



Design and synthesis of a simplified inhibitor for XIAP-BIR3 domain

Fernando R. Pinacho Crisóstomo, Yongmei Feng, Xuejun Zhu, Kate Welsh, Jing An, John C. Reed, Ziwei Huang*

The Burnham Institute for Medical Research, 10901 N. Torrey Pines Rd., La Jolla, CA 92037, USA

ARTICLE INFO

Article history:

Received 27 July 2009

Revised 4 September 2009

Accepted 14 September 2009

Available online 17 September 2009

Keywords:

XIAP-BIR3 domain

Molecular simplification

Small molecule

ABSTRACT

Based on tetrapeptide AVPI, we were able to design and synthesize a new simplified scaffold to inhibit the BIR3 domain of the XIAP protein at low micromolar range. The uncomplicated synthesis and the binding activity of the molecule disclosed here represent an attractive alternative to develop new compounds targeting the protein–protein interaction of XIAP/caspase9.

© 2009 Elsevier Ltd. All rights reserved.

Apoptosis plays a crucial role in the homeostasis and development of living organisms. Deregulation in this mechanism is associated with many diseases including several types of cancer. In the apoptosis pathway, the inhibitors of apoptosis proteins (IAPs) are one of the mechanisms used by tumor cells to evade programmed cell death.¹

The XIAP is the most potent caspase inhibitors among IAPs protein family. This protein interacts with initiator caspase 9 and executioner caspase 3 and 7 through its BIR3 and BIR2 domains respectively.² The search for new compounds able to disrupt the XIAP-caspase interaction has attracted the attention of scientific community as a promising strategy for cancer treatment.

The natural inhibitor of XIAP is a protein (SMAC/DIABLO) released from the mitochondria into the cytosol in response to apoptotic stimuli. SMAC removes XIAP inhibition of caspase 9 by binding to the BIR3 domain of XIAP through AVPI tetrapeptide present in the N-terminal part of SMAC. This interaction (AVPI/BIR3) has been determined unequivocally by X-ray crystallography.³

Using the AVPI structure, Fesik and co-workers have performed a comprehensive study to determine which amino acids could be substituted without compromising its binding affinity. The authors have determined the essential amino acids residues to preserve the activity of this tetrapeptide to be the alanine (first amino acid) and proline (third amino acid)⁴ (Fig. 1).

Based on precedents in the literature,⁵ it is possible to rationalize about some structural features for peptidomimetic derivatives and postulate general 'structural guidelines' to design new com-

pounds based on the AVPI structure. As common features, the analogs should contain: (1) an alanine residue or a *N*-methyl alanine residue, (2) the presence of a rigid core (3) an aromatic residue as a surrogate of the isoleucine, and (4) the molecules should adopt a 'U-conformation' for a suitable interaction with the protein (Fig. 2). Most of the compounds with biological activity in vitro at nano molar range follow this pattern.

Structural simplification represents an efficient drug design strategy to shorten synthetic routes while keeping or enhancing the biological activity of complex compounds.⁶ Combining the molecular simplification concept with the guidelines highlighted previously, we report here a series of simplified compounds inspired by the Smac-AVPI tetrapeptide.

Preserving the alanine residue, we proposed a molecular simplification where the second and third amino acids were substituted by thiazole ring fused to a carbocycle with different sizes as rigid

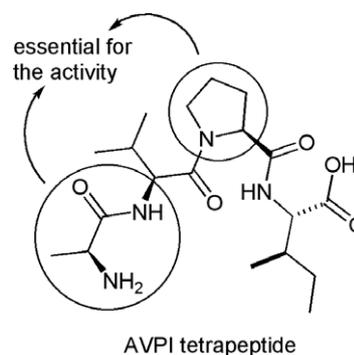


Figure 1. Tetrapeptide from the N-terminal part of SMAC protein.

* Corresponding author. Current address: Department of Pharmacology, SUNY Upstate Cancer Research Institute, State University of New York (SUNY), Upstate Medical University, Syracuse, New York 13210, USA.

E-mail address: huangz@upstate.edu (Z. Huang).

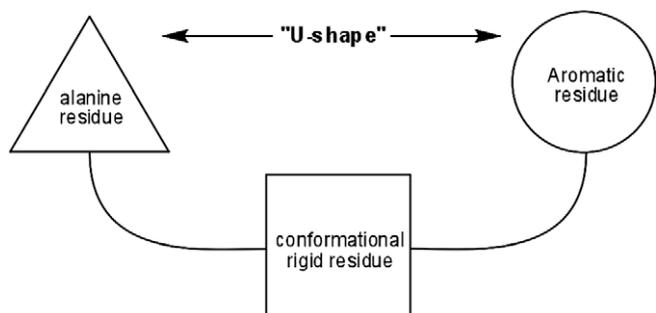


Figure 2. Common structural features found in most of XIAP-BIR3 domain inhibitors.

central core. In this approach, we eliminated one chiral center while at same time conferring more rigidity to the molecule. Finally, different aromatic moieties linked through an amide bond to the rigid portion provided analogs structurally less complex (Fig. 3).

The retrosynthetic analysis of these molecules is depicted in the Scheme 1. It is important to mention that the final compounds were predicted to adopt the necessary 'U-conformation' for a suitable interaction with the protein based on molecular modeling studies.⁷

After docking analysis of the molecules containing different sizes in the central core with the BIR3 domain of the XIAP protein (RCSB PDB ID 2jk7), one of the suitable candidates to initiate the synthesis was the compound containing the seven member ring

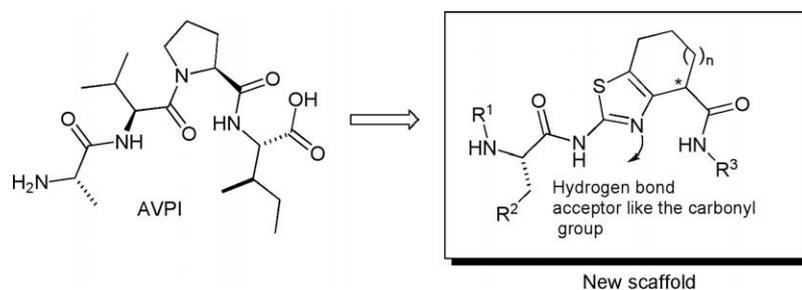
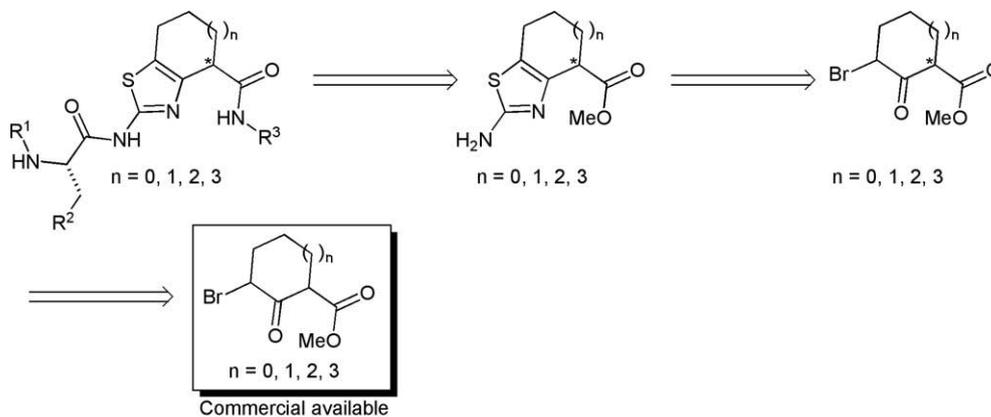


Figure 3. New synthetic scaffold using a thiazole ring fused to a carbocycle as peptide surrogate.



Scheme 1. Retrosynthesis and conformational analysis of the proposed compounds.

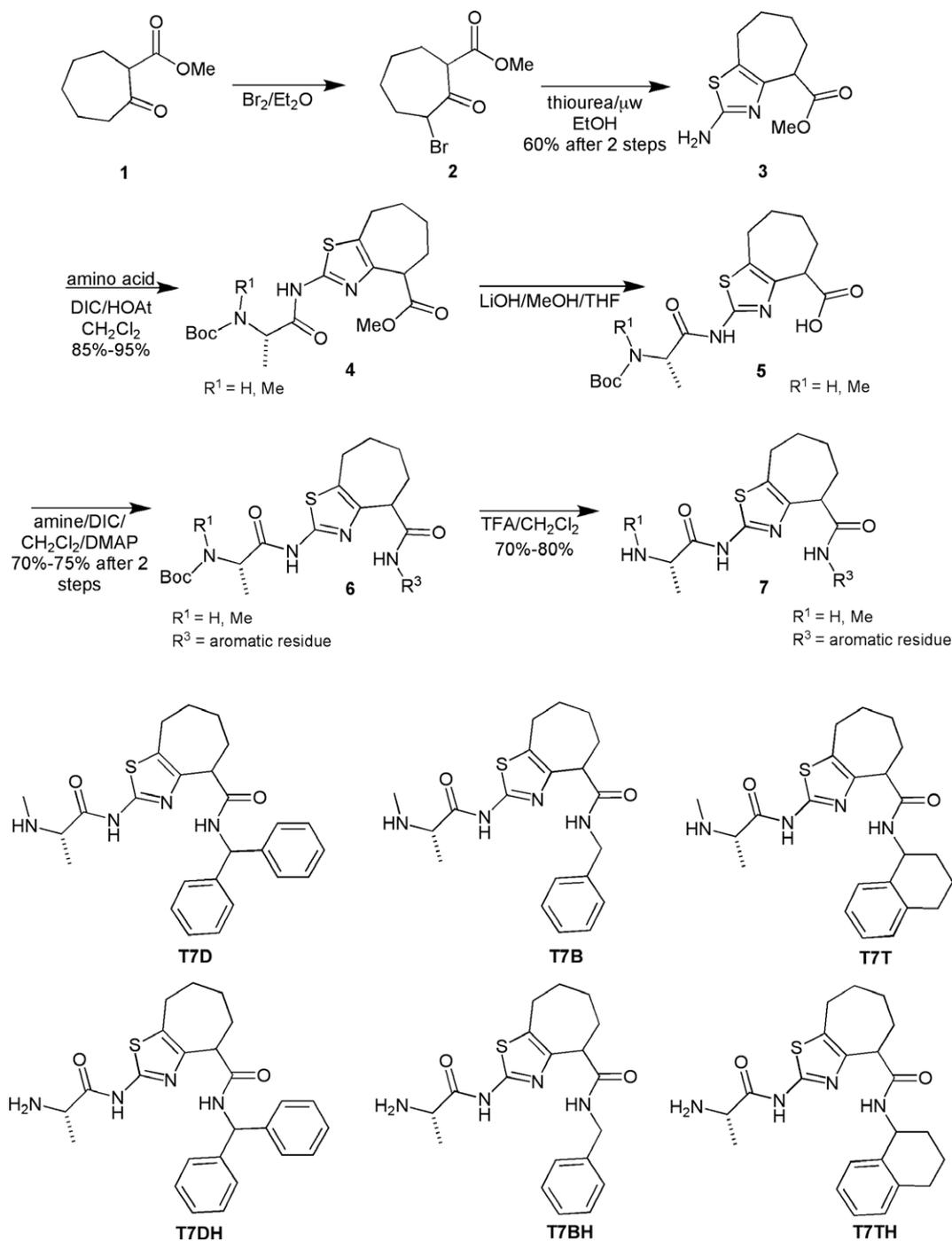
carbocycle.⁸ Furthermore, examples containing a seven member ring fused to a five member ring have been reported in the literature with excellent biological activity.^{5b,7}

The synthesis started with the bromination of the commercially available compound methyl-2-oxo-1-cycloheptanecarboxylate (**1**).¹⁰ The product **2** was used without purification in the thiazole formation assisted by microwave irradiation affording the fused bicyclic compound **3**. The peptide coupling with the amino acids was achieved using standard conditions (DIC/HOAt/CH₂Cl₂) leading to compound **4** in good yields (85–95%). The methyl ester **4** was hydrolyzed using LiOH and the product **5** was used in amide formation using different commercial available amines. Finally,

deprotection of the Boc group present in the amino acid residue was performed using a solution of 10%TFA (v/v) in CH₂Cl₂ providing the first set of molecules. (Scheme 2).

These compounds were synthesized to identify which amino acid residue (R¹) and aromatic moieties (R³) would provide convenient substitution patterns for this scaffold. Their binding affinities to XIAP-BIR3 domain were evaluated using competitive fluorescence polarization BINDING assay (FPA).

Despite the preliminary docking result, which indicated the thiazole ring fused with a seven member carbocycle as a promising candidates, these molecules did not exhibit relevant activity.¹¹



Scheme 2. Synthesis of the thiazole ring fused to a seven member ring carbocycle.

Focusing our attention in the central part of the proposed scaffold, we synthesized another group of molecules containing a five member ring in the rigid portion. This modification was envisioned to confer more rigidity to system and at same time probe any unfavorable steric interactions in the binding pocket of the protein.

The synthesis of these compounds followed the same procedure as described above although the starting material was the commercially available compound cyclopentanone-2-carboxylic acid methyl ester (Fig. 4).

The results obtained in the fluorescence polarization assay (FPA) experiment prompted us to make some considerations about the structure–activity relationship (SAR) of this set of molecules (Fig. 5).

As shown in Figure 5, all the compounds with the *N*-methyl alanine residue exhibited better activity than those containing the alanine. On the other hand, the compounds with 1,2,3,4-tetrahydronaphthalene moiety showed better results when compared with others with different aromatic substitution patterns.

The promising $K_i = 37 \mu\text{M}$ obtained with the compound **T5T**¹² has provided valuable information about this scaffold, which includes: (1) the amino acid residue must contain an *N*-methyl group; (2) the aromatic moiety should be the 1,2,3,4-tetrahydronaphthalene; and (3) probably because of unfavorable steric interactions, the thiazole ring in the central core should be fused with a five member ring.

Another important aspect for the activity of the small molecules targeting the BIR3 domain is the correct configuration of the chiral centers present in the molecule. This is not unexpected, considering that the binding pocket is a chiral environment and many examples support the importance of the correct diastereoisomer.^{5b,9,13}

In our initial studies we wanted to determine the suitable substitution pattern and also the convenient central core for BIR3 binding activity. Nonetheless, the chiral aspect of the molecules was not considered. As can be observed, only one of the three chiral centers (the amino acid residue) present in the structure is defined while the other two are in the racemic form. Thus, the compound

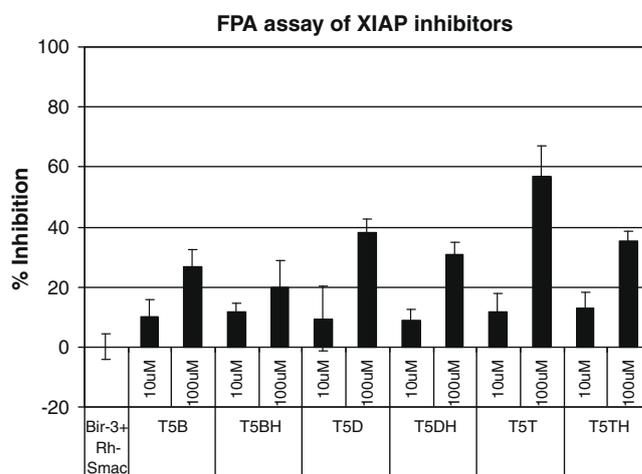


Figure 5. FPA result obtained with the thiazole fused to a five member ring carbocycle derivatives.

T5T exists in a diastereoisomeric mixture of four compounds (Fig. 6).

To determine which diastereoisomer was responsible for the inhibitory activity, we performed a docking study of these different diastereoisomers. The result obtained indicated that the only diastereoisomer suitable for the interaction with the XIAP-BIR3 domain is **T5TR1** with the configuration *S,R,R*, highlighted in the Figure 6. The docked conformation of this diastereoisomer is shown in Figure 7.

To verify experimentally that **T5TR1** is indeed the diastereoisomer responsible for XIAP-BIR3 binding, we conducted synthesis of the four diastereoisomers using the same conditions as depicted in the Scheme 2. However, in the coupling with the aromatic moiety, the commercially available enantiopure compounds (*R*) and (*S*)-1,2,3,4-tetrahydronaphthalene were used. The diastereoisomeric mixture from the coupling with (*R*)-1,2,3,4-tetrahydronaphthalene

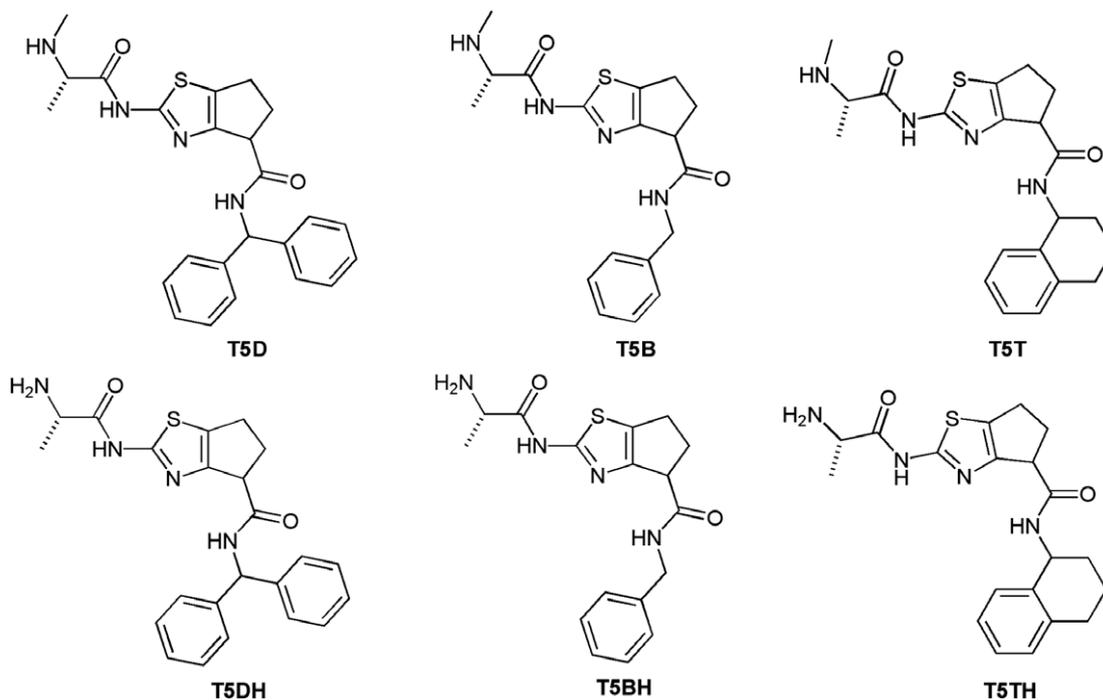


Figure 4. Set of molecules containing thiazole ring fused to five member ring carbocycle.

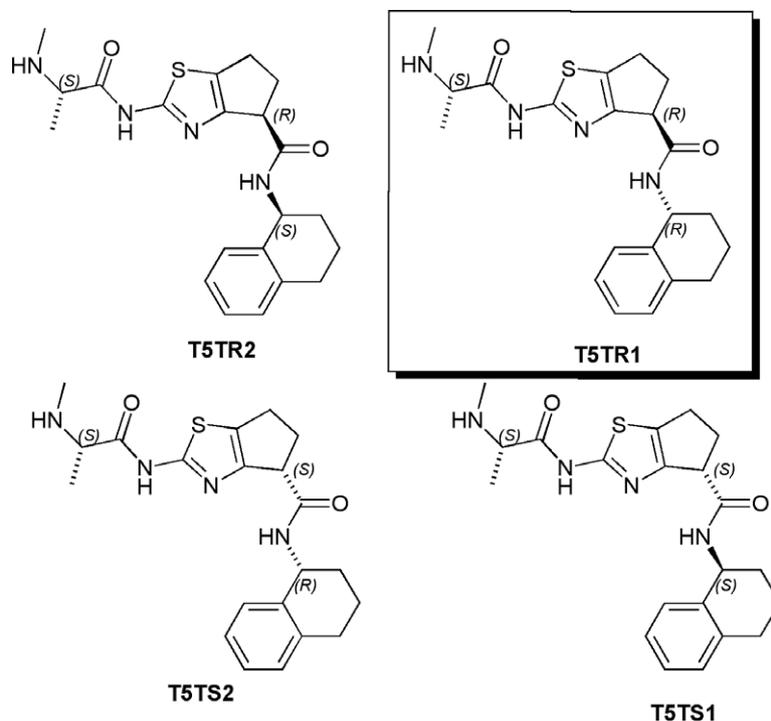


Figure 6. Diastereoisomers present in the **T5T** mixture.

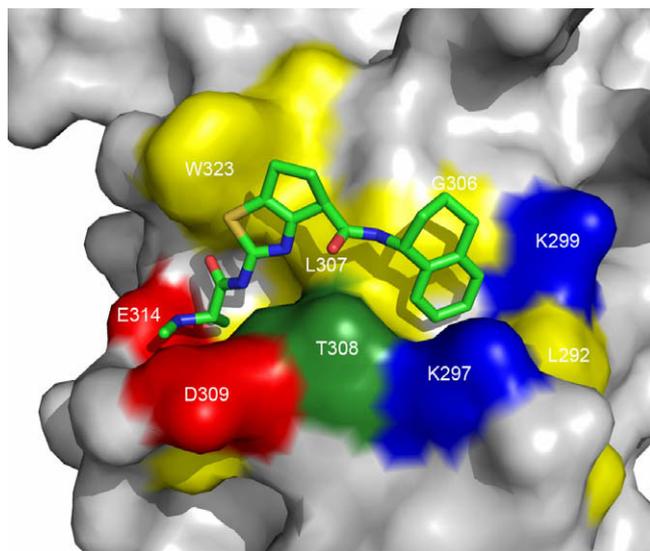


Figure 7. The docked conformation of the diastereoisomer **T5TR1** (shown in Fig. 6) in XIAP-BIR3 domain (RCSB PDB ID 2jk7) as predicted by molecular docking calculation using the program SYBYL 8.0.

was possible to separate in a regular (silica gel) chromatographic column, affording the compounds **T5TR1** and **T5TR2**. In the docking analysis, the isomers from the coupling with (*S*)-1,2,3,4-tetrahydronaphthalene (**T5TS1** and **T5TS2**) did not exhibit interaction with protein. We therefore tested these compounds as diastereoisomeric mixture (**T5TSM**) (Fig. 8).

As a control experiment we have used the known compound **BV6**, which was determined to have a K_i value of 0.90 μM for BIR3 domain in our FPA assay.¹⁴ The K_i obtained for **T5TR1** was 5.58 μM while the other diastereoisomers did not exhibit significant binding activity, supporting the docking result and demonstrating

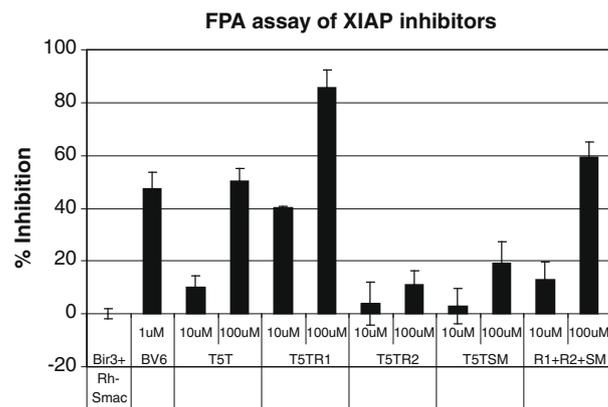


Figure 8. FPA result obtained with diastereoisomers of compound **T5T**.

that **T5TR1** is indeed the active diastereoisomer responsible for XIAP-BIR3 binding (Fig. 9).

Although the K_i value for the compound **T5TR1** was approximately seven times less potent compared to the control **BV6**, the uncomplicated synthesis (six steps) and the structural simplicity of our designed scaffold make it an interesting starting point to develop new analogs.

Precedents in the literature have in their structures between 4 and 5 chiral centers (as in the case of the original AVPI peptide), and in some examples the synthesis has up to 10 chemical steps to achieve the final product.^{5c,11}

In conclusion, we have designed a simplified analog using the AVPI tetrapeptide as template. The expedient synthetic route (six steps) combined with binding activity of this new compound (**T5TR1**) represents an alternative to other more complex compounds attacking the XIAP-BIR3 domain. This lead compound presents the structural novelty of a thiazole ring fused to five member ring carbocycle (2-amino-5,6-dihydro-4*H*-cyclopenta[*d*]-

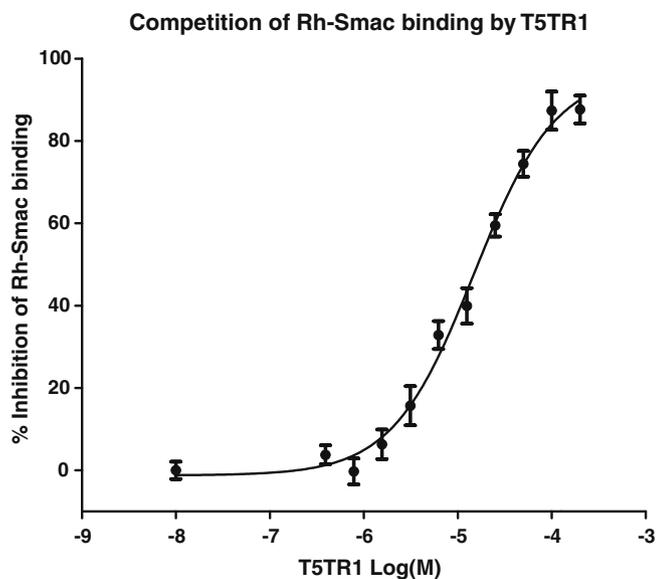


Figure 9. Titration curve to determine the K_i value of compound **T5TR1**.

thiazole-4-carboxylic acid) as a peptide surrogate. New analogs inspired by this scaffold are undergoing evaluation in our laboratory.

Acknowledgment

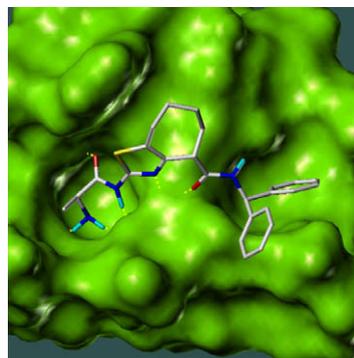
This work was supported by the NIH PO1 grants CA055164, U01-CA113318.

References and notes

1. Reed, J. *Nat. Rev. Drug Disc.* **2002**, *1*, 111.
2. Shiozaki, E. N.; Chai, J.; Rigotti, D. J.; Riedl, S. J.; Li, P.; Srinivasula, S. M.; Alnemri, E. S.; Fairman, R.; Shi, Y. *Mol. Cell.* **2003**, *11*, 519.
3. Liu, Z.; Sun, C.; Olejniczak, E. T.; Meadows, R. P.; Betz, S. F.; Oost, T.; Hermann, J.; Wu, J. C.; Fesik, S. W. *Nature* **2000**, *408*, 1004.
4. Oost, T. K.; Sun, C.; Armstrong, R. C.; Assad, A.-S.; Betz, S.; Deckwerth, T. L.; Ding, H.; Elmore, S. W.; Meadows, R. P.; Olejniczak, E. T.; Oleksijew, A.

Oltersdorf, T.; Rosenberg, S.; Shoemaker, A.; Tomaselli, K.; Zou, H.; Fesik, S. W. *J. Med. Chem.* **2004**, *47*, 4417.

5. (a) Park, C. M.; Sun, C.; Olejniczak, E. T.; Wilson, A.; Meadows, R.; Betz, S.; Elmore, S.; Fesik, S. W. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 771; (b) Sun, H.; Coleska, Z. N.; Yang, C. Y.; Xu, L.; Liu, M.; Tomita, Y.; Pan, H.; Yoshika, Y.; Krajewski, K.; Roller, P.; Wang, S. *J. Am. Chem. Soc.* **2004**, *126*, 16686; (c) Sun, H.; Coleska, Z. N.; Yang, C. Y.; Lu, J.; Meagher, J. L.; Yang, C. Y.; Qiu, S.; Tomita, Y.; Ueda, Y.; Jiang, S.; Krajewski, K.; Roller, P.; Stuckey, J.; Wang, S. *J. Am. Chem. Soc.* **2007**, *129*, 15279; (d) Wist, A. D.; Gu, L.; Riedl, S.; Shi, Y.; McLendon, G. L. *Bioorg. Med. Chem.* **2007**, *15*, 2935.
6. Wender, P. A.; Verma, V. A.; Paxton, T. J.; Pillow, T. H. *Acc. Chem. Res.* **2008**, *41*, 40.
7. The structure was minimized using AM1 semi-empirical method in the program HyperChem 7.5 for windows.
8. Docking result for the thiazole ring fused to a seven member ring carbocycle using SYBYL 8.0 and the XIAP-BIR3 domain crystal structure with RCSB PDB ID 2jk7.



9. Zobel, K.; Wang, L.; Varfolomeev, E.; Franklin, M. C.; Elliott, L. O.; Wallweber, H. J. A.; Okawa, D. C.; Flygare, J. A.; Vucic, D.; Fairbrother, W. J.; Deshayes, K. *ACS Chem. Biol.* **2006**, *8*, 525.
10. Hedberg, C.; Kallstrom, K.; Brandt, P.; Hansen, L. K.; Andersson, P. G. *J. Am. Chem. Soc.* **2006**, *128*, 2995.
11. These compounds did not exhibit activity at concentration of 100 μ M.
12. The K_i value was calculated according to Nikolovska-Coleska, Z.; Wang, R.; Fang, X.; Pan, H.; Tomita, Y.; Li, P.; Roller, P. P.; Krajewski, K.; Saito, N. G.; Stuckey, J. A.; Wang, S. *Anal. Biochem.* **2004**, *332*, 261.
13. Sun, W.; Coleska, Z. N.; Qin, D.; Sun, H.; Yang, C. Y.; Bai, L.; Qiu, S.; Wang, Y.; Ma, D.; Wang, S. *J. Med. Chem.* **2009**, *52*, 593.
14. Varfolomeev, E.; Blankenship, J. W.; Wayson, S. M.; Fedorova, A. V.; Kayagaki, N.; Garg, P.; Zobel, K.; Elliot, L. O.; Wallweber, H. J. A.; Flygare, J. A.; Fairbrother, W. J.; Deshayes, K.; Dixit, V. M.; Vucic, D. *Cell* **2007**, *131*, 669.