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Synthesis, structure, and antimycobacterial activity of 6-[1(3*H*)-isobenzofuranylidenemethyl]purines and analogs

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1. Introduction

Tuberculosis (TB) still claims ca. two-million deaths per year world-wide and resistance to existing drugs is a growing problem.¹ We have previously reported selective antibacterial activity against Mycobacterium tuberculosis (Mtb) for certain 9-benzylpurines.² Fig. 1 shows examples of purines with profound antimycobacterial activity as well as a summary of SAR knowledge. Previous results have led us to believe that an aryl group in the purine 6-position is a requirement for significant antimycobacterial effect,^{2a} and among the 6-aryl- and 6-heteroarylpurines examined, especially high activities are found for 6-(2-furyl)purines,^{2e} for instance compounds 1a-1d (Fig. 1). The more bulky 6-(benzofur-2-yl)purine 2a is less active than the corresponding furylpurine **1a**.^{2d} However, we recently found MIC values for antimycobacterial activity of 6-[1(3H)-isobenzofuranylidenemethyl]purines 3a and 4a only one titer step higher than 6-furylpurine **1a**.³ Since we have shown that compound **4a** easily isomerizes into the Z-isomer **3a**,³ it may be that compound 4a also isomerizes into compound 3a in the antimycobacterial assay.

The profound, and somewhat unexpected, effect on *Mtb* growth found for the *Z*-6-[1(3H)-isobenzofuranylidenemethyl]purine **3a**, prompted us to synthesize and determine antimycobacterial activity for the related compound **3b** (Fig. 1) where the substitution pattern elsewhere in the molecule is optimized according to our current SAR knowledge, as well as the optimized 6-(benzofur-2-

ABSTRACT

6-Benzofuryl-, styryl, benzyl, and furfurylpurines as well as 6-[1(3H)-isobenzofuranylidenemethyl]purines have been synthesized and their activities against*Mycobacterium tuberculosis*(*Mtb*) determined. Several compounds displayed profound antimycobacterial activity in combination with lowtoxicity towards mammalian cells. NMR and X-ray crystallography were employed to determine thedetailed structures and the results were supported by quantum chemical calculations.

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yl)purine **2b** (Fig. 1) and the styryl- **5**, benzyl- **6**, and furfurylpurine **7**. The structural relationship between compounds **5–7** and the (isobenzofuranylidenemethyl)purine **3b**, is illustrated in Figure 2. It was previously shown that compound **3a** exists only as a (isobenzofuranylidenemethyl)purine, and not as an aromatic, but quinoid, benzo[c]furylmethylpurine.³ On the other hand, the simple furyl derivative **7**, may exist as tautomer **7b** rather than **7a**. Hence, we included a structural study of the synthesized compounds employing NMR, X-ray, and quantum chemical calculations in order to reveal their detailed structures and broaden our SAR knowledge.

2. Synthesis

Z-6-[1(3*H*)-Isobenzofuranylidenemethyl]purine **3b** was synthesized essentially as reported for the preparation of compound **3a** before.³ When the dichloropurine **8** was reacted with the alkyne **9** under Sonogashira coupling conditions, cyclization took place to give mainly the *E*-isomer **4b**. The mixture of isomers was treated with TFA to give the pure *Z*-6-[1(3*H*)-isobenzofuranylidenemethyl]purine **3b** (Scheme 1).

The other target molecules, **2b** and **5–7**, were available by regioselective Stille- or Negishi-coupling on the dichloropurine **8** (Scheme 2, Table 1) a synthetic strategy employed by us in the synthesis of several 6-substituted purines.^{1.4} Furfuryl halides are regarded as unstable and difficult to utilize for synthetic purposes.⁵ However, we found that the desired furfurylzinc chloride could be generated from furfuryl chloride and Rieke zinc following the protocol previously used for generation of benzylzinc chloride.⁶



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Figure 1. Structures of antimycobacterial purines and previously reported MIC values,^{2a,d,e,3} summary of SAR for antimycobacterial 9-benzylpurines, and target molecules in the study.

This constitutes the first formation and synthetic application of a furfurylmetal reagent carrying no stabilizing groups on the furyl ring.

4. Antimycobacterial activity

3. Structure

In line with our previous observations on compound 3a,³ no quinoid benzo[c]furylmethylpurine tautomer could be detected in the ¹H NMR spectrum of the *Z*-6-[1(3*H*)-isobenzofuranylide-nemethyl]purine **3b** or the isomer **4b**. On the other hand, ¹H NMR revealed that compound **7** was present only as the furfurylpurine **7b**. Electronic structure calculations supported these findings (see Supplementary data). The furfurylpurine **7** also crystallized as the **7b** tautomer (Fig. 3). The benzyl group in crystalline compound **7b** is oriented quite different from what is previously found by X-ray crystallography for the benzylic substituent in the 6-furylpurine **1d**⁷ and related structures,⁸ for further discussions, see Supplementary data.

The novel purine derivatives **2b**, **3b**, **5**, **6**, and **7b** were screened for antibacterial activity against *M. tuberculosis* $H_{37}Rv$ in vitro and the IC_{90} and IC_{50} values are presented in Table 2 together with comparable data for the compounds **1d** and **3a** synthesized earlier. Previously determined MIC values (*Mtb*) for compounds **1a–1d**, **2a**, **3a**, and **4a** are displayed in Figure 1.

Both the 6-furyl- **1d** and 6-benzofurylpurine **2b** were found to be highly potent inhibitors of *Mtb* ($IC_{90} < 0.20 \ \mu g/mL$, $< 0.6 \ \mu M$). As expected, both (isobenzofuranylidenemethyl)purines **3a** and **3b** also displayed considerable growth inhibition ($IC_{90} \ 3-5 \ \mu g/mL$, $8-12 \ \mu M$). However, the furfurylpurine **7b** was essentially inactive ($IC_{90} > 100 \ \mu g/mL$) whereas the benzyl- **6** and styrylpurine **5** were only slightly weaker inhibitors ($IC_{90} \ 6-11 \ \mu g/mL$, $16-30 \ \mu M$) compared to the (isobenzofuranylidenemethyl)purines **3**. None of the novel compounds described herein could compete with 6-furyl- **1d** or 6-benzofurylpurine **2b** where the aryl substituent is connected



Figure 2. Target molecules 5-7 and their structural relationship with target molecule 3b.



Scheme 1. Reagents and conditions: (a) (Ph₃P)₂PdCl₂, CuI, (*i*-Pr)₂NH, DMF, 60 °C; (b) TFA, CH₂Cl₂.



Scheme 2. Reagents and conditions: (a) R-Met, Pd-cat, 50 $^{\circ}\text{C},$ for more details, see Table 1.

 Table 1

 Reaction conditions applied in synthesis of compounds 5–7

-Met	Pd-catalyst	Solvent	Time (h)	Yield (%)
–SnBu ₃ –SnBu ₃ –ZnCl –ZnCl	[(2-Furyl) ₃ P] ₄ Pd [(2-Furyl) ₃ P] ₄ Pd (Ph ₃ P) ₄ Pd (Ph ₃ P) ₄ Pd	DMF DMF THF THF	8 7 5 3	78, 2b 77, 5 69, 6 59, 7

directly to C-6 in the purine, with respect to inhibition of *Mtb*. However, and in contrast to our previous believe (Fig. 1),² also other substituents (i.e., styryl- or isobenzofuranylidenemethyl-) in the 6position may result in purines with a considerable antimycobacterial activity. The compounds displaying IC₉₀ values against *Mtb* lower than 10 µg/mL were also screened for toxicity towards mammalian cells (VERO cells, Table 2). All compounds examined, except **3a**, showed IC₅₀ against VERO cells >40 µg/mL. We have previously reported that compound **1c** (Fig. 1) showed virtually no cross resistance against a panel of drug-resistant *Mtb* strains.^{2b} We assume that the closely related structures reported herein act by the same, currently unknown, mechanism of action as purine **1c**, so we have reasons to believe that cross resistance will not be an issue for the novel compounds described herein.

5. Experimental

The ¹H NMR spectra were recorded at 300 MHz with a Bruker Avance DPX 300 instrument, and the ¹H decoupled ¹³C NMR spec-

tra were recorded 75 MHz using the same instrument. Mass spectra under electron impact conditions (EI) were recorded at 70 eV ionizing voltage with a VG Prospec instrument, and are presented as m/z (% rel. int.). Elemental analyses were performed by Ilse Beetz Mikroanalytisches Laboratorium, Kronach, Germany. Melting points were determined with a C. Reichert melting point apparatus or a Büchi Melting Point B-545 apparatus and are uncorrected. DMF was distilled from BaO and stored over 4 Å mol. sieve, dichloromethane was distilled from CaH₂, and THF from Na/benzophenone. Antimycobacterial activity was determined as previously reported.² The following compounds were prepared according to literature procedures: 2-chloro-6-(2-furyl)-9-(4-methoxyphenylm-1d,^{2d} (Z)-9-benzyl-6-[isobenzofuran-1(3H)ethyl)-9H-purine ylidenemethyl]-9*H*-purine $\mathbf{3a}_{,2}^{2}$ 2,6-dichloro-9-(4-methoxyphe-nylmethyl)-9*H*-purine $\mathbf{8}_{,2}^{2d}$ (*E*)-tributyl(phenylethenyl)stannane,⁹ tributyl(benzo[b]fur-2-yl)stannane,¹⁰ and furfuryl chloride.¹¹ Benzylzinc chloride was generated from benzyl chloride and Rieke zinc¹² according to a published procedure⁶ and the concentration of the solution was determined by hydrolysis and iodolysis.¹³ Furfurylzinc chloride was generated from furfuryl chloride following the same protocol.

5.1. X-ray crystallographic analysis for compound 7b

Crystals of **7b** suitable for X-ray crystallography were obtained from benzene solution at +4 °C. X-ray data were collected on a Siemens SMART CCD diffractometer¹⁴ using graphite monochromated Mo K α radiation (λ = 0.71073 Å). Data collection method: ω -scan, step 0.3°, crystal to detector distance 5 cm. Data reduction and cell determination were carried out with the SAINT and XPREP programs. Absorption corrections were applied by the use of the sadabs program.¹⁵ The structure was determined and refined using the SHELX program package.¹⁶ The non-hydrogen atoms were refined with isotropic thermal parameters; H atoms were positioned geometrically and allowed to ride and rotate (for the CH₃ group) on their carrier atoms, with C-H bond lengths of 0.95 (aromatic C-H), 0.99 (CH₂) or 0.98 Å (CH₃) and with $U_{iso}(H) = 1.2U_{eq}(C)$ for CH₂ and aromatic C-H or 1.5U_{eq}(C) for CH₃. Crystal structure data for **7b** are available from the Cambridge Crystallographic Data Center, CCDC no. 643827.



Figure 3. X-ray structure of compound 7b, the gas-phase equilibrium structure of compound 7b, and X-ray structure of compound 1d.⁷

Table 2

Activity against *M. tuberculosis* for purines **1d**, **2b**, **3a**, **3b**, **5**, **6**, and **7b** as well as cytotoxicity against VERO cells for selected compounds^a

Compound	IC ₉₀ <i>M. tuberculosis</i>	IC ₅₀ <i>M. tuberculosis</i>	IC ₅₀ VERO
	H ₃₇ Rv (μg/mL), μM	H ₃₇ Rv (μg/mL), μM	cells (μg/
	values in brackets ^b	values in brackets ^b	mL) ^c
1d	<0.20 (<0.59)	<0.20 (<0.59)	>40
2b	<0.20 (<0.51)	<0.20 (<0.51)	>40
3a	2.7 (7.9)	1.5 (4.3)	17
3b	4.9 (12)	<0.20 (<0.48)	>40
5	6.1 (16)	3.0 (8.0)	>40
6	11 (30)	7.4 (20)	n.d.
7b	>100 (>280)	>100 (>280)	n.d.

^a Structures of compounds are shown in Schemes 1 and 2.

 b IC_{90} amicain 0.13 $\mu g/mL$ (0.22 $\mu M)$ and IC_{50} amicain 0.07 $\mu g/mL$ (0.12 $\mu M).$

 $^{c}~\text{EC}_{50}$ hyamine 0.01 $\mu\text{g/mL}.$

5.2. Crystal data for $C_{18}H_{15}CIN_4O_2$ 7b

M = 354.79, monoclinic, *P*2(1)/*n a* = 4.5528 (6) Å, *b* = 18.867(3) Å, *c* = 18.955(3) Å, β = 96.486(2)°, *V* = 1617.8(4) Å³, *Z* = 4, *Dx* = 1.457 Mg m⁻³, μ = 0.26 mm⁻¹, *T* = 105(2) K, measured 16,774 reflections in 2θ range 6.2–60.6°, *R*_{int} = 0.032. To 128 parameters refined against 4518 *F*², *R* = 0.0333 for 3293 *I*₀ > 2 σ (*I*₀) and 0.0553 for all data.

5.3. Antimycobacterial data

The purines were screened for antimycobacterial activities essentially as described before.^{2,17} Compounds were tested in 10 twofold dilutions, from 100 to 0.19 µg/mL, against *M. tuberculosis* H_{37} Rv (ATCC 27294) in BACTEC 12B medium using the Microplate Alamar Blue Assay (MABA). The IC₉₀ and IC₅₀ values are determined from the dose–response curve as the IC₉₀ using the curve fitting program XLFIT, formula 205.

5.4. Activity against VERO cells

The compounds were screened for mammalian cell cytotoxicity to VERO cells essentially as described before;² after 72 h exposure, viability is assessed using the CellTiter 96[®] Non-Radioactive Cell Proliferation Assay (MTT) reagent from Promega. Cytotoxicity is determined from the dose–response curve as the EC₅₀ using the curve fitting program XLFIT, formula 205.

5.5. 6-(Benzofuran-2-yl)-2-chloro-9-(4-methoxyphenylmeth-yl)-9*H*-purine (2b)

A mixture of tris(dibenzylideneacetone)dipalladium, Pd₂dba₃ (28 mg, 0.030 mmol), and tri(2-furyl)phosphine (51 mg, 0.22 mmol) in dry DMF (4 mL) was stirred at ambient temperature for 5 min, before a solution of 2,6-dichloro-9-(4-methoxyphenylmethyl)-9H-purine 8 (309 mg, 1.0 mmol) in DMF (4 mL) was added. After an additional 5 min, tributyl(benzo[b]fur-2-yl)stannane (575 mg, ca. 1.20 mmol, ca. 85% pure) was introduced, the resulting mixture stirred at 50 °C for 8 h and evaporated in vacuo. A satd solution of potassium fluoride in methanol (40 mL) was added to the residue, the mixture was stirred over night and evaporated in vacuo together with a small amount of silica gel. The residue was added on top of a silica gel column, and the product purified by flash chromatography eluting with CH₂Cl₂-acetone (29:1), followed by CH₂Cl₂-acetone (19:1); yield 303 mg (78%), mp 189 °C, colorless crystals. ¹H NMR (300 MHz, CDCl₃) δ = 3.75 (s, 3H, OCH₃), 5.32 (s, 2H, NCH₂), 6.87 (d, J = 8.7 Hz, 2H, Ar), 7.24-7.30 (m, 3H, Ar and benzofuryl), 7.38-7.43 (m, 1H, benzofuryl), 7.67-7.70 (m, 2H, benzofuryl), 8.02 (s, 1H, 8-H) and 8.28 (s, 1H, 3-H in benzofuryl) ppm. ¹³C NMR (75 MHz, CDCl₃) δ = 47.1 (NCH₂), 55.3 (OCH₃), 112.4 (CH in benzofuryl), 114.6 (CH in Ar), 115.2 (CH in benzofuryl), 122.4 (CH in benzofuryl), 123.6 (CH in benzofuryl), 126.4 (C-1 in Ar), 127.2 (CH in benzofuryl), 128.1 (C in benzofuryl), 128.4 (C-5), 129.6 (CH in Ar), 145.1 (C-8), 147.6 (C-6/C-2), 149.7 (C-2 in benzofuryl), 153.7 (C-4), 154.5 (C-2/C-6), 156.0 (C in benzofuryl) and 159.9 (C-4 in Ar) ppm. MS (EI): m/z (%): 392/390 (15/42) [M]⁺, 234 (1), 195 (1), 181 (2), 122 (9), 121 (100). HRMS (EI): calcd for C₂₁H₁₅ClN₄O₂ 390.0884, found 390.0886. C₂₁H₁₅ClN₄O₂ (390.8): C, 64.54; H, 3.87; N, 14.34. Found: C, 64.50; H, 3.93; N, 14.20.

5.6. 2-Chloro-6-[(*Z*)-1(3*H*)-isobenzofuranylidenemethyl]-9-(methoxyphenylmethyl)-9*H*-purine (3b)

2,6-Dichloro-9-(4-methoxyphenylmethyl)-9H-purine 8 (155 mg, 0.50 mmol) was added to a stirring solution of CuI (9.5 mg, 0.050 mmol), bis(triphenylphosphine)palladium(II) chloride (18 mg, 0.025 mmol), and dry diisopropylamine (420 µL, 3.00 mmol) in dry DMF (2.5 mL) under N₂. The solution was heated to 60 °C before 2ethynylbenzenemethanol 9 (79 mg, 0.60 mmol) dissolved in DMF (1 mL) was added dropwise over 1 h. After stirring for additional 4 h at 60 °C, the reaction mixture evaporated in vacuo with a small amount of silica. The residue was added on top of a silica gel column, and the product, as a ca. 7:3 E/Z mixture was isolated by flash chromatography eluting with CH₂Cl₂-acetone (19:1) followed by CH₂Cl₂-acetone (9:1). The isomeric mixture was dissolved in CH₂Cl₂ (30 mL) and trifluoroacetic acid (74 µL, 0.96 mmol) was added. After stirring for 1 h at ambient temperature the reaction mixture was washed with satd aq NaHCO₃ (2×20 mL), water (20 mL), brine (20 mL), dried (MgSO₄) and evaporated in vacuo. The product was purified by flash chromatography on silica gel eluting with CH₂Cl₂-acetone (9:1); yield 101 mg (50%), pale yellow crystals. ¹H NMR (300 MHz, CDCl₃) δ = 3.76 (s, 3H, OCH₃), 5.28 (s, 2H, NCH₂), 5.71 (s, 2H, OCH₂), 6.72 (s, 1H, =CH), 6.87 (d, J = 8.7 Hz, 2 H, Ar), 7.25 (d, J = 8.7 Hz, 2H, Ar), 7.40–7.48 (m, 3H, Ar'), 7.79–7.82 (m, 1H, Ar') and 7.84 (s, 1H, 8-H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ = 46.8 (NCH₂), 55.3 (OCH₃), 77.2 (OCH₂), 88.6 (=CH), 114.5 (CH in Ar), 121.3 (CH in Ar'), 121.8 (CH in Ar'), 126.9 (C-1 in Ar), 128.5 (CH in Ar'), 129.0 (C-5), 129.6 (CH in Ar), 131.1 (CH in Ar'), 133.9 (C in Ar'), 141.5 (C in Ar'), 142.7 (C-8), 152.1 (C-4), 154.6 (C-2/C-6), 156.2 (C-6/C-2), 159.8 (C-4 in Ar) and 165.7 (=C) ppm. MS (EI): m/ z (%): 406/404 (9/25) [M]⁺, 285 (3), 284 (2), 283 (8), 140 (2), 121 (100). HRMS (EI): calcd for C₂₂H₁₇ClN₄O₂ 404.1040, found 404.1030. C₂₂H₁₇ClN₄O₂ (404.9): C, 65.27; H, 4.23; N, 13.84. Found: C, 64.94; H, 4.16; N, 13.81.

5.7. (*E*)-2-Chloro-9-(4-methoxybenzyl)-6-(2-phenylethenyl)-9*H*-purine (5)

The product was formed by Stille coupling between 2,6-dichloro-9-(4-methoxyphenylmethyl)-9H-purine 8 (309 mg, 1.0 mmol) and (*E*)-tributyl(phenylethenyl)stannane (1.12 g, 1.20 mmol, ca. 42% purity) as described for compound **3b** above. The reaction time was 7 h, and the product purified by flash chromatography on silica gel eluting with CH₂Cl₂-acetone (29:1), followed by CH₂Cl₂-acetone (19:1) and finally CH_2Cl_2 -acetone (9:1); yield 292 mg (77%), colorless foam. ¹H NMR (300 MHz, CDCl₃) δ = 3.75 (s, 3H, OCH₃), 5.30 (s, 2H, NCH₂), 6.88 (d, J = 8.8 Hz, 2H, Ar), 7.26 (d, J = 8.8 Hz, 2H, Ar), 7.32-7.42 (m, 3H, Ph), 7.59 (d, I = 16.1 Hz, 1H, CH=), 7.66-7.69 (m, 2H, Ph), 7.94 (s, 1H, 8-H) and 8.42 (d, J = 16.1 Hz, 1H, CH=) ppm. ¹³C NMR (75 MHz, CDCl₃) δ = 47.0 (NCH₂), 55.3 (OCH₃), 114.5 (CH in Ar), 121.4 (CH=), 126.6 (C-1 in Ar), 128.1 (CH in Ph), 128.8 (CH in Ph), 129.6 (CH in Ar), 129.8 (CH in Ph), 130.0 (C-5), 135.7 (C-1 in Ph), 141.7 (CH=), 144.3 (C-8), 153.4 (C-4), 154.3 (C-2), 155.8 (C-6) and 159.9 (C-4 in Ar) ppm. MS (EI): *m/z* (%): 378/376 (13/36) [M]⁺, 257 (1), 255 (4), 122 (9), 121 (100). HRMS (EI): calcd for C₂₁H₁₇ClN₄O 376.1091, found 376.1093. C21H17CIN4O(376.8): C, 66.93; H, 4.55; N, 14.87. Found: C, 66.76; H, 4.46; N, 14.71.

5.8. 6-Benzyl-2-chloro-9-(4-methoxyphenylmethyl)-9*H*-purine (6)

2,6-Dichloro-9-(4-methoxyphenylmethyl)-9H-purine 8 (309 mg, 1.0 mmol) was added to a solution of tetrakis(triphenylphosphine)palladium(0)[generated in situ from tris(diphenylmethylideneacetone)dipalladium chloroform adduct (23 mg, 0.025 mmol) and triphenylphosphine (53 mg, 0.20 mmol)] in dry THF (4 mL) at ambient temperature under N₂. After 10 min, a solution of benzylzinc chloride (1.83 mL, 1.52 mmol, 0.83 M) in THF was added and the mixture stirred at 50 °C for 5 h. Satd aq NH₄Cl (10 mL) was added and the resulting mixture was extracted with EtOAc (4×25 mL). The combined organic extracts were washed with brine (2 \times 20 mL), dried (MgSO₄), and evaporated in vacuo with a small amount on silica. The residue was added on top of a silica gel column, and the product purified by flash chromatography eluting with CH₂Cl₂-acetone (29:1), followed by CH₂Cl₂-acetone (19:1) and finally CH₂Cl₂acetone (9:1); yield 250 mg (69%), colorless oil. ¹H NMR (300 MHz, $CDCl_3$) δ = 3.74 (s, 3H, OCH₃), 4.43 (CH₂), 5.25 (s, 2H, NCH₂), 6.85 (d, J = 8.7 Hz, 2H, Ar), 7.11-7.27 (m, 2H, Ar and 3H, Ph), 7.45-7.48 (m, 2H, Ph) and 7.92 (s, 1H, 8-H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ = 39.5 (CH₂), 47.0 (NCH₂), 55.3 (OCH₃), 114.5 (CH in Ar), 126.4 (C-1 in Ar), 126.7 (CH in Ph), 128.5 (CH in Ph), 129.3 (CH in Ph), 129.6 (CH in Ar), 131.7 (C-5), 137.0 (C-1 in Ph), 144.4 (C-8), 152.8 (C-4), 154.1 (C-2), 159.9 (C-4 in Ar) and 162.8 (C-6) ppm. MS (EI): m/z (%): 366/364 (24/60) [M]⁺, 245 (9), 243 (27), 207 (2), 180 (3), 153 (2), 121 (100). HRMS (EI): calcd for C₂₀H₁₇ClN₄O 364.1091, found 364.1090. C₂₀H₁₇ClN₄O (363.8): C, 65.84; H, 4.70; N, 15.36. Found: C, 65.83; H, 4.84; N, 15.26.

5.9. 2-Chloro-6-(furan-2-ylmethyl)-9-(4methoxyphenylmethyl)-9*H*-purine (7b)

The product was formed by Negishi coupling between 2,6-dichloro-9-(4-methoxyphenylmethyl)-9H-purine 8 (309 mg, 1.0 mmol) and furfurylzinc chloride (3.9 mL, 1.60 mmol, 0.41 M) as described for compound **6** above. The reaction time was 3 h, and the product purified by flash chromatography on silica gel eluting with CH₂Cl₂-acetone (29:1), followed by CH₂Cl₂-acetone (19:1) and finally CH₂Cl₂-acetone (9:1); yield 208 mg (59%), mp 135 °C, colorless crystals. ¹H NMR (300 MHz, CDCl₃) δ = 3.74 (s, 3H, OCH₃), 4.46 (s, 2H, CH₂), 5.27 (s, 2H, NCH₂), 6.22 (dd, *J* = 3.2 and 0.7 Hz, 1H, 3-H in furyl), 6.25 (dd, *J* = 3.2 and 1.9 Hz, 1H, 4-H in furyl), 6.85 (d, J = 8.7 Hz, 2H, Ar), 7.25 (d, J = 8.7 Hz, 2H, Ar), 7.28 (dd, J = 1.8 and 0.8 Hz, 1H, 5-H in furyl) and 7.94 (s, 1H, 8-H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ = 32.2 (CH₂), 47.0 (NCH₂), 55.3 (OCH₃), 107.6 (C-3 in furyl), 110.5 (C-4 in furyl), 114.5 (CH in Ar), 126.4 (C-1 in Ar), 129.6 (CH in Ar), 131.7 (C-5), 141.9 (C-5 in furyl), 144.7 (C-8), 149.9 (C-2 in furyl), 152.9 (C-4), 154.1 (C-2), 159.8 (C-6) and 159.9 (C-4 in Ar) ppm. MS (EI): *m/z* (%): 356/354 (15/43) [M]⁺, 121 (100), 91 (3), 78 (5), 77 (5). HRMS (EI): calcd for

 $C_{18}H_{15}CIN_4O_2$ 354.0884, found 354.0873. $C_{18}H_{15}CIN_4O_2$ (354.8): C, 60.94; H, 4.26; N, 15.79. Found: C, 60.92; H, 4.28; N, 15.71.

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bmc.2009.08.012.

References and notes

- 1. See for instance: Bhowruth, V.; Dover, L. G.; Besra, G. S. *Prog. Med. Chem.* **2007**, 45, 169. and references therein.
- (a) Bakkestuen, A. K.; Gundersen, L.-L.; Langli, G.; Liu, F.; Nolsøe, J. M. N. Bioorg. Med. Chem. Lett. 2000, 10, 1207; (b) Gundersen, L.-L.; Nissen-Meyer, J.; Spilsberg, B. J. Med. Chem. 2002, 45, 1383; (c) Andresen, G.; Gundersen, L.-L.; Nissen-Meyer, J.; Rise, F.; Spilsberg, B. Bioorg. Med. Chem. Lett. 2002, 12, 567; (d) Bakkestuen, A. K.; Gundersen, L.-L.; Utenova, B. T. J. Med. Chem. 2005, 48, 2710; (e) Brændvang, M.; Gundersen, L.-L. Bioorg. Med. Chem. 2005, 13, 6360; (f) Brændvang, M.; Gundersen, L.-L. Bioorg. Med. Chem. 2007, 15, 7144.
- Berg, T. C.; Bakken, V.; Gundersen, L.-L.; Petersen, D. Tetrahedron 2006, 62, 6121.
- 4. Langli, G.; Gundersen, L.-L.; Rise, F. Tetrahedron 1996, 52, 5625.
- 5. Takanishi, K.; Hirokazu, U.; Kuwajima, I. Tetrahedron Lett. 1987, 28, 2281.
- 6. Zhu, L.; Wehmeyer, R. M.; Rieke, R. D. J. Org. Chem. 1991, 56, 1445.
- (a) Brændvang, M.; Gundesen, L.-L. Acta. Crystallogr., Sect. C 2007, 63, 0274– 0276; (b) Brændvang, M.; Gundersen, L.-L. Acta. Crystallogr., Sect. E 2007, 63, 03036.
- Mazumdar, P. A.; Das, A. K.; Bakkestuen, A. K.; Gundersen, L.-L.; Bertolasi, V. Acta. Crystallogr., Sect. E 2001, 57, o1052.
- 9. Labadie, J. W.; Tueting, D.; Stille, J. K. J. Org. Chem. 1983, 48, 4634.
- 10. Liebeskind, L. S.; Wang, J. J. Org. Chem. 1993, 58, 3550.
- 11. Chaudhari, S. S.; Akamanchi, K. G. Synlett 1999, 1763.
- (a) Chen, T.-A.; Wu, X.; Rieke, R. D. J. Am. Chem. Soc. 1995, 117, 233–244; (b) Rieke, R. D.; Hanson, M. V.; Brown, J. D.; Niu, Q. J. J. Org. Chem. 1996, 61, 2726.
- Knochel, P.; Jones, P.; Langer, F. In Organozinc Reagents: A Practical Approach; Knochel, P., Jones, P., Eds.; Oxford University Press: Oxford, 1999; p 14.
 SMART (Version 5.625) and SAINT+ (Version 6.02). Area-Detector Control and
- Integration Software, Bruker Analytical X-ray Instruments, Madison, WI, 1998. 15. Sheldrick, G. M. sadabs. Program for Empirical Correction of Area Detector Data,
- Sheldrer, G. M. Subass, Frightan for Empirical contection of Area Detector Data, University of Göttingen, Germany, 1996.
 Cheldreich, G. M. Subass, Frightan for Subass, Germany, 1996.
- Sheldrick, G. M. SHELXL (release 97-2)—Programs for Crystal Structure Analysis Structure, University of Göttingen, Germany, 1997.
- 17. Collins, L. A.; Franzblau, S. G. Antimicrob. Agents Chemother. 1997, 41, 1004.