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Orally bioavailable dual MMP-1/MMP-14 sparing, MMP-13 selective α -sulfone hydroxamates

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ABSTRACT

A series of phenyl piperidine α -sulfone hydroxamate derivatives has been prepared utilizing a combination of solution-phase and resin-bound library technologies to afford compounds that are potent and highly selective for MMP-13, are dual-sparing of MMP-1 and MMP-14 (MT1-MMP) and exhibit oral bioavailability in rats.

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The matrix metalloproteinases (MMPs) are a family of zincdependent enzymes that degrade all components of the extracellular matrix.^{1,2} There are at least 24 isozymes in the MMP family, and they are roughly classified on the basis of their substrate specificity: collagenases (MMP-1, -8, -13 and -18), gelatinases (MMP-2, and-9), stromelysins (MMP-3, -10 and -11) and membrane-type MMPs (MMP-14, -15, -16, -17, and -24) and others (MMP-7, -11, -12, -19, -20, -21, -22, and -23). Clinical experience with the pan MMP inhibitor Marimastat has revealed a constellation of adverse effects collectively referred to as musculoskeletal syndrome (MSS).³ We have hypothesized that this is predominantly a result of inhibiting both MMP-1 and MMP-14, and that MMP inhibitors sparing these two isozymes should be devoid of MSS.⁴ Toward obtaining efficacy in mitigating damage suffered in osteoarthritic patients, it has been reported that MMP-13 mRNA levels are increased in osteoarthritic cartilage.⁵ Thus, we envisioned the development of a selective inhibitor of MMP-13 sparing both MMP-1 and MMP-14 as a safe means of treating osteoarthritis (OA) which we refer to as the dual-sparing hypothesis.

In the preceding paper,⁴ we demonstrated that the N-substituted phenyl isonipecotamide hydroxamic acid template **1** (Fig. 1) yielded

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potent and selective MMP-13 inhibitors. Figure 2 shows a sequence comparison for a set of MMP family members focused on the S'_1 loop. There are differences in both amino acid identity and in length of the loop, as MMP-1, -2 and -9 are two residues shorter than MMP-13, -8 and -14. Interaction of the amide N-substituents of **1** deep in the S'_1 pocket was expected to affect isozyme selectivity across the MMP family. In an effort to further investigate SAR in this region, we designed templates **2** and **3**, where the distal phenyl group resides in approximately the same region as the amide N-substituents of **1**. To explore the effect of substituents on the distal aryl rings of **2** and **3**, we undertook a parallel synthetic approach with the goal of optimizing MMP-13 potency and selectivity.

A solid-phase parallel synthesis approach was used to create a small library of *N*-arylpiperazine α -sulfone hydroxamic acid derivatives (**2**) from commercially available *N*-aryl piperazines **5**. Nucle-ophilic aromatic substitution of the previously reported polymerbound aryl fluoride **4**⁴ with *N*-arylpiperazines was found to require a 10-fold excess of **5** and presence of 2 equiv of cesium carbonate in *N*-methylpyrrolidinone (NMP) at 100 °C over night to achieve good conversions. Acidic deprotection with trifluoroacetic acid afforded α -sulfone hydroxamic acids **2** in good yields (Scheme 1).

Alternatively, a solution phase approach was developed to synthesize analogs with a basic amine in the α -heterocycle (**8**). THPprotected hydroxymate **7** was obtained by reaction of carboxylic acid **6** with THP-protected hydroxylamine using the water-soluble carbodiimide EDC.⁶ Subsequent nucleophilic aromatic substitution

⁰⁹⁶⁰⁻⁸⁹⁴X/\$ - see front matter @ 2010 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2010.04.130

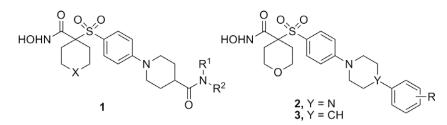


Figure 1. 4-Substituted piperidine/piperazine sulfone hydroxamic acids.

Figure	
	*
MMP13	DPGALMFPIYTGKSHFML
MMP-8	DPGALMYPNYAF <mark>RE</mark> TSNYSL
MMP-2	DPGALMAPIYTYTKNFRL
MMP-9	VPEALMYPMYRFTEGPPL
MMP-14	DPSAIMAPFYQWMDTENFVL
MMP-1	DIGALMYPSYTFSGDVQL

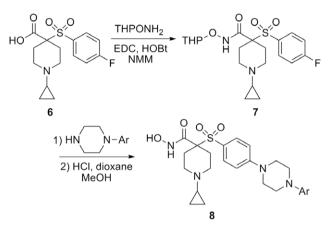
Figure 2. S₁ loop sequence variation across selected MMP family members.

with the requisite *N*-arylpiperazines followed by acidic deprotection afforded α -sulfone hydroxamic acids **6** in high yields.

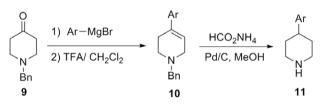
For the synthesis of 4-arylpiperidine phenyl sulfones **3**, the required 4-aryl piperidines **11** were purchased commercially or prepared either via the Grignard route (Scheme 3) or via Suzuki coupling (Scheme 4). For the Grignard route, similar to the method of Burns,⁷ the appropriate aryl Grignard was added to *N*-benzyl piperidine-4-one followed by dehydration with TFA in methylene chloride. Reduction then gave 4-arylpiperidines **11**. Alternatively, enol triflate **12** was reacted with the requisite arylboronic acid with a catalytic amount of tetrakistriphenylphosphine palladium to afford *N*-Boc-4-aryl tetrahydropyridine **13**, following the procedure of Wustrow and Wise⁸ (Scheme 2). Catalytic hydrogenation of **13** followed by removal of the *N*-Boc protecting group afforded 4-arylpiperidines **11**.

Assembly of the *N*-arylpiperidine sulfone hydroxamic acids **3** was accomplished by a solution phase approach (Scheme 5) similar to that described above. Carboxylic acid **14**⁶ was coupled with THP-protected hydroxylamine, then aromatic nucleophilic substitution with **11** followed by acidic deprotection afforded α -sulfone hydroxamic acids **3** in good yields.

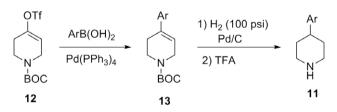
The MMP inhibitory potency values for *N*-aryl piperazine α -sulfone hydroxamic acids (**2** and **8**) are summarized in Table 1. All compounds were determined to have no measurable potency at MMP-1 (IC₅₀ >10,000 nM), hence selectivity for MMP-13 over MMP-1 is >1000× in most cases. The *N*-aryl piperazines (**2a–2j** and **8a–8b**) exhibited mostly single-digit nanomolar potency for MMP-13, but most were also potent for MMP-2, resulting in only very modest selectivity for MMP-13 over MMP-13 over MMP-2 with a number of compounds which were nearly equipotent. *N*-Phenyl piperazine **2a** was very potent for MMP-13 (IC₅₀ = 1.7 nM) with a 14-fold selectivity versus



Scheme 2. Solution phase synthesis of *N*-arylpiperazine sulfone hydroxamic acids 8.

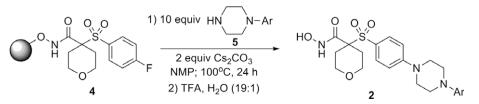


Scheme 3. Syntheses of 4-arylpiperidines via the Grignard route.

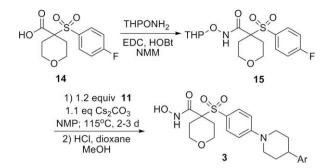


Scheme 4. Syntheses of 4-arylpiperidines via Suzuki coupling.

MMP-2, and nearly 6000-fold selectivity versus MMP-14, whereas the corresponding α -piperidine **8a** was nearly equipotent at MMP-13 and MMP-2 (IC₅₀ = 3.3 and 5.4 nM, respectively.) The reduction in selectivity due to the change from X = 0 to X = *N*-cyclopropyl was unexpected given the continuity between α -tetrahydropyran



Scheme 1. Solid phase synthesis of N-arylpiperazine sulfone hydroxamates 2.



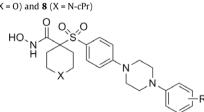
Scheme 5. Solution phase synthesis of 4-arylpiperidine sulfone hydroxamates.

and α -piperidine analogs in our earlier MMP-1 sparing series,⁶ although this single pair does not necessarily constitute a trend. The *ortho*-fluorinated derivative **2b** exhibited similar potency to the *N*-phenyl parent **2a**, whereas the bulkier *ortho*-methyl and chloro derivatives 2c-2d dropped five-fold in potency for MMP-13, and the ortho-methoxy derivative **2e** dropped 76-fold in potency. The meta-derivatives 2f and 2g suffered a similar loss in potency for MMP-13. On the other hand, para-substituted derivatives maintained high potency and selectivity for MMP-13, in particular *para*-methoxy derivative **2h** with an $IC_{50} = 0.5$ nM for MMP-13, a 40-fold selectivity versus MMP-2, and the highest selectivity observed versus MMP-14 (>20,000). para-Methyl and para-trifluoromethyl derivatives 2i and 8b exhibited good potency for MMP-13 (1.9 and 2.4 nM, respectively) and selectivity versus MMP-14 (both >4000×), noting that **8b** is an α -piperidine. The more sterically demanding 2,4-dimethylphenylpiperazine 2j suffered a drop in potency at MMP-13 (IC₅₀ = 28.6 nM), although selectivity against MMP-2 was the highest of all N-arylpiperazines at approximately 50×.

Table 2 shows MMP inhibitory potencies of 4-aryl piperidine α sulfone hydroxamates (**3**). 4-Phenylpiperidine **3a** was $3 \times$ less potent at MMP-13 than *N*-phenylpiperazine **2a** but its potency for MMP-2 increased to 4.4 nM, making **3a** equipotent for MMP-13 and MMP-2. A substantial boost in MMP-13 selectivity was achieved by the presence of an *ortho*-methoxy substituent (**3b**). Potency of **3b** for MMP-13 dropped threefold from the parent compound (**3a**), while MMP-2 potency dropped 840-fold, generating a selectivity ratio of 211X. On the other hand, *para*-chloro analog

Table 1

MMP inhibitory potency of *N*-aryl piperazine α -sulfones **2** (X = O) and **8** (X = N-cPr)



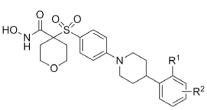
Compd	Х	R		IC ₅₀ (nM)	Ratio MMP 2/13		
			2	9	13	14	
2a	0	Н	23.5	450	1.7	>10,000	13.8
2b	0	2-F	21.1	356	2.7	>10,000	7.8
2c	0	2-Me	88.0	2860	7.7	>10,000	11.4
2d	0	2-Cl	40	2090	9.0	>10,000	4.4
2e	0	2-MeO	370	-	130	_	2.8
2f	0	3-MeO	48.5	163	8.0	2860	6.1
2g	0	3-CF ₃	330	-	20	-	16.5
2h	0	4-MeO	18.4	722	0.63	>10,000	29
2i	0	4-Me	35.0	682	1.9	8220	18.4
2j	0	2,4-diMe	1400	-	28.6	-	49.0
8a	N-cPr	Н	5.4	_	3.3	-	1.6
8b	N-cPr	4-CF ₃	32.6	>10,000	2.4	>10,000	13.6

3c was found to have an increased potency relative to parent compound 3a at both MMP-13 and -2. The substantial effect of ortho substitution on selectivity prompted further evaluation of additional ortho-substituted analogs (3d-i). Generally, MMP-13 potencies were similar and reduced compared to **3a**, but IC₅₀'s for MMP-2 (and thus the MMP-2/13 selectivity ratio) corresponded approximately to the size of the substituent, with methoxy being optimal: $H < Cl, OH < CH_3, CF_3 < OMe, OEt, 4-F-C_6H_4$. The effect of an additional substituent was explored in an attempt to increase potency for MMP-13 while maintaining micromolar affinity for MMP-2. The 1-naphthyl derivative 3j was slightly more potent than the orthomethoxy analog 3b at MMP-13, but potency at MMP-2 increased 7-fold. Other disubstituted analogs (3k-n) showed a similar trend, except for **3n** with a 2-methoxy and a 5-isopropyl substitution where MMP-13 potency dropped fourfold. Presumably the decreased affinity for MMP-13 was due to steric reasons. Comparison of the MMP-2/13 selectivity for ortho-methoxy substituted N-arvl piperazine 2e (2.8-fold) with that of the 4-arylpiperidine analog 3b (211-fold) is noteworthy. Presumably, 3b adopts a conformation where the aryl group is orthogonal to the piperidine ring, evidenced by the substantial effect of ortho-substitution on selectivity. The energetic penalty for an N-aryl piperazine to adopt such a conformation would be high, which is likely responsible for the reduced potency of 2e at MMP-13 and the increased potency at MMP-2 relative to 3b.

Based on the superior MMP-13 potency and dual MMP-1 and -14 sparing profiles of the *p*-substituted *N*-aryl piperazines, additional analogs were prepared for more thorough enzyme and PK evaluation (Table 3). Potent MMP-13 inhibition was observed for compounds **2l**, **2m** and **8c** with IC₅₀'s of 0.6, 1.0, 0.5 nM, respectively. Selectivity versus other MMP family members was generally \gg 100-fold except for MMP-2 (4–20-fold) and MMP-3 (58–500fold). Rat PK for these three compounds showed low to moderate values for half-life and bioavailability. Aryl piperazine 8c had an acceptable BA of 20.7%, but a very short $t_{1/2}$ of only 0.24 h. Aryl piperidine 21 exhibited a modest bioavailability of 16%, but a much improved half-life of 2.59 h, which we attribute to the trifluoromethylphenyl moiety in P'_1 , which has enhanced the PK of other series as well. 4-Chlorophenyl piperidine 2m possessed a longer half-life but a disappointing BA of only 7.4%. Included for comparison is broader-spectrum, MMP-1 sparing α -sulfone SC-276.⁶ Compound SC-276 lacks selectivity among MMPs, only significantly

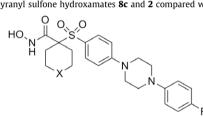
Table 2

MMP inhibitory potency of *N*-aryl piperidine α -sulfones (3)



Compd	\mathbb{R}^1	R ²		Ratio MMP 2/13			
			2	9	13	14	
3a	Н	Н	4.4	265	6.0	>10,000	0.7
3b	MeO	Н	3700	6700	17.5	>10,000	211
3c	Н	4-Cl	2.0	350	0.7	>10,000	2.9
3d	Cl	Н	82	-	42.5	-	1.9
3e	Me	Н	700	-	32.7	_	21.4
3f	CF ₃	Н	900	-	28.8	_	31.3
3g	EtO	Н	3100	-	35.0	_	88.6
3h	OH	Н	70	>10,000	11.3	>10,000	6.2
3i	$4-F-C_6H_4$	Н	5400	-	33.7	_	160
3j	2,3-(CH=CH) (naphthyl)		500	2501	11.4	>10,000	43.9
3k	Me	4-MeO	505	>10,000	7	>10,000	72.1
31	MeO	4-diMeO	1750	>10,000	11	>10,000	159
3m	MeO	5-diMeO	800	-	11.4	_	70.2
3n	MeO	5- <i>i</i> Pr	>10,000	-	70	-	143

Table 3 MMP inhibitory potency of *N*-aryl piperazine α-tetrahydropyranyl sulfone hydroxamates **8c** and **2** compared with **SC-276**



Compd	Х	\mathbb{R}^1	IC ₅₀ at MMP-X (nM)								cLog P	C _{max}	C_{6h}	$t_{1/2}(h)$	BA (%)
			1	2	3	7	8	9	13	14					
21	0	CF ₃	>10000	12.1	300	>10000	670	5000	0.6	>10000	1.76	4210	10	2.59	16.3
2m	0	Cl	>10000	3.7	130	_	1000	270	1.0	5000	1.46	49	36	1.91	7.4
8c	cPr-N	OCH_3	>10000	5.3	24.5	_	59.2	729	0.42	>10000	1.32	751	19	0.24	20.7
SC-276 ^a			8660	0.33	13.0	>10000	1.8	1.5	0.40	19	2.04	13630	281	1.1	28

^a Data for **SC-276** from Ref. 6, except for MMP-7 and MMP-14 were not previously reported.

sparing MMP-1, yet this compound has the very high exposure in the rat that is compelling for development, consistent with its potent and efficacious antitumor activity.⁶

In summary, the related series of compounds described herein have demonstrated single-digit to sub-nanomolar potency for MMP-13 combined with exceptional selectivity versus MMP-1 and MMP-14 of typically >100 \times up to 20,000 \times . Selectivities versus other MMPs when tested varied for MMP-3 (40–500 \times), MMP-8 (140– $2500\times$) and for MMP-9 (20 to >4000×). Selectivity for MMP-13 over MMP-2 ranged from equipotency to 200×, thus selectivities were somewhat lower relative to the related isonipecotate α -sulfone hydroxamate series.⁴ Rat PK for selected members of these series demonstrated bioavailabilities of up to 20.7% (8c) and a half lives of up to 2.6 h (21) yet individual compounds lacked a compelling complete package to initiate development. Undoubtedly the high molecular weights of these analogs (551 a.u. for 2m) plays a role given the recommendations of Lipinski although the limited number of rotatable bonds⁹ favors the limited bioavailability that was observed. We therefore turned our attention to lower molecular

weight species, while applying our learnings about P'₁ manipulations toward optimizing MMP-13 selectivity.

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