.
- 10 T. Hayashita, D. Fujita, T. Hashimoto, A. Endo, H. Zhao, K. Odagiri, S. Takahashi, M. M. Mohamad and N. Teramae, *ARKIVOC*, 2010, vii, 203; T. Hayashita, A. Yamauchi, A.-J. Tong, J. C. Lee, B. D. Smith and N. Teramae, *J. Inclusion Phenom.*, 2004, 50, 87; A. Yamauchi, T. Hayashita, A. Kato, S. Nishizawa, M. Watanabe and N. Teramae, *Anal. Chem.*, 2000, 72, 5841; A. Yamauchi, T. Hayashita, S. Nishizawa, M. Watanabe and N. Teramae, *J. Am. Chem. Soc.*, 1999, 121, 2319.
- 11 Y. Kurishita, T. Kohira, A. Ojida and I. Hamachi, J. Am. Chem. Soc., 2012, **134**, 18779; L. Yan, Z. Ye, C. Peng and S. Zhang, *Tetrahedron*, 2012, **68**, 2725; H. T. Ngo, X. Liu and K. A. Jolliffe, *Chem. Soc. Rev.*, 2012, **41**, 4928, and references therein.
- B. Mayer, X. Zhang, W. M. Nau and G. Marconi, J. Am. Chem. Soc., 2001, 123, 5240; M. Kodaka, J. Am. Chem. Soc., 1993, 115, 3702; M. Kodaka, J. Phys. Chem., 1991, 95, 2110; K. Harata and H. Uedaira, Bull. Chem. Soc. Jpn., 1975, 48, 375; I. Tinoco Jr., Adv. Chem. Phys., 1962, 4, 113.