Angiogenesis Inhibitor Epoxyquinol A: Total Synthesis and Inhibition of Transcription Factor NF-*k*B

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ABSTRACT



The asymmetric synthesis of the natural product (+)-epoxyquinol A (1) and related epoxyquinoid dimers, employing a cascade oxidation/ electrocyclization/Diels–Alder dimerization sequence, is reported. In addition, we show that 1 and related molecules inhibit activation of the transcription factor NF- κ B.

Angiogenesis, new blood vessel formation, involves a number of distinct processes, including endothelial cell migration, proliferation, and capillary tube formation, and is believed to be a key requirement for tumor growth and metastasis.¹ Angiogenesis inhibition thus represents an important approach to cancer chemotherapy, and several agents, including those derived from natural product leads, are now entering clinical development.² The pentaketide dimer (+)-epoxyquinol A (1) was recently isolated from an uncharacterized fungus by Osada and co-workers³ and was shown to have potent antiangiogenic activity. The relative stereochemistry of 1 was determined by X-ray crystallography. We have previously reported syntheses of the dimeric epoxyquinon natural product torreyanic acid⁴ and two monomeric epoxyquinol natural products (cycloepoxy-



don (inset, Figure 1)⁵ and jesterone).⁶ Thus far, chemical synthesis of an epoxyquinol dimer has not been reported. In this Letter, we report the synthesis and absolute stereochemical assignment of (+)-epoxyquinol A and four related epoxyquinoid dimers. In addition, we show that **1** and related molecules inhibit activation of transcription factor NF- κ B.

A retrosynthetic analysis for (+)-epoxyquinol A is depicted in Figure 1. Compound 1 may be prepared by an

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Figure 1. Retrosynthetic analysis for epoxyquinol A.

oxidation/electrocyclization/Diels—Alder dimerization of diastereomeric 2*H*-pyran monomers 2/2'.^{4,6} In this biomimetic, *endo*-selective⁷ Diels—Alder heterodimerization, the methyl groups of the pyran are *anti* to one another and the dienophile approaches the diene *anti* to the epoxide moiety. Monomers 2/2' may be derived from selective primary oxidation of epoxyquinol **3** (RK-302, isolated from the same fermentation broth as **1**³) followed by 6π -electrocyclic ring closure of the resulting dienal **4**. Compound **3** may be prepared from readily available chiral epoxide **5**, which is a known intermediate in our previous synthesis of cycloepoxydon.⁵

The synthesis was initiated by Stille coupling of **5** with (*E*)-tributyl-1-propenyl-stannane to afford α -propenyl enone **6** in near quantitative yield (Scheme 1).⁸ Sequential hydrolysis of the cyclic acetal and silyl ether protecting groups (aqueous HF) afforded the quinone monoepoxide **7**. Regioand stereoselective reduction of **7** using Kiyooka's conditions⁹ (2 equiv of DIBAL-H, THF, -78 °C) afforded **3**¹⁰ (72%) as the major diastereomer.

In line with our previous studies,^{4,6} treatment of **3** with Dess-Martin periodinane afforded a crude product mixture, which was subjected to column chromatography to provide two major fractions. NMR analysis indicated that the major fraction contained 2H-pyrans 2/2' and aldehyde **4** (cf. Figure 1), as well as the desired dimer **1**. The minor fraction comprised quinone epoxide **7**, the related quinone 2H-pyran monomers **8/8'**, and quinone dimer **9** (Scheme 1).¹¹ This result indicates that undesired oxidation of the secondary allylic alcohol was occurring under these conditions.¹² To



^{*a*} Reagents and conditions: (a) (*E*)-tributyl-1-propenyl-stannane, Pd₂dba₃, AsPh₃, PhCH₃, 110 °C, 2 h (98%); (b) 48% HF, CH₃CN, rt, 1 h (84%); (c) DIBAL-H, THF, -78 °C, 10 min (72%); (d) O₂ (1 atm), TEMPO, CuCl, DMF, 3 h; (e) 40% MeOH–H₂O, 10 h, **1** (55%), **10** (14%); (f) Dess–Martin periodinane, CH₂Cl₂, 1.5 h; SiO₂, 2 h, 67%. DIBAL-H = diisobutylaluminum hydride, TEMPO = 2,2,6,6-tetramethyl-1-piperidinyloxy.

promote the desired electrocyclization/Diels-Alder dimerization, the major fraction was treated with silica gel in CHCl₃ for 20 h.⁴ Epoxyquinol A was obtained in 26% yield after silica gel chromatography. Synthetic (+)-epoxyquinol A was confirmed to be identical to natural (+)-epoxyquinol A by ¹H and ¹³C NMR, mass spectrum, $[\alpha]_D$ (+66° (c =0.23, CHCl₃)), and TLC R_f values in three different solvent systems.

Alternative oxidation methods were next evaluated in order to obtain higher selectivity for oxidation of the primary vs secondary allylic alcohol of 3 (Scheme 1). We found that selective oxidation could be achieved using conditions reported by Semmelhack (CuCl (cat.), O2, TEMPO, DMF).13 Under these conditions, no oxidation of the secondary allylic alcohol or overoxidation to the carboxylic acid was detected. After oxidation, the crude product mixture¹⁴ was dissolved in CDCl₃, and ¹H NMR analysis was used to monitor the disappearance of aldehyde and 2H-pyran. Use of this slightly acidic condition for dimerization afforded endo dimer 1 (36%) along with a related dimer 10 (30%) after column chromatography. In contrast, Diels-Alder dimerization in 40% aqueous MeOH (10 h) afforded epoxyquinol A and 10 in 55% and 14% yield, respectively.¹⁵ The structure of 10 was determined to be an exo-dimer of two identical 2H-pyran monomers¹⁶ by X-ray crystal structure analysis (Figure 2).

⁽⁷⁾ In refs 3 and 4, epoxyquinol and epoxyquinone dimers were referred to as "*exo*." However, it is more appropriate to refer to the dimers as "*endo*" with respect to the carbonyl substituent on the 2*H*-pyran dienophile.

⁽⁸⁾ Attempted Stille coupling using Pd(PPh₃)₄ as catalyst afforded the undesired 1,3-diketone as the main product; cf.: Suzuki, M.; Watanabe, A.; Noyori, R. J. Am. Chem. Soc. **1980**, 102, 2095.

⁽⁹⁾ Kiyooka, S.; Kuroda, H.; Shimasaki, Y. Tetrahedron Lett. 1986, 27, 3009.

⁽¹⁰⁾ Synthetic **3** has the same HPLC retention time and the same R_f by TLC with the natural compound RK-302 (see Supporting Information).

⁽¹¹⁾ *Endo* quinone dimer **9** was also produced in 67% yield by Dess– Martin periodinane oxidation of **7**, followed by treatment with silica gel (Scheme 1). See Supporting Information for details.

⁽¹²⁾ It is also conceivable that internal hydride transfer of dienal **4** to a quinone epoxide may occur. For a recent example of an internal hydride transfer, see: Szendi, Z.; Forgó, P.; Tasi, G.; Böcskel, Z.; Nyerges, L.; Sweet, F. *Steroids* **2002**, *67*, 31.

⁽¹³⁾ Semmelhack, M. F.; Schmid, C. R.; Cortes, D. A.; Chou, C. S. J. Am. Chem. Soc. **1984**, *106*, 3374.

^{(14) &}lt;sup>1</sup>H NMR (CDCl₃) analysis of the crude reaction mixture indicates a ratio of dienal 4:2H-pyran 2:dimer 1:dimer 10 of 1:4.9:7.9:7.7.

⁽¹⁵⁾ For enhancement of *endo/exo* selectivity by water in Diels-Alder reactions, see: Otto, S.; Engberts, J. B. F. N. *Pure Appl. Chem.* **2000**, *72*, 1365.



Figure 2. X-ray structure of dimer 10.

With diastereomeric epoxyquinol dimers **1** and **10** in hand, we next investigated their reactivity to thermolysis. Surprisingly, both dimers were found to be stable at 80 °C in CDCl₃. Microwave irradiation was next used as a method to establish rapid thermodynamic equilibration of the compounds.¹⁷ Treatment of **1** in the CEM Discover microwave system (chlorobenzene, 180 °C, 5 min) afforded *exo* dimer **10** (14%), dimer **11** (43%) (Figure 3), and **12** (10%). Extended



Figure 3. Microwave irradiation of epoxyquinol A and AM1 equilibrium geometries of 11 and 12 (PC Spartan Pro, v. 1.0.7).

microwave irradiation of 1 (180 °C, 2×5 min) led to the production of *exo* dimer 10 in higher yield (36%) along with 11 (25%) and 12 (10%). However, when dimer 10 was irradiated using the same conditions, starting material was recovered, indicating higher stability of 10, possibly derived from intramolecular hydrogen bonding.¹⁸ The structure of 11 was assigned by NOE difference NMR spectroscopy to be an *endo* dimer in which the dienophile *approaches the*

diene syn to the epoxide. Dimer 12 was similarly determined to be an exo dimer in which the dienophile approaches the diene syn to the epoxide. Interestingly, dimer 11 is derived from Diels-Alder reaction of diastereomeric 2H-pyran monomers (cf. 1), and 12 from identical 2*H*-pyran monomers (cf. 10). Evidently, 11 and 12 are not kinetically favored products and may be obtained under microwave irradiation as a result of the higher activation energy provided under these conditions. On the basis of these results, three underlying rules for Diels-Alder dimerizations of 2H-pyran epoxyquinol monomers are thus apparent: (1) two identical or diastereomeric 2H-pyran monomers may undergo endoor exo-Diels-Alder cycloaddition; (2) the methyl groups of the diene and dienophile tend to orient themselves away from one another, presumably to avoid steric interactions; and (3) kinetically, the dienophile approaches the diene anti to the epoxide moiety (cf. 1 and 10),¹⁹ whereas thermodynamically (by higher activation energy with microwave irradiation), the dienophile can approach the diene syn to the epoxide moiety.

Because other epoxyquinoids, including both natural^{5,20} and synthetic²¹ molecules, have been shown to inhibit activation of transcription factor NF- κ B,²² we determined whether epoxyquinol A and three synthetic derivatives also had this ability. As shown in Table 1 and Figure 4, dimers

Table 1. Inhibition of NF- κ B DNA Binding by Epoxyquinol A (1) and Related Compounds

compound	IC ₅₀ (µM)
epoxyquinol A (1) quinone dimer (9) quinol monomer (3) quinone monomer (7)	$egin{array}{c} 11^a \ 10^a \ 20^b \ 2.3^b \end{array}$

^a Average of four experiments. ^b Average of two experiments.

1 and **9** and monomers **3** and **7** all inhibit tumor necrosis factor (TNF)-induced activation of NF- κ B DNA binding in mouse 3T3 cells. In these assays, monomer **7** was approximately 5–10 times more effective than the others, with an IC₅₀ of 2.3 μ M. Of note, transcription of the gene encoding vascular endothelial growth factor (VEGF), one of the primary mediators of angiogenesis, is known to be controlled, at least in part, by NF- κ B.²³ Indeed, the concentrations of **1**

⁽¹⁶⁾ For *endo* Diels-Alder dimerization of two identical 2*H*-pyran monomers, see ref 4.

⁽¹⁷⁾ For select examples of microwave irradiation of natural products, see: (a) Salmoria, G. V.; Dall'Oglio, E. L.; Zucco, C. Synth. Commun. **1997**, 29, 4335. (b) Das, B.; Venkataiah, B. Synth. Commun. **1999**, 29, 863. (c) Das, B.; Madhusudhan, P.; Venkataiah, B. Synth. Commun. **2000**, 30, 4001. (d) Das, B.; Venkataiah, B.; Kashinatham, A. Tetrahedron **1999**, 55, 6585.

⁽¹⁸⁾ Dimer **10** shows a distance of 2.7–2.8 Å between the two hydroxyl oxygens (X-ray, two conformers), as well as an anomalous downfield shift for one hydroxyl proton (¹H NMR). For a recent example of intramolecular OH–OH hydrogen bonding, see: Vasquez, T. E., Jr.; Bergset, J. M.; Fierman, M. B.; Nelson, A.; Roth, J.; Khan, S. I.; O'Leary, D. J. *J. Am. Chem. Soc.* **2002**, *124*, 2931.

⁽¹⁹⁾ For an example of Diels–Alder cycloaddition with π -facial selectivity *anti* to the diene epoxide moiety, see: Gillard, J. R.; Newlands, M. J.; Bridson, J. N.; Burnell, D. J. *Can. J. Chem.* **1991**, *69*, 1337.

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⁽²²⁾ For a review of natural products as modulators of the NF- κ B signaling pathway, see: Bremner, P.; Heinrich, M. J. Pharm. Pharmacol. **2002**, *54*, 453.

⁽²³⁾ For recent reports on suppression of angiogenesis by inhibition of NF- κ B in cancer cells, see: (a) Huang, S.; Robinson, J. B.; DeGuzman, A.; Bucana, C. D.; Fidler, I. J. *Cancer Res.* **2000**, *60*, 5334. (b) Huang, S.; Pettaway, C. A.; Uehara, H.; Bucana, C. D.; Fidler, I. J. *Oncogene* **2001**, *20*, 4188.



Figure 4. Compound **7** inhibits tumor necrosis factor-induced activation of NF- κ B. Mouse 3T3 cells were untreated (lane 1) or were treated with TNF- α (lanes 2–6), and NF- κ B DNA-binding activity was analyzed in an electrophoretic mobility shift assay. In lanes 3–6, cells were preincubated with 0.675, 1.25, 2.5, and 5 μ M, respectively, **7** prior to treatment with TNF- α ; in lanes 1 and 2, cells were preincubated with solvent (CH₃OH). The arrow indicates the induced NF- κ B (p50-RelA) complex.

reported to inhibit cell migration induced by VEGF³ (ED₁₀₀ = 7 μ M) and activation of NF- κ B (Table 1) are similar. Thus, our demonstration that epoxyquinol A and related compounds inhibit induction of NF- κ B may explain the molecular basis for inhibition of angiogenesis by epoxyquinol A. If so, one would predict that **7**, the most effective inhibitor of TNF-induced activation of NF- κ B, would also be the most effective blocker of angiogenesis.

In summary, we have completed an asymmetric synthesis of epoxyquinol A employing a [4 + 2] dimerization of

epoxyquinol 2*H*-pyran monomers. Additional epoxyquinone and epoxyquinol dimers have been produced as part of these studies, which further clarifies the underlying rules for Diels—Alder dimerizations of epoxide-containing 2*H*-pyran monomers. Notably, microwave irradiation of epoxyquinol A has been used to produce novel stereoisomers of **1** that are not accessible at ambient temperature. Future biological experiments will be aimed at identifying the molecular target(s) for epoxyquinol A mediated inhibition of NF- κ B activation and angiogenesis. Similarly, it will be of interest to determine whether epoxyquinol A and related compounds also inhibit other biological processes, including inflammation and the growth of certain tumor cells, that are dependent on NF- κ B.

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Supporting Information Available: Experimental procedures and characterization data for all new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

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