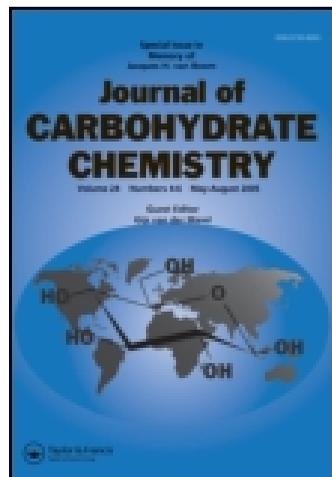


This article was downloaded by: [Akdeniz Universitesi]

On: 25 December 2014, At: 14:18

Publisher: Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information:

<http://www.tandfonline.com/loi/lcar20>

Synthesis and Antigenic Property of a Novel Sialyl 6-O-Sulfo Lewis X Neo-glycolipid Containing Lactamized Neuraminic Acid

Masanori Yamaguchi ^a, Hideharu Ishida ^a, Akiko Kanamori ^b, Reiji Kannagi ^{b c} & Makoto Kiso ^{a c}

^a Department of Applied Bio-organic Chemistry, Gifu University, Gifu, 501-1193, Japan

^b Department of Molecular Pathology, Research Institute, Aichi Cancer Center, Nagoya, Japan

^c CREST, Japan Science and Technology Agency (JST), Japan
Published online: 17 Aug 2006.

To cite this article: Masanori Yamaguchi, Hideharu Ishida, Akiko Kanamori, Reiji Kannagi & Makoto Kiso (2004) Synthesis and Antigenic Property of a Novel Sialyl 6-O-Sulfo Lewis X Neo-glycolipid Containing Lactamized Neuraminic Acid, *Journal of Carbohydrate Chemistry*, 23:4, 201-215, DOI: [10.1081/CAR-200030039](https://doi.org/10.1081/CAR-200030039)

To link to this article: <http://dx.doi.org/10.1081/CAR-200030039>

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms &

Conditions of access and use can be found at <http://www.tandfonline.com/page/terms-and-conditions>

Synthesis and Antigenic Property of a Novel Sialyl 6-O-Sulfo Lewis X Neo-glycolipid Containing Lactamized Neuraminic Acid[#]

Masanori Yamaguchi,¹ Hideharu Ishida,^{1,*} Akiko Kanamori,²
Reiji Kannagi,^{2,3} and Makoto Kiso^{1,3,*}

¹Department of Applied Bio-organic Chemistry, Gifu University, Gifu, Japan

²Department of Molecular Pathology, Research Institute,
Aichi Cancer Center, Nagoya, Japan

³CREST, Japan Science and Technology Agency (JST), Japan

CONTENTS

ABSTRACT	202
I. INTRODUCTION	202
II. RESULTS AND DISCUSSION	203
III. EXPERIMENTAL	207
A. General Methods	207
ACKNOWLEDGMENTS	214
REFERENCES	214

[#]Synthetic Studies on Sialoglycoconjugates, Part 136. For part 135, see Ref.^[1].

*Correspondence: Hideharu Ishida and Makoto Kiso, Department of Applied Bio-organic Chemistry, Gifu University, Gifu 501-1193, Japan; E-mail: ishida@cc.gifu-u.ac.jp or kiso@cc.gifu-u.ac.jp.

ABSTRACT

Synthesis and antigenic property of a novel 6-*O*-sulfated sLe^x neo-glycolipid containing lactamized neuraminic acid are described. Coupling of methyl (methyl 4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-5-trifluoroacetamido-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-4,6-di-*O*-acetyl-2-*O*-benzoyl-1-thio- β -*D*-galactopyranoside (**3**) with 2-(tetradecyl)hexadecyl (2,3,4-tri-*O*-benzyl- α -*L*-fucopyranosyl)-(1 \rightarrow 3)-2-acetamido-2-deoxy-6-*O*-4-methoxyphenyl- β -*D*-glucopyranoside (**7**) gave a protected sLe^x tetrasaccharide glycolipid (**8**). Removal of all the acyl protecting groups and subsequent lactamization afforded the lactamized sLe^x derivative(**10**), which was converted to the target compound (**14**) by selective removal of the 4-methoxyphenyl group and 6-*O*-sulfation of the GlcNAc residue, and removal of all protective groups under the basic conditions furnished the target molecule. The antigenic property of the synthesized neo-glycolipid was examined by TLC-immunostaining with G159 monoclonal antibody.

Key Words: Selectin; Sialic acid; Sialyl Lewis X; Glycolipid.

INTRODUCTION

L-selectin mediates the attachment of lymphocytes to specialized high endothelial venules (HEV) in the course of their migration from the blood to lymphoid tissues.^[2] We have shown that the sialyl 6-*O*-sulfo Lewis X serves as the major endogenous ligand for L-selectin on HEV in human lymph nodes,^[3,4] and proposed the ligand processing pathway involving "activation" by de-*N*-acetylation of sialic acid and "inactivation" by lactamization (Fig. 1). The G159 mAb has newly been cloned as the monoclonal antibody which specifically recognizes the lactamized-sialyl 6-*O*-sulfo Le^x ganglioside.^[5]

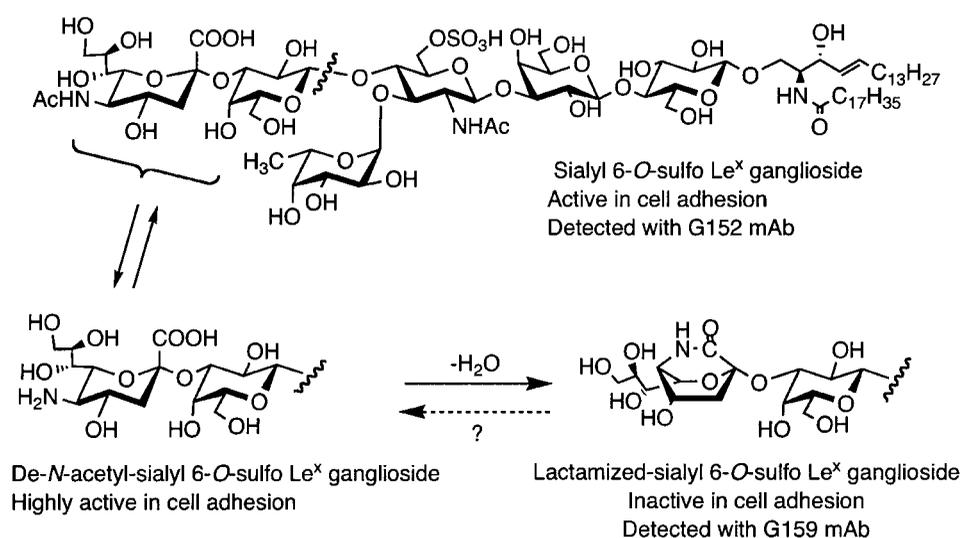


Figure 1. Proposed ligand processing pathway and binding reactivity for human L-selectin.

To investigate the structure required for recognition by G159 monoclonal antibody, we have synthesized lactamized GM4^[6] and sLe^x^[7] analogs containing lactamized neuraminic acid. However, these compounds were not recognized by G159 mAb, suggesting that the antibody may specifically recognize the structure involving the 6-*O*-sulfated GlcNAc residue. We here describe the synthesis of lactamized-sialyl 6-*O*-sulfo Lewis X tetrasaccharide glycolipid (**14**) containing both lactamized neuraminic acid and the 6-*O*-sulfated GlcNAc residue being expected as the minimal structure defined by G159 mAb (Fig. 2).

RESULTS AND DISCUSSION

The main problems to be solved in the synthesis of the title compound **14** are (i) suitable protection of the amino group at C-5 of neuraminic acid and 6-OH of the GlcNAc residue, which undergo lactamization and sulfation, respectively, and (ii) efficient introduction of the lipid part and fucose. To solve the first problem, we designed the suitably protected sialyl Lewis X tetrasaccharide glycolipid **8** (Sch. 1) as the key intermediate,

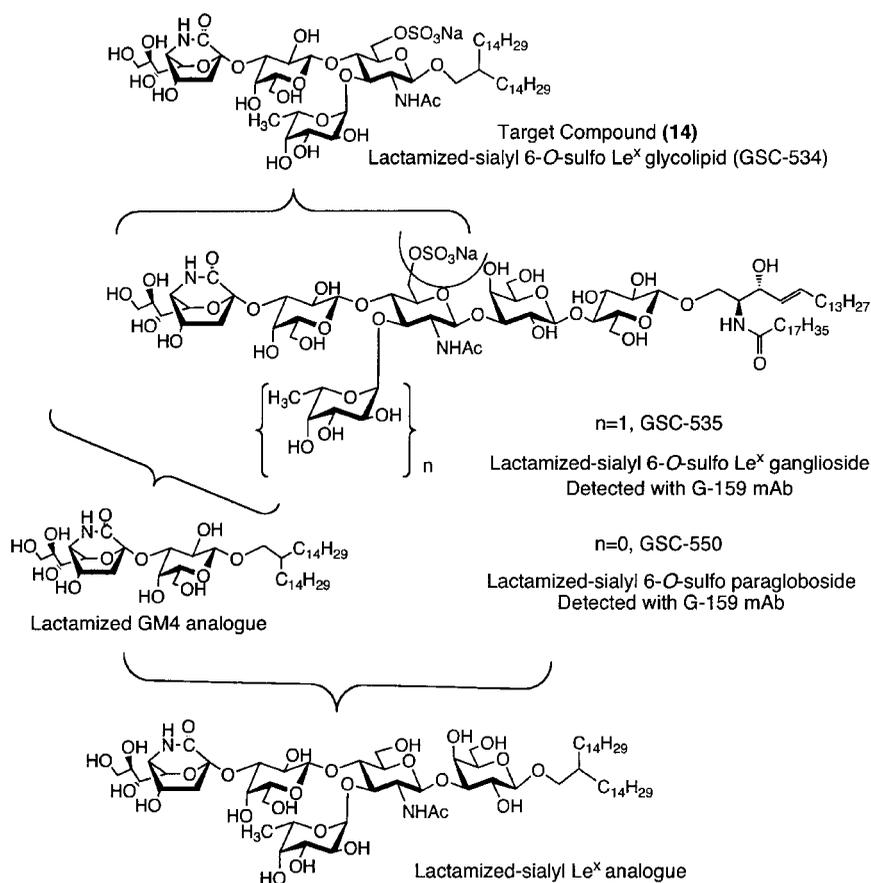
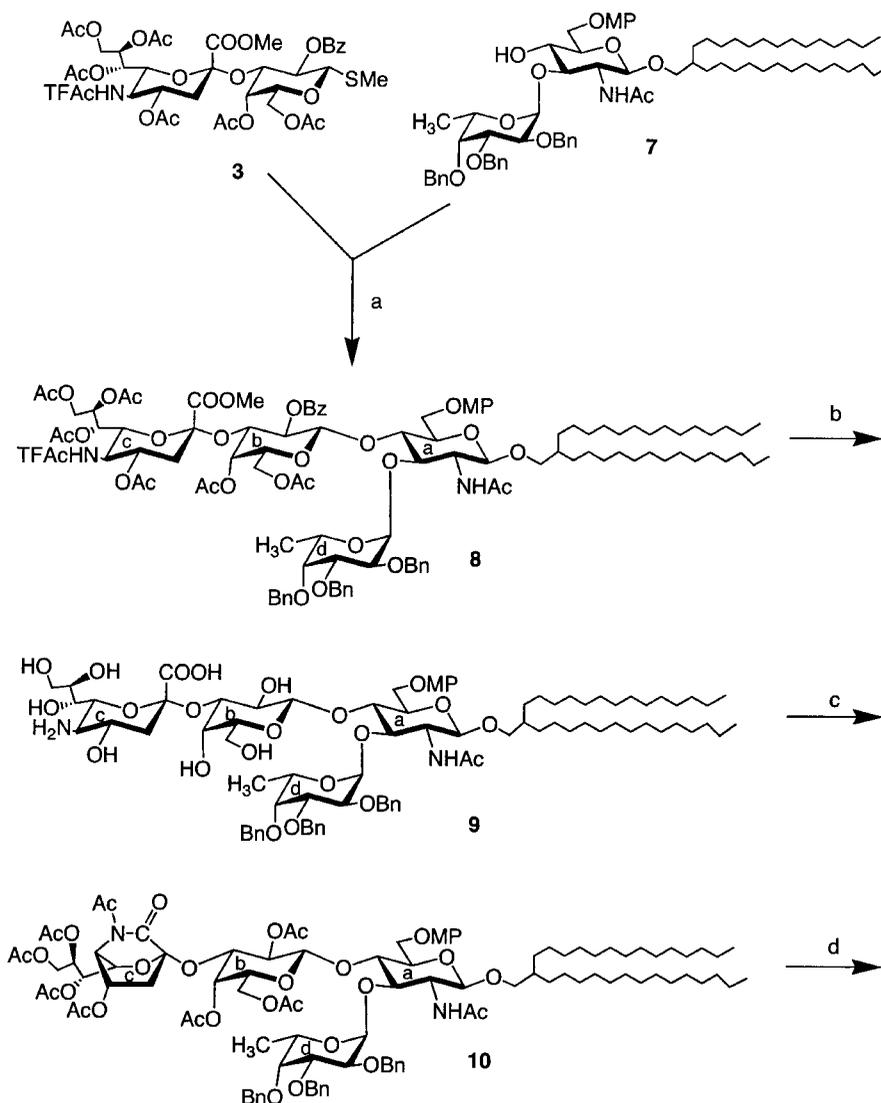
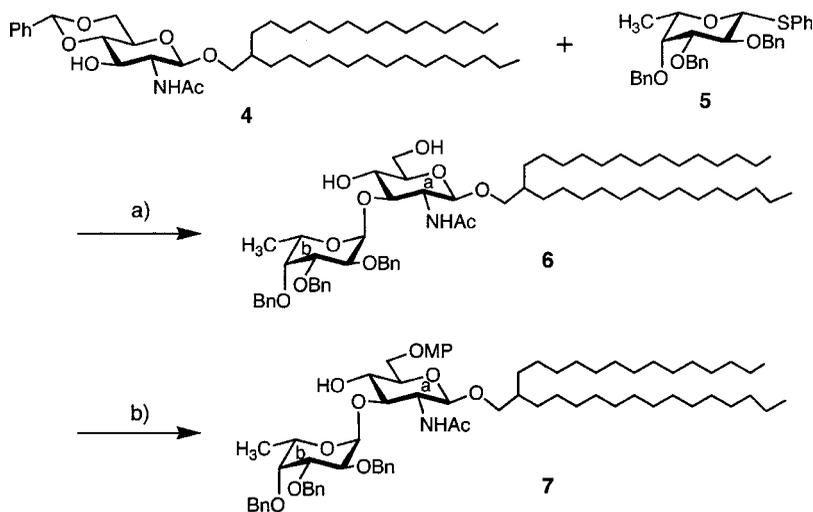


Figure 2. Lactamized-sialyl Le^x analog.



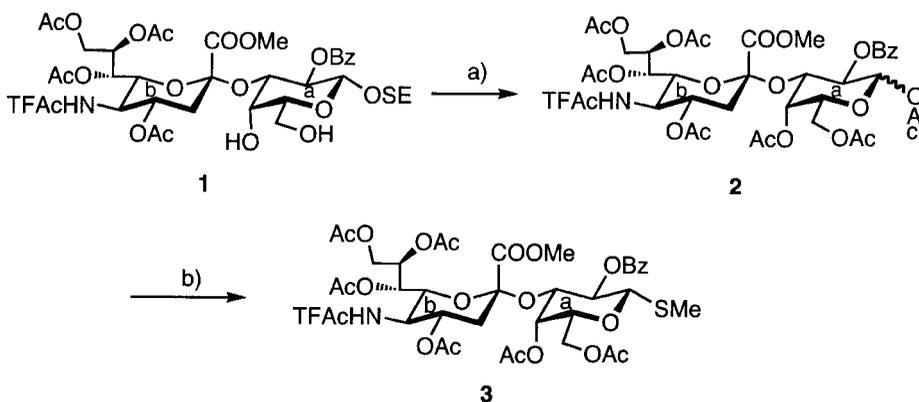
Scheme 1. (a) DMTST/ CH_2Cl_2 , MS4A, 0°C , 57%; (b) NaOMe/MeOH, then H_2O , 45°C , 95%; (c) 1, HBTU, HOBT/DMF, 65°C , 2, Ac_2O /Pyr, 76.2% (two steps); (d) 1, H_2 , $\text{Pd}(\text{OH})_2/\text{EtOH}$, 2, Ac_2O /Pyr, 93% (two steps).

in which the amino group of neuraminic acid and 6-OH of the GlcNAc residue are protected by the trifluoroacetyl (TFAc) and 4-methoxyphenyl (MP) groups, respectively. The TFAc protected sialic acid is equivalent to the *N*-deacetylated and lactamized sialic acids, and the MP group can be chemoselectively cleaved by ceric ammonium nitrate (CAN). For the second problem, we examined the efficient construction of 2-(tetradecyl)hexadecyl (2,3,4-tri-*O*-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)-2-acetamido-6-*O*-4-methoxyphenyl-2-deoxy- β -D-glucopyranoside (**7**) (Sch. 2).



Scheme 2. (a) 1, NIS, TfOH/benzene, 7°C; 2, 80% AcOHaq, 45°C, 51%, two steps and (b) MPOH, PPh₃, DEAD/THF, 80°C, 88%.

The suitably protected sialyl galactose donor **3** was prepared by treatment of **1**^[8] with boron trifluoride etherate in dichloromethane–acetic anhydride, giving the acetate **2** (97%, $\alpha/\beta = 5/1$),^[9] and successive treatment of **2** with methylthiotrimethylsilane (TMSSMe) and trimethylsilyl trifluoromethanesulfonate (TMSOTf) in dichloroethane for 72 hr at 50°C (Sch. 3). For the synthesis of acceptor **7**, compound **4** was first synthesized by using the established method,^[10] and then coupled with the fucose donor **5**.^[11] The resulting crude disaccharide was treated with 80% AcOH at 45°C to afford **6** in 51% yield in two steps (Sch. 2). The low yield of this reaction was owing to a cleavage of the tri-*O*-benzylated fucoside during the acidic hydrolysis of the benzylidene acetal. In the ¹H NMR spectrum of **6**, the anomeric proton of fucose appeared at δ 4.94 (d, $J_{1,2} = 3.8$ Hz, H-1b), indicating the newly formed glycoside linkage to be α . The regioselective 4-methoxyphenylation

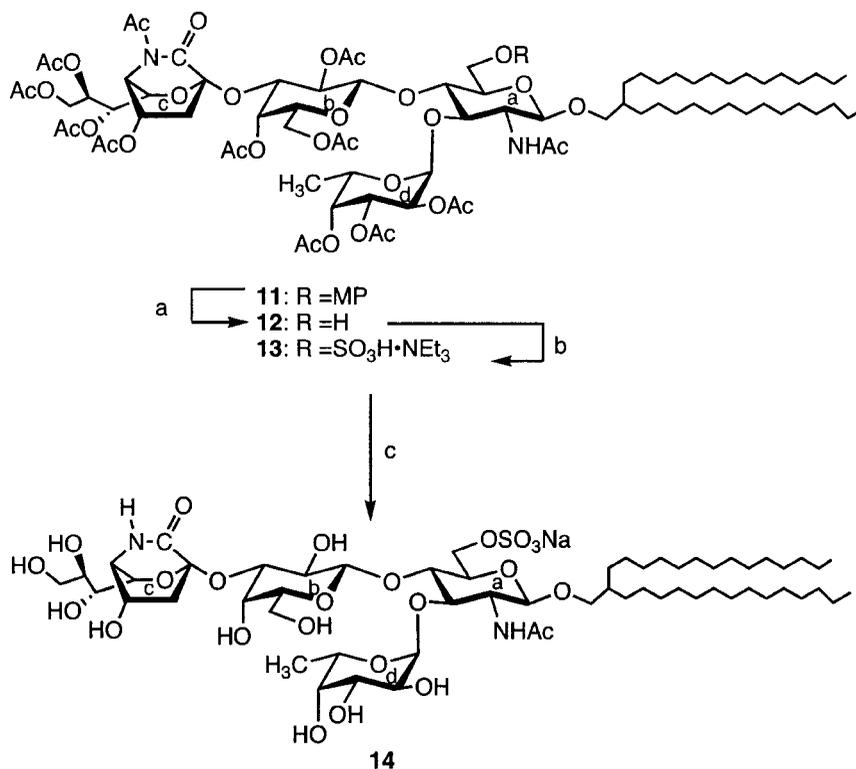


Scheme 3. (a) BF₃·OEt₂, Ac₂O/CH₂Cl₂, 97% and (b) TMSSMe, TMSOTf/CH₂ClCH₂Cl, 50°C, 89%.

at *O*-6 of **6** was carried out by treatment with *p*-methoxyphenol (MPOH), PPh₃, and diethylazodicarboxylate (DEAD) in THF,^[12,13] to give **7** in 88% yield.

Coupling of **3** and **7**, promoted by dimethyl(methylthio)sulfonium triflate (DMTST),^[14,15] gave the desired tetrasaccharide glycolipid **8** in 57% yield (Sch. 1). The partial *N,O*-deacylation of **8** with sodium methoxide in methanol for 72 hr at 45°C, and subsequent saponification of the methylester group afforded *N*-deacetylated sLe^x derivative **9** in 95% yield, which upon treatment with HBTU^[16] and HOBt in DMF at 65°C, followed by acetylation to afford the desired **10** in good yield (76%, two steps). The ¹H NMR spectrum of **10** showed signals at δ 2.32 (dd, 1H, *J*_{3β,4} = 10.1 Hz, *J*_{gem} = 14.6 Hz, H-3cβ), and δ 2.41 (dd, 1H, *J*_{3α,4} = 5.5 Hz, *J*_{gem} = 14.6 Hz, H-3cα), which are characteristic of the *B*^{5,2} boat conformation of sialic acid.^[6] Hydrogenolytic removal of the benzyl (Bn) groups in **10** and the following acetylation gave **11** (93%, two steps). Selective cleavage of the MP group in **11** and the subsequent 6-*O*-sulfation of **12** with a sulfur trioxide–pyridine (SO₃·Pyr.) complex in DMF, followed by an addition of triethylamine to stabilize the sulfate group during the column chromatography, gave **13** in good yield. Removal of all protective groups in **13** under alkaline conditions furnished the target compound **14** (GSC-534) in high yield (Sch. 4).

As shown in Fig. 3, the compound **14** (GSC-534) was not stained with G159 antibody in TLC-immunostaining, in contrast with the lactamized-sialyl 6-*O*-sulfo Le^x ganglioside



Scheme 4. (a) CAN/CH₃CN, H₂O, 0°C, 75%; (b) SO₃·Pyr complex/DMF, then Et₃N, r.t., 78%; and (c) NaOMe/MeOH, r.t., 98%.

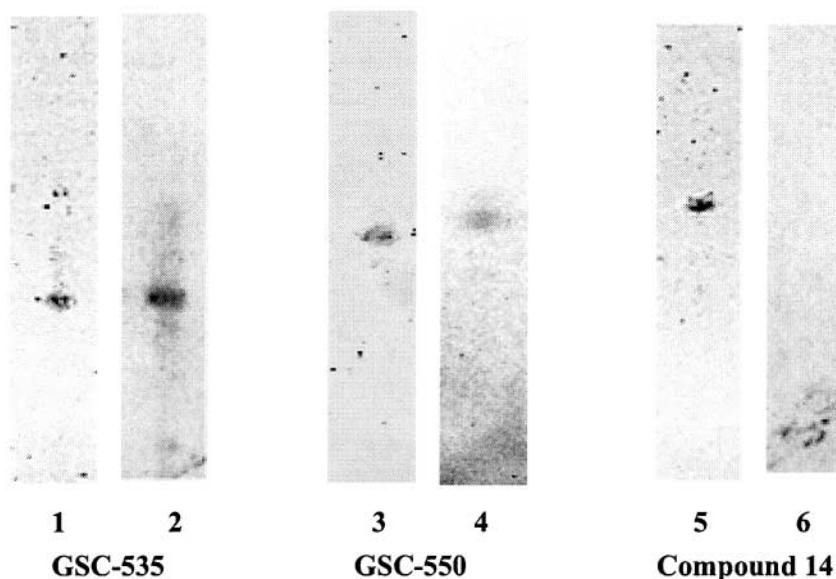


Figure 3. TLC-immunostaining patterns of synthesized glycolipids. Lanes 1, 3, and 5 are Orcinol/ H_2SO_4 staining. Lanes 2, 4, and 6 are G159 mAb staining. Lactamized-sialyl 6-*O*-sulfo Lewis X ganglioside (GSC-535) and lactamized-sialyl 6-*O*-sulfo paragloboside (GSC-550) were detected with G159 mAb, but compound **14** was not.

(GSC-535)^[1] as well as the corresponding paragloboside derivative (GSC-550)^[1] (Fig. 2). These results suggest that the lactose moiety could be critical for the recognition by G159 mAb either as the essential component of the G159-defined determinant or as a spacer to keep the determinant apart from the lipid moiety adsorbed on TLC sheets. The difference in the structures of the ceramide and the artificial ceramide (B30) also may affect the recognition by G159 mAb. A further study to elucidate the details of the recognition mapping defined by G159 mAb is now under investigation.

In summary, we have successfully synthesized a novel 6-*O*-sulfated sLe^x neo-glycolipid, which was designed as a probe to determine the minimum structure required for the recognition by G159 mAb. Utilizing this synthetic probe, the significance of the lactose moiety for the recognition by G159 mAb was suggested.

EXPERIMENTAL

General Methods

TLC was conducted on E. Merck silica gel 60 F-254 aluminum plate. Compounds were visualized by exposure to UV light or by spraying with a solution of 10% H_2SO_4 in ethanol. Column chromatography on silica gel (Fuji Silysia Co., 300 mesh) was performed with the solvent systems (v/v) specified. Specific rotations were determined with a Horiba SEPA-300 high-sensitive polarimeter at 25°C. ^1H NMR and ^{13}C NMR spectra were recorded

at 300 K with a Varian Unity Inova 500 (500 MHz) or Varian Unity Inova 400 (100.6 MHz) spectrometer, respectively. The values (ppm) are given relative to Me₄Si as the internal standard. MALDI-TOF mass spectra were recorded on a BRUKER Daltonics Autoflex-G MALDI-TOF Mass spectrometer instrument using a 2,5-dihydroxybenzoic acid matrix. Dichloromethane, methanol, ethanol, benzene, and DMF were kept dry over 4 Å MS, while pyridine and acetonitrile were kept dry over 3 Å MS.

Methyl (methyl 4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-5-trifluoroacetamido-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-4,6-di-*O*-acetyl-2-*O*-benzoyl-1-thio- β -*D*-galactopyranoside (3). To a solution of **1** (108 mg, 0.118 mmol) in Ac₂O (0.5 mL) and dry CH₂Cl₂ (1 mL) was added BF₃·OEt₂ (130 μ L, 0.47 mmol), and the mixture was stirred for 24 hr at room temperature and extracted with chloroform. The extract was successively washed with NaHCO₃ and water, dried (Na₂SO₄), and concentrated. Column chromatography (70:1 CHCl₃-MeOH) of the residue on silica gel gave **2** (108.1 mg, 97.3%) as an amorphous mass ($\alpha/\beta = 5/1$).

To a solution of **2** (108.1 mg, 0.12 mmol) in dry CH₂ClCH₂Cl (1 mL) were added TMSSMe (57.4 μ L, 0.45 mmol) and TMSOTf (77.8 μ L, 0.4 mmol). The mixture was stirred for 72 hr at 50°C. After dilution with chloroform, the extract was successively washed with 1 M sodium carbonate and water, dried (Na₂SO₄), and concentrated. Column chromatography (100:1 CHCl₃-MeOH) of the residue on silica gel afforded **3** (94.5 mg, 88.6%) as an amorphous mass; $[\alpha]_D + 31.2^\circ$ (c 0.18, CHCl₃); ¹H NMR (CDCl₃) δ 1.42 (s, 3H, MeS), 1.71 (t, 1H, $J_{gem} = J_{3ax,4} = 12.5$ Hz, H-3 $_{\text{bax}}$), 1.94, 2.05, 2.056, 2.12, 2.16, 2.18 (6s, 18H, 6AcO), 2.57 (dd, 1H, $J_{3eq,4} = 4.5$ Hz, $J_{gem} = 12.5$ Hz, H-3 $_{\text{beq}}$), 3.84 (s, 3H, COOMe), 4.34 (dd, 1H, $J_{8,9} = 2.8$ Hz, $J_{gem} = 12.5$ Hz, H-9b), 4.69 (d, 1H, $J_{1,2} = 9.8$ Hz, H-1a), 4.76 (dd, $J_{2,3} = 9.6$ Hz, $J_{3,4} = 3.2$ Hz, H-3a), 4.96 (m, 1H, H-4b), 5.03 (d, 1H, H-4a), 5.18 (dd, $J_{6,7} = 2.0$, $J_{7,8} = 8.9$ Hz, H-7b), 5.35 (t, 1H, H-2a), 5.52 (m, 1H, H-8b), 6.48 (d, 1H, $J_{5,NH} = 9.1$ Hz, NH of b), 7.46–8.14 (m, 5 H, 1Ph).

¹³C NMR (CDCl₃) δ 11.40, 11.99, 20.45, 20.70, 20.74, 20.79, 21.48, 37.29, 49.63, 53.29, 62.10, 62.24, 66.30, 67.95, 68.48, 68.62, 71.10, 72.20, 74.36, 83.22, 96.84, 128.46, 130.06, 130.26, 133.53, 157.67, 165.53 (C=O), 167.82 (C=O), 169.77 (C=O), 170.22 (C=O), 170.52 (C=O), 170.73 (C=O), 170.74 (C=O), 170.79 (C=O).

Anal. Calcd for C₃₈H₄₆F₃NO₂₀S (925.83): C, 49.30; H, 5.01; N, 1.51. Found: C, 49.12; H, 4.83; N, 1.25.

2-(Tetradecyl)hexadecyl (2,3,4-tri-*O*-benzyl- α -*L*-fucopyranosyl)-(1 \rightarrow 3)-2-acetamido-2-deoxy- β -*D*-glucopyranoside (6). To a solution of **4** (105 mg, 0.14 mmol) and **5** (113.6 mg, 0.21 mmol) in dry benzene (5 mL) was added molecular sieves 4 Å (300 mg), and the mixture was stirred for 6 hr at room temperature, then cooled to 0°C. *N*-Iodosuccinimide (NIS; 145 mg, 0.64 mmol) and TMSOTf (8.3 μ L, 42 μ mol) were added to the mixture, and the resultant mixture was stirred for 2 hr at 7°C and neutralized with Et₃N. After dilution with chloroform, the precipitate was filtered off, and washed with chloroform. The filtrate and washings were combined, and successively washed with 1 M sodium carbonate and sodium thiosulfate, dried (Na₂SO₄), and concentrated.

The residue was dissolved in AcOH (8 mL) and water (2 mL), and the reaction mixture was stirred for 48 hr at 45°C, and then concentrated. Column chromatography (200:1 CHCl₃-MeOH) of the residue on silica gel gave **6** (82.8 mg, 51%, two steps) as an amorphous mass; $[\alpha]_D - 26.8^\circ$ (c 0.4, CHCl₃); ¹H NMR (CDCl₃) δ 0.87 (t, 6H, $J = 6.6$ Hz, 2CH₃), 1.16 (d, 3H, $J_{5,6} = 6.4$ Hz, H-6b), 1.22–1.52 (m, 53H, 26CH₂ and CH), 1.58 (s, 3H, AcN), 3.25 (dd, 1H, $J_{1,2} = 7.8$ Hz, $J_{2,3} = 9.2$ Hz, H-2a), 3.31

(dd, 1H, OCH₂C of alkyl part), 3.41 (dd, 1H, H-4a), 3.68 (s, 1H, H-4b), 3.74–3.79 (m, 3H, H-5a, H-6'a and OCH₂C of alkyl part), 3.86–3.90 (m, 2H, H-3a and H-6a), 3.94 (dd, 1H, $J_{2,3} = 10.5$ Hz, $J_{3,4} = 2.1$ Hz, H-3b), 4.09 (dd, 1H, $J_{1,2} = 3.8$ Hz, $J_{2,3} = 10.5$ Hz, H-2b), 4.84 (d, 1H, $J_{1,2} = 7.8$ Hz, H-1a), 4.94 (d, 1H, $J_{1,2} = 3.8$ Hz, H-1b), 5.52 (d, 1H, $J_{\text{NH},2} = 7.3$ Hz, NH), 7.25–7.40 (m, 15H, 3Ph).

¹³C NMR (CDCl₃) δ 14.14, 16.71, 22.69, 23.05, 26.63, 26.79, 29.37, 29.66, 29.71, 30.10, 30.16, 30.87, 31.13, 31.92, 38.09, 56.12, 62.73, 68.14, 70.98, 72.86, 73.08, 74.08, 74.72, 75.00, 75.94, 76.73, 79.06, 83.96, 99.27, 100.62, 127.46, 127.61, 127.67, 127.75, 127.98, 128.21, 128.28, 128.35, 128.46, 128.57, 138.22, 138.28, 138.43, 170.58 (C=O).

Anal. Calcd for C₆₅H₁₀₃NO₁₀ (1057.76): C, 73.75; H, 9.81; N, 1.32. Found: C, 73.60; H, 9.58; N, 1.23.

2-(Tetradecyl)hexadecyl (2,3,4-tri-*O*-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)-2-acetamido-2-deoxy-6-*O*-4-methoxyphenyl- β -D-glucopyranoside (7). To a solution of **6** (82.8 mg, 0.078 mmol) in THF (5 mL) were added PPh₃ (102.1 mg, 0.39 mmol), DEAD (123.1 μ L, 0.28 mmol), and MPOH (58.8 mg, 0.47 mmol), and the mixture was stirred under reflux for 17 hr. After completion of the reaction, the mixture was concentrated. Column chromatography (200 : 1 CHCl₃–MeOH) of the residue on silica gel afforded **7** (80.1 mg, 88%) as an amorphous mass; $[\alpha]_{\text{D}} -24.0^\circ$ (c 0.86, CHCl₃); ¹H NMR (CDCl₃) δ 0.87 (t, 6H, $J = 6.9$ Hz, 2CH₃), 1.15 (d, 3H, $J_{5,6} = 6.4$ Hz, H-6b), 1.21–1.52 (m, 53H, 26CH₂ and CH), 1.59 (s, 3H, AcN), 3.27 (dd, 1H, $J_{\text{gem}} = 9.6$ Hz, $J_{\text{vic}} = 6.6$ Hz, OCH₂C of alkyl part), 3.32 (dd, 1H, $J_{2,3} = 10.0$ Hz, H-2a), 3.49 (t, 1H, H-4a), 3.61 (m, 1H, H-5a), 3.69 (s, 1H, H-4b), 3.75 (s, 3H, MeOPh), 3.79 (dd, 1H, OCH₂C of alkyl part), 3.95 (m, 2H, H-3a and H-3b), 4.09 (dd, 1H, $J_{1,2} = 3.6$ Hz, $J_{2,3} = 10.2$ Hz, H-2b), 4.09 (m, 1H, H-5b), 4.10–4.14 (m, 2H, H-6a and H-6'a), 4.86 (d, 1H, $J_{1,2} = 8.5$ Hz, H-1a), 4.98 (d, 1H, $J_{1,2} = 3.6$ Hz, H-1b), 5.51 (d, 1H, $J_{\text{NH},2} = 7.1$ Hz, NH), 6.79–7.40 (m, 19H, MeOPh, 3Ph).

¹³C NMR (CDCl₃) δ 11.34, 14.38, 20.39, 20.83, 24.34, 24.47, 27.07, 27.37, 27.42, 27.83, 27.88, 28.62, 28.87, 29.63, 35.73, 53.37, 54.08, 65.75, 66.28, 68.03, 70.52, 70.60, 71.66, 71.80, 72.68, 73.62, 74.41, 74.73, 74.94, 75.05, 76.78, 81.68, 96.80, 97.94, 112.17, 113.89, 125.18, 125.37, 125.44, 125.62, 125.97, 126.06, 126.16, 126.24, 135.95, 136.02, 136.15, 150.94, 151.62, 168.20 (C=O).

Anal. Calcd for C₇₂H₁₀₉NO₁₁ (1163.80): C, 74.25; H, 9.43; N, 1.20. Found: C, 74.09; H, 9.18; N, 1.01.

2-(Tetradecyl)hexadecyl (methyl 4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-(4,6-di-*O*-acetyl-2-*O*-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[(2,3,4-tri-*O*-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2-acetamido-2-deoxy-6-*O*-4-methoxyphenyl- β -D-glucopyranoside (8). To a solution of **3** (44.8 mg, 0.048 mmol) and **7** (37 mg, 0.031 mmol) in dry CH₂Cl₂ (0.6 mL) was added molecular sieves 4 Å (38 mg), and the mixture was stirred for 3 hr at room temperature, then cooled to 0°C. DMTST (89.5 mg, 0.18 mmol) was added to the mixture, and the resultant mixture was stirred for 3 days at 0°C and neutralized with Et₃N. After dilution with chloroform, the precipitate was filtered off, and washed with chloroform. The filtrate and washings were combined, and successively washed with 1 M sodium carbonate and water, dried (Na₂SO₄), and concentrated. Column chromatography (80 : 1 CHCl₃–MeOH) of the residue on silica gel afforded **8** (37.1 mg, 57.2%) as an amorphous mass; $[\alpha]_{\text{D}} +10.5^\circ$ (c 0.7, CHCl₃); ¹H NMR

(CDCl₃) δ 0.88 (t, 6H, $J = 6.9$ Hz, 2CH₃), 1.05 (d, 3H, $J_{5,6} = 6.4$ Hz, H-6d), 1.14–1.41 (m, 53H, 26CH₂ and CH), 1.51 (s, 3H, AcN), 1.71 (t, 1H, $J_{\text{gem}} = J_{3\text{ax},4} = 12.5$ Hz, H-3*cax*), 1.957, 1.959, 1.96, 1.98, 2.06, 2.18 (6s, 18H, 6AcO), 2.59 (dd, 1H, $J_{3\text{eq},4} = 4.6$ Hz, $J_{\text{gem}} = 12.5$ Hz, H-3*ceq*), 2.82 (dd, 1H, $J_{\text{gem}} = 8.7$, $J_{\text{vic}} = 6.6$ Hz, OCH₂C of alkyl part), 3.34 (s, 1H, H-4d), 3.43 (dd, 1H, $J_{\text{gem}} = 9.4$ Hz, $J_{\text{vic}} = 5.7$ Hz, OCH₂C of alkyl part), 3.63 (dd, 1H, $J_{2,3} = 9.8$ Hz, $J_{3,4} = 2.5$ Hz, H-3d), 3.68 (s, 3H, MeOPh), 3.74 (t, 1H, H-2a), 3.87 (s, 3H, COOMe), 3.95–3.98 (m, 2H, H-9c and H-5d), 4.00 (dd, 1H, H-4d), 4.36 (dd, 1H, H-9'*c*), 4.80 (dd, 1H, $J_{2,3} = 10.0$ Hz, $J_{3,4} = 3.4$ Hz, H-3b) 4.93 (d, 1H, $J_{1,2} = 8.0$ Hz, H-1b), 4.99 (m, 1H, H-4c), 5.04 (d, 1H, H-4b), 5.08 (d, 1H, $J_{1,2} = 3.6$ Hz, H-1d), 5.20 (dd, 1H, $J_{6,7} = 2.1$ Hz, $J_{7,8} = 9.2$ Hz, H-7c), 5.28 (dd, 1H, $J_{1,2} = 8.0$ Hz, $J_{2,3} = 10.0$ Hz, H-2b), 5.58 (m, 1H, H-8c), 6.10 (d, 1H, $J_{\text{NH},5} = 8.9$ Hz NH of c), 6.21 (d, 1H, $J_{\text{NH},2} = 9.15$ Hz, NH), 6.71–8.15 (m, 24H, 4Ph, MeOPh).

¹³C NMR (CDCl₃) δ 14.13, 16.54, 20.16, 20.45, 20.61, 20.59, 20.70, 21.43, 22.70, 23.12, 26.72, 26.89, 29.38, 29.67, 29.72, 29.79, 30.23, 30.92, 31.11, 31.93, 37.97, 49.74, 53.38, 55.57, 61.62, 62.06, 66.38, 66.74, 67.48, 67.61, 67.87, 68.50, 70.88, 70.93, 71.06, 71.14, 72.66, 73.08, 73.40, 74.31, 74.50, 96.89, 97.28, 99.39, 100.41, 114.51, 115.37, 127.21, 127.31, 127.43, 127.51, 127.93, 128.14, 128.27, 128.29, 128.40, 128.62, 129.93, 130.12, 133.41, 138.57, 138.76, 139.02, 152.79, 153.93, 166.10 (C=O), 167.73 (C=O), 169.54 (C=O), 169.80 (C=O), 170.11 (C=O), 170.27 (C=O), 170.54 (C=O), 170.60 (C=O), 170.69 (C=O).

Anal. Calcd for C₁₀₉H₁₅₁F₃N₂O₃₁ (2041.03): C, 64.10; H, 7.45; N, 1.37; O, 24.28. Found: C, 63.83; H, 7.41; N, 1.33.

2-(Tetradecyl)hexadecyl (5-amino-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-(β -D-galactopyranosyl)-(1 \rightarrow 4)-[(2,3,4-tri-O-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2-acetamido-2-deoxy-6-O-4-methoxyphenyl- β -D-glucopyranoside (9). To a solution of **8** (35.8 mg, 71 μ mol) in methanol (4 mL) was added catalytic amount of 28% sodium methoxide in methanol, and the mixture was stirred for 72 hr at 45°C. Water (0.1 mL) was added and the mixture was stirred for 48 hr at room temperature. The mixture was neutralized with Amberlite IR-120 (H⁺) resin and filtered. The resin was washed with methanol, and the combined filtrate and washings were concentrated. Column chromatography (MeOH) of the residue on Sephadex LH-20 gave **9** (26.3 mg, 95%) as an amorphous mass; $[\alpha]_{\text{D}} -43.2^\circ$ (c 0.52, CHCl₃); ¹H NMR (CD₃OD) δ 0.79 (t, 6H, $J = 6.9$ Hz, 2CH₃), 1.04 (d, 3H, $J_{5,6} = 6.4$ Hz, H-6d), 1.10–1.42 (m, 53H, 26CH₂ and CH), 1.60 (t, 1H, $J_{\text{gem}} = J_{3\text{ax},4} = 12.1$ Hz, H-3*cax*), 1.87 (s, 3H, AcN of a), 2.72 (dd, 1H, H-3*ceq*), 2.93 (t, 1H, H-5c), 3.14 (dd, 1H, $J_{6,7} = 2.9$ Hz, $J_{7,8} = 8.0$ Hz H-7c), 3.19 (dd, 1H, $J = 9.6$ Hz, $J_{\text{vic}} = 6.2$ Hz, OCH₂C of alkyl part), 3.41 (dd, 1H, $J_{1,2} = 7.8$ Hz, $J_{2,3} = 9.4$ Hz, H-2b), 3.42 (m, 1H, H-8c), 3.60 (m, 1H, H-4c), 3.65 (s, 3H, MeOPh), 3.80–3.83 (m, 2H, H-6c and H-3b), 3.86 (dd, 1H, $J_{1,2} = 3.7$ Hz, $J_{2,3} = 10.3$ Hz, H-2d), 4.01 (dd, 1H, $J_{2,3} = 10.3$ Hz, $J_{3,4} = 2.9$ Hz, H-3d), 4.25 (d, 1H, $J_{1,2} = 7.8$ Hz, H-1b), 4.71 (m, 1H, H-5d), 5.22 (d, 1H, $J_{1,2} = 3.7$ Hz, H-1d), 6.76–7.31 (m, 19H, 3Ph, MeOPh).

¹³C NMR (CD₃OD) δ 13.09, 15.51, 22.22, 22.37, 26.37, 26.54, 29.11, 29.39, 29.74, 30.68, 30.81, 31.71, 37.98, 40.55, 52.82, 54.83, 62.01, 66.41, 69.85, 71.98, 72.27, 74.01, 74.34, 75.04, 78.43, 95.62, 97.81, 99.78, 100.74, 114.48, 115.89, 123.84, 126.95, 127.07, 127.77, 127.90, 128.06, 128.23, 136.32, 138.85, 154.26, 157.04, 171.50 (C=O), 173.03 (C=O).

Anal. Calcd for C₈₇H₁₃₄N₂O₂₃ (1574.94): C, 66.30; H, 8.57; N, 1.78. Found: C, 66.05; H, 8.54; N, 1.55.

2-(Tetradecyl)hexadecyl (5-acetylamino-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosyl 1,5-lactam)-(2 \rightarrow 3)-(2,4,6-tri-*O*-acetyl- β -*D*-galactopyranosyl)-(1 \rightarrow 4)-[(2,3,4-tri-*O*-benzyl- α -*L*-fucopyranosyl)-(1 \rightarrow 3)]-2-acetamido-2-deoxy-6-*O*-4-methoxyphenyl- β -*D*-glucopyranoside (10). To a solution of **9** (26.3 mg, 16.6 μ mol) in DMF (3 mL) were added HBTU (19.5 mg, 50 μ mol) and HOBt (6.83 mg, 50 μ mol), and the mixture was stirred for 4 hr at 65°C, and then concentrated. Column chromatography (MeOH) of the residue on Sephadex LH-20 gave the lactamized sLe^x derivative. The residue was treated with acetic anhydride (1.5 mL) and pyridine (3 mL) for 12 hr at room temperature, then cooled to 0°C. Methanol (1 mL) was added and the mixture was concentrated, and the residue was extracted with chloroform and successively washed with cold 2 M hydrochloric acid and water, dried (Na₂SO₄), and concentrated. Column chromatography (100:1 CHCl₃-MeOH) of the residue on silica gel gave **10** (24.1 mg, 76.2%, two steps) as an amorphous mass; [α]_D -17.6° (*c* 0.48, CHCl₃); ¹H NMR (CDCl₃) δ 0.88 (t, 6H, *J* = 6.9 Hz, 2CH₃), 0.98 (d, 3H, *J*_{5,6} = 6.4 Hz, H-6d), 1.19–1.50 (m, 53H, 26CH₂ and CH), 1.97, 1.98, 2.06, 2.07, 2.10, 2.13, 2.14, 2.21 (8s, 7OAc, AcN of a), 2.32 (dd, 1H, *J*_{3 β ,4} = 10.1 Hz, *J*_{gem} = 14.6 Hz, H-3c β), 2.41 (dd, 1H, *J*_{3 β ,4} = 5.5 Hz, *J*_{gem} = 14.6 Hz, H-3c α), 2.57 (s, 3H, AcN of c), 3.28 (dd, 2H, OCH₂C of alkyl part and H-4d), 3.57 (dd, 1H, *J*_{2,3} = 10.0 Hz, *J*_{3,4} = 3.0 Hz, H-3d), 3.63 (dd, 1H, *J*_{gem} = 9.8 Hz, *J*_{vic} = 6.6 Hz, OCH₂C of alkyl part), 3.68 (s, 3H, *MeOPh*), 3.80 (m, 1H, H-5d), 3.94 (dd, 1H, *J*_{1,2} = 3.4 Hz, *J*_{2,3} = 10.1 Hz, H-2d), 4.13–4.16 (m, 2H, H-2a and H-3b), 4.19 (dd, 1H, *J*_{8,9} = 5.3 Hz, *J*_{gem} = 11.8 Hz, H-9c), 4.27 (dd, 1H, *J*_{8,9'} = 6.4 Hz, *J*_{gem} = 11.2 Hz, H-9'c), 4.49 (d, 1H, *J*_{1,2} = 8.0 Hz, H-1a), 4.55 (t, 1H, H-3a), 4.63 (d, 1H, H-4b), 4.85 (m, 1H, H-4c), 5.03 (d, 1H, *J*_{1,2} = 3.4 Hz, H-1d), 5.12 (dd, 1H, *J*_{1,2} = 8.0 Hz, *J*_{2,3} = 10.6 Hz, H-2b), 5.36 (m, 1H, H-5c), 5.43 (m, 1H, H-8c), 5.77 (dd, 1H, *J*_{6,7} = 3.7 Hz, *J*_{7,8} = 9.8 Hz, H-7c), 6.40 (d, 1H, *J*_{NH,2} = 9.2 Hz, NH), 6.74–7.39 (m, 19H, 3Ph, *MeOPh*).

¹³C NMR (CDCl₃) δ 14.14, 16.47, 20.62, 20.67, 20.72, 20.87, 21.03, 22.69, 23.09, 26.33, 26.84, 26.95, 29.38, 29.67, 29.72, 30.26, 31.04, 31.15, 31.93, 35.69, 38.19, 47.93, 55.21, 61.29, 61.80, 66.81, 67.55, 68.59, 69.44, 69.88, 71.24, 72.58, 72.95, 73.27, 74.50, 76.02, 79.59, 95.84, 97.70, 99.64, 100.64, 114.62, 115.30, 127.13, 127.31, 127.43, 127.51, 127.78, 128.12, 128.26, 128.32, 128.46, 138.48, 138.77, 138.98, 152.46, 154.07, 164.80 (C=O), 168.96 (C=O), 169.04 (C=O), 169.56 (C=O), 169.78 (C=O), 169.84 (C=O), 170.13 (C=O), 170.36 (C=O), 170.58 (C=O).

Anal. Calcd for C₁₀₃H₁₄₈N₂O₃₀ (1893.01): C, 65.31; H, 7.88; N, 1.48. Found: C, 65.04; H, 7.87; N, 1.43.

2-(Tetradecyl)hexadecyl (5-acetylamino-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosyl 1,5-lactam)-(2 \rightarrow 3)-(2,4,6-tri-*O*-acetyl- β -*D*-galactopyranosyl)-(1 \rightarrow 4)-[(2,3,4-tri-*O*-acetyl- α -*L*-fucopyranosyl)-(1 \rightarrow 3)]-2-acetamido-2-deoxy-6-*O*-4-methoxyphenyl- β -*D*-glucopyranoside (11). A solution of **10** (20.1 mg, 10.6 μ mol) in ethanol (2 mL) was vigorously stirred over Pd(OH)₂ (40 mg) for 24 hr at room temperature under hydrogen. The catalyst was collected and washed with methanol. The combined filtrate and washings were concentrated, and the residue was treated with acetic anhydride (2.5 mL) and pyridine (4 mL) for 12 hr at room temperature, then cooled to 0°C. Methanol (1.5 mL) was added and the mixture was concentrated, and the residue was extracted with chloroform and successively washed with cold 2 M

hydrochloric acid and water, dried (Na_2SO_4), and concentrated. Column chromatography (70:1 CHCl_3 -MeOH) of the residue on silica gel gave **10** (17.3 mg, 93%) as an amorphous mass; $[\alpha]_{\text{D}} -17.1^\circ$ (c 0.35, CHCl_3), ^1H NMR (CDCl_3) δ 0.88 (t, 6H, $J = 6.7$ Hz, 2CH_3), 1.09 (d, 3H, $J_{5,6} = 6.4$ Hz, H-6d), 1.24–1.52 (m, 53H, 26CH_2 and CH), 1.96, 1.98, 2.04, 2.06, 2.07, 2.11, 2.12, 2.13, 2.16, 2.17, 2.20 (11s, 33H, 10OAc, AcN of a), 2.24 (dd, 1H, $J_{3\beta,4} = 10.5$ Hz, $J_{\text{gem}} = 13.9$ Hz, H-3c β), 2.29 (dd, 1H, $J_{3\alpha,4} = 5.3$ Hz, $J_{\text{gem}} = 13.9$ Hz, H-3c α), 2.56 (s, 3H, AcN of c), 3.22 (dd, 1H, $J_{\text{gem}} = 9.4$ Hz, $J_{\text{vic}} = 6.2$ Hz, OCH_2C of alkyl part), 3.71 (dd, 1H, $J_{\text{gem}} = 9.4$ Hz, $J_{\text{vic}} = 5.3$ Hz, OCH_2C of alkyl part), 3.76 (s, 3H, *MeO*Ph), 3.93 (dd, 1H, $J_{2,3} = 10.3$ Hz, $J_{3,4} = 3.4$ Hz, H-3b), 4.18 (dd, 1H, $J_{8,9} = 6.2$ Hz, $J_{\text{gem}} = 11.8$ Hz, H-9c), 4.20 (d, 1H, H-6c), 4.30 (dd, 1H, $J_{8,9'} = 5.7$ Hz, $J_{\text{gem}} = 11.4$ Hz, H-9'c), 4.50 (d, 1H, $J_{1,2} = 8.2$ Hz, H-1b), 4.84 (m, 1H, H-4c), 5.02 (dd, 1H, H-2d), 5.07 (dd, 1H, $J_{1,2} = 8.2$ Hz, $J_{2,3} = 10.3$ Hz, H-2b), 5.18 (d, 1H, $J_{1,2} = 3.4$ Hz, H-1d), 5.24 (d, 1H, H-4b), 5.34–5.37 (m, 2H, H-5c and H-3d), 5.43 (m, 1H, H-8c), 5.69 (d, 1H, $J_{\text{NH},2} = 9.6$ Hz, NH), 5.74 (dd, 1H, $J_{6,7} = 3.7$ Hz, $J_{7,8} = 9.6$ Hz, H-7c), 6.85–6.93 (m, 4H, *MeO*Ph).

^{13}C NMR (CDCl_3) δ 14.14, 15.78, 20.63, 20.69, 20.77, 20.83, 20.99, 22.70, 23.31, 26.32, 26.67, 26.89, 29.37, 29.67, 29.72, 30.12, 30.19, 30.90, 31.14, 31.93, 35.51, 38.09, 47.92, 55.55, 61.33, 61.96, 64.46, 67.39, 67.65, 68.08, 68.60, 69.40, 69.98, 71.23, 71.43, 71.73, 72.16, 72.67, 73.54, 73.96, 74.47, 95.86, 96.04, 99.88, 100.85, 114.82, 115.97, 152.39, 154.31, 164.84 (C=O), 169.05 (C=O), 169.37 (C=O), 169.47 (C=O), 169.69 (C=O), 169.75 (C=O), 169.83 (C=O), 170.18 (C=O), 170.42 (C=O), 170.48 (C=O), 170.64 (C=O), 170.68 (C=O), 171.03 (C=O).

Anal. Calcd for $\text{C}_{88}\text{H}_{136}\text{N}_2\text{O}_{33}$ (1748.90): C, 60.40; H, 7.83; N, 1.60. Found: C, 60.10; H, 7.69; N, 1.46.

2-(Tetradecyl)hexadecyl (5-acetylamino-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosyl 1,5-lactam)-(2 \rightarrow 3)-(2,4,6-tri-*O*-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[(2,3,4-tri-*O*-acetyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2-acetamido-2-deoxy- β -D-glucopyranoside (12**).** To a solution of **11** (17.3 mg, 9.9 μmol) in acetonitrile (1.8 mL) and water (0.2 mL) was added CAN (16.8 mg, 0.03 mmol), and the mixture was stirred for 2.5 hr at 0°C and extracted with chloroform. The extract was successively washed with 1 M sodium carbonate and water, dried (Na_2SO_4), and concentrated. Column chromatography (100:1 CHCl_3 -MeOH) of the residue on silica gel gave **12** (12.1 mg, 74.6%) as an amorphous mass; $[\alpha]_{\text{D}} -8.2^\circ$ (c 0.24, CHCl_3); ^1H NMR (CDCl_3) δ 0.88 (t, 6H, $J = 6.6$ Hz, 2CH_3), 1.22–1.52 (m, 56H, 26CH_2 , CH and H-6d), 1.94, 1.95, 2.02, 2.05, 2.08, 2.09, 2.10, 2.14, 2.16, 2.18, 2.21 (11s, 33H, 10OAc, AcN of a), 2.27 (dd, 1H, $J_{3\beta,4} = 10.5$ Hz, $J_{\text{gem}} = 14.4$ Hz, H-3c β), 2.38 (dd, 1H, $J_{3\alpha,4} = 5.4$ Hz, $J_{\text{gem}} = 14.4$ Hz, H-3c α), 2.56 (s, 3H, AcN of c), 3.20 (dd, 1H, $J_{\text{gem}} = 9.3$ Hz, $J_{\text{vic}} = 6.6$ Hz, OCH_2C of alkyl part), 3.28 (br-d, 1H, H-6a), 3.73 (dd, 1H, OCH_2C of alkyl part), 3.85 (dd, 1H, H-2a), 4.13 (dd, 1H, $J_{2,3} = 9.8$ Hz, $J_{3,4} = 3.6$ Hz, H-3b), 4.18 (dd, 1H, $J_{8,9} = 5.5$ Hz, $J_{\text{gem}} = 11.6$ Hz, H-9c), 4.22 (br-d, 1H, H-6c), 4.26 (dd, 1H, $J_{8,9'} = 6.4$ Hz, $J_{\text{gem}} = 11.6$ Hz, H-9'c), 4.37 (d, 1H, $J_{1,2} = 7.6$ Hz, H-1a), 4.60 (d, 1H, $J_{1,2} = 8.2$ Hz, H-1b), 4.85 (m, 1H, H-4c), 4.97–5.00 (m, 2H, H-2d and H-5d), 5.05 (dd, 1H, $J_{1,2} = 8.2$ Hz, $J_{2,3} = 10.0$ Hz, H-2b), 5.21 (dd, 1H, $J_{2,3} = 10.9$ Hz, $J_{3,4} = 3.2$ Hz, H-3d), 5.29 (d, 1H, H-4b), 5.36 (m, 1H, H-5c), 5.43 (m, 2H, H-8c and H-1d), 5.76 (dd, 1H, $J_{6,7} = 3.8$ Hz, $J_{7,8} = 8$ Hz, H-7c).

^{13}C NMR (CDCl_3) δ 14.13, 15.77, 20.45, 20.62, 20.69, 20.70, 20.77, 20.83, 20.98, 22.70, 23.30, 26.63, 26.79, 26.89, 29.37, 29.68, 29.71, 30.13, 30.18, 30.91, 31.13,

31.92, 35.71, 39.12, 48.29, 61.58, 62.02, 64.52, 67.68, 68.28, 68.66, 69.47, 69.82, 71.28, 71.35, 71.46, 71.77, 72.18, 72.54, 72.62, 73.58, 73.62, 73.99, 74.39, 75.62, 96.88, 97.27, 99.48, 100.72, 169.21 (C=O), 168.37 (C=O), 169.12 (C=O), 169.58 (C=O), 169.84 (C=O), 170.12 (C=O), 170.27 (C=O), 170.42 (C=O), 170.51 (C=O), 170.55 (C=O), 170.63 (C=O), 170.77 (C=O), 171.05 (C=O).

Anal. Calcd for C₈₁H₁₃₀N₂O₃₂ (1642.86): C, 59.18; H, 7.97; N, 1.70. Found: C, 59.10; H, 7.85; N, 1.63.

2-(Tetradecyl)hexadecyl (5-acetylamino-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosyl 1,5-lactam)-(2 \rightarrow 3)-(2,4,6-tri-*O*-acetyl- β -*D*-galactopyranosyl)-(1 \rightarrow 4)-[(2,3,4-tri-*O*-acetyl- α -*L*-fucopyranosyl)-(1 \rightarrow 3)]-2-acetamido-2-deoxy-6-*O*-sulfo- β -*D*-glucopyranoside triethylammonium salt (13). To a solution of **12** (12.1 mg, 7.4 μ mol) in DMF (1 mL) was added sulfur trioxide pyridine complex (7.1 mg, 44 μ mol), and the mixture was stirred for 6 hr at room temperature. Triethylamine (0.1 mL) was added and the mixture was concentrated. Column chromatography (1 : 1 CHCl₃-MeOH) of the residue on Sephadex LH-20 gave the crude sulfated product, and this was purified by column chromatography (30:1 CHCl₃-MeOH) on silica gel to afford **13** (10.5 mg, 78%) as an amorphous mass; [α]_D -28.9° (*c* 0.04, CHCl₃); ¹H NMR (CDCl₃) δ 0.88 (t, 6H, *J* = 6.6 Hz, 2CH₃), 1.22 (d, 3H, *J*_{5,6} = 6.4 Hz, H-6d), 1.25–1.30 (m, 52H, 26CH₂), 1.40 (t, 9H, NCH₂CH₃), 1.45 (m, 1H, CH), 1.94, 1.95, 2.05, 2.06, 2.07, 2.091, 2.094, 2.14, 2.160, 2.165, 2.23 (11s, 33H, 10OAc, AcN of a), 2.29 (dd, 1H, *J*_{3 β ,4} = 10.8 Hz, *J*_{gem} = 14.3 Hz, H-3c β), 2.46 (dd, 1H, *J*_{3 α ,4} = 5.3 Hz, *J*_{gem} = 14.3 Hz, H-3c α), 2.56 (s, 3H, AcN of c), 3.16 (m, 7H, NCH₂CH₃ and OCH₂C of alkyl part), 3.68 (dd, 1H, *J*_{gem} = 9.6 Hz, *J*_{vic} = 5.5 Hz, OCH₂C of alkyl part), 3.96 (t, 1H, H-2a), 4.20 (br-d, 1H, H-6c), 4.24–4.28 (m, 3H, H-9c, H-9'c and H-3b), 4.85 (m, 2H, H-4c and H-5d), 4.89 (d, 1H, *J*_{1,2} = 8.2 Hz, H-1b), 5.02 (dd, 1H, *J*_{1,2} = 3.4 Hz, *J*_{2,3} = 10.5 Hz, H-2d), 5.04 (dd, 1H, *J*_{1,2} = 8.2 Hz, *J*_{2,3} = 10.1 Hz, H-2b), 5.24 (dd, 1H, *J*_{2,3} = 10.5 Hz, H-3d), 5.34 (m, 1H, H-8c), 5.37 (d, 1H, *J*_{1,2} = 3.2 Hz, H-1d), 5.43 (d, 1H, H-4b), 5.51 (d, 1H, *J*_{NH,2} = 8.5 Hz, NH), 5.77 (dd, 1H, *J*_{6,7} = 4.1 Hz, *J*_{7,8} = 8.6 Hz, H-7c).

Anal. Calcd for C₈₇H₁₄₅N₃O₃₅S (1823.94): C, 57.25; H, 8.01; N, 2.30. Found: C, 56.98; H, 7.72; N, 2.25.

2-(Tetradecyl)hexadecyl (5-amino-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosyl 1,5-lactam)-(2 \rightarrow 3)-(β -*D*-galactopyranosyl)-(1 \rightarrow 4)-[(α -*L*-fucopyranosyl)-(1 \rightarrow 3)]-2-acetamido-2-deoxy-6-*O*-sulfo- β -*D*-glucopyranoside sodium salt (14). To a solution of **13** (5.6 mg, 3.1 μ mol) in methanol (3 mL) and dioxane (1 mL) was added catalytic amount of 28% sodium methoxide in methanol, and the mixture was stirred for 30 hr at room temperature and then concentrated. Column chromatography (1 : 1 CHCl₃-MeOH) of the residue on Sephadex LH-20 gave the target molecule **14** (3.9 mg, 98%) as an amorphous mass; [α]_D +3.2° (*c* 0.07, 1 : 1 CHCl₃-MeOH); ¹H NMR (CD₃OD) δ 0.86 (t, 6H, *J* = 7.6 Hz, 2CH₃), 1.07 (d, 3H, *J*_{5,6} = 6.6 Hz, H-6d), 1.19–1.43 (m, 53H, 26CH₂ and CH), 1.84 (s, 3H, AcN of a), 2.01 (dd, 1H, *J*_{3 α ,4} = 5.0 Hz, *J*_{gem} = 13.6 Hz, H-3c α), 2.30 (dd, 1H, *J*_{3 β ,4} = 10.3 Hz, *J*_{gem} = 13.4 Hz, H-3c β), 3.43 (m, 1H, H-8c), 3.46 (m, 1H, H-5c), 3.48 (dd, 1H, *J*_{1,2} = 7.5 Hz, *J*_{2,3} = 9.6 Hz, H-2b), 3.54 (dd, 1H, H-2d), 3.58 (d, 1H, *J*_{1,2} = 8.0 Hz, H-1a), 3.61 (m, 2H, H-9c and H-4d), 3.64 (dd, 1H, *J*_{gem} = 14.4 Hz, H-9'c), 3.70 (t, 1H, *J*_{1,2} = 8.0 Hz, H-2a), 3.78 (dd, 1H, *J*_{2,3} = 10.0 Hz, *J*_{3,4} = 3.2 Hz, H-3d), 3.98 (m, 2H, H-3b and H-4c), 4.10 (br-d, 1H, H-4b), 4.37 (br-d, 1H, H-6c), 4.56 (d, 1H, *J*_{1,2} = 7.5 Hz, H-1b), 4.74 (m, 1H, H-5d), 4.92 (d, 1H, *J*_{1,2} = 4.1 Hz, H-1d).

MALDI-TOF MS (positive-ion mode, DHB, MeCN) Calcd for $C_{59}H_{107}N_2NaO_{24}S$ $[M + Na]^+$ 1282.6832. Found: 1282.5850.

ACKNOWLEDGMENTS

This work was partly supported by Japan Society for the Promotion of Science (Grant-in-Aid for Scientific Research No. 12306007 to M.K.) and CREST of Japan Science and Technology Corporation (JST). We thank Ms. Kiyoko Ito for the technical assistance.

REFERENCES

1. Yamaguchi, M.; Ishida, H.; Kanamori, A.; Kannagi, R.; Kiso, M. Studies on the endogenous L-selectin ligands: systematic and highly efficient total synthetic routes to lactamized-sialyl 6-*O*-sulfo Lewis X and other novel gangliosides containing lactamized neuraminic acid. *Carbohydr. Res.* **2003**, *338*, 2793–2812.
2. Springer, T.A. Traffic signals for lymphocyte recirculation and leukocyte emigration: the multi-step paradigm. *Cell.* **1994**, *76*, 301–304.
3. Galustian, C.; Lawson, A.M.; Komba, S.; Ishida, H.; Kiso, M.; Feizi, T. Sialyl-Lewis X sequence 6-*O*-sulfated at *N*-acetylglucosamine rather than at galactose is preferred ligand for L-selectin and de-*N*-acetylation of the sialic acid enhances the binding strength. *Biochem. Biophys. Res. Commun.* **1997**, *240*, 748–751.
4. Mitsuoka, C.; Sawada-Kasugai, M.; Ando-Furui, K.; Izawa, M.; Nakanishi, H.; Nakamura, S.; Ishida, H.; Kiso, M.; Kannagi, R. Identification of major carbohydrate capping group of the L-selectin ligand on high endothelial venules in human lymph nodes as 6-sulfo sialyl Lewis X. *J. Biol. Chem.* **1998**, *273*, 11225–11233.
5. Mitsuoka, C.; Ohmori, K.; Kimura, N.; Kanamori, A.; Komba, S.; Ishida, H.; Kiso, M.; Kannagi, R. Regulation of selectin binding activity by cyclization of sialic acid moiety of carbohydrate ligands on human leukocytes. *Proc. Natl Acad. Sci. USA* **1999**, *96*, 1597–1602.
6. Otsubo, N.; Ishida, H.; Kiso, M. Synthesis of novel ganglioside GM4 analogues containing *N*-deacetylated and lactamized sialic acid: probes for searching new ligand structures for human L-selectin. *Carbohydr. Res.* **2001**, *330*, 1–5.
7. Otsubo, N.; Yamaguchi, M.; Ishida, H.; Kiso, M. The first, efficient synthesis of novel sLe^x neoglycolipids containing *N*-deacetylated and lactamized sialic acid: key ligand structures for selectin binding. *J. Carbohydr. Chem.* **2001**, *20*, 329–334.
8. Komba, S.; Galustian, C.; Ishida, H.; Feizi, T.; Kannagi, R.; Kiso, M. First total synthesis of 6-sulfo de-*N*-acetyl-sialyl Lewis X ganglioside: a superior ligand for human L-selectin. *Angew. Chem. Int. Ed. Engl.* **1999**, *38*, 1131–1133.
9. Jansson, K.; Ahlfors, S.; Frejd, T.; Kihlberg, J.; Magnusson, G.; Dahmen, J.; Noori, G.; Stenvall, K. 2-(Trimethylsilyl)ethyl glycosides. Synthesis, anomeric deblocking, and transformation into 1,2-trans 1-*O*-acyl sugars. *J. Org. Chem.* **1988**, *53*, 5629–5647.
10. Gelhausen, M.; Besson, F.; Chierici, S.; Lafont, D.; Boullanger, P.; Roux, B. Lectin recognition of liposomes containing neoglycolipids. Influence of their lipidic anchor and spacer length. *Colloids Surfaces B: Biointerfaces* **1998**, *10*, 395–404.

11. Komba, S.; Ishida, H.; Kiso, M.; Hasegawa, A. Synthesis and biological activities of three sulfated sialyl Le^x ganglioside analogues for clarifying the real carbohydrate ligand structure of L-selectin. *Bioorg. Med. Chem.* **1996**, *4*, 1833–1847.
12. Mitsunobu, O. The use of diethyl azodicarboxylate and triphenylphosphine in synthesis and transformation of natural products. *Synthesis* **1981**, 1–28.
13. Fukuyama, T.; Lard, A.A.; Hotchkiss, L.A. *p*-Anisyl group: a versatile protecting group for primary alcohols. *Tetrahedron Lett.* **1985**, *26*, 6291–6292.
14. Fügedi, P.; Garegg, P.J. A novel promoter for the efficient construction of 1,2-trans linkages in glycoside synthesis, using thioglycosides as glycosyl donors. *Carbohydr. Res.* **1986**, *149*, c9–c12.
15. Kanie, O.; Kiso, M.; Hasegawa, A. Glycosylation using methylthioglycosides of *N*-acetylneuraminic acid and dimethyl (methylthio) sulfonium triflate. *J. Carbohydr. Chem.* **1988**, *7*, 501–506.
16. Knorr, R.; Trzeciak, A.; Bannwarth, W.; Gillessen, D. New coupling reagents in peptide chemistry. *Tetrahedron Lett.* **1989**, *30*, 1927–1930.

Received February 2, 2004

Accepted March 22, 2004