

Preliminary communication

Synthesis of α - and β -(2 $\rightarrow$ 9)-linked disialylglycerolipids*

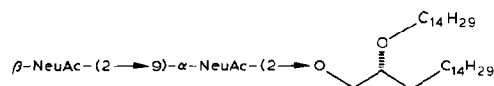
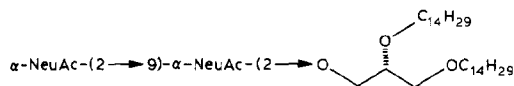
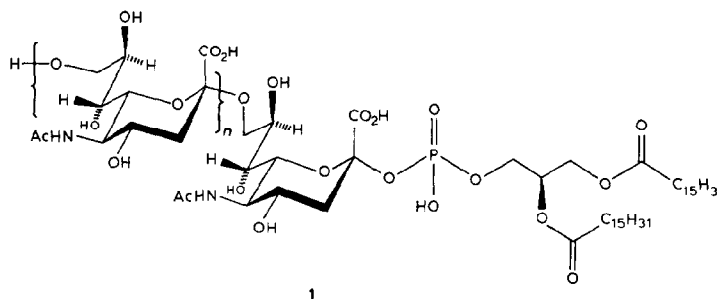
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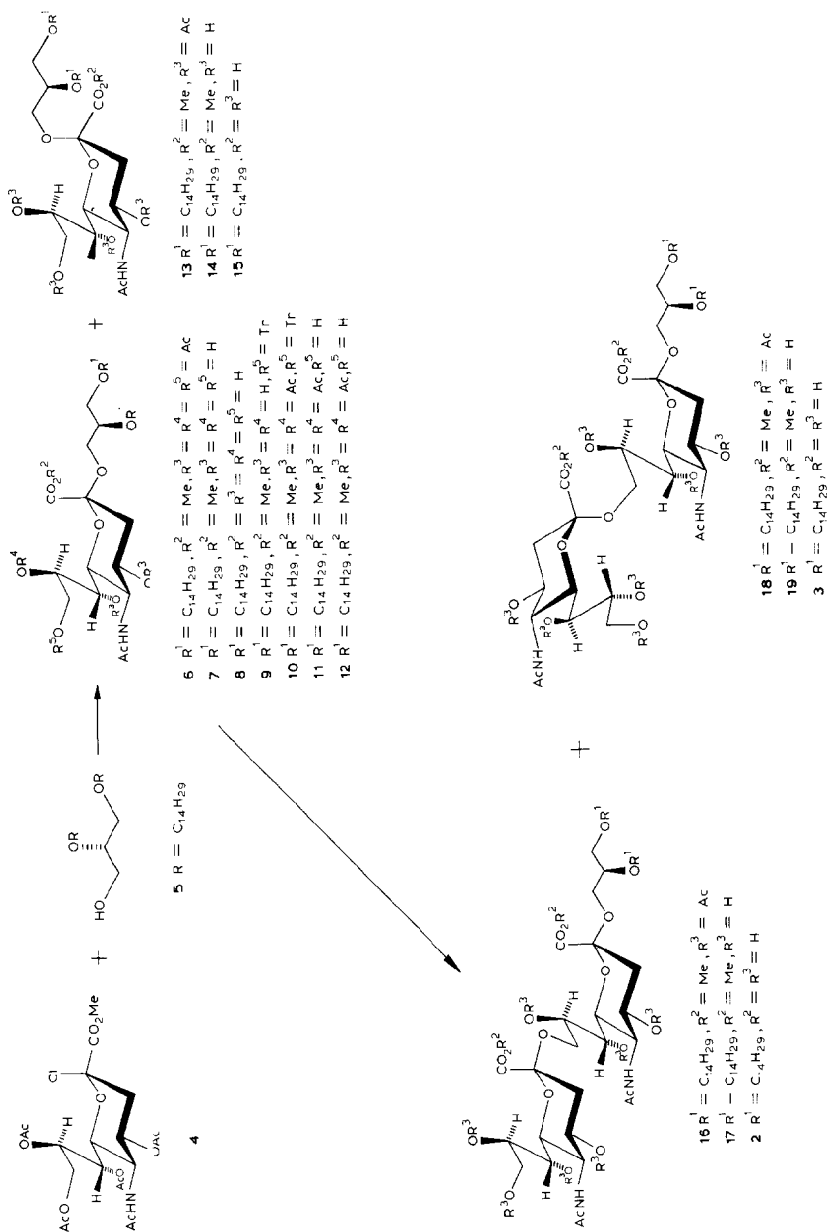
Oligosialyl structures² have been reported to occur in glycoproteins, glycolipids, and bacterial polysaccharides. In surface antigens of pathogenic bacteria, two types of sialic acid-containing polymers are known: α -(2 $\rightarrow$ 8)- and α -(2 $\rightarrow$ 9)-linked sialic acid³. In 1981, the α -(2 $\rightarrow$ 9)-linked polysialyl phospholipid structure 1 was proposed for the group C meningococcal polysaccharide⁴.

As part of a project on the synthesis of glyceroglycolipids⁵, we report here a regioselective synthesis of the α - and β -(2 $\rightarrow$ 9)-linked disialyl glycerolipids 2 and 3 as model glycolipids for 1.



*Synthetic Studies on Cell-surface Glycans, Part XXVIII. For Part XXVII, see ref. 1.

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Scheme 1

Glycosidation of the di-*O*-alkylated glycerol **5** (ref. 6) with the glycosyl donor **4** (ref. 7) in the presence of 1:1:2 Hg(CN)₂–HgBr₂–powdered molecular sieves **4A** in Cl(CH₂)₂Cl for 48 h at 20° gave a mixture of the anomers. Separation by flash chromatography⁸ over silica gel C-300 in toluene–EtOAc afforded **6** and **13** in 37 and 24% yield, respectively: **6**, [α]_D^{*} –9.3° (c 1.04); R_F 0.26 in 1:1 toluene–EtOAc; δ_H (CDCl₃): 4.85 (m, 1 H, H-4), 2.60 (q, 1 H, *J* 4.80 and 13.10 Hz, H-3e), and 1.97 (t, 1 H, *J* 12.8 Hz, H-3a); **13**, [α]_D –13.0° (c 1.00); R_F 0.36 in 1:1 toluene–EtOAc; δ_H (CDCl₃): 5.23 (m, 1 H, H-4), 2.45 (q, 1 H, *J* 4.8 and 12.5 Hz, H-3e), and 1.90 (t, 1 H, *J* 12.2 Hz, H-3a).

The anomeric configurations of **6** and **13** were assigned by the presence in the ¹H-n.m.r. spectrum of the more deshielded signal for H-4 of **13** compared with that of **6**, according to the observation reported by Paulsen and Tietz⁹.

Deprotection of **6** and **13** [(i) NaOMe–MeOH, (ii) 0.1M NaOH–THF] afforded **8** and **15** in 43 and 58% overall yield, respectively, *via* **7** and **14**: **8**, R_F 0.51 in 2:1:1 BuOH–EtOH–H₂O; δ_H (CD₃OD): 2.75 (q, 1 H, *J* 4.4 and 12.7 Hz, H-3e), 2.00 (s, 3 H, Ac), and 1.71 (t, 1 H, *J* 12.5 Hz, H-3a); **15**, R_F 0.67 in 2:1:1 BuOH–EtOH–H₂O; δ_H (CD₃OD): 2.40 (bm, 1 H, H-3e) and 2.02 (s, 3 H, Ac).

In order to synthesize the target disialyl glycerolipids **2** and **3**, compound **7** {[α]_D –5;30° (c 1.0); R_F 0.26 in 1:9 MeOH–EtOAc} was transformed into acetate **11** in 52% overall yield, in 3 steps: (i) TrCl–pyridine, (ii) Ac₂O–pyridine, (iii) 90% aq. AcOH for 2 h at 50–60°; **9**: [α]_D –5.20° (c 1.0); R_F 0.17 in 1:1 toluene–EtOAc; δ_H (CDCl₃): 7.5–7.1 (m, 15 H, CPh₃), 3.76 (s, 3 H, OMe), and 1.94 (s, 3 H, Ac); δ_C (CDCl₃): 98.79 (C-2) and 86.60 (CPh₃). Compound **10**: [α]_D +2.92° (c 1.20); R_F 0.48 in 1:1 toluene–EtOAc; δ_H (CDCl₃): 2.59 (q, 1 H, *J* 4.7 and 12.7 Hz, H-3e), and 1.90 (t, 1 H, *J* 12.4 Hz, H-3a); δ_C (CDCl₃): 98.64 (C-2) and 86.51 (CPh₃). Compound **11**: [α]_D –17.2° (c 0.50); R_F 0.52 in 20:1 EtOAc–MeOH; δ_H (CDCl₃): 5.17 (m, 1 H, H-8), 5.10 (bd, 1 H, *J* 9.8 Hz, H-7), 3.31 (q, 1 H, *J* 5.6 and 10.3 Hz, H-9), 3.00 (q, 1 H, *J* 4.8 and 10.3 Hz, H-9'), 2.60 (q, 1 H, *J* 4.5 and 12.7 Hz, H-3e), and 2.00 (t, 1 H, *J* 12.5 Hz, H-3a). Besides **11**, a product of acetyl migration was also isolated in 14% yield, and the structure was assigned to be **12** {[α]_D –12.3° (c 0.61); R_F 0.62 in 20:1 EtOAc–MeOH} according to the ¹H-n.m.r. data, which showed a deshielded signal at δ 5.14 for H-7, but not for H-8 (ref. 10).

Glycosidation of **11** with **4**, as for **5**, afforded a mixture of **16** and **18** in 41% yield. Flash chromatography of the product on silica gel C-300 in 5:3 CCl₄–acetone gave **16** and **18** in 17.1 and 9.2% yield, respectively: **16**, [α]_D –13.14° (c 1.41); R_F 0.47 in 1:1 CCl₄–acetone; δ_H 4.88 (oct, 1 H, H-4), 4.84 (oct, 1 H, H-4'), 3.80 (s, 6 H, 2 MeO), 2.60 (q, 1 H, *J* 4.6 and 12.7 Hz, H-3e), 2.57 (q, 1 H, *J* 4.6 and 12.7 Hz, H-3'e), 1.96 (t, 1 H, *J* 12.6 Hz, H-3a), and 1.93 (t, 1 H, *J* 12.4 Hz, H-3'a); **18**, [α]_D –4.26° (c 0.47); R_F 0.56 in 1:1 CCl₄–acetone; δ_H (CDCl₃): 5.09 (sex, 1 H, H-4b), 4.88 (oct, 1 H, H-4a), 3.80 (s, 3 H, MeO), 3.79 (s, 3 H, MeO), 2.60 (q, 1 H, *J* 4.6 and 12.9 Hz, H-3a-e), 2.45 (q, 1 H, *J* 4.9 and 12.9 Hz, H-3b-e), 1.95 (t, 1 H, *J* 12.7 Hz, H-3a-a), and 1.82 (q, 1 H, *J* 11.7 and 12.9 Hz, H-3b-a).

Finally, both **16** and **18** were deacetylated (MeONa–MeOH) and saponified (NaOH–aq. MeOH), to give **2** and **3**, respectively, *via* **17** (R_F 0.57 in 2:1:1 BuOH–EtOH–

*Values of [α]_D were measured for CHCl₃ solutions at 25°, unless noted otherwise. Compounds having [α]_D recorded gave satisfactory data for elemental analyses.

H₂O) and 19 (R_F 0.67 in 2:1:1 BuOH–EtOH–H₂O); 2: $[\alpha]_D +2.22^\circ$ (c 0.45, MeOH); R_F 0.50 in 2:1:1 BuOH–EtOH–H₂O; δ_H (CD₃OD): 2.80 (bq, 2 H, J 4.7 and 12.0 Hz, 2 H-3e), 2.01 (s, 3 H, Ac), and 2.00 (s, 3 H, Ac); 3: $[\alpha]_D -8.5^\circ$ (c 0.34, MeOH); R_F 0.54 in 2:1:1 BuOH–EtOH–H₂O; δ_H (CD₃OD): 2.73 (q, 1 H, J 4.5 and 12.0 Hz, H-3a-e), 2.41 (q, 1 H, J 4.5 and 12.1 Hz, H-3b-e), and 2.01 (s, 6 H, 2 Ac).

In conclusion, by use of a stepwise approach, the disialylglycerolipids 2 and 3 were synthesized, and their ¹H-n.m.r. data were in good agreement¹¹ with the anomeric configurations assigned.

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REFERENCES

- 1 T. Kitajima, M. Sugimoto, T. Nukada, and T. Ogawa, *Carbohydr. Res.*, 127 (1984) C1–C4.
- 2 A.P. Corfield and R. Schauer, in R. Schauer (Ed.), *Sialic Acids*, Springer-Verlag, Wien, 1982, pp. 5–50.
- 3 A. K. Bhattacharjee, H. J. Jennings, C. P. Kenny, A. Martin, and I. C. P. Smith, *J. Biol. Chem.*, 250 (1975) 1926–1932; H. J. Jennings and A. K. Bhattacharjee, *Carbohydr. Res.*, 55 (1977) 105–112; W. Egan, T.-Y. Lui, D. Dorow, J. S. Cohen, J. D. Robbins, E. C. Gotschlich, and J. B. Robbins, *Biochemistry*, 16 (1977) 3687–3692.
- 4 E. C. Gotschlich, B. A. Fraser, O. Nishimura, J. B. Robbins, and T.-Y. Lui, *J. Biol. Chem.*, 256 (1981) 8915–8921.
- 5 T. Ogawa and K. Beppu, *Carbohydr. Res.*, 101 (1982) 271–277.
- 6 E. Baer and N. Z. Stanacev, *J. Biol. Chem.*, 240 (1965) 44–48; M. Kates, T. H. Chan, and N. Z. Stanacev, *Biochemistry*, 2 (1963) 394–397; T. Ogawa and K. Beppu, *Agric. Biol. Chem.*, 46 (1982) 255–262.
- 7 R. Kuhn, P. Lutz, and D. L. MacDonald, *Chem. Ber.*, 99 (1966) 611–617.
- 8 W. C. Still, M. Kahn, and A. Mitra, *J. Org. Chem.*, 43 (1978) 2923–2925.
- 9 H. Paulsen and H. Tietz, *Angew. Chem., Int. Ed. Engl.*, 21 (1982) 927–928.
- 10 J. Haverkamp, H. van Halbeek, L. Dorland, J. F. G. Vliegthart, R. Pfeil, and R. Schauer, *Eur. J. Biochem.*, 122 (1982) 305–311.
- 11 E. B. Brown, W. S. Brey, Jr., and W. Weltner, Jr., *Biochim. Biophys. Acta*, 399 (1975) 124–130; U. Dabrowski, H. Friebohn, R. Brossmer, and M. Supp, *Tetrahedron Lett.*, (1979) 4637–4640; J.-M. Beau, R. Schauer, J. Haverkamp, L. Dorland, J. F. G. Vliegthart, and P. Sinaÿ, *Carbohydr. Res.*, 82 (1980) 125–129; L. W. Jaques, B. F. Riesco, and W. Weltner, Jr., *ibid.*, 83 (1980) 21–32.