# 2-Aryl-dibenzo-1,2-oxaphosphorine as a Ligand in Borane and in Pt(II) Complexes\*

György Keglevich,<sup>1</sup> Helga Szelke,<sup>2</sup> Andrea Kerényi,<sup>1</sup> Tímea Imre,<sup>3</sup> Krisztina Ludányi,<sup>3</sup> József Dukai,<sup>4</sup> Ferenc Nagy,<sup>4</sup> and Péter Arányi<sup>4</sup>

<sup>1</sup>Department of Organic Chemical Technology, Budapest University of Technology and Economics, 1521 Budapest, Hungary

<sup>2</sup>*Research Group of the Hungarian Academy of Sciences at the Department of Organic Chemical Technology, Budapest University of Technology and Economics, 1521 Budapest, Hungary* 

<sup>3</sup>Hungarian Academy of Sciences, Chemical Research Center, 1525 Budapest, Hungary

<sup>4</sup>NIKE 2000, 8175 Balatonfüzfö, Hungary

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ABSTRACT: Optimum conditions for the synthesis of aryl-dibenzo-oxaphosphorines (2) from the corresponding phosphonous chloride (1) were explored to avoid the formation of by-products (e.g., **4–6**). The aryl-dibenzo-oxaphosphorines (2) were converted to the P-oxides (3), and were utilized in the synthesis of phosphinite-boranes **7** and Pt(II) complexes **8**. Depending on the aryl substituent, the Pt(II) complex (**8**) was formed with cis and trans geometry. © 2004 Wiley Periodicals, Inc. Heteroatom Chem 15:459–463, 2004; Published online in Wiley InterScience (www.interscience.wiley.com). DOI 10.1002/hc.20042

## INTRODUCTION

The P-ligands are widely applied in transition metal complexes that may be useful catalysts. The heterocyclic P-ligands including five- and six-membered

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P-cycles form a special class [1]. The dibenzo-1,2oxaphosphorines with phosphorous function represent a group that attracted much attention [2–4]. The P-aryl derivatives were not, however, studied in detail, mainly their oxides were described [5]. For this, we decided to examine the synthesis and complexing properties of aryl-dibenzo-1,2-oxaphosphorines.

The chloro derivative (1) available via the cyclization of ortho-hydroxybiphenyl by phosphorus trichloride [2] was first reacted with phenylmagnesium bromide in tetrahydrofuran at  $0 \rightarrow 26^{\circ}C$ under nitrogen. After the work-up procedure including hydrolysis, a mixture of phosphinite 2a (83%) and 2-diphenylphosphino-2'-hydroxybiphenyl 4 (17%) formed by ring opening with a second equivalent of Grignard reagent was obtained according to <sup>31</sup>P NMR and mass spectroscopic analysis of the crude mixture. The two components were obtained in a pure form as their oxides (3a and 5, respectively) after oxidation and purification by column chromatography (Scheme 1). A similar side reaction was also observed in the reaction of the P-oxide of the chlorodibenzooxaphosphorine with methylmagnesium bromide [5]. The substitution was extended to the preparation of triisopropylphenyl phosphinite **2b** (Scheme 1). The product (**2b**) was formed in a neat reaction, under the conditions of the hydrolysis

 $<sup>^{*}\</sup>mbox{IUPAC}$  name of 2-aryl-dibenzo-1,2-oxaphosphorine is 1,2-oxaphosphinine.

*Correspondence to:* György Keglevich; e-mail: gkeglevich@mail. bme.hu.

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Ar =  $C_6H_5$  (**a**), 2,4,6-tri <sup>*i*</sup>PrC<sub>6</sub>H<sub>2</sub> (**b**)

SCHEME 1





applying 0.15 M hydrochloric acid; the oxaphosphorine ring of **2b** was, however, opened up partially to afford 2-aryl-H-phosphinoxido-2'-biphenyl 6-2 (as the predominant component of a tautomeric equilibrium). According to <sup>31</sup>P NMR and MS, the crude mixture consisted of 80% of phosphinite (2b) and 20% of by-product 6-2. Performing the hydrolysis in the absence of hydrochloric acid, the ring opening side reaction could be avoided. It can be seen that the oxaphosphorine ring of the triisopropylphenyl compound (2b) is more sensitive under the conditions of acidic hydrolysis than that of the phenyl derivative (2a) is. At same time, the oxaphosphorine ring in 2 is more vulnerable toward PhMgBr than to tri<sup>*i*</sup>PrC<sub>6</sub>H<sub>2</sub>MgBr, as the by-product from double arylation was formed only in the first case (4).

To prepare a more stable derivative, phosphinite **2b** was converted to phosphinate **3b** by oxidation and was isolated in 66% after column chromatography (Scheme 1).

Phosphinites **2a** and **2b** and phosphinates **3a** and **3b** were characterized by <sup>31</sup>P and <sup>13</sup>C NMR, as well as mass spectroscopy, the P-oxides (**3a** and **3b**) also by <sup>1</sup>H NMR spectral data. By-products **4**, **5**, and **6-1** were identified by <sup>31</sup>P NMR and MS.

The P-ligands are usually protected against oxidation as borane complexes [6]. The arylphosphinites (**2a** and **2b**) were reacted with dimethylsulfide borane to give the phosphiniteboranes (**7a** and **7b**, respectively) that were characterized after purification by <sup>31</sup>P, <sup>11</sup>B, and <sup>13</sup>C NMR spectral parameters (Scheme 2). The phosphinite boranes (**7a** and **7b**) were of a ca. 95% purity.

Finally, arylphosphinites **2a** and **2b** were reacted with dichlorodibenzonitrile platinum in boiling benzene. On cooling, the products were precipitated and identified as complexes **8a** and **8b** (Scheme 2).

The <sup>31</sup>P NMR spectra of the complexes **8a** and **8b** revealed a signal at  $\delta$  76.8 with a  $J_{Pt-P}$  of 4240 Hz and at  $\delta$  84.6 with a  $J_{Pt-P}$  of 2900 Hz, respectively. Both



#### SCHEME 2

signals were accompanied by a pair of satellites resulting from the presence of the <sup>195</sup>Pt isotope. The *cis* and *trans* orientation of the ligands in complexes **8a** and **8b**, respectively, is in accord with the stereospecific  $J_{Pt-P}$  couplings detected. It is known that within a *cis* and *trans* diastereomer pair, the larger coupling belongs to the *cis* isomer, while the smaller one to the *trans* form [7]. The outcome of the complexation can be explained well with the role of steric factors; with the smaller phenyl group, the orientation of the ligands in the usual *cis*, while with the sterically more demanding tri*iso*propylphenyl substituent, the ligands are in the more favorable *trans* relationship. A similar situation was reported for the corresponding phospholes [7,8].

Complex **8a** was also characterized by <sup>1</sup>H NMR spectroscopic data. The solubility of complexes **8a** and **8b** in chloroform is very poor. Moreover, compound **8b** was found to be unstable in CHCl<sub>3</sub> (CDCl<sub>3</sub>) solution.

In summary, two arylphosphines together with the corresponding boranes and the two kinds of platinum complexes based on the dibenzooxaphosphorine skeleton were prepared and characterized.

### EXPERIMENTAL SECTION

The <sup>31</sup>P, <sup>11</sup>B, <sup>13</sup>C, and <sup>1</sup>H NMR spectra were taken on a Bruker DRX-500 spectrometer operating at 202.4, 160.4 125.7, and 500 MHz, respectively. Positive chemical shifts are downfield relative to 85%  $H_3PO_4$ ,  $F_3B \cdot OEt_2$  or TMS. The coupling constants are given in Hz. Mass spectrometry was performed on a ZAB-2SEQ instrument.

# *The Preparation of Phenyl-oxaphosphorines* **2a**, **3a**, *and* **7a**

To 1.0 g (4.26 mmol) of chlorooxaphosphorine **1** in 10 mL of tetrahydrofuran was added dropwise 5.54 mmol of phenylmagnesium bromide in 10 mL of diethyl ether (prepared from 0.86 g (5.54 mmol) of bromobenzene and 0.14 g (5.54 mmol) of magnesium in 10 mL of ether) at 0°C with stirring. After addition was complete, content of the flask was stirred at 26°C for 20 h under nitrogen. Solvent was evaporated and the residue taken up in the mixture of 20 mL of chloroform and 5 mL of water. The organic phase was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. 0.82 g crude product containing 99% of **2a** and 1% of **4** was obtained.

**2a:** Yield: 70%; <sup>31</sup>P NMR (CDCl<sub>3</sub>)  $\delta$  84.6; <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  120.8 (CH=), 123.1 (CH=), 124.0 (<sup>3</sup>*J* = 5.8, C<sub>1a</sub>)<sup>a</sup>, 124.1 (<sup>3</sup>*J* = 1.5, CH=), 125.0 (CH=), 127.5 (CH=), 128.1 (<sup>3</sup>*J* = 4.2, C<sub>3'</sub>), 129.4 (CH=), 129.6 (CH=), 130.7 (CH=), 130.9 (C<sub>2'</sub>), 131.5 (C<sub>4'</sub>), 131.9 (<sup>1</sup>*J* = 13.9, C<sub>6a</sub>)<sup>b</sup>, 133.5 (C<sub>10a</sub>)<sup>a</sup>, 138.9 (<sup>1</sup>*J* = 30.1, C<sub>1'</sub>)<sup>b</sup>, 151.3 (<sup>2</sup>*J* = 10.8, C<sub>4a</sub>), <sup>a,b</sup>may be reversed; FAB-MS, 277 (M+H); (M+H)<sup>+</sup><sub>found</sub> = 277.0782, C<sub>18</sub>H<sub>14</sub>OP requires 277.0755.

**4:** <sup>31</sup>P NMR (CDCl<sub>3</sub>) δ–11.8; FAB-MS, 355 (M+H).

The crude product was taken up in 20 mL of chloroform and was treated with 1.3 mL (12.8 mmol)

of 30% hydrogen peroxide at 0°C for 2 h on stirring. The organic phase was washed with  $3 \times 10$  mL of water and dried (Na<sub>2</sub>SO<sub>4</sub>), and finally concentrated. The main component was obtained by column chromatography (silica gel, 3% methanol in chloroform) to afford 0.69 g (80%) of **3a** as a white solid.

**3a:** m.p. 156–158°C (acetone); <sup>31</sup>P NMR (CDCl<sub>3</sub>)  $\delta$  25.0 (99%); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  120.4 (<sup>2</sup>J = 6.0, C<sub>7</sub>)<sup>a</sup>, 121.8 (<sup>3</sup>J = 11.1, C<sub>1a</sub>)<sup>b</sup>, 123.5 (<sup>3</sup>J = 9.6, C<sub>10</sub>)<sup>a</sup>, 124.5 (C<sub>3</sub>)<sup>c</sup>, 124.7 (<sup>1</sup>J = 128.1, C<sub>6a</sub>)<sup>d</sup>, 124.9 (C<sub>1</sub>)<sup>c</sup>, 128.1 (<sup>3</sup>J = 14.1, C<sub>4</sub>)<sup>a</sup>, 128.4 (<sup>3</sup>J = 14.2, C<sub>3'</sub>)<sup>e</sup>, 129.3 (<sup>1</sup>J = 143.6, C<sub>1'</sub>)<sup>d</sup>, 130.3 (C<sub>4'</sub>), 130.7 (<sup>3</sup>J = 12.2, C<sub>8</sub>)<sup>a</sup>, 131.9 (<sup>2</sup>J = 11.0, C<sub>2'</sub>)<sup>e</sup>, 132.8 (C<sub>9</sub>)<sup>c</sup>, 132.9 (C<sub>2</sub>)<sup>c</sup>, 135.5 (<sup>2</sup>J = 5.3, C<sub>10a</sub>)<sup>b</sup>, 148.9 (<sup>2</sup>J = 8.5, C<sub>4a</sub>), <sup>a-e</sup>tentative assignment; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  7.27–8.07 (m, 13H, Ar); FAB-MS, 293 (M+H); M<sup>+</sup><sub>found</sub> = 292.0601, C<sub>18</sub>H<sub>14</sub>O<sub>2</sub>P requires 292.0653.

**5:** <sup>31</sup>P NMR (CDCl<sub>3</sub>) δ 32.7 (1%); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 6.46–8.04 (m, 18H, Ar), 9.03 (s, 1H, OH); FAB-MS, 371 (M+H).

To the 20 mL dichloromethane solution of 4.26 mmol of phosphine 2a was added 2.6 mL (5.12 mmol) of 2 M tetrahydrofuran solution of dimethylsulfide borane at room temperature under nitrogen and the mixture was stirred for 24 h. Evaporation of the volatile components led to 7a as pale yellow solid. Yield: ~100%.

**7a:** <sup>31</sup>P NMR (CDCl<sub>3</sub>)  $\delta$  93.7 (broad); <sup>11</sup>B NMR (CDCl<sub>3</sub>)  $\delta$  -36.2 (broad); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  120.8 (<sup>2</sup>*J* = 4.5, C<sub>7</sub>)<sup>a</sup>, 123.1 (<sup>1</sup>*J* = 59.9, C<sub>1'</sub>)<sup>b</sup>, 123.4 (<sup>3</sup>*J* = 10.0, C<sub>1a</sub>)<sup>c</sup>, 124.3 (<sup>3</sup>*J* = 6.0, C<sub>10</sub>)<sup>a</sup>, 124.8 (C<sub>3</sub>)<sup>d</sup>, 125.4 (C<sub>1</sub>)<sup>d</sup>, 128.7 (<sup>3</sup>*J* = 10.0, C<sub>3'</sub>)<sup>e</sup>, 128.8 (<sup>3</sup>*J* = 13.3, C<sub>4</sub>)<sup>a</sup>, 129.8 (<sup>1</sup>*J* = 41.0, C<sub>6a</sub>)<sup>b</sup>, 130.7 (C<sub>4'</sub>), 131.3 (<sup>2</sup>*J* = 11.5, C<sub>2'</sub>)<sup>e</sup>, 131.7 (<sup>3</sup>*J* = 18.9, C<sub>8</sub>)<sup>a</sup>, 132.4 (C<sub>9</sub>)<sup>d</sup>, 133.3 (C<sub>2</sub>)<sup>d</sup>, 135.0 (C<sub>10a</sub>)<sup>c</sup>, 149.4 (<sup>2</sup>*J* = 12.6, C<sub>4a</sub>), <sup>a-e</sup>tentative assignment; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  7.06–7.96 (m, 13H, Ar); FAB-MS, 277 (M–BH<sub>3</sub>+H).

## *The Preparation of Triisopropylphenyl-Oxaphosphorines* **2b**, **3b**, *and* **7b**

**2b**, **3b**, and **7b** were prepared from **1** as described for the  $1 \rightarrow 2a \rightarrow 3a \rightarrow 7a$  transformation.

**2b:** Yield: 1.4 g (82%); <sup>31</sup>P NMR (CDCl<sub>3</sub>)  $\delta$  112.0; <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  23.3, 23.7, 26.0 (CH(*CH*<sub>3</sub>)<sub>2</sub>), 31.4 (C<sub>2'</sub>-*CH*Me<sub>2</sub>), 34.7 (C<sub>4'</sub>-*CH*Me<sub>2</sub>), 120.9 (CH=), 122.3 (CH=), 122.6 (<sup>3</sup>J = 3.6, C<sub>3'</sub>), 123.2 (CH=), 123.6 (CH=), 125.3 (CH=), 125.8 (<sup>3</sup>J = 9.7, C<sub>1a</sub>)<sup>a</sup>, 128.0 (CH=), 129.1 (CH=), 129.7 (<sup>1</sup>J = 31.3, C<sub>1'</sub>)<sup>b</sup>, 130.1 (CH=), 134.5 (<sup>2</sup>J = 12.6, C<sub>10a</sub>)<sup>a</sup>, 138.6 (<sup>1</sup>J = 26.4, C<sub>6a</sub>)<sup>b</sup>, 152.5 (C<sub>4a</sub>), 152.9 (C<sub>4'</sub>), 156.5 (C<sub>2'</sub>), <sup>a,b</sup>may be reversed; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.12 (d, <sup>3</sup>J<sub>HH</sub> = 6.5, 6H, CH(*CH*<sub>3</sub>)<sub>2</sub>), 1.26 (d, <sup>3</sup>J<sub>HH</sub> = 7.0, 6H, CH(*CH*<sub>3</sub>)<sub>2</sub>), 1.37 (d, <sup>3</sup>J<sub>HH</sub> = 7.0, 6H, CH(*CH*<sub>3</sub>)<sub>2</sub>), 2.97 (sept, <sup>3</sup>J<sub>HH</sub> = 7.0, 1H,  $C_{4'}$ -*CH*Me<sub>2</sub>), 3.99 (m, 2H,  $C_{2'}$ -*CH*Me<sub>2</sub>), 6.85-8.15 (m, 10H, Ar); FAB-MS, 403 (M+H); (M + H)<sup>+</sup><sub>found</sub> = 403.2124,  $C_{27}H_{31}$ OP requires 403.2191.

**3b:** Yield: 1.11 g (76%); could be separated as a crystalline compound, m.p. 155–157°C (acetone); <sup>31</sup>P NMR (CDCl<sub>3</sub>) δ 25.6; <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 23.8, 24.4, 25.3 (CH( $CH_3$ )<sub>2</sub>), 30.4 (<sup>3</sup>J = 3.6, C<sub>2</sub>– $CHMe_2$ ), 34.6  $(C_{4'}-CHMe_2)$ , 118.7 (<sup>1</sup>J = 140.7, C')<sup>c</sup>, 120.9 (<sup>2</sup>J = 5.9,  $C_7$ )<sup>a</sup>, 121.7 (<sup>3</sup>J = 11.3,  $C_{1a}$ )<sup>d</sup>, 123.0 (<sup>3</sup>J = 13.4,  $C_{3'}$ ), 123.7 (<sup>3</sup>J = 8.9,  $C_{10}$ )<sup>a</sup>, 124.2 ( $C_{3}$ )<sup>b</sup>, 125.0 ( $C_{1}$ )<sup>b</sup> 127.0 ( ${}^{3}J = 14.6, C_{4}$ )<sup>a</sup>, 129.1 ( ${}^{1}J = 125.3, C_{6a}$ )<sup>c</sup>, 129.4  $({}^{3}J = 13.9, C_{8})^{a}, 130.2 (C_{2})^{b}, 132.0 (C_{9})^{b}, 133.8 ({}^{2}J =$ 4.3,  $C_{10a}$ )<sup>d</sup>, 148.4 (<sup>2</sup>J = 8.2,  $C_{4a}$ ), 153.9 ( $C_{4'}$ ), 156.5  $(^{2}J = 13.3, C_{2'})$ , <sup>a-d</sup>tentative assignment; <sup>1</sup>H NMR  $(\text{CDCl}_3) \delta 1.00 \text{ (d, } {}^3J_{\text{HH}} = 7.0, 6\text{H}, \text{CH}(CH_3)_2), 1.22 \text{ (d,}$  ${}^{3}J_{\rm HH} = 6.5, 6\text{H}, CH(CH_{3})_{2}), 1.31 (d, {}^{3}J_{\rm HH} = 7.0, 6\text{H},$ CH( $CH_3$ )<sub>2</sub>), 2.96 (sept,  ${}^{3}J_{HH} = 7.0$ , 1H, C<sub>4'</sub>-CHMe<sub>2</sub>), 3.89 (m, 2H, C<sub>2'</sub>-CHMe<sub>2</sub>), 7.20-8.07 (m, 10H, Ar); FAB-MS, 419 (M+H);  $M_{found}^+ = 418.1982$ ,  $C_{27}H_{31}O_2P$ requires 418.2062.

**6-2:** <sup>31</sup>P NMR (CDCl<sub>3</sub>) δ 15.3; FAB-MS, 420 (M+H)

**7b:** Yield: ~100%; <sup>31</sup>P NMR (CDCl<sub>3</sub>)  $\delta$  101.9 (broad); <sup>11</sup>B NMR (CDCl<sub>3</sub>)  $\delta$  -30.0 (broad); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ ; 23.9, 24.6, 25.7 (CH(*CH*<sub>3</sub>)<sub>2</sub>), 31.0 (<sup>3</sup>*J* = 6.3, C<sub>2'</sub>-*CH*Me<sub>2</sub>), 34.5 (C<sub>4'</sub>-*CH*Me<sub>2</sub>), 120.9 (<sup>2</sup>*J* = 5.4, C<sub>7</sub>)<sup>a</sup>, 121.5 (<sup>1</sup>*J* = 50.5, C<sub>1'</sub>)<sup>b</sup>, 123.2 (<sup>3</sup>*J* = 9.8, C<sub>1a</sub>)<sup>c</sup>, 123.4 (<sup>3</sup>*J* = 8.8, C<sub>3'</sub>), 124.0 (<sup>3</sup>*J* = 6.4, C<sub>10</sub>)<sup>a</sup>, 124.2 (C<sub>3</sub>)<sup>d</sup>, 125.1 (C<sub>1</sub>)<sup>d</sup>, 128.2 (<sup>3</sup>*J* = 11.7, C<sub>4</sub>)<sup>a</sup>, 128.6 (<sup>1</sup>*J* = 55.5, C<sub>6a</sub>)<sup>b</sup>, 130.0 (<sup>3</sup>*J* = 14.7, C<sub>8</sub>)<sup>a</sup>, 130.6 (C<sub>2</sub>)<sup>d</sup>, 131.8 (C<sub>9</sub>)<sup>d</sup>, 134.2 (C<sub>10a</sub>)<sup>c</sup>, 149.2 (<sup>2</sup>*J* = 9.1, C<sub>4a</sub>), 153.7 (C<sub>4'</sub>), 156.1 (<sup>2</sup>*J* = 12.0, C<sub>2'</sub>), <sup>a-d</sup>tentative assignment; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.02 (d, <sup>3</sup>*J*<sub>HH</sub> = 6.5, 6H, CH(*CH*<sub>3</sub>)<sub>2</sub>), 1.10 (d, <sup>3</sup>*J*<sub>HH</sub> = 7, 6H, CH(*CH*<sub>3</sub>)<sub>2</sub>), 1.23 (d, <sup>3</sup>*J*<sub>HH</sub> = 7.0, 6H, CH(*CH*<sub>3</sub>)<sub>2</sub>), 2.87 (m, 1H, C<sub>4'</sub>-*CH*Me<sub>2</sub>), 3.82 (m, 2H, C<sub>2'</sub>-*CH*Me<sub>2</sub>), 7.08-7.85 (m, 10H, Ar); FAB-MS, 417 (M+H), 403 (M-BH<sub>3</sub>+H).

### The Preparation of Pt-Complexes 8a and 8b

To 0.21 mmol of phosphinite 2 in 20 mL of benzene, 0.10 g (0.21 mmol) of dichlorodibenzonitrileplatinum was added and the mixture was stirred at reflux for 1.5 h under nitrogen. Fractional crystallization from the benzene solution furnished 8 as pale yellow powder-like crystal.

**8a:** Yield: 60 mg (44%); in a purity of ca. 90%; <sup>31</sup>P NMR (CDCl<sub>3</sub>)  $\delta$  76.8 ( $J_{Pt-P} = 4239$ ); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  6.11–7.97 (m, 18H, Ar).

**8b:** Yield: 50 mg (31%); in a purity of ca. 75%; <sup>31</sup>P NMR (CDCl<sub>3</sub>)  $\delta$  84.6 ( $J_{Pt-P} = 2900$ ); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.90 (d, <sup>3</sup> $J_{HH} = 6.5$ , 6H, CH(*CH*<sub>3</sub>)<sub>2</sub>), 1.02 (d, <sup>3</sup> $J_{HH} = 6.0$ , 6H, CH(*CH*<sub>3</sub>)<sub>2</sub>), 1.15 (d, <sup>3</sup> $J_{HH} = 6.5$ , 6H, CH(*CH*<sub>3</sub>)<sub>2</sub>), 2.77 (sept, <sup>3</sup> $J_{HH} = 7.0$ , 1H, C<sub>4</sub>'–*CH*Me<sub>2</sub>), 4.28 (m, 2H, C<sub>2'</sub>–*CH*Me<sub>2</sub>), 6.93–8.03 (m, 15H, Ar).

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