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## Novel non-peptidic and small-sized BACE1 inhibitors

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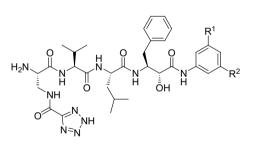
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Abstract—Recently, we reported substrate-based  $\beta$ -secretase (BACE1) inhibitors with a hydroxymethylcarbonyl (HMC) isostere as a substrate transition-state mimic. These inhibitors showed potent BACE1 inhibitory activities ( $\sim 1.2 \text{ nM IC}_{50}$ ). In order to improve in vivo enzymatic stability and permeability across the blood-brain barrier, these penta-peptidic inhibitors would need to be further optimized. On the other hand, non-peptidic inhibitors possessing isophthalic residue at the P2 position were reported from other research groups. We selected isophthalic-type aromatic residues at the P2 position and an HMC isostere at the P1 position as lead compounds. On the basis of the design approach focused on the conformer of docked inhibitor in BACE1, we found novel nonpeptidic and small-sized BACE1 inhibitors possessing a 2.6-pyridinedicarboxylic, chelidamic or chelidonic residue at the P<sub>2</sub> position. © 2008 Elsevier Ltd. All rights reserved.

Amyloid  $\beta$  (A $\beta$ ) peptide is a main component of senile plaques in the brains of Alzheimer's disease (AD) patients, that seems to cause ADs pathologies. According to the amyloid hypothesis,<sup>1</sup>  $\beta$ -secretase [BACE1:  $\beta$ -site APP (amyloid precursor protein) cleaving enzyme] appears promising as a molecular target for therapeutic intervention in AD,<sup>2-6</sup> because BACE1 triggers  $A\beta$  peptide formation by cleaving APP at the N-terminus of the A $\beta$  domain.<sup>7–12</sup> Recently, we reported potent penta-pep-tidic BACE1 inhibitors<sup>13–16</sup> containing a hydroxymethylcarbonyl isostere (HMC) as a substrate transitionstate mimic<sup>17</sup> as shown in Figure 1. Among them, KMI-429<sup>14</sup> exhibited effective inhibition of BACE1 activity in cultured cells, and significant reduction of Aß production in vivo (by direct administration into the hippocampuses of APP transgenic and wild-type mice).<sup>14b</sup> However, these inhibitors contained some natural amino acids that seemed to be required to improve enzymatic stability in vivo and permeability across the



**KMI-420** (R<sup>1</sup> = -H, R<sup>2</sup> = -COOH) IC<sub>50</sub> = 8.2 nM  
**KMI-429** (R<sup>1</sup> = -COOH, R<sup>2</sup> = -COOH) IC<sub>50</sub> = 3.9 nM  
**KMI-570** (R<sup>1</sup> = -H, R<sup>2</sup> = 
$$\bigvee_{\substack{N=N \\ N=N}}^{N}$$
) IC<sub>50</sub> = 4.8 nM  
**KMI-684** (R<sup>1</sup> =  $\bigvee_{\substack{N=N \\ N=N}}^{N}$ NH, R<sup>2</sup> =  $\bigvee_{\substack{N=N \\ N=N}}^{N}$ NH ) IC<sub>50</sub> = 1.2 nM

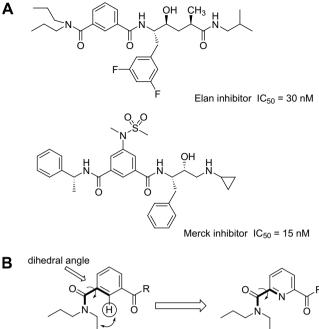
Figure 1. BACE1 inhibitors consisting of penta-peptides.

blood-brain barrier, so as to be practical anti-ADs drugs. Non-peptidic BACE1 inhibitors possessing isophthalic residue such as Elan's and Merck's inhibitors as shown in Figure 2A, were reported from other re-search groups.<sup>18,19</sup> Herein, we selected isophthalic-type aromatic residues at the P<sub>2</sub> position and an HMC isoste-

Keywords: Alzheimer's disease; BACE1; ß-Secretase; Non-peptidic B-ACE1 inhibitor.

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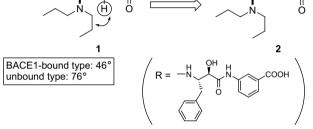
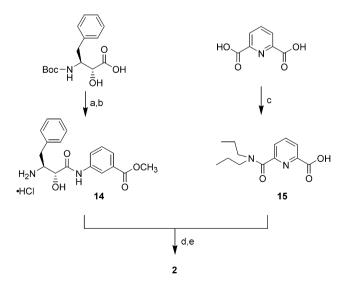


Figure 2. (A) Non-peptidic BACE1 inhibitors. (B) Design of smallsized BACE1 inhibitor 2 possessing heterocyclic ring at the  $P_2$  position.

re at the  $P_1$  position as lead compounds. Based on the design approach that a conformer of docked inhibitor in BACE1 was stabilized, we designed novel BACE1 inhibitors.

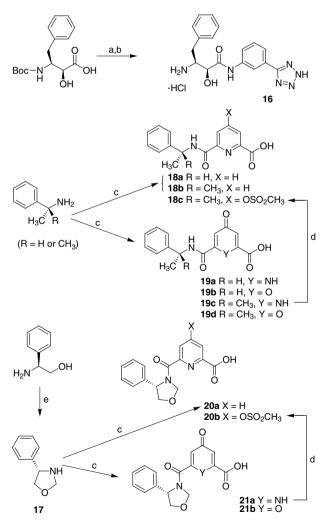
BACE1 inhibitors 2–13 were synthesized to connect in tandem the blocks corresponding to the  $P_3$ – $P_2$  residues



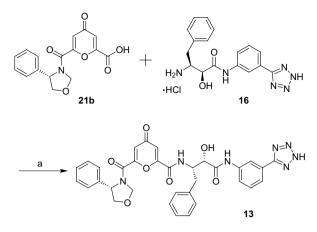
Scheme 1. Reagents and conditions: (a) methyl 3-aminobenzoate, EDC·HCl, HOBt/DMF, 85%; (b) anisole, 4 N HCl/dioxane, 99%; (c) dipropylamine, EDC·HCl, HOBt/DMF, 55%; (d) EDC·HCl, HOBt/ DMF, 91%; (e) 1 N NaOH/MeOH, 99% (crude).

and  $P_1-P_1'$  residues, respectively. Amide bonds were formed by traditional solution-phase synthesis methods using 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide·HCl (EDC·HCl) in the presence of 1-hydroxybenzotriazole (HOBt) as coupling agents. As shown in Scheme 1, after amide-bond formation, inhibitor 2 was synthesized by deprotection using alkaline hydrolysis. Synthesis of the blocks for the preparation of inhibitors 3-13 is shown in Scheme 2. 4-Methanesulfonyloxypyridine-2,5dicarboxlic derivatives 18c and 20b were synthesized from chelidamic derivatives 19c and 21a by tautomerization and an ensuing methanesulfonation using methanesulfonyl chloride in the presence of triethylamine. Blocks 20-21 possessing a 5-membered ring were synthesized from 17 that was prepared from (S)-2-phenylglycinol by cyclization with formaldehyde. As an example, the synthesis of inhibitor 13 is shown in Scheme 3. All of the inhibitors were purified by preparative RP-HPLC.

BACE1 inhibitory activity of the inhibitors was determined by enzymatic assay using a recombinant human



Scheme 2. Reagents and conditions: (a) 5-(3-aminophenyl)tetrazole, EDC·HCl, HOBt/DMF, 83%; (b) anisole, 4 N HCl/dioxane, 99%; (c) 2,6-pyridinedicarboxylic acid, chelidamic acid or chelidonic acid, EDC·HCl, HOBt/DMF, 55–65%; (d) methanesulfonyl chloride  $Et_3N/$ THF, 60–78%; (e) HCHO, water, 86%.

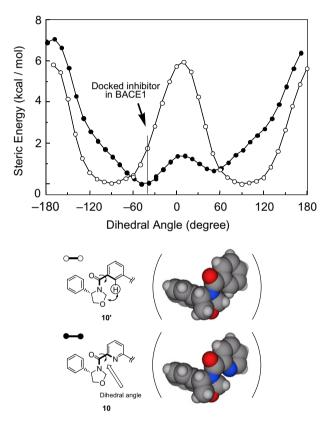


Scheme 3. Reagents: (a) EDC·HCl, HOBt/DMF, 90%.

BACE1 and FRET (fluorescence resonance energy transfer) substrate as previously reported.<sup>5</sup>

We first envisioned virtual inhibitor 1 possessing an isophthalic residue at the P<sub>2</sub> position and an HMC isostere, phenylnorstatine [Pns: (2R,3S)-3-amino-2-hydroxy-4phenylbutyric acid], at the P<sub>1</sub> position. A stable conformation docked in BACE1 (PDB ID: 1W51) was calculated using docking simulation software, MOE (Chemical Computing Group Inc., Canada). Second, a stable conformation of unbound inhibitor 1 was searched by calculating the steric energies of conformers around the bond of the  $P_3$  amide and  $P_2$  isophthalic residue in MM2 force field. The ideal dihedral-angles between the  $P_3$  amide and  $P_2$  isophthalic residue of inhibitor 1 in the BACE1-bound and unbound conformers were found to be quite different (BACE1-bound type: 46°, unbound type:  $76^{\circ}$ ). We thought that high-active inhibitors could be found by resolving this variation. Herein, we designed inhibitor 2 possessing a  $P_2$  heterocyclic residue, which lacks a hydrogen atom that would induce steric hindrance between the  $P_3$  moiety and  $P_2$  aromatic ring, as shown in Figure 2B. However, synthesized inhibitor 2 showed no BACE1 inhibitory activity.

Next, we designed inhibitors 3–5 possessing (R)- $\alpha$ -methylbenzylamino group at the P<sub>3</sub> position and allophenylnorstatine [Apns: (2S,3S)-3-amino-2-hydroxy-4-phenyl butyric acid], with syn-stereochemistry similar to Merck's BACE1 inhibitor (Fig. 2A), at the P<sub>1</sub> position. **3–5** showed moderate BACE1 inhibitory activity. We noticed the importance of the  $\alpha$ -methyl group at the P<sub>3</sub> position for BACE1 inhibitory activity. Inhibitors **6–9** possessing an  $\alpha, \alpha$ -dimethylbenzylamino group at the P<sub>3</sub> position were designed and showed improved inhibitory activity (Table 1). According to the crystal structure of isophthalic-type Merck's inhibitor (PDB ID: 2B8L, Fig. 3B) and the coordinates of docked inhibitors in BACE1 by our docking simulation study, inhibitors possessing a P<sub>3</sub> benzylamino-type residue displayed



**Figure 4.** Conformational study of BACE1 inhibitor **10** (KMI-1023). **10**' is virtual isophthalic analogue of inhibitor **10**.

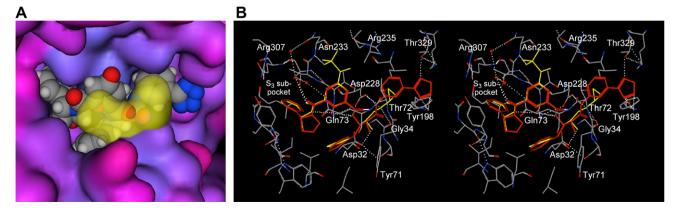


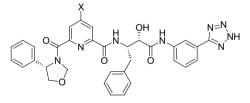
Figure 3. (A) Inhibitor 13 (KMI-1027, space-filling model) docked in BACE1. Molecular surface model indicates BACE1. The yellow transparent part in the center indicates a flap domain. (B) Stereoview of superimposed inhibitor 10 (KMI-1023, red lines) and Merck inhibitor (yellow lines) in BACE1(PDB ID: 2B8L). White dashed lines and red balls indicate hydrogen bond interactions and water molecules, respectively.

folding conformers between the P<sub>2</sub> ring and P<sub>3</sub> benzene rings to bind to the S<sub>3</sub> sub-pocket<sup>19</sup> in BACE1. We assumed that the two methyl groups stabilized this folding conformer by *gem*-dimethyl effect. However,  $\alpha,\alpha$ -dimethylbenzylamino group is labile under acidic conditions<sup>20</sup> similar to trityl group. Interestingly, HIV protease inhibitors with  $\alpha,\alpha$ -dimethylbenzyl amide showed low oral bioavailability, because of their acidinstability.<sup>21</sup> We designed inhibitors **10–13** introducing a 5-membered ring at the P<sub>3</sub> position, in order to improve acid-resistance and fix the folding pose between the P<sub>2</sub> and P<sub>3</sub> rings. Inhibitors **10–13** showed potent BACE1 inhibitory activity (Table 2). Modeled structures of docked inhibitors **10** and **13** in BACE1 (PDB ID: 2B8L) by docking simulation using MOE are shown in Figure 3. The P<sub>3</sub> benzene ring was confirmed to take



		$ \begin{array}{c}                                     $	H OH H N=N N NH	
	$\mathbb{R}^1$	$\mathbb{R}^2$	Х	BACE1 inhibition % at 2 $\mu$ M
3 6 7	CH <sub>3</sub> CH <sub>3</sub> CH <sub>3</sub>	H CH <sub>3</sub> CH <sub>3</sub>	H H OSO <sub>2</sub> CH <sub>3</sub>	66% 69% 84%
		$ \begin{array}{c}                                     $		
4 5 8 9	CH <sub>3</sub> CH <sub>3</sub> CH <sub>3</sub> CH <sub>3</sub>	H H CH <sub>3</sub> CH <sub>3</sub>	NH O NH O	51% 70% 65% 81%

Table 2. BACE1 inhibitors with a 5-membered ring at the P<sub>3</sub> position and their BACE1 inhibitory activities



Compound (KMI No.)	X	BACE1 inhibition %		IC <sub>50</sub> (nM)
		at 2 µM	at 0.2 µM	
<b>10</b> (KMI-1023)	Н	93%	63%	140
11 (KMI-1036)	$OSO_2CH_3$	96%	73%	96
			Ň	
12 (KMI-1030)	NH	83%	36%	360
13 (KMI-1027)	0	96%	78%	50

the form of a folding conformer against the  $P_2$  ring and tightly occupy the  $S_3$  sub-pocket of BACE1 (Fig. 3A). In addition, both ideal conformers around the bond of the  $P_3$  amide and  $P_2$  ring of unbound and BACE1-bound inhibitor **10** were confirmed to coincide. Interestingly, virtual isophthalic analogue **10'** in BACE1 showed high steric energy state caused by the interaction of the  $P_3$ moiety and ring's hydrogen atom (Fig. 4).

In conclusion, on the basis of the design approach focused on the conformer of a docked inhibitor in complex with BACE1, BACE1 inhibitors possessing a heterocyclic ring at the  $P_2$  position and 5-membered ring at the  $P_3$  position were designed. We found novel small-sized and non-peptidic BACE1 inhibitors. These inhibitors are expected improved membrane permeability and bioavailability as practical anti-ADs drugs. This design approach based on a conformer of enzyme-/receptorbound inhibitor/ligand, which was predicted in silico, was showed the usefulness of the drug discovery research, in contrast to exhaustive combinatorial synthesis.

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## **References and notes**

- (a) Hong, L.; Koelsch, G.; Lin, X.; Wu, S.; Terzyan, S.; Ghosh, A. K.; Zhang, X. C.; Tang, J. Science 2000, 290, 150; (b) Sipe, J. D. Annu. Rev. Biochem. 1992, 61, 947; (c) Selkoe, D. J. Ann. N. Y. Acad. Sci. 2000, 924, 17; (d) Steiner, H.; Capell, A.; Leimer, U.; Haass, C. Eur. Arch. Psychiatry Clin. Neurosci. 1999, 249, 266; (e) Selkoe, D. J. Ann. Med. 1989, 21, 73.
- (a) Ghosh, A. K.; Shin, D.; Downs, D.; Koelsch, G.; Lin, X.; Ermolieff, J.; Tang, J. J. Am. Chem. Soc. 2000, 122, 3522; (b) Ghosh, A. K.; Bilcer, G.; Harwood, C.; Kawahara, R.; Shin, D.; Hussain, K. A.; Hong, L.; Loy, J. A.; Nguyen, C.; Koelsch, G.; Ermolieff, J.; Tang, J. J. Med. Chem. 2001, 44, 2865.
- Tung, J. S.; Davis, D. L.; Anderson, J. P.; Walker, D. E.; Mamo, S.; Jewett, N.; Hom, R. K.; Sinha, S.; Thorsett, E. D.; John, V. J. Med. Chem. 2002, 45, 259.
- Tamamura, H.; Kato, T.; Otaka, A.; Fujii, N. Org. Biomol. Chem. 2003, 1, 2468.
- Shuto, D.; Kasai, S.; Kimura, T.; Liu, P.; Hidaka, K.; Hamada, T.; Shibakawa, S.; Hayashi, Y.; Hattori, C.; Szabo, B.; Ishiura, S.; Kiso, Y. *Bioorg. Med. Chem. Lett.* 2003, 13, 4273.
- (a) Ziora, Z.; Kimura, T.; Kiso, Y. Drugs Future 2006, 31, 53; (b) Nguyen, J.-T.; Yamani, A.; Kiso, Y. . Curr. Pharm. Des. 2006, 12, 4309.
- Vassar, R.; Bennett, B. D.; Babu-Khan, S.; Kahn, S.; Mendiaz, E. A.; Denis, P.; Teplow, D. B.; Ross, S.; Amarante, P.; Loeloff, R.; Luo, Y.; Fisher, S.; Fuller, J.;

Edenson, S.; Lile, J.; Jarosinski, M. A.; Biere, A. L.; Curran, E.; Burgess, T.; Louis, J. C.; Collins, F.; Treanor, J.; Rogers, G.; Citron, M. *Science* **1999**, *286*, 735.

- Yan, R.; Bienkowski, M. J.; Shuck, M. E.; Miao, H.; Tory, M. C.; Pauley, A. M.; Brashier, J. R.; Stratman, N. C.; Mathews, W. R.; Buhl, A. E.; Carter, D. B.; Tomasselli, A. G.; Parodi, L. A.; Heinrikson, R. L.; Gurney, M. E. *Nature* 1999, 402, 533.
- Sinha, S.; Anderson, J. P.; Barbour, R.; Basi, G. S.; Caccavello, R.; Davis, D.; Doan, M.; Dovey, H. F.; Frigon, N.; Hong, J.; Jacobson-Croak, K.; Jewett, N.; Keim, P.; Knops, J.; Lieberburg, I.; Power, M.; Tan, H.; Tatsuno, G.; Tung, J.; Schenk, D.; Seubert, P.; Suomensaari, S. M.; Wang, S.; Walker, D.; Zhao, J.; McConlogue, L.; John, V. *Nature* 1999, 402, 537.
- Hussain, I.; Powell, D.; Howlett, D. R.; Tew, D. G.; Meek, T. D.; Chapman, C.; Gloger, I. S.; Murphy, K. E.; Southan, C. D.; Ryan, D. M.; Smith, T. S.; Simmons, D. L.; Walsh, F. S.; Dingwall, C.; Christie, G. Mol. Cell. Neurosci. 1999, 14, 419.
- 11. Selkoe, D. J. Nature 1999, 399, A23.
- 12. Sinha, S.; Lieberburg, I. Proc. Natl. Acad. Sci. U.S.A. 1999, 96, 11049.
- Kimura, T.; Shuto, D.; Kasai, S.; Liu, P.; Hidaka, K.; Hamada, T.; Hayashi, Y.; Hattori, C.; Asai, M.; Kitazume, S.; Saido, T. C.; Ishiura, S.; Kiso, Y. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 1527.
- (a) Kimura, T.; Shuto, D.; Hamada, Y.; Igawa, N.; Kasai, S.; Liu, P.; Hidaka, K.; Hamada, T.; Hayashi, Y.; Kiso, Y. *Bioorg. Med. Chem. Lett.* 2005, *15*, 211; (b) Asai, M.; Hattori, C.; Iwata, N.; Saido, T. C.; Sasagawa, N.; Szabó, B.; Hashimoto, Y.; Maruyama, K.; Tanuma, S.; Kiso, Y.; Ishiura, S. J. Neurochem. 2006, *96*, 533.
- Kimura, T.; Hamada, Y.; Stochaj, M.; Ikari, H.; Nagamine, A.; Abdel-Rahman, H.; Igawa, N.; Hidaka, K.; Nguyen, J.-T.; Saito, K.; Hayashi, Y.; Kiso, Y. *Bioorg. Med. Chem. Lett.* 2006, 16, 2380.
- Hamada, Y.; Igawa, N.; Ikari, H.; Ziora, Z.; Nguyen, J.-T.; Yamani, A.; Hidaka, K.; Kimura, T.; Saito, K.; Hayashi, Y.; Ebina, M.; Ishiura, S.; Kiso, Y. *Bioorg. Med. Chem. Lett.* 2006, 16, 4354.
- (a) Mimoto, T.; Imai, J.; Tanaka, S.; Hattori, N.; Takahashi, O.; Kisanuki, S.; Nagano, Y.; Shintani, M.; Hayashi, H.; Akaji, K.; Kiso, Y. *Chem. Pharm. Bull.* 1991, *39*, 2465; (b) Mimoto, T.; Imai, J.; Tanaka, S.; Hattori, N.; Kisanuki, S.; Akaji, K.; Kiso, Y. *Chem. Pharm. Bull.* 1991, *39*, 3088.
- Hom, R. K.; Gailunas, A. F.; Mamo, S.; Fang, L. Y.; Tung, J. S.; Walker, D. E.; Davis, D.; Thorsett, E. D.; Jewett, N. E.; Moon, J. B.; John, V. J. Med. Chem. 2004, 47, 158.
- Stachel, S. J.; Coburn, C. A.; Steele, T. G.; Jones, K. G.; Loutzenhiser, E. F.; Gregro, A. R.; Rajapakse, H. A.; Lai, M.-T.; Crouthamel, M.-C.; Xu, M.; Tugusheva, K.; Lineberger, J. E.; Pietrak, B. L.; Espeseth, A. S.; Shi, X.-P.; Chen-Dodson, E.; Holloway, M. K.; Munshi, S.; Simon, A. J.; Kuo, L.; Vacca, J. P. J. Med. Chem. 2004, 47, 6447.
- α,α-Dimethylbenzyl (cumyl) group is recognized as a Nprotecting group of amides that is cleavable by acid, such as TFA or formic acid. e.g. 'Wuts, P. G. M.; Greene, T. W. *Greene's PROTECTIVE GROUPS in ORGANIC SYS-THESIS*, 4th ed., John Wiley & Sons, Inc, 2007, p. 907.
- Reich, S.; Melnick, M.; Davies, J. F., II; Appelt, K.; Lewis, K. K.; Fuhry, M. A.; Pino, M.; Trippe, A. J.; Nguyen, D.; Dawson, H.; Wu, B.-W.; Musick, L.; Kosa, M.; Kahil, D.; Webber, S.; Gehlhaar, D. K.; Andrada, D.; Shetty, B. Proc. Natl. Acad. Sci. U.S.A. 1995, 92, 3298.