

## Note

## Enantioselective Synthesis of Aspergillide B

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**The enantioselective synthesis of aspergillide B, a 14-membered macrocyclic cytotoxin, was achieved in a 49% yield via 7 steps from a synthetic intermediate of aspergillide C. The spectroscopic data and specific rotation value for the synthetic material matched those of natural aspergillide B.**

**Key words:** aspergillide; cytotoxic; macrolide; enantioselective synthesis

Aspergillides A–C, which exhibit significant cytotoxicity against mouse lymphocytic leukemia cells (L1210), were recently isolated by Kusumi and co-workers from a bromine-modified 1/2PD (potato-dextrose) culture medium of the marine-derived fungus, *Aspergillus ostianus* strain 01F313, and their structures were proposed to be heptaketidic 14-membered macrolides based on extensive spectroscopic analyses.<sup>1)</sup> The structural proposal for aspergillides A and B was, however, revealed to be incorrect by the synthesis of the proposed structures by Uenishi and co-workers.<sup>2)</sup> They concluded that the genuine structure of aspergillide B must be represented by structure **1** (Fig. 1), and the real structure of aspergillide A should be reinvestigated.<sup>2)</sup> We also took an interest in the unique structures of aspergillides A–C proposed by Kusumi *et al.*, which featured a 14-membered macrolide structure incorporating a 2,6-*trans*-substituted tetrahydro- or dihydropyran ring,<sup>3)</sup> and embarked on their total synthesis. We recently completed the enantioselective total synthesis of the proposed structure of aspergillide C (**2**), and confirmed its structure.<sup>3)</sup> In this note, we describe a new synthesis for aspergillide B (**1**) from a synthetic intermediate of **2**.

As shown in Scheme 1, our synthesis of **1** began with the saponification of **5** and subsequent *in-situ* iodolactonization of the resulting carboxylate salt to give **6**; olefinic ester **5** in turn was prepared according to our previously reported procedure, using **3** and **4** as chiral sources.<sup>3)</sup> Reductive elimination of iodolactone **6** proceeded smoothly, affording **7** in a 90% yield from **5**. Hydrolysis of the lactone moiety of **7** with lithium hydroxide in aqueous THF gave a mixture containing the corresponding hydroxy carboxylate. The mixture was concentrated to dryness, dissolved in DMF, and treated with TBSOTf, imidazole and DMAP to give a bis-silylated intermediate, the TBS ester group of which

was then selectively hydrolyzed by directly adding water to the reaction mixture to afford **8** in an 89% yield from **7**.<sup>3)</sup> Oxidative deprotection of the PMB group with DDQ gave hydroxy carboxylic acid **9** in a 77% yield. Finally, seco acid **9** was subjected to the known two-step sequence involving Yamaguchi lactonization and TBS deprotection to afford aspergillide B (**1**) in an 80% yield.<sup>2)</sup> The <sup>1</sup>H- and <sup>13</sup>C-NMR spectra of **1**, obtained as a microcrystalline solid (mp 82.5–83.5 °C), were identical with those reported for natural aspergillide B, and the specific rotation of **1** {[α]<sup>25</sup><sub>D</sub> –108 (c 0.175, MeOH)} showed good agreement with reported data {[α]<sup>31</sup><sub>D</sub> –97.2 (c 0.27, MeOH),<sup>1)</sup> [α]<sup>20</sup><sub>D</sub> –90.0 (c 0.10, MeOH)<sup>2)</sup>

## Experimental

IR spectra were recorded by a Jasco FT/IR-4100 spectrometer, using an ATR (ZnSe) attachment. NMR spectra were recorded with TMS as an internal standard in CDCl<sub>3</sub> by a Varian Gemini 2000 spectrometer (300 MHz for <sup>1</sup>H and 75 MHz for <sup>13</sup>C), unless otherwise stated. Optical rotation values were measured with a Horiba Septa-300 polarimeter, and mass spectra were obtained with a Jeol JMS-700 spectrometer. Merck silica gel 60 (70–230 mesh) was used for column chromatography.

(3*a*S,5*R*,7*a*S)-5-[(1*E*,6*S*)-6-(4-Methoxybenzyloxy)-1-heptenyl]hexahydrofuro[3,2-*b*]pyran-2-one (**7**). To a stirred solution of **5** (30.7 mg, 79.0 μmol) in THF (0.15 ml) was added a solution of NaOH (9.7 mg, 0.23 mmol) in water (50 μl) at room temperature. The mixture was stirred at 40 °C for 5 h and then cooled to room temperature. To the mixture were successively added a solution of NaHCO<sub>3</sub> (67.9 mg, 0.808 mmol) in water (1 ml) and a solution of I<sub>2</sub> (26.8 mg, 0.106 mmol) and KI (67.8 mg, 0.408 mmol) in water (1 ml). After being stirred overnight in the dark, the mixture was quenched with satd. Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> aq. and extracted with CHCl<sub>3</sub>. The resulting extract was washed with brine, dried (MgSO<sub>4</sub>), and concentrated *in vacuo* to give **6** (42.5 mg) as a yellow oil which was then taken up in toluene (2.0 ml). To the solution was successively added Bu<sub>3</sub>SnH (30.0 μl, 0.108 mmol) and a solution of Et<sub>3</sub>B (1 M in hexane, 45.0 μl, 45 μmol) at –30 °C. After being stirred at –30 °C for 30 min under an oxygen atmosphere, the mixture was quenched with satd. NaHCO<sub>3</sub> aq. and extracted with EtOAc. The resulting extract was washed with brine, dried (MgSO<sub>4</sub>), and concentrated *in vacuo*. The residue was purified by silica gel column chromatography (hexane/EtOAc = 3:1) to give 26.6 mg (90%) of **7** as a pale yellow oil. [α]<sup>23</sup><sub>D</sub> –34.5 (c 1.18, CHCl<sub>3</sub>); IR *v*<sub>max</sub>: 1782 (vs), 1612 (m), 1513 (s), 1246 (s), 1034 (s); <sup>1</sup>H-NMR δ: 1.18 (3H, d, *J* = 6.0 Hz, 7'-H<sub>3</sub>), 1.37–1.61 (5H, m), 1.97–2.11 (5H, m), 2.50 (1H, dd, *J* = 17.3, 1.1 Hz, 3-H), 2.65 (1H, dd, *J* = 17.3, 4.4 Hz, 3-H), 3.46–3.55 (1H, m, 6'-H), 3.80 (3H, s, OCH<sub>3</sub>), 4.33–4.40 (3H, m, 3*a*-H, 7*a*-H, 5-H), 4.37 (1H, d, *J* = 11.3 Hz, Ar-CH), 4.51 (1H, d, *J* = 11.3 Hz, Ar-CH), 5.57 (1H, ddt, *J* = 15.7, 4.9, 1.4 Hz, 1'-H), 5.71 (1H, ddt, *J* = 15.7, 1.4, 6.6 Hz, 2'-H), 6.84–6.90 (2H, m, Ar-H), 7.23–

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Abbreviations: PMB, *p*-methoxybenzyl; TBS, *tert*-butyldimethylsilyl; TBSOTf, *tert*-butyldimethylsilyl trifluoromethanesulfonate; imid, imidazole; DMAP, 4-(dimethylamino)pyridine; DDQ, 2,3-dichloro-5,6-dicyano-1,4-benzoquinone

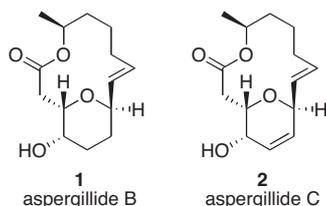
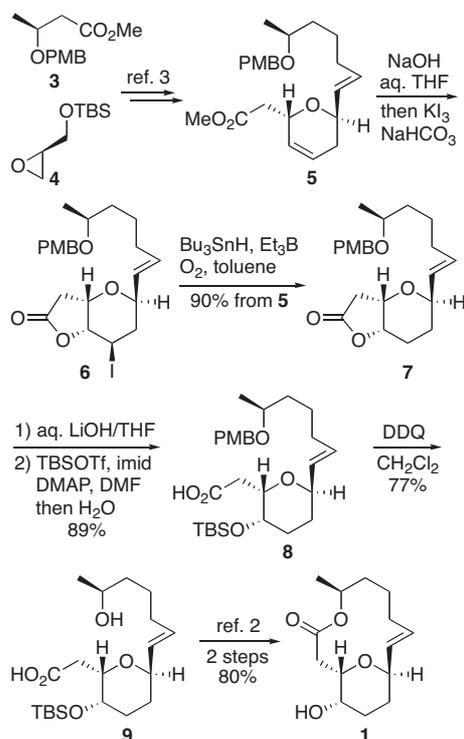


Fig. 1. Structures of Aspergillides B and C.



Scheme 1. Synthesis of Aspergillide B.

7.29 (2H, m, Ar-H);  $^{13}\text{C-NMR}$   $\delta$ : 19.5, 20.6, 22.1, 24.9, 32.4, 36.1, 38.0, 55.2, 67.2, 69.9, 71.7, 74.2, 76.9, 113.8 (2C), 127.4, 129.2 (2C), 131.2, 134.4, 159.2, 176.2; HRMS (EI)  $m/z$ : calcd. for  $\text{C}_{22}\text{H}_{30}\text{O}_5$ , 374.2093; found, 374.2095 ( $\text{M}^+$ ).

*{(2S,3S,6R)-3-(tert-Butyldimethylsilyloxy)-6-[(1E,6S)-6-(4-methoxybenzyloxy)-1-heptenyl]tetrahydropyran-2-yl}acetic acid (8)*. To a stirred solution of **7** (26.2 mg, 70.0  $\mu\text{mol}$ ) in THF (0.10 ml) was added a solution of LiOH·H<sub>2</sub>O (3.4 mg, 77  $\mu\text{mol}$ ) in water (30  $\mu\text{l}$ ) at room temperature. After 2 h, the mixture was concentrated *in vacuo* to give a lithium carboxylate salt as a pale yellow solid which was then dissolved in DMF (0.2 ml). To the solution were successively added a solution of imidazole (26.7 mg, 0.392 mmol) and DMAP (6.0 mg, 49  $\mu\text{mol}$ ) in DMF (0.25 ml) and TBSOTf (68  $\mu\text{l}$ , 0.290 mmol) at room temperature. After 2 h, water (2.0  $\mu\text{l}$ ) was added, and the mixture was stirred for 1 h. The mixture was diluted with water and extracted with  $\text{CH}_2\text{Cl}_2$ . The extract was washed with brine, dried ( $\text{MgSO}_4$ ), and concentrated *in vacuo*. The residue was purified by silica gel column chromatography (hexane/EtOAc = 2:1) to give 31.5 mg (89%) of **8** as a pale yellow oil.  $[\alpha]_D^{23}$   $-11.3$  ( $c$  0.780,  $\text{CHCl}_3$ ); IR  $\nu_{\text{max}}$ : 3000 (br m), 1711 (s), 1513 (m), 1248 (s), 835 (s);  $^1\text{H-NMR}$   $\delta$ : 0.05 (3H, s,  $\text{SiCH}_3$ ), 0.06 (3H, s,  $\text{SiCH}_3$ ), 0.89 (9H, s,  $\text{Si}(\text{C}(\text{CH}_3)_3)$ ), 1.17 (3H, d,  $J = 6.3$  Hz,

$7''\text{-H}_3$ ), 1.32–1.68 (6H, m), 1.72–1.83 (1H, m), 1.85–1.96 (1H, m), 2.01 (2H, br q,  $J = 6.6$  Hz), 2.61 (1H, dd,  $J = 15.7, 4.9$  Hz, 2-H), 2.73 (1H, dd,  $J = 15.7, 8.8$  Hz, 2-H), 3.43–3.53 (1H, m,  $6''\text{-H}$ ), 3.76–3.86 (1H, m,  $3'\text{-H}$ ), 3.80 (3H, s,  $\text{OCH}_3$ ), 4.13–4.20 (1H, m,  $2'\text{-H}$ ), 4.22–4.29 (1H, m,  $6'\text{-H}$ ), 4.37 (1H, d,  $J = 11.4$  Hz, Ar-CH), 4.49 (1H, d,  $J = 11.4$  Hz, Ar-CH), 5.45 (1H, br dd,  $J = 15.7, 5.5$  Hz,  $1'\text{-H}$ ), 5.66 (1H, ddt,  $J = 15.7, 1.1, 6.6$  Hz,  $2''\text{-H}$ ), 6.84–6.90 (2H, m, Ar-H), 7.23–7.29 (2H, m, Ar-H);  $^{13}\text{C-NMR}$   $\delta$ :  $-5.1, -4.9, 17.9, 19.5, 24.8, 25.7$  (3C), 27.2, 27.3, 32.3, 33.4, 36.0, 55.2, 67.6, 69.9, 70.9, 72.2, 74.3, 113.8 (2C), 129.3 (2C), 129.4, 131.2, 133.3, 159.2, 176.4; HRMS (EI)  $m/z$ : calcd. for  $\text{C}_{28}\text{H}_{46}\text{O}_6\text{Si}$ , 506.3064; found, 506.3069 ( $\text{M}^+$ ).

*{(2S,3S,6R)-3-(tert-Butyldimethylsilyloxy)-6-[(1E,6S)-6-hydroxy-1-heptenyl]tetrahydropyran-2-yl}acetic acid (9)*. To a stirred mixture of **8** (22.5 mg, 44.4  $\mu\text{mol}$ ) in  $\text{CH}_2\text{Cl}_2$  (0.4 ml)/1 M phosphate buffer (pH 7.0, 0.2 ml) was added DDQ (22.2 mg, 94.9  $\mu\text{mol}$ ) at room temperature. After 12 h, additional DDQ (15.9 mg, 67.9  $\mu\text{mol}$ ) was added, and the mixture was stirred for 4 h. The mixture was extracted with  $\text{CH}_2\text{Cl}_2$ , and the extract was washed with brine, dried ( $\text{MgSO}_4$ ), and concentrated *in vacuo*. The residue was purified by silica gel column chromatography (hexane/EtOAc = 2:1–1:0) to give 13.2 mg (77%) of **9** as a pale yellow oil.  $[\alpha]_D^{22}$   $-27.9$  ( $c$  0.815,  $\text{CHCl}_3$ ); IR  $\nu_{\text{max}}$ : 3402 (br w), 3000 (br w), 1713 (s), 1104 (s), 836 (s);  $^1\text{H-NMR}$   $\delta$ : 0.05 (3H, s,  $\text{SiCH}_3$ ), 0.06 (3H, s,  $\text{SiCH}_3$ ), 0.89 (9H, s,  $\text{Si}(\text{C}(\text{CH}_3)_3)$ ), 1.18 (3H, d,  $J = 6.0$  Hz,  $7''\text{-H}_3$ ), 1.36–1.52 (5H, m), 1.57–1.69 (1H, m), 1.74–1.84 (1H, m), 1.88–1.97 (1H, m), 2.00–2.10 (2H, m,  $3''\text{-H}_2$ ), 2.56 (1H, dd,  $J = 15.6, 4.4$  Hz, 2-H), 2.72 (1H, dd,  $J = 15.6, 9.3$  Hz, 2-H), 3.75–3.85 (2H, m,  $3'\text{-H}, 6''\text{-H}$ ), 4.17–4.29 (2H, m,  $2'\text{-H}, 6'\text{-H}$ ), 5.48 (1H, br dd,  $J = 15.7, 5.8$  Hz,  $1''\text{-H}$ ), 5.68 (1H, ddt,  $J = 15.7, 1.1, 6.6$  Hz,  $2''\text{-H}$ );  $^{13}\text{C-NMR}$   $\delta$ :  $-5.1, -4.8, 17.9, 23.2, 24.9, 25.7$  (3C), 27.0, 27.2, 32.1, 33.8, 38.4, 67.7, 68.0, 70.9, 72.1, 129.5, 133.3, 176.3; HRMS (FAB)  $m/z$ : calcd. for  $\text{C}_{20}\text{H}_{39}\text{O}_5\text{Si}$ , 387.2566; found, 387.2564 ( $[\text{M} + \text{H}]^+$ ).

*(1S,5S,9E,11R,14S)-14-Hydroxy-5-methyl-4,15-dioxabicyclo[9.3.1]-pentadec-9-en-3-one (1)*. Compound **1** was prepared from **9** in an 80% yield via 2 steps according to the procedure reported in ref. 2. Mp 82.5–83.5  $^\circ\text{C}$ ;  $[\alpha]_D^{25}$   $-108$  ( $c$  0.175, MeOH); IR  $\nu_{\text{max}}$ : 3437 (br m), 2930 (s), 1724 (vs), 1183 (m), 1023 (s);  $^1\text{H-NMR}$  ( $\text{C}_6\text{D}_6$ )  $\delta$ : 0.99 (1H, dddd,  $J = 14.0, 4.9, 2.5, 1.2$  Hz, 6-H), 1.06 (3H, d,  $J = 6.3$  Hz, 5- $\text{CH}_3$ ), 1.27–1.44 (3H, m), 1.46–1.68 (3H, m), 1.68–1.91 (2H, m), 1.86–1.96 (1H, br, OH), 1.98–2.09 (1H, m, 8-H), 2.12 (1H, dd,  $J = 13.7, 1.9$  Hz, 2-H), 2.72 (1H, dd,  $J = 13.7, 11.5$  Hz, 2-H), 3.21 (1H, br s, 14-H), 4.08 (1H, br d,  $J = 11.5$  Hz, 1-H), 4.28–4.34 (1H, m, 11-H), 5.04–5.14 (1H, m, 5-H), 5.39 (1H, br dd,  $J = 15.7, 4.4$  Hz, 10-H), 6.19 (1H, dddd,  $J = 15.7, 10.9, 4.9, 1.9$  Hz, 9-H);  $^{13}\text{C-NMR}$  ( $\text{C}_6\text{D}_6$ )  $\delta$ : 19.0, 22.4, 25.1, 27.6, 30.6, 31.9, 39.7, 67.2, 69.5, 69.7, 71.4, 129.0, 138.2, 169.9; HRMS (EI)  $m/z$ : calcd. for  $\text{C}_{14}\text{H}_{22}\text{O}_4$ , 254.1518; found, 254.1520 ( $\text{M}^+$ ).

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