# ACS Medicinal Chemistry Letters

Letter

Subscriber access provided by UNIV OF TASMANIA

# Optimization of ADME Properties for Sulfonamides Leading to the Discovery of a T-Type Calcium Channel Blocker ABT-639

Qingwei Zhang, Zhiren Xia, Shailen Joshi, Victoria E Scott, and Michael F Jarvis

ACS Med. Chem. Lett., Just Accepted Manuscript • DOI: 10.1021/acsmedchemlett.5b00023 • Publication Date (Web): 28 Apr 2015 Downloaded from http://pubs.acs.org on April 29, 2015

## **Just Accepted**

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a free service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are accessible to all readers and citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



ACS Medicinal Chemistry Letters is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

# **Optimization of ADME Properties for Sulfonamides Leading to the Discovery of a T-Type Calcium Channel Blocker ABT-639**

Qingwei Zhang,\* Zhiren Xia, Shailen Joshi, Victoria E. Scott and Michael F. Jarvis Neuroscience Research, AbbVie, 1 North Waukegan Rd., North Chicago, IL, 60064 USA **KEYWARDS**: Ca<sub>y</sub>3.2, T-type calcium channel, pain, Sulfonamides, ADME

ABSTRACT: The discovery of a novel peripherally acting and selective Ca, 3.2 T-type calcium channel blocker, ABT-639 is described. HTS hits 1 and 2 which have poor metabolic stability were optimized to obtain 4, which has improved stability and oral bioavailability. Modification of 4 to further improve ADME properties led to the discovery of ABT-639. Following oral administration, ABT-639 produces robust antinociceptive activity in experimental pain models at doses that do not significantly alter psychomotor or hemodynamic function in the rat.

Voltage-gated calcium channels (VGCC) play an important role in the regulation of calcium influx into cells in response to change membrane conductance, thereby activating various physiological functions, such as neurotransmitter release, cellular excitability, muscle contraction and many others.<sup>1</sup> These channels can be classified into low-voltage activated Ttype and high-voltage activated L-type and P/Q-, N- and Rtypes calcium channels. N-type calcium channels are found primarily at presynaptic terminals and are involved in neurotransmitter release.<sup>2,3</sup> T-type channels are primarily involved in postsynaptic excitability.<sup>4</sup> Recent studies have shown that T-type calcium channels may be important therapeutic targets for the treatment of several neurophysiological disorders, including as epilepsy,<sup>5</sup> pain,<sup>6,7,8</sup> hypertension,<sup>9</sup> sleep architecture,<sup>10</sup> tremor,<sup>11</sup> and Parkinson's disease.<sup>12, 13</sup>

 $Ca_v 3.2$  is the predominant T-type calcium channel isoform in sensory nerves that modulate nociception, and is expressed in dorsal root ganglion (DRG) neurons, peripheral receptive fields, spinal cord dorsal horn and brain.<sup>14</sup> Bourinet et al demonstrated that silencing of Cav3.2 channel strongly reduced acute and neuropathic nociception.<sup>15</sup> Intrathecal administration or local injection of Ca<sub>v</sub>3.2-specific, but not and Ca<sub>v</sub>3.3-specific antisense oligonucleotides Ca. 3.1 produces a significant knockdown of Ca<sub>v</sub>3.2 T-type currents in nociceptive DRG neurons, and robust long-lasting and reversible mechanical and thermal antinociceptive effects. Jagodic et al. demonstrated that following chronic constriction injury (CCI) of the sciatic nerve induced upregulation of Ttype calcium channel currents in small rat DRG.16 Modulation of the  $Ca_v 3.2$  ( $\alpha 1H$ ) channel controls the sensitization of nociceptors, the peripheral pain-sensing neurons.<sup>17</sup> These results further support Ca<sub>v</sub>3.2 T-type channels as a mechanism for modulating nociceptive sensitivity.

High throughput screening (HTS) generated a number of sulfonamides hits including 1 (IC<sub>50</sub> = 3  $\mu$ M) and 2 (IC<sub>50</sub> = 5 μM) (Figure 1) against Ca<sub>v</sub>3.2 T-type channel in a FLIPR based  $Ca^{2+}$  flux assay.<sup>18,19</sup> However, HTS hits 1 and 2 are metabolically unstable in rats and both have very poor oral bioavailability (F = 0.5% and 1.9\%, respectively) (Table 1).



Since the potency of 1 and 2 were in a similar range to other previously described T/N-type calcium channel blockers, lead optimization efforts were focused on improving ADME properties for these hits.<sup>20</sup> Modification of both sulfonamide and amide sides of these hits led to identification of a new lead 3, which has a lower cLogP than 1 and 2. Compound 3 afforded 29% oral bioavailability in rats. Unfortunately, the half-life  $(t_{1/2})$  of **3** following i.v. dosing at 5  $\mu$ mol/kg was only 0.31 hr, mainly due to the high plasma clearance rate (CLp) of 1.66 L/hr/kg and a low volume distribution ( $V_{\beta}$ ) of 0.75 L/kg (Table 1).

and 4								
	RLM <sup>a</sup>	cLogP	$V_{\beta}$	CLp	t <sub>1/2</sub>	F (%)		
	(%)		(L/kg)	(L/hr/kg)	(hr)	(p.o.)		
1	0.3	4.4	4.94	1.90	1.80	0.5 <sup>b</sup>		
2	0.1	5.3	11.1	3.47	2.16	1.9 <sup>b</sup>		
3	46	3.1	0.75	1.66	0.31	29.0 °		

Table 1. RLM and PK Parameters of Compounds 1, 2, 3

<sup>a</sup>Rat Liver Microsomal stability. Percentage remaining after 30 min at  $1\mu$ M.<sup>b</sup>3  $\mu$ M/kg iv and po.<sup>c</sup>5  $\mu$ M/kg iv and 30  $\mu$ M/kg po. Oral formulation: PEG400: Cremophor EL: Oleic Acid (10:10:80, by weight 2 ml/kg).

1.79

0.25

31.3 °

0.65

After evaluating several diamines, a rigid bicyclic diamine was identified to replace dimethylmorpholine in 3. Compound

4

68

2.8

4 demonstrated better stability in rat liver microsomes compared to 1, 2 and 3. However, the plasma clearance rate of 4 is still high (1.79 L/hr/kg) with a volume distribution ( $V_{\beta}$ ) of 0.65 L/kg in rats. In order to further improve the ADME properties and PK profile of 4, compounds 5 to 9 and the (S)enantiomers 4b to 6b with different R<sub>2</sub> groups which have electron-withdrawing substituents on anilines were investigated to compare their plasma clearance (CLp) and oral bioavailability (Table 2). We observed that the plasma clearance (CLp) rate of the (R)-enantiomers (4, 5, and 6) is lower than the (S)-enantiomers (4b, 5b and 6b), and the oral bioavailability in rats is improved for the (R)-enantiomers. i.e. compound 4b, the (S)-enantiomer of 4 shows higher plasma clearance (3.36 L/hr/kg) than (R)-enantiomer 4. Compound 7 with 2-chloroaniline had increased CLp, and decreased bioavailability compared to 4, but compounds 5, 8, 6 and 9 with 2.3-. 2,6-difluoroaniline, 4and 2-(trifluoromethyl)aniline had lower CLp and higher bioavailability. Compounds 6 and 6b have an excellent PK profile with CLp of 0.45 - 0.81 L/hr/kg and oral bioavailability of 76 - 100%. However, compound 4 is the only one in the Table 2 which shows  $IC_{50} = 10.6 \mu M$  potency, other compounds in Table 2 are weak Cav3.2 T-type calcium channel blockers (<30% inhibition @10 µM). Our next attempt was to add the halogen atoms to the central aromatic ring since it is possible that the high plasma clearance rate was also due to the metabolic oxidation of the central aromatic ring.

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22 23

24

25

26

27

28 29

30

31

32 33

44

45

46

47

48 49

50

51

52

53 54 55

56

57

58

59

60

 Table 2.<sup>a</sup>
 Pharmacokinetic Parameters of Compounds 4 to

 9 and 4b to 6b

Compound	$\mathbb{R}^1$	R <sup>2</sup>	Plasma clearance CLp (L/hr/kg)	bioavailability (p.o.) F (%)
4	<b>N</b> ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓	rot F	1.79	31
4b		<sup>3</sup> <sup>3</sup> <sup>4</sup> <sup>N</sup> − F	3.36	28
5	N H R	P <sup>AR</sup> N F	1.21	44
5b		P <sup>Ar</sup> N F	2.28	29
6	N H N prie	r <sup>k</sup> N H CF₃	0.45	100
6b	K N N N N N N N N N N N N N N N N N N N	r <sup>ser</sup> N − CF <sub>3</sub>	0.81	76
7	N H N P	A L C	1.93	5.8
8	H Roke	F F NH F	0.92	54
9	H R R R R R R R R R R R R R R R R R R R	P <sup>A</sup> N CF <sub>3</sub>	0.74	57

 $<sup>^{</sup>a}5~\mu M/kg$  iv and 30  $\mu M/kg$  po. Oral formulation: PEG400: Cremophor EL: Oleic Acid (10:10:80, by weight 2 ml/kg).

Substituent on aromatic ring can influence the microsomal stability and pharmacokinetic properties of the compounds. Introduction of the F or Cl atom to the central aromatic ring of 4 led to the discovery of ABT-639 (Figure 2), which has a significantly decreased plasma clearance rate of 0.55 L/hr/kg. The volume distribution  $(V_{\beta})$  was increased to 2.7 L/kg, and the half-life  $(t_{1/2})$  of ABT-639 was improved to 3.3 hr in rats. The oral bioavailability in rats was also significant improved (F = 73%). The increase in volume distribution in rat and monkey may be due to the increase of tissue-binding or partitioning into fat since the cLogP (3.8) of ABT-639 is larger than the cLogP (1.79) of  $4^{.22}$  Compound 10 with dichloro-substitutes on the central aromatic ring was prepared and showed weaker potency (IC<sub>50</sub> = 19  $\mu$ M) against Ca<sub>v</sub>3.2 Ttype calcium channel with 44% oral bioavailability. Addition of a methyl group to the sulfonamide side of ABT-639 gives compound 11 (Figure 2). Remarkably, compound 11 had decreased stability in rat (RLM) and human liver microsomes (HLM) from 81 - 95% to less than 0.01%. The (S)-enantiomer of ABT-639 was also synthesized, it has 57% oral bioavailability in rats.



Scheme 1. Synthesis of ABT-639 and Compounds 4 -11<sup>a</sup>



<sup>a</sup>Reagents and conditions: (a) oxalyl chloride, CH<sub>2</sub>Cl<sub>2</sub>, RT, overnight; (b) (R)-octahydropyrrolo[1,2-*a*]pyrazine, Na<sub>2</sub>CO<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, RT, 5-18 hrs; (c) 2-fluoroaniline or other substitutedanilines, RT, overnight. (d) 2-fluoro-*N*-methylaniline, RT, overnight.

Synthesis of ABT-639 and its analogs is outlined in Scheme 1. ABT-639 was obtained in 77% overall yield in 3 steps with one step purification from commercially available starting materials. 2-chloro-5-(chlorosulfonyl)-4-fluorobenzoyl chloride was prepared by reaction of 2-chloro-5-(chlorosulfonyl)-4-fluorobenzoic acid with oxalyl chloride at room temperate in the presence of DMF as a catalyst.

**ACS Paragon Plus Environment** 

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28 29

30

31

32

33 34 35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55 56

57

58

59

60

Addition of one equivalent of (*R*)-octahydropyrrolo[1,2*a*]pyrazine to this resulting benzoyl chloride slowly over 1 hr generated the amide intermediate, (*R*)-4-chloro-2-fluoro-5-(octahydropyrrolo[1,2-a]pyrazine-2-carbonyl)benzene-1-

sulfonyl chloride. Since the amide formation is at a much faster rate than the sulfonamide formation, very little or no side products were detected by LC-MS. Subsequent sulfonamide formation was completed overnight by addition of 2-fluoroaniline to afford ABT-639 after purification by chromatography. Compound 10 was prepared starting from available 2.4-dichloro-5commercially (chlorosulfonyl)benzoic acid by following the same synthetic route of preparation of ABT-639. Compound 11 was obtained in 71% overall yield by using the same procedures. 2-fluoro-*N*-methylaniline was used at the last step (Scheme 1, step d). Compounds 3 to 9 and 4b to 6b were synthesized in 50 - 89% vield by a one-pot reaction from the commercially available 3-(chlorosulfonyl)benzoyl chloride by following the same procedures (Scheme 1, steps b and c).

ABT-639 is a selective voltage-dependent Ca<sub>v</sub>3.2 T-type calcium channel blocker. It blocks human T-type (Ca<sub>v</sub>3.2) channels with IC<sub>50</sub> = 2.3  $\mu$ M, and also blocks low voltage activated currents in native rat DRG neurons (IC<sub>50</sub> = 7.6  $\mu$ M).<sup>17</sup> ABT-639 shows little or no activity at other calcium channels (L-type, N-type, and P/Q-type) and is inactive (IC<sub>50</sub> > 10  $\mu$ M) across a wide array of cell surface receptors and ion channels.<sup>18</sup>

Table 3.Pharmacokinetic profile of ABT-639 acrossspecies

species	CLp	V <sub>β</sub>	t <sub>1/2</sub>	F	Microsomal		
	(L/hr/kg)	(L/kg)	(hr)	(%) <sup>a</sup>	Stability (%)		
rat <sup>b</sup>	0.55	2.7	3.3	73	81		
dog <sup>c</sup>	0.045	0.3	4.9	88	100		
monkey <sup>c</sup>	0.11	1.35	8.3	95	88		
<sup>a</sup> Oral formulation: PEG400: Cremophor EL: Oleic Acid							
(10:10:80, by weight 2 ml/kg). $^{b}5 \mu$ M/kg iv and 30 $\mu$ M/kg po. $^{c}1$							
mg/kg iv and po.							

The pharmacokinetic profile of ABT-639 was evaluated in rat, dog and monkey, respectively. The data are shown in Table 3. ABT-639 exhibits moderate to low plasma clearance (CLp) ranging from 0.55 L/hr/kg in rat to 0.045 L/hr/kg in dog. It also demonstrates moderate to low-moderate volume distribution values in these three animal species. The half-life (3.3, 4.9 and 8.3 hrs) and high oral bioavailability (73, 88 and 95%) in rat, dog and monkey are in agreement with the liver microsomal stability data (81-100% remaining after 30 min). ABT-639 shows good aqueous solubility of 489 µM in phosphate buffer (pH = 7.4) and over 9.5 mM in 0.1 HClsolution. In rats, the plasma concentration of ABT-639 was increased proportionally in dose escalation at 30, 100 and 300 mg/kg. ABT-639 has low protein binding (88.9% in rat and 85.2% in human). The brain to plasma concentration ratio was 1:20 in rats. ABT-639 is a not a competitive inhibitor of CYP1A2, 2C9, 2C19, 2D6 and 3A4 (IC<sub>50</sub> > 10 µM). ABT-639 showed no CYP3A4 (PXR) induction (EC<sub>50</sub> >10  $\mu$ M), no CYP1A2 mRNA induction (EC<sub>50</sub> >10  $\mu$ M).

### ASSOCIATED CONTENT

#### Supporting Information

Experimental and characterization data for all compounds and the Capsiacin-induced secondary mechanical hyperalgesia assay ABT-639 dose-dependently attenuates nociception in a capsaicin-induced secondary mechanical hyperalgesia model (Cap-SMH) (Figure 3). The antinociceptive activity of ABT-639 in this model is consistent with its dose-dependent antinociceptive activity in multiple models of neuropathic pain.<sup>18, 23</sup> Additionally, ABT-639 did not produce any decrement in balance or motor performance in the rat Edge (ED<sub>50</sub> > 300 mg/kg or rat plasma 114 µg/ml, p.o.). In rat cardiovascular (CV) studies, intravenous administration (i.v.) of ABT-639 yielded negligible changes from vehicle control on mean arterial pressure (MAP), heart rate (HR), left ventricular contractility (dP/dt<sub>50</sub>), and vascular resistance (VR) at concentrations (30 mg/kg, plasma concentration of 43.9 µg/ml).<sup>18</sup>



Figure 3. ABT-639 dose dependently reduces tactile allodynia in the rat Cap-SMH model. ABT-639 was administered 1 hr before behavioral testing. Gabapentin (Gaba, 500  $\mu$ M/kg, i.p.) was included as a positive control for assay sensitivity.

We have described here the discovery of a novel selective Ttype calcium channel blocker, ABT-639. Starting from HTS hits 1 and 2 with poor metabolic stability, we replaced the 2methylindoline with aniline to improve the rat (RLM) and human liver (HLM) microsomal stability. Subsequently, we optimized the new lead 3 by incorporating a novel bicyclic fused diamine. We then introduced the F and Cl atoms to the central aromatic ring to improve the oral bioavailability, metabolic stability, decrease the plasma clearance rate, increase the half-life  $(t_{1/2})$ , and led to discovery of a novel Ttype calcium channel blocker ABT-639. ABT-639 displayed good selectivity against N-type, P/Q-type, L-type calcium channel and hERG channel. ABT-639 has an excellent PK profile with high oral bioavailability in all species. In vivo, ABT-639 dose-dependently reduces nociception in a chronic pain model, with no significant cardiovascular effects at analgesic doses.

are provided. This material is available free of charge via the Internet at <u>http://pubs.acs.org</u>.

#### AUTHOR INFORMATION

# **Corresponding Author**

\*E-mail: henry.zhang@abbvie.com.

# **Author Contributions**

The manuscript was written through contributions of all authors. All authors have given approval to the final version.

# Funding Sources

QZ, ZX, SJ, VES and MFJ are employees of AbbVie. The research described herein is solely funded by AbbVie. AbbVie contributed to the study design, research and interpretation of data.

# ABBREVIATIONS

HTS, high throughput screening; VGCC, Voltage-gated calcium channels, RLM, rat liver microsomes; HLM, human liver microsomes; HT-ADME, high throughput-absorption, distribution, metabolism, and excretion; FLIPR, fluorescent imaging plate reader; EP, electrophysiology; DMF, N, Ndimethylformamide, DRG, dorsal root ganglion; PK, pharmacokinetics.

# REFERENCES

(1) Catterall, W. A. Voltage-gated calcium channels. Cold *Spring Harb Perspect Biol.* **2011**, *3(8)*, a00394.

(2) Yamamoto, T.; Takahara, A. Recent updates of N-type calcium channel blockers with therapeutic potential for neuropathic pain and stroke. Curr. Top. Med. Chem. 2009, 9, 377-395.

(3) McGivern J. G. Targeting N-type and T-type calcium channels for the treatment of pain. Drug Disco Today 2006, 11.245-253.

(4) Shin, H. S., Cheong, E. J.; Choi, S.; Lee, J.; Na, H. S. T-type Ca<sup>2+</sup> channels as therapeutic targets in the nervous system. Curr. Opin. in Pharmacology 2008, 8, 33-41. (5) Khosravani, H.; Zamponi, G. W. Voltage-gated calcium channels and idiopathic generalized epilepsies. Physiol. Rev. 2006, 86(3), 941-966. (6) Nelson, M. T.; Todorovic, S. M.; Perez-Reyes, E. The role of T-type calcium channels in epilepsy and pain. Curr. Pharm. Des. 2006, 12(18), 2189-2197. (7) Dogrul, A.; Gardell, L. R.; Ossipov, M. H.; Tulunay, F. C.; Lai, J.; Porreca, F. Reversal of experimental neuropathic pain by T-type calcium channel blockers. Pain 2003, 105, 159 - 169(8) Belardetti, F.: Zamponi, G. W. Linking calciumchannel isoforms to potential therapies. Curr. Opin. Invest.Drugs 2008, 9, 707-715. (9) Oshima, T.; Ozono, R.; Yano, Y.; Higashi, Y.; Teragawa, H.; Miho, N.; Ishida, T.; Ishida, M.; Yoshizumi, M.; Kambe, M. Beneficial effect of T-type calcium channel blockers on endothelial function in patients with essential

hypertension. Hypertens Res. 2005, 28(11), 889-894. (10) Yang, Z. O.; Schlegel, K. S.; Shu, Y.; Reger, T. S.; Cube, R.; Mattern, C.; Coleman, P. J.; Small, J.; Hartman, G.

- D.; Ballard, J.; Tang, C.; Kuo, Y.; Prueksaritanont, T.; Nuss,
- C. E.; Doran, S.; Fox, S. V.; Garson, S. L.; Li, Y.; Kraus, R. L.; Uebele, V. N.; Taylor, A. B.; Zeng, W.; Fang, W.;
- Chavez-Eng, C.; Troyer, M. D.; Luk, J. Ann; Laethem, T.;
- Cook, W. O.; Renger, J. J.; Barrow, J. C. Short-Acting T-
- Type Calcium Channel Antagonists Significantly Modify
- Sleep Architecture in Rodents. ACS Med. Chem. Lett. 2010,
  - 1(9), 504-509.

(11) Miwa, H.; Kondo, T. T-type calcium channel as a new therapeutic target for tremor. Cerebellum. 2011, 10(3), 563-569

(12) Miwa, H.; Koh, J.; Kajimoto, Y.; Kondo, T. Effects of T-type calcium channel blockers on a parkinsonian tremor model in rats. Pharmacol Biochem Behav. 2011, 97(4), 656-659

(13) Yang, Z. Q.; Barrow, J. C.; Shipe, W. D.; Schlegel, K.A.; Shu, Y.; Yang, F. V.; Lindsley, C. W.; Rittle, K. E.; Bock, M. G.; Hartman, G.D.; Uebele, V. N.; Nuss, C. E.; Fox. S. V.: Kraus. R. L.: Doran. S. M.: Connolly. T. M.: Tang, C.; Ballard, J. E.; Kuo, Y.; Adarayan, E. D.; Prueksaritanont, T.; Zrada, M. M.; Marino, M. J.; Graufelds, V. K.; DiLella, A. G.; Reynolds, I. J.; Vargas, H. M.; Bunting, P. B.; Woltmann, R. F.; Magee, M. M.; Koblan, K. S.; Renger, J. J. Discovery of 1,4-substituted piperidines as potent and selective inhibitors of T-type calcium channels. J. Med. Chem. 2008, 51, 6471–6477.

(14) Perez-Reyes, E. Molecular physiology of low-voltageactivated t-type calcium channels. Physiol Rev 2003, 83, 117-161.

(15) Bourinet, E.; Alloui, A.; Monteil, A.; Barrère, C.; Couette, B.; Poirot, O.; Pages, A.; McRory, J.; Snutch, T. P.; Eschalier, A.; Nargeot, J. Silencing of the Ca<sub>v</sub>3.2 T-type calcium channel gene in sensory neurons demonstrates its major role in nociception. EMBO J 2005, 24, 315-324.

(16) Jagodic, M. M.; Pathirathna, S.; Joksovic, P. M.; Lee, W.; Nelson, M. T.; Naik, A. K.; Su, P.; Jevtovic-Todorovic, V.; Todorovic, S. M. Upregulation of the T-type calcium current in small rat sensory neurons after chronic constrictive injury of the sciatic nerve. J Neurophysiol 2008, 99, 3151-3156.

(17) Nelson, M. T.; Woo, J.; Kang, H. W.; Vitko, I.; Barrett, P.Q.; Perez-Reyes, E.; Lee, J. H.; Shin, H. S.; Todorovic, S. M. Reducing agents sensitize C-type nociceptors by relieving high-affinity zinc inhibition of Ttype calcium channels. J Neurosci 2007, 27, 8250-8260. (18) Jarvis, M. F.; Scott, V. E.; McGaraughty, S.; Chu, K. L.; Xu, J.; Niforatos, W.; Milicic, I.; Joshi, S.; Zhang, Q.; Xia, Z. A peripherally acting, selective T-Type calcium channel blocker, ABT-639, effectively reduces nociceptive and neuropathic pain in rats. Biochem Pharmacol. 2014, 89, 536-544.

(19) Vortherms, T. A.; Swensen, A. M.; Niforatos, W.; Limberis, J. T.; Neelands, T. R.; Janis, R. S.; Thimmapaya, R.; Donnelly-Roberts, D. L.; Namovic, M. T.; Zhang, D.; Brent Putman, C.; Martin, R. L.; Surowy, C. S.; Jarvis, M. F.; Scott, V. E. Comparative analysis of inactivated-state block of N-type (Cav2.2) calcium channels. Inflamm Res 2011, 60, 683-693.

(20) Scott, V. E.; Vortherms, T. A.; Niforatos, W.; Swensen, A. M.; Neelands, T.; Milicic, I.; Banfor, P. N.; King, A.; Zhong, C.; Simler, G.; Zhan, C.; Bratcher, N.; Boyce-Rustay, J. M.; Zhu, C. Z.; Bhatia, P.; Doherty, G.; Mack, H.; Stewart, A. O.; Jarvis, M. F. A-1048400 is a novel, orally active, state-dependent neuronal calcium channel blocker that produces dose-dependent antinociception without altering hemodynamic function in rats. Biochem Pharmacol 2012, 83, 406-418.

(21) Dossetter, A. G. A statistical analysis of in vitro human microsomal metabolic stability of small phenyl group substituents, leading to improved design sets for parallel SAR exploration of a chemical series. Bioorg. Med. Chem. 2010, 18, 4405-4414.

1

(22) Rodgers T.; Leahy, D.; Rowland. M. Physiologically based pharmacokinetic modeling 1: Predicting the tissue distribution of moderate-to-strong bases. *J Pharm Sci* **2005**, *94*, 1259-1267.

(23) Serra, J.; Jones, M.; Sumalla, M.; Jarvis, M. F. ABT-639, a selective, peripherally acting T-type calcium channel blocker, inhibits spontaneous activity of C-nociceptors in a rat model of neuropathic pain. *Society for Neuroscience*. **2013**, 829.02/H.



146x70mm (300 x 300 DPI)