

## THE STRUCTURE OF TWO NEW COUMARINS FROM THE ROOTS OF *LOMATIUM COLUMBIANUM*\*

BENT EICHSTEDT NIELSEN† and ELIN JENSEN

The Royal Danish School of Pharmacy, Chemical Institute BC, DK-2100 Copenhagen Ø, Denmark

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**Key Word Index**—*Lomatium columbianum*; Umbelliferae; coumarins; *trans-p*-coumaroyllomatin; *cis-p*-coumaroyllomatin; columbianadin; selinidin; lomatin; dihydrooroselol.

**Abstract**—Two new coumarins, obtained from the roots of *Lomatium columbianum* are shown to be *trans-p*-coumaroyllomatin and *cis-p*-coumaroyllomatin. Furthermore, the roots afforded columbianadin, selinidin, lomatin, (+)-dihydrooroselol, and a mixture of monoesters of khellactone.

### INTRODUCTION

*Lomatium columbianum* Mathias et Const. has previously been an object of chemical investigations. In 1956 Call and Green [1] investigated this plant and from a petroleum ether extract they isolated a crystalline substance named colubianin. No structural studies were reported on this substance. In 1964 Willette and Soine [2] isolated columbianadin (3) and a glycoside, columbianin (8) [3,4] from petroleum ether and alcoholic extracts, respectively.

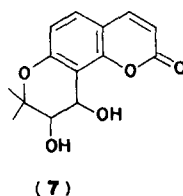
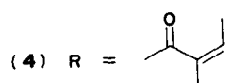
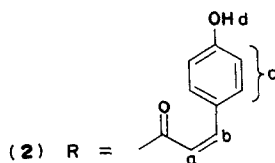
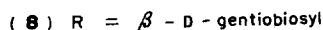
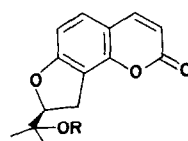
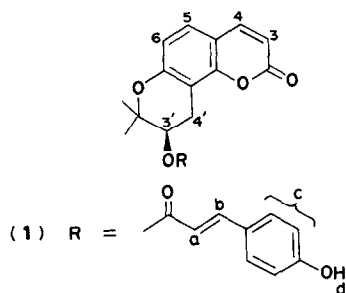
### RESULTS AND DISCUSSION

In our hands the ether extract of the roots, in addition to columbianadin (3), afforded two new coumarins (1) and (2), selinidin (4), lomatin (5), (+)-dihydrooroselol (6), and a mixture of polar coumarins believed to be monoesters of khellactone (7), the acid moieties being angelic, senecioic, and isovaleric acid.

All attempts to crystallize the coumarins (1) and (2) were unsuccessful. (1) appears as a colourless, glassy substance whereas (2) is a colourless, amorphous powder. From the UV, IR, and especially <sup>1</sup>H NMR spectra (1) and (2) appeared to be esters of lomatin (5) with *trans-p*-coumaric acid and *cis-p*-coumaric acid, respectively. The proton signals arising from the acid moieties of (1) and (2) were shown to be concordant with those from the

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† Present address: National Health Service Laboratory, Frederikssundsvej 378, DK-2700 Brønshøj, Denmark.



acid moieties of the methyl esters of authentic *trans-p*-coumaric acid and *cis-p*-coumaric acid, respectively. The *cis-p*-coumaric acid was obtained by photochemical inversion of the *trans* isomer.

Furthermore, saponification of (1) and (2) afforded lomatin (5) and in addition (1) yielded *trans-p*-coumaric acid whereas (2) gave a mixture of *cis*- and *trans-p*-coumaric acid.

The absolute configuration of lomatin (5) has been shown to be (*R*) [5] and accordingly the coumarins (1) and (2) are 3'(*R*)-3'-O-((*E*)-(3-(4-hydroxyphenyl)propenyl)-3',4'-dihydroreselin and 3'(*R*)-3'-O-((*Z*)-(3-(4-hydroxyphenyl)propenyl)-3',4'-dihydroreselin, respectively.

The possibility of (1) or (2) being an artifact formed by inversion of the other isomer during the isolation procedure has been considered. TLC of a newly prepared CH<sub>2</sub>Cl<sub>2</sub> extract of the roots disclosed that the extract contained both compounds. Accordingly (1) and (2) are believed to be naturally occurring.

The coumarins columbianadin (3), selinidin (4), lomatin (5), and (+)-dihydrooroselol (6) were identified by comparison with authentic materials.

#### EXPERIMENTAL

Mp's, IR and <sup>1</sup>H NMR spectra were determined as in a previous paper [6]. UV spectra were recorded in ethanol. TLC was carried out using Si gel GF<sub>254</sub>, Merck, as the adsorbent. Si gel (Merck, 0.05–0.20 mm) used for column chromatography was treated as earlier described [6].

*Plant material.* Dried and ground roots of *Lomatium columbianum* Mathias et Const. were delivered by professor T. O. Soine (see lit. [2]).

*Isolation and gross fractionation of the coumarin mixture.* The dried and ground roots (800 g) on extraction with Et<sub>2</sub>O and subsequent evaporation of the solvent afforded 24 g of an oily residue, which was dissolved in 90% MeOH and freed of lipids by extraction with petrol. The defatted extract (10 g) was chromatographed on Si gel (250 g). The eluent was CH<sub>2</sub>Cl<sub>2</sub>-CCl<sub>4</sub> (2:1) to which EtOAc was gradually added in amounts increasing to 20% after which MeOH (0.5 5%) was added. The gross fractionation divided the defatted extract into four main fractions, A, B, C, and D mentioned in the order eluted.

*Fraction A* (2.1 g) was chromatographed on Si gel using C<sub>6</sub>H<sub>6</sub> to which increasing amounts of EtOAc were added. The following compounds were obtained: a. Columbianadin (3) (1.1 g). Recrystallized from Et<sub>2</sub>O mp 118.5–119.0. [ $\alpha$ ]<sub>D</sub><sup>25</sup> + 225° (CHCl<sub>3</sub>; *c* 1.0) (lit. [7] mp 118.5–119.0. [ $\alpha$ ]<sub>D</sub><sup>25</sup> + 227°). The <sup>1</sup>H NMR and IR spectra were concordant with those of an authentic sample. b. A mixture (120 mg) of two blue fluorescent compounds. This fraction was mixed with fraction B. *Fraction B* (250 mg) was chromatographed several times on Si gel. The eluents were C<sub>6</sub>H<sub>6</sub>-EtOAc mixtures and CH<sub>2</sub>Cl<sub>2</sub>-CCl<sub>4</sub> (2:1) to which increasing amounts of EtOAc were added. In addition to columbianadin (3), selinidin (4) (86 mg) was obtained. Mp 94.5–96.0. [ $\alpha$ ]<sub>D</sub><sup>25</sup> - 18.0 (CHCl<sub>3</sub>; *c* 0.8) (lit. [8] mp 97–98°, lit. [9] [ $\alpha$ ]<sub>D</sub><sup>25</sup> - 17.8). The <sup>1</sup>H NMR and IR spectra were concordant with those of an authentic sample. *Fraction C* (ca 1 g) TLC on Si gel using CCl<sub>4</sub>-MeOH (8:2) as the eluent divided fraction C into 3 blue fluorescent spots. However, all attempts to separate this mixture by column chromatography (Si gel or acetylated cellulose, 20%) or by PLC were unsuccessful. Only mixtures of at least three different compounds were obtained. <sup>1</sup>H NMR spectra of fraction C and of the fractions obtained in the attempts to separate fraction C disclosed that all fractions were mixtures of monoesters of khellactone (7), the acid moieties being angelic, senecioic, and isovaleric acid. *Fraction D* (1.8 g) was chromatographed on acetylated cellulose (Mach-

ery-Nagel 2100 AC/ca 20%). The eluent was MeCOEt-Me<sub>2</sub>CO-H<sub>2</sub>O (1:3:5) and the sample/column packing ratio 1:150. Hereby fraction D was divided into two fractions D<sub>1</sub> and D<sub>2</sub>. D<sub>1</sub> was chromatographed on Si gel impregnated with 3% of H<sub>2</sub>O using CH<sub>2</sub>Cl<sub>2</sub>-CCl<sub>4</sub> (2:1) to which increasing amounts of EtOAc were added as the eluent. The following compounds were obtained: a. Lomatin (5) (18 mg); mp 182.5–183.5. [ $\alpha$ ]<sub>D</sub><sup>25</sup> + 50 (EtOH; *c* 0.2) (lit. [5] mp 182.5–183.5. [ $\alpha$ ]<sub>D</sub><sup>25</sup> + 52). The IR and <sup>1</sup>H NMR spectra were identical with those of an authentic sample. b. (+)-dihydrooroselol (6) (14 mg); mp 162.5–163.5. [ $\alpha$ ]<sub>D</sub><sup>25</sup> + 264 (MeOH; *c* 0.2) (lit. [5] mp 163.5–164.0. [ $\alpha$ ]<sub>D</sub><sup>25</sup> + 273). The IR and <sup>1</sup>H NMR spectra were identical with those of an authentic sample. D<sub>2</sub> was chromatographed in the same manner as described for D<sub>1</sub> followed by rechromatography of appropriate fractions on Si gel using CCl<sub>4</sub>-MeOH (97:3) as the eluent. The following substances were obtained: *c. trans-p*-coumaroyllomatin (1) (226 mg); a colourless glassy substance. [ $\alpha$ ]<sub>D</sub><sup>25.8</sup> + 370° (CHCl<sub>3</sub>; *c* 0.7).  $\lambda_{\text{max}}^{\text{OH}}$  nm (log  $\epsilon$ ) 318 (4.54), 302 (*sh*) (4.43). <sup>1</sup>H NMR (60 M Hz, CDCl<sub>3</sub>;  $\delta$  values; internal standard TMS): 1. The coumarin moiety: 6.18 (1 H, *d*  $J_{3,4}$  = 9.5 Hz, C-3), 7.62 (1 H, *d*,  $J_{3,4}$  = 9.5 Hz, C-4), 7.25 (1 H, *d*,  $J_{5,6}$  = 8.5 Hz, C-5), 6.78 (1 H, *d*,  $J_{5,6}$  = 8.5 Hz, C-6), 5.0–5.3 (1 H, slightly broadened *t*, C-3'), 2.9–3.2 (2 H, *m*, C-4'), 1.35 and 1.33 (6 H, two *s*, *gem* dimethyl protons). 2. The acid moiety: 6.15 (1 H, *d*,  $J_{\text{ab}}$  = 16 Hz, H<sub>a</sub>), 7.60 (1 H, *d*,  $J_{\text{ab}}$  = 16 Hz, H<sub>b</sub>), 7.35 and 6.82 (4 H, two *d*,  $J$  = 9 Hz, H<sub>c</sub>), 6.50 (1 H, broad *s*, H<sub>d</sub>). The signal at 6.50 disappeared on deuteration. *d. cis-p*-coumaroyllomatin (2) (16 mg), an amorphous powder, melting range 171.5–176.0. [ $\alpha$ ]<sub>D</sub><sup>25.3</sup> - 12° (CHCl<sub>3</sub>; *c* 1.0).  $\lambda_{\text{max}}^{\text{OH}}$  nm (log  $\epsilon$ ) 318 (4.50), 302 (*sh*) (4.41). <sup>1</sup>H NMR (60 M Hz, CDCl<sub>3</sub>;  $\delta$  values with reference to TMS): 1. The coumarin moiety: 6.20 (1 H, *d*,  $J_{3,4}$  = 9.5 Hz, C-3), 7.62 (1 H, *d*,  $J_{3,4}$  = 9.5 Hz, C-4), 7.23 (1 H, *d*,  $J_{5,6}$  = 8.5 Hz, C-5), 6.72 (1 H, *d*,  $J_{5,6}$  = 8.5 Hz, C-6), 5.0–5.3 (1 H, slightly broadened *t*, C-3'), 2.8–3.1 (2 H, *m*, C-4'), 1.31 and 1.28 (6H, two *s*, *gem* dimethyl protons). 2. The acid moiety: 5.73 (1 H, *d*,  $J_{\text{ab}}$  = 13 Hz, H<sub>a</sub>), 6.85 (1 H, *d*,  $J_{\text{ab}}$  = 13 Hz, H<sub>b</sub>), 7.50 and 6.77 (4 H, two *d*,  $J$  = 9 Hz, H<sub>c</sub>), 6.6 (1 H, broad *s*, H<sub>d</sub>). The signal at 6.6 disappeared on deuteration. Comparison of the <sup>1</sup>H NMR spectrum of (2) with that of *cis-p*-coumaric acid methylester confirmed the interpretation of the spectrum of (2) with respect to the acid moiety.

*Saponification of (1).* 31 mg of (1) were saponified by treatment with 0.5 N methanolic NaOH at 50° for 7 hr and worked up as previously described [10]. The yield of lomatin (5) was 15 mg, mp 183.5–184.5. [ $\alpha$ ]<sub>D</sub><sup>25.2</sup> + 53.7° (EtOH; *c* 0.2) (lit. [5], mp 182.5–183.5. [ $\alpha$ ]<sub>D</sub><sup>25</sup> + 52). The IR spectrum was identical with that of an authentic sample. In addition 4 mg of *trans-p*-coumaric acid, mp 207.5–209.5° (lit. [11] mp 210–213°) were obtained. The IR spectrum of the acid isolated was identical with that of an authentic sample.

*Saponification of 2.* 15 mg of (2) were saponified with 0.5 N methanolic NaOH at 50° for 21 hr as described above. The yield of lomatin (5) was 5 mg; mp 182.5–183.5. [ $\alpha$ ]<sub>D</sub><sup>25.7</sup> + 50.6° (EtOH; *c* 0.3). The identity was established by IR spectroscopy. The acid phase upon chromatography on Si gel (1 g) using CH<sub>2</sub>Cl<sub>2</sub>-CCl<sub>4</sub>-MeOH-HCOOH (120:70:10:1) as the eluent afforded 1 mg of a crystalline material with a melting range 125–195° (lit. [12] *cis-p*-coumaric acid mp 126–127°). IR spectroscopy of this material showed it to be a mixture of *cis*- and *trans-p*-coumaric acid.

*Synthesis of cis-p-coumaric acid.* The synthesis was carried out in analogy with the method used by Bregman *et al.* [13] in their synthesis of *p*-methoxy-*cis*-cinnamic acid from the *trans* isomer. 0.3 g of *trans-p*-coumaric acid were dissolved in 0.5% aqueous Na<sub>2</sub>CO<sub>3</sub>. The soln was deaerated with a stream of N<sub>2</sub> and subsequently irradiated for 200 min with a high pressure 125 W Hg immersion lamp. The reaction mixture was acidified with 4N HCl and extracted with Et<sub>2</sub>O. Chromatography on Si gel (25 g) using CH<sub>2</sub>Cl<sub>2</sub>-CCl<sub>4</sub>-MeOH-HCOOH (120:70:10:1) as the eluent afforded 80 mg of *cis-p*-coumaric acid, mp 127–128° recrystallized from C<sub>6</sub>H<sub>6</sub>)

(lit. [12] mp 126–127°). The compound gave satisfactory IR and <sup>1</sup>H NMR spectra.

*Methyl esters of cis- and trans-p-coumaric acid.* The esters were prepared in the usual manner using CH<sub>2</sub>N<sub>2</sub>. *Cis-p-coumaric acid methylester* mp 85.5–86° (Not described in lit.); *trans-p-coumaric acid methylester*, mp 136.5–137° (lit. [14] mp 136–137°).

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#### REFERENCES

1. Call, T. G. and Green, J. (1956) *Proc. Montana Acad. Sci.* **16**, 49.
2. Willette, R. E. and Soine, T. O. (1964) *J. Pharm. Sci.* **53**, 275.
3. Shipchandler, M. and Soine, T. O. (1967) *J. Pharm. Sci.* **56**, 661.
4. Shipchandler, M. and Soine, T. O. (1968) *J. Pharm. Sci.* **57**, 747.
5. Lemmich, J. and Nielsen, B. E. (1969) *Tetrahedron Letters* **3**.
6. Lemmich, E., Lemmich, J. and Nielsen, B. E. (1970) *Acta Chem. Scand.* **24**, 2893.
7. Nielsen, B. E. and Lemmich, J. (1964) *Acta Chem. Scand.* **18**, 1379.
8. Seshadri, T. R., Sood, M. S., Handa, K. L. and Vishwapaul (1964) *Tetrahedron Letters* 3367.
9. Lee, K. and Soine, T. O. (1968) *J. Pharm. Sci.* **57**, 865.
10. Lemmich, J., Lemmich, E. and Nielsen, B. E. (1966) *Acta Chem. Scand.* **20**, 2497.
11. Power, F. B. and Salway, A. H. (1914) *J. Chem. Soc.* **105**, 770.
12. Roth, W. A. and Stoermer, R. (1913) *Ber. Deut. Chem. Ges.* **46**, 260.
13. Bregman, J., Osaki, K., Schmidt, G. M. J. and Sonntag, F. J. (1964) *J. Chem. Soc.* 2021.
14. Konek, F. von and Pascu, E. (1918) *Ber. Deut. Chem. Ges.* **51**, 855.