Tetrahedron: Asymmetry 22 (2011) 367-372

Contents lists available at ScienceDirect

Tetrahedron: Asymmetry

journal homepage: www.elsevier.com/locate/tetasy





First asymmetric synthesis of the oxylipin, (6S,9R,10S)-6,9,10-trihydroxyoctadeca-7*E*-enoic acid

Sucheta Chatterjee, Seema V. Kanojia, Subrata Chattopadhyay, Anubha Sharma*

Bio-Organic Division, Bhabha Atomic Research Centre, Mumbai 400085, India

ARTICLE INFO

Article history: Received 10 January 2011 Accepted 10 February 2011 Available online 14 March 2011

ABSTRACT

A brief and facile synthesis of the title compound has been developed using cyclohexylideneglyceraldehyde as a chiral template. The key steps in the synthesis were: (i) two highly diastereoselective organometallic addition reactions to the aldehyde to furnish the required synthons with the appropriate stereogenic centers, and (ii) their cross metathesis to give the *E*-olefin geometry of the target compound. © 2011 Elsevier Ltd. All rights reserved.

1. Introduction

Araceae is one of the most dominant tropical families amongst herbs and vines, and many of the species are used as traditional remedies or food. The plant, Dracontium loretense Engl., which belongs to this family (subfamily Lasioideae), is widely distributed in the Peruvian Amazon, where it is known as 'jergón sacha'. There are various preparations of jergón sacha such as dried powder, tincture, alcoholic, and its aqueous-alcoholic extracts, which are popular amongst current Peruvian herbal medicines. The infusion of its corms is considered an immune-booster in Peruvian folk medicine. The combination of the extracts of D. loretense and Uncaria tomentosa is also used by AIDS patients to reinforce the immune system.^{1a-c} Very recently, four novel oxylipins 1a-d (Fig. 1) were isolated from the n-butanol extract of D. loretense corms and their structures were elucidated by extensive spectrocopy.² The absolute configurations of the three stereogenic carbinol centers of 1d remain unresolved, although the relative stereochemistry of the C-9 and C-10 centers was ascertained by NMR spectrometry. The *n*-butanol extract and some of its fractions $(10 \,\mu\text{g/mL})$, as well as compound **1d** $(10 \,\mu\text{M})$, showed potential immunostimulatory effects as revealed by a ³H-thymidine incorporation assay with human peripheral blood mononuclear cells (PBMCs). However, at a higher concentration, these were toxic to the cells. Amongst the constituent oxylipins of D. loretense, only compound 1d exhibited proliferation activity on the OKT3activated PBMCs, while its C-10 epimer 1c was inactive.² This emphasized the crucial role of the C-10 stereochemistry for biological activity.

The primary motivation of the present work stems from our own interest in aliphatic polyhydroxy acids as immunomodulatory, anti-inflammatory, and anti-neoplastic agents.³ In particular,

* Corresponding author. *E-mail address:* anubhas@barc.gov.in (A. Sharma). the reported immunomodulatory activity of **1d** was very attractive because many such agents show impressive anti-ulcer, antiinflammatory, and anti-neoplastic properties. Similar polyhydroxy acids and the macrolides derived from them are of wide occurrence in various natural sources and show diverse biological properties.^{4a-c} These factors and the lack of any synthesis of **1d** prompted us to develop the first asymmetric synthesis of the (6*S*,9*R*,10*S*)-stereomer of compound **1d**.

2. Results and discussion

For the past several years, we have been involved in devising simple and efficient asymmetric syntheses of various bioactive target compounds. To this end, we have found the inexpensive and easily available aldehyde 2 to be a versatile chiral template for the synthesis of a diverse array of natural compounds.⁵ The aldehyde 2 is amenable to various diastereoselective transformations using commonly available, inexpensive reagents.⁶ Herein, a convergent strategy toward the synthesis of the C₁₈ fatty acid 1d was conceived using a cross metathesis reaction between the chiral building blocks, 'A' and 'B' that would also provide the desired 7E-olefin function of the target oxylipin (Fig. 1). Different versions of the olefin metathesis reaction have turned out to be promising strategies for the construction of C-C bonds.⁷ We have also successfully utilized the cross metathesis of terminal alkenes to formulate efficient syntheses of three hydroxy fatty acids.^{8a-c} It was envisaged that the building blocks, 'A' and 'B' can be constructed by carrying out diastereoselective additions of alkyl/allyl organometallic reagents to aldehyde 2. This would provide the required stereogenic diol/carbinol moieties of the 'A' and 'B' units, while the cyclohexylidenedioxy group can be suitably functionalized to provide the terminal alkene functionality.

Very recently, we have found that the Ga-mediated crotylation of aldehyde **2** at room temperature in the ionic liquid, [bmim][Br] proceeded with excellent diastereoselectivity.⁹ For the synthesis

^{0957-4166/\$ -} see front matter \odot 2011 Elsevier Ltd. All rights reserved. doi:10.1016/j.tetasy.2011.02.008





(Scheme 1) of the 'A' unit, a similar protocol was employed using allyl bromide. This furnished the homoallylic alcohol 3 almost exclusively (anti/syn = 95:5). The pure anti-isomer was easily separated by column chromatography. This was silylated with tert-butyldiphenylsilyl chloride (TBDPSCl) in the presence of 4-dimethylaminopyridine (DMAP) to furnish 4. The alkene function of 4 was regioselectively hydroborated with BH₃-Me₂S (BMS) and the resultant product oxidized with H₂O₂ to afford alcohol **5**. This upon oxidation with pyridinium chlorochromate (PCC) afforded the aldehyde 6. Its base-catalyzed Horner-Emmons reaction with triethyl phosphonoacetate produced the conjugated ester 7. Although inconsequential to the present synthesis, the 2E-geometry of the ester 7 can be easily confirmed from the ¹H NMR spectrum. Catalytic hydrogenation of **7** gave the ester **8**, which upon treatment with aqueous trifluoroacetic acid (TFA) furnished the diol ester 9. Its reaction with excess *p*-toluenesulphonyl chloride (*p*-TsCl) in the presence of pyridine furnished the ditosylate, which on heating with NaI and Zn-dust in DMF afforded the alkene ester 10 in an unusually clean manner.

As per our synthetic plan, the synthesis of the other building block 15 (B equivalent) required two key steps. For the synthesis of the chiral diol segment of compound 15, we resorted to our own methodology of alkyl lithium addition to aldehyde 2.^{5d} The cyclohexylidenedioxy moiety of the resultant product was then converted to the one-carbon homologated allylic alcohol using sulfur-ylide chemistry. Despite its tremendous potential, the latter reaction has not been utilized extensively in organic synthesis. Thus, aldehyde 2 was reacted with CH₃(CH₂)₇Li to furnish the anti-triol derivative 11 almost exclusively (dr = 95:5). In contrast, using the corresponding Grignard reagent produced a 29:71 mixture of *syn/anti* isomers. The *anti*-compound **11**¹⁰ could be easily isolated as a pure enantiomer by column chromatography. The anti-stereochemistry of **11** was confirmed from the ¹H NMR resonances of the carbinol protons that appeared at δ 3.70–3.78 (1H) and δ 3.86–3.97 (m, 3H). For the corresponding syn-isomer, these appear as 1:1:2 multiplets at δ 3.46–3.48, δ 3.68–3.72 and δ 3.94–4.02.¹⁰ Silvlation of alcohol **11** with TBDPSCI/DMAP in CH₂Cl₂ produced compound 12, which upon treatment with aqueous TFA furnished diol 13. Its reaction with p-TsCl/pyridine proceeded regioselectively at the primary carbinol function to furnish the corresponding monotosylate. This was converted to epoxide 14 by

treatment with K_2CO_3 in MeOH. The reaction of the sulphorane, generated by the base (*n*-BuLi)-catalyzed deprotonation of trimethylsulphonium iodide (Me₃SI) with epoxide **14** proceeded regioselectively at the C-1 position to afford allylic alcohol **15**.

In the final sequence of reactions, alcohol **15** was subjected to a cross-metathesis reaction with ester **10** in the presence of Grubbs' 2nd generation catalyst to furnish the desired alcohol **16** (63%, based on **10**) along with the unreacted substrates **10** and **15**. The homo-dimerized product of **10** was also obtained in 10–12% yields. We used alcohol **15** in excess, as its dimerized product would produce a highly polar product, which can be easily separated from alcohol **16**. Generally, increasing the steric bulk via the addition of a hydroxyl protection reduces the cross-metathesis reactivity of the alkenols.^{7f} Hence we used directly the unprotected alcohol **15** for the metathesis. Desilylation of the compound **16** with Bu₄NF afforded the ester **17**, which on alkaline hydrolysis furnished the title acid **1d**.

3. Experimental

3.1. General experimental details

Chemicals (Fluka and Lancaster) were used as received. Other reagents were of AR grade. All anhydrous reactions were carried out under an Ar atmosphere, using freshly dried solvents. The organic extracts were dried over anhydrous Na₂SO₄. The IR spectra as thin films were scanned with a Jasco model A-202 FT-IR spectrometer. The ¹H NMR (200 MHz) and ¹³C NMR (50 MHz) spectra were recorded with a Bruker AC-200 spectrometer in CDCl₃. The optical rotations were recorded with a Jasco DIP 360 digital polarimeter.

3.1.1. (2*R*,3*S*)-3-*tert*-Butyldiphenylsilyloxy-1,2-cyclohexylidenedioxyhex-5-ene 4

To a stirred and cooled $(-30 \,^{\circ}\text{C})$ solution of the mixture of **3** (1.48 g, 6.95 mmol), imidazole (0.80 g, 11.80 mmol) and DMAP (catalytic) in CH₂Cl₂ (25 mL) was dropwise added TBDPSCl (2.86 g, 10.4 mmol) in CH₂Cl₂ (10 mL). After stirring the mixture for 10 h at room temperature, it was poured into ice-cold water (25 mL). The organic layer was separated and the aqueous portion



Scheme 1. Reagents and conditions: (i) Allyl bromide/Ga/[bmim][Br]/4 h; (ii) TBDPSCl/imidazole/DMAP/CH₂Cl₂/25 °C/10 h; (iii) BMS/THF/0 °C/3 h; aqueous NaOH/H₂O₂/0-25 °C/15 h; (iv) PCC/NaOAc/CH₂Cl₂/0 °C/3 h; (v) NaH/THF/(EtO)₂P(O)CH₂CO₂Et/0-25 °C/18 h; (vi) H₂/10% Pd-C/EtOH/25 °C/22 h; (vii) aqueous 80% TFA/CH₂Cl₂/0 °C/3 h/25 °C/18 h; (viii) PTSCl/pyridine/0-25 °C/24 h (89%); Zn/DMF/80 °C/4 h; (ix) CH₃(CH₂)₇Li/THF/-78 °C to 25 °C/3 h; (x) PTSCl/pyridine/0 °C/18 h; K₂CO₃/MeOH/25 °C/3 h; (xi) Me₃Sl/BuLi/THF/-40 °C/4 h/25 °C/12 h; (xiii) **10**/Grubbs' 2nd generation catalyst (5 mol %)/CH₂Cl₂/25 °C/22 h; (xii) Bu₄NF/THF/0 °C/3 h; (xiii) aqueous-ethanolic KOH/25 °C/4 h.

extracted with CHCl₃ (2 × 15 mL). The combined organic extracts were washed with water (2 × 10 mL) and brine (1 × 5 mL), and dried. Removal of solvent in vacuo followed by purification of the residue by column chromatography (silica gel, 0–5% EtOAc/hexane) afforded pure **4**. Yield: 2.75 g (88%); colorless oil; $[\alpha]_D^{24} = +16.8$ (*c* 1.13, CHCl₃); IR: 3070, 1639 cm⁻¹; ¹H NMR: δ 1.07 (s, 9H), 1.38–1.44 (m, 2H), 1.50–1.64 (m, 8H), 2.12–2.21 (m, 2H), 3.68–3.79 (m, 1H), 3.89–3.96 (m, 2H), 4.04–4.09 (m, 1H), 4.87–4.99 (m, 2H), 5.74–5.78 (m, 1H), 7.36–7.44 (m, 6H), 7.67–7.76 (m, 4H); ¹³C NMR: δ 19.3, 23.8, 23.9, 25.2, 26.9, 34.8, 36.1, 38.7, 65.8, 73.1, 77.3, 109.2, 117.4, 127.4, 127.5, 129.6, 129.7, 133.5, 133.8, 133.9, 135.9. Anal. Calcd for C₂₈H₃₈O₃Si: C, 74.62; H, 8.50. Found: C, 74.48; H, 8.36.

3.1.2. (4*S*,5*R*)-4-*tert*-Butyldiphenylsilyloxy-5,6-cyclohexylidenedioxyhexan-1-ol 5

To a stirred and cooled (0 °C) solution of **4** (3.52 g, 7.82 mmol) in THF (20 mL) was added BMS (0.52 mL, 5.2 mmol), and the mixture stirred for 3 h at the same temperature. Aqueous 3 M NaOH (3.13 mL) was then added, followed by H_2O_2 (30%, 3.13 mL) at 0 °C. After stirring for 3 h at 0 °C and 12 h at room temperature, the mixture was extracted with EtOAc (3×15 mL). The combined organic extracts were washed with H_2O (2 \times 10 mL), aqueous 10% HCl (1 \times 10 mL), H₂O (2 \times 10 mL) and brine (1 \times 5 mL), and dried. Solvent removal followed by column chromatography (silica gel, 0-15% EtOAc/hexane) of the residue furnished 5. Yield: 3.39 g (93%); colorless oil; $[\alpha]_D^{25} = +3.4$ (*c* 1.13, CHCl₃); IR: 3433 cm⁻¹; ¹H NMR: δ 1.04 (s, 9H), 1.32–1.39 (m, 2H), 1.41–1.52 (m, 13H), 3.39 (t, J = 6.2 Hz, 2H), 3.64–3.72 (m, 1H), 3.78–3.83 (m, 1H), 3.92-4.13 (m, 2H) 7.32-7.47 (m, 6H), 7.64-7.77 (m, 4H); ¹³C NMR: *δ* 19.4, 23.8, 23.9, 25.1, 26.9, 27.2, 30.2, 34.8, 36.1, 60.4, 62.7, 66.8, 73.6, 109.5, 127.5, 129.7, 133.5, 133.9, 135.8, 135.9. Anal. Calcd for C₂₈H₄₀O₄Si: C, 71.75; H, 8.60. Found: C, 71.64; H, 8.78.

3.1.3. (4S,5R)-4-tert-Butyldiphenylsilyloxy-5,6-cyclohexylidenedioxyhexanal 6

To a cooled (0 °C) and stirred suspension of PCC (1.90 g, 8.82 mmol) and NaOAc (10 mol %) in CH₂Cl₂ (20 mL) was added alcohol **5** (2.70 g, 5.77 mmol) in one portion. After stirring for 3 h, the reaction mixture was diluted with Et₂O (30 mL) and the supernatant passed through a pad of silica gel (2" × 1"). Removal of solvent in vacuo followed by column chromatography of the residue (silica gel, 0–10% EtOAc/hexane) furnished pure **6**. Yield: 2.19 g (81%); colorless oil; $[\alpha]_D^{24} = +7.7$ (*c* 1.24, CHCl₃); IR: 2714, 1729 cm⁻¹; ¹H NMR: δ 1.04 (s, 9H), 1.32–1.36 (m, 2H), 1.45–1.52 (m, 8H), 1.81–1.88 (m, 2H), 2.25–2.54 (m, 2H), 3.48–3.62 (m, 1H), 3.67–3.77 (m, 1H), 3.88–4.00 (m, 2H), 7.34–7.43 (m, 6H), 7.62–7.68 (m, 4H), 9.14 (t, *J* = 1.2 Hz, 1H); ¹³C NMR: δ 19.4, 23.8, 25.1, 26.3, 26.9, 34.7, 36.1, 38.8, 67.2, 73.3, 77.6, 109.7, 127.7, 129.8, 133.3, 133.5, 135.9, 202.2. Anal. Calcd for C₂₈H₃₈O₄Si: C, 72.06; H, 8.21. Found: C, 72.21; H, 8.18.

3.1.4. Ethyl (6S,7R)-6-*tert*-butyldiphenylsilyloxy-7,8-cyclohexylidenedioxyoct-2*E*-enoate 7

To a stirred suspension of pentane-washed NaH (0.444 g, 9.24 mmol, 50% suspension in oil) in THF (20 mL) was added triethyl phosphonoacetate (1.85 mL, 9.24 mmol) in THF (5 mL). After 15 min, when the solution became clear, the mixture was cooled to 0 °C, and the aldehyde **6** (2.15 g, 4.61 mmol) in THF (5 mL) was dropwise added into it. After stirring at room temperature for 18 h, the mixture was poured into ice-water and extracted with Et₂O (3 × 20 mL). The ether layer was washed with H₂O (2 × 10 mL) and brine (1 × 5 mL), dried, and concentrated in vacuo to give a residue, which upon column chromatography (silica gel, 0–10% Et₂O/hexane) furnished pure **7**. Yield: 2.25 g (91%); colorless oil; $[\alpha]_D^{24} = +12.7$ (*c* 1.07, CHCl₃); IR: 1716, 997 cm⁻¹; ¹H NMR: δ 1.04 (s, 9H), 1.22 (t, *J* = 7.2 Hz, 3H), 1.33–1.37 (m, 2H), 1.45–1.59 (m, 8H), 1.65–1.76 (m, 2H), 2.12–2.21 (m, 2H),

3.58–3.66 (m, 1H), 3.70–3.76 (m, 1H), 3.90–4.05 (m, 2H), 4.14 (q, J = 7.2 Hz, 2H), 5.66 (d, J = 16.0 Hz, 1H), 6.71–6.79 (m, 1H), 7.32–7.43 (m, 6H), 7.62–7.68 (m, 4H); ¹³C NMR: δ 14.2, 19.3, 23.7, 23.8, 25.0, 26.9, 32.1, 34.7, 36.0, 59.9, 67.0, 73.5, 77.4, 109.5, 121.1, 127.5, 129.7, 133.3, 133.6, 135.7, 135.8, 148.7, 166.5. Anal. Calcd for C₃₂H₄₄O₅Si: C, 71.60; H, 8.26. Found: C, 71.44; H, 8.48.

3.1.5. Ethyl (6S,7R)-6-*tert*-butyldiphenylsilyloxy-7,8-cyclohexylidenedioxyoctanoate 8

A mixture of **7** (1.3 g, 2.42 mmol) and 10% Pd/C (0.05 g) in EtOH (10 mL) was magnetically stirred at 25 °C for 22 h under a positive pressure of H₂. The mixture was diluted with Et₂O (30 mL) and passed through a small pad of silica gel. Removal of solvent in vacuo furnished pure **8**. Yield: 1.25 g (~quant.); colorless oil; $[\alpha]_D^{24} = +7.7$ (*c* 1.06, CHCl₃); IR: 1735 cm⁻¹; ¹H NMR: δ 1.04 (s, 9H), 1.23 (t, *J* = 7.2 Hz, 3H), 1.20–1.48 (m, 8H), 1.50–1.60 (m, 8H), 2.07–2.15 (m, 2H), 3.63–3.78 (m, 2H), 3.89–3.97 (m, 2H), 4.08 (q, *J* = 7.2 Hz, 2H), 7.32–7.42 (m, 6H), 7.63–7.70 (m, 4H); ¹³C NMR: δ 14.1, 19.3, 23.6, 23.7, 23.8, 24.8, 25.1, 26.9, 33.6, 34.1, 34.7, 36.0, 60.0, 66.5, 73.5, 77.5, 109.3, 127.5, 129.6, 133.5, 134.0, 135.8, 173.5. Anal. Calcd for C₃₂H₄₆O₅Si: C, 71.33; H, 8.61. Found: C, 71.34; H, 8.47.

3.1.6. Ethyl (6*S*,7*R*)-6-*tert*-butyldiphenylsilyloxy-7,8-dihydroxy-octanoate 9

A mixture of 8 (1.20 g, 2.23 mmol) in CH₂Cl₂ (15 mL) and aqueous 80% TFA (8 mL) was stirred at 0 °C for 3 h and then at 25 °C until completion of the reaction (TLC, 24 h). Most of the solvent was removed in vacuo. The residue was diluted with H₂O (30 mL) and extracted with EtOAc (3 \times 20 mL). The combined organic extracts were washed successively with H_2O (3 × 10 mL), aqueous 10% NaHCO₃ (2 \times 10 mL), H₂O (2 \times 10 mL) and brine (1 \times 5 mL), and dried. Solvent removal followed by column chromatography (silica gel, 0–5% MeOH/CHCl₃) of the residue gave pure 9. Yield: 0.830 g (81%); colorless oil; $[\alpha]_D^{25} = +26.5$ (*c* 1.08, CHCl₃); IR: 3455, 1731 cm⁻¹; ¹H NMR: δ 1.06 (s, 9H), 1.23 (t, *J* = 7.2 Hz, 3H), 1.32– 1.43 (m, 6H), 2.04-2.12 (m, 2H), 2.43 (br s, 2H), 3.63-3.72 (m. 3H), 3.80-3.84 (m, 1H), 4.07 (q, J = 7.2 Hz, 2H), 7.36-7.47 (m, 6H), 7.64–7.70 (m, 4H); ¹³C NMR: δ 14.2, 19.4, 24.7, 27.0, 29.7, 32.5, 33.9, 60.2, 63.1, 73.4, 75.1, 127.6, 127.8, 129.9, 133.0, 133.5, 135.8, 173.6. Anal. Calcd for C₂₆H₃₈O₅Si: C, 68.08; H, 8.35. Found: C, 67.86; H, 8.53.

3.1.7. Ethyl (6S)-6-tert-butyldiphenylsilyloxyoct-7-enoate 10

To a cooled (0 °C) and stirred solution of **9** (0.820 g, 1.79 mmol) in pyridine (8 mL) was added p-TsCl (0.750 g, 3.94 mmol). The reaction mixture was brought to room temperature and stirred for 24 h. It was then poured onto ice-cold water (30 mL) and extracted with EtOAc (2×15 mL). The combined organic extracts were washed successively with H_2O (2 × 10 mL), aqueous 2 M HCl (2 \times 10 mL), H₂O (2 \times 10 mL) and brine (1 \times 5 mL), and dried. Solvent removal followed by column chromatography (silica gel, 0-10% EtOAc/hexane) of the residue gave the pure ditosylate as a mixture of rotamers. Yield: 1.22 g (89%); colorless oil; $[\alpha]_{D}^{24} = +20.9$ (c 1.03, CHCl₃); IR: 1370, 1177 cm⁻¹; ¹H NMR: δ 1.00 (m, 9H), 1.23-1.51 (m, 7H), 1.60-1.80 and 1.90-2.10 (m, 2H), 2.45 (s, 6H), 3.80-3.88 (m, 1H), 4.03-4.28 (m, 3H), 4.34-4.43 (m, 1H), 4.56-4.70 (m, 1H), 7.30-7.48 (m, 8H), 7.50-7.69 (m, 10H); ¹³C NMR: δ 14.2, 19.3, 21.7, 24.3, 26.8, 33.5, 33.8, 60.2, 67.0, 73.7, 80.8, 127.5, 127.7, 127.9, 128.1, 129.9, 130.0, 135.7, 136.1, 144.9, 145.1, 172.7.

A mixture of the above compound (1.22 g, 1.59 mmol), NaI (1.43 g, 9.53 mmol) and Zn-dust (0.650 g, 10.0 mmol) in DMF (10 mL) was heated at 90 °C for 8 h (TLC). The mixture was brought to room temperature, poured in H₂O (30 mL) and extracted with Et₂O (3 × 15 mL). The combined organic extracts were washed

successively with 10% aqueous Na₂S₂O₃ (1 × 10 mL), H₂O (3 × 10 mL) and brine (1 × 5 mL), and dried. Solvent removal followed by column chromatography (silica gel, 0–10% Et₂O/hexane) of the residue gave pure **10**. Yield: 0.565 g (84%); colorless oil; $[\alpha]_D^{24} = +21.3$ (*c* 1.01, CHCl₃); IR: 1735, 997 cm⁻¹; ¹H NMR: δ 1.05 (s, 9H), 1.23 (t, *J* = 7.2 Hz, 3H), 1.39–1.58 (m, 6H), 2.16 (t, *J* = 7.0 Hz, 2H), 4.08 (q, *J* = 7.2 Hz, 2H), 4.92–5.01 (m, 3H), 5.70–5.80 (m, 1H), 7.33–7.38 (m, 6H), 7.61–7.68 (m, 4H); ¹³C NMR: δ 14.2, 19.3, 23.9, 24.8, 26.9, 34.2, 37.0, 60.1, 74.3, 114.3, 127.3, 127.4, 129.4, 129.5, 134.1, 134.3, 135.8, 135.9, 140.6, 173.6. Anal. Calcd for C₂₆H₃₆O₃Si: C, 73.54; H, 8.54. Found: C, 73.78; H, 8.71.

3.1.8. (2R,3S)-1,2-Cyclohexylidenedioxyundecan-3-ol 11

To a cooled $(-78 \,^{\circ}\text{C})$ and stirred solution of C_8H_{17} Li [prepared from 1-bromooctane (11.60 g, 60.0 mmol) and Li (0.890 g, 127.1 mmol)] in THF (80 mL) was added 2 (5.10 g, 30.0 mmol) in THF (20 mL). After stirring the mixture for 3 h at -78 °C, it was gradually brought to room temperature and treated with aqueous saturated NH₄Cl. The supernatant was decanted and the residue washed with Et_2O (2 × 20 mL). The combined organic extracts were washed with aqueous saturated NH₄Cl (1×10 mL), dried, and concentrated in vacuo to give a residue, which upon column chromatography (silica gel, 0-15% EtOAc/hexane) furnished 11. Yield: 7.32 g (86%); colorless oil; $[\alpha]_D^{24} = +12.6$ (*c* 1.02, CHCl₃), {lit.¹⁰ $[\alpha]_D^{24} = +6.2$ (c 1.15, CHC1₃)}; IR: 3457 cm⁻¹; ¹H NMR: δ 0.86 (t, J = 6.4 Hz, 3H), 1.17–1.43 (m containing a s at δ 1.21, 16H), 1.56-1.79 (m, 8H), 1.94 (br s, 1H), 3.70-3.78 (m, 1H), 3.86-3.97 (m, 3H); ¹³C NMR: δ 13.9, 22.5, 23.7, 23.9, 25.0, 25.7, 29.1, 29.4, 29.5, 31.7, 32.5, 34.8, 36.0, 63.9, 70.5, 78.2, 109.3. Anal. Calcd for C₁₇H₃₂O₃: C, 71.79; H, 11.34. Found: C, 71.94; H, 11.50.

3.1.9. (2R,3S)-3-tert-Butyldiphenylsilyloxy-1,2-cyclohexylidenedioxyundecane 12

As described earlier, silylation of **11** (2.87 g, 10.10 mmol) with TBDPSCI (3.53 g, 12.85 mmol), imidazole (0.87 g, 12.85 mmol) and DMAP (catalytic) in CH₂Cl₂ (40 mL), followed by work up and column chromatography (silica gel, 0–5% EtOAc/hexane) afforded pure **12**. Yield: 4.54 (86%); colorless oil; $[\alpha]_D^{24} = +15.5$ (*c* 1.06, CHCl₃); IR: 3072, 3050 cm⁻¹; ¹H NMR: δ 0.89 (t, *J* = 6.4 Hz, 3H), 1.14–1.38 (m containing a s at δ 1.06, 25H), 1.48–1.72 (m, 8H), 3.66–3.75 (m, 2H), 3.80–3.87 (m, 1H), 3.93–4.10 (m, 1H), 7.34–7.47 (m, 6H), 7.67–7.75 (m, 4H); ¹³C NMR: δ 14.1, 19.4, 22.6, 23.8, 23.9, 24.2, 25.2, 26.9, 29.1, 29.3, 29.6, 31.8, 34.1, 34.8, 36.1, 66.4, 73.7, 77.7, 109.3, 127.4, 129.5, 133.8, 134.3, 135.8, 135.9. Anal. Calcd for C₃₃H₅₀O₃Si: C, 75.81; H, 9.64. Found: C, 75.74; H, 9.48.

3.1.10. (2*R*,3*S*)-3-*tert*-Butyldiphenylsilyloxyundecane-1,2-diol 13

As described earlier, compound **12** (2.87 g, 5.50 mmol) was deacetalized in CH₂Cl₂ (20 mL) with aqueous 80% TFA (8.0 mL). Usual work up and column chromatography (silica gel, 0–5% MeOH/CHCl₃) afforded pure **13**. Yield: 2.0 g (84%); colorless oil; $[\alpha]_D^{24} = +21.0$ (*c* 1.04, CHCl₃); IR: 3398 cm⁻¹; ¹H NMR: δ 0.89 (t, *J* = 6.8 Hz, 3H), 1.09–1.35 (m containing a s at δ 1.10, 21H), 1.45–1.55 (br m, 4H), 3.65–3.72 (m, 3H), 3.78–3.89 (m, 1H), 7.35–7.49 (m, 6H), 7.68–7.75 (m, 4H); ¹³C NMR: δ 14.1, 19.4, 22.6, 24.8, 27.0, 29.1, 29.2, 29.4, 31.7, 32.9, 63.1, 73.6, 75.5, 127.5, 127.7, 129.7, 129.8, 133.1, 133.7, 135.9. Anal. Calcd for C₂₇H₄₂O₃Si: C, 73.25; H, 9.56. Found: C, 73.14; H, 9.72.

3.1.11. (2*R*,3*S*)-3-*tert*-Butyldiphenylsilyloxy-1,2-epoxyundecane 14

To a cooled (0 °C) stirred solution of **13** (1.10 g, 2.49 mmol) in pyridine (8 mL) was added PTSCl (0.475 g, 2.49 mmol) in portions. After stirring for 18 h at the same temperature, the mixture was

poured into ice-water and extracted with EtOAc ($2 \times 20 \text{ mL}$). The combined organic extracts were washed with H₂O ($3 \times 15 \text{ mL}$) and brine ($1 \times 5 \text{ mL}$), dried, and concentrated in vacuo. The residue was purified by column chromatography (silica gel, 0–10% EtOAc/hexane) to furnish the monotosylate. Yield: 1.30 g (84%); colorless oil; [α]_D²⁴ = +19.5 (*c* 1.16, CHCl₃); IR: 3528, 1362, 1177 cm⁻¹; ¹H NMR: δ 0.86 (t, *J* = 6.8 Hz, 3H), 1.02 (s, 9H), 1.25–1.42 (m, 14H), 2.18 (br s, 1H), 2.45 (s, 3H), 3.75–3.87 (m, 2H), 3.98–4.04 (m, 1H), 4.21–4.27 (m, 1H), 7.31–7.45 (m, 8H), 7.61–7.66 (m, 4H), 7.78 (t, *J* = 8.0 Hz, 2H); ¹³C NMR: δ 14.0, 19.3, 21.5, 22.5, 24.5, 26.9, 28.9, 29.2, 29.3, 31.7, 32.7, 71.3, 71.5, 74.2, 127.4, 127.6, 127.9, 129.8, 132.5, 132.9, 133.5, 135.7, 144.8.

A mixture of the above compound (1.20 g, 2.01 mmol) and anhydrous K_2CO_3 (0.63 g, 4.58 mmol) in MeOH (10 mL) was stirred for 3 h at room temperature. The supernatant was decanted, and the solid residue washed with EtOAc (20 mL) and the combined organic extracts concentrated in vacuo. The residue was taken in EtOAc (30 mL) washed with H₂O (3 × 15 mL) and brine (1 × 5 mL), dried, and concentrated in vacuo. Column chromatography (silica gel, 0–5% EtOAc/hexane) of the residue gave pure **14**. Yield: 0.770 g (90%); colorless oil; $[\alpha]_D^{24} = +20.2$ (*c* 1.14, CHCl₃); IR: 1110 cm⁻¹; ¹H NMR: δ 0.89 (t, *J* = 6.4 Hz, 3H), 1.08 (s, 9H), 1.22–1.40 (m, 12H), 1.53–1.59 (m, 2H), 2.11–2.16 (m, 1H), 2.43–2.48 (m, 1H), 2.86–2.91 (m, 1H), 3.37–3.43 (m, 1H), 7.32–7.48 (m, 6H), 7.64–7.72 (m, 4H); ¹³C NMR: δ 13.9, 19.2, 22.5, 24.1, 26.7, 29.0, 29.2, 29.5, 31.7, 35.2, 45.9, 54.1, 73.1, 127.3, 127.4, 129.4, 129.5, 133.6, 133.7, 135.7. Anal. Calcd for C₂₇H₄₀O₂Si: C, 76.36; H, 9.49. Found: C, 76.42; H, 9.27.

3.1.12. (3R,4S)-4-tert-Butyldiphenylsilyloxydodec-1-en-3-ol 15

To a cooled (-40 °C) and stirred suspension of Me₃SI (1.25 g, 6.13 mmol) in THF (20 mL) was added n-BuLi (3.0 mL, 1.5 M in hexane, 4.91 mmol). After stirring for 1 h, compound 14 (0.522 g, 1.23 mmol) in THF (5.0 mL) was injected into the mixture and stirring was continued at -40 °C for 3 h and at room temperature for 12 h. Next, H₂O (15 mL) was added to the mixture. The organic layer was separated, and the aqueous layer extracted with EtOAc $(2 \times 10 \text{ mL})$. The combined organic extracts were washed with H_2O (1 × 10 mL), and brine (1 × 5 mL), and dried. Solvent removal followed by column chromatography (silica gel, 0-15% EtOAc/hexane) of the residue gave pure 15. Yield: 0.436 g (81%); colorless oil; $[\alpha]_{D}^{24} = +16.5$ (*c* 1.08, CHCl₃); IR: 3475, 997, 925 cm⁻¹; ¹H NMR: δ 0.86 (t, J = 6.4 Hz, 3H), 1.08 (s, 9H), 1.14-1.39 (m, 14H), 1.96 (br s, 1H), 3.48-3.52 (m, 1H), 4.01-4.07 (m, 1H), 4.93-5.23 (m, 2H), 5.74–5.95 (m, 1H), 7.34–7.43 (m, 6H), 7.61–7.65 (m, 4H); ¹³C NMR: 8 14.1, 19.3, 22.6, 25.4, 26.9, 29.1, 29.2, 29.4, 31.8, 31.9, 74.3, 77.9, 117.6, 127.4, 127.5, 127.6, 129.7, 129.8, 133.4, 133.6, 135.5, 135.7, 135.9, 136.4. Anal. Calcd for C₂₈H₄₂O₂Si: C, 76.66; H, 9.65. Found: C, 76.84; H, 9.48.

3.1.13. Ethyl (6*S*,9*R*,10*S*)-6,10-di-*tert*-butyldiphenylsilyloxy-9hydroxyoctadeca-7*E*-enoate 16

A mixture of **10** (0.097 g, 0.23 mmol), **15** (0.153 g, 0.35 mmol) and Grubbs' 2nd generation catalyst (5 mol %) in CH₂Cl₂ (5 mL) was stirred for 22 h. After concentrating the mixture in vacuo, the residue was subjected to column chromatography (silica gel, 0–15% EtOAc/hexane) to give pure **16**. Yield: 0.120 g (63% based on **10**); colorless oil; $[\alpha]_D^{25} = +16.3$ (*c* 1.02, CHCl₃); IR: 3583, 1736 cm⁻¹; ¹H NMR: δ 0.86 (t, *J* = 6.8 Hz, 3H), 1.19–1.44 (m containing a s at δ 1.12, 36H), 1.48–1.74 (m, 6H), 2.16 (t, *J* = 7.2 Hz, 2H), 3.41–3.48 (m, 1H), 4.01–4.14 (m containing a q at δ 4.07, *J* = 7.2 Hz, 4H), 5.68–5.72 (m, 2H), 7.35–7.41 (m, 12H), 7.61–7.68 (m, 8H); ¹³C NMR: δ 14.1, 14.2, 19.3, 22.6, 23.9, 24.8, 25.5, 26.9, 29.2, 29.4, 31.8, 31.9, 34.2, 37.0, 60.1, 74.2, 74.4, 78.0, 114.3, 127.3, 127.4, 127.6, 129.4, 129.5, 129.7, 129.8, 133.4, 133.7,

134.1, 134.4, 135.5, 135.8, 135.9, 140.6, 173.6. Anal. Calcd for C₅₂H₇₄O₅Si₂: C, 74.77; H, 8.93. Found: C, 74.98; H, 9.09.

3.1.14. Ethyl (6S,9R,10S)-6,9,10-trihydroxyoctadeca-7*E*-enoate 17

To a cooled (0 °C) and stirred solution of **16** (0.300 g, 0.36 mmol) in THF (5 mL) was added Bu₄NF (0.57 mL, 1 M in THF, 0.57 mmol). The reaction mixture was brought to room temperature and stirred until the reaction was complete (TLC, 3 h). The mixture was poured into ice-cold water (15 mL) and extracted with EtOAc (2×10 mL). The organic extract was washed with water (2 \times 10 mL) and brine (1 \times 5 mL), and dried. Removal of solvent followed by column chromatography of the residue (silica gel, 0-15% EtOAc/hexane) furnished 17. Yield: 0.112 g (87%); colorless oil; $[\alpha]_{D}^{24} = +3.3$ (*c* 1.22, CHCl₃); IR: 3435, 1729 cm⁻¹; ¹H NMR: δ 0.88 (t, J = 6.4 Hz, 3H), 1.22 (t, J = 7.2 Hz, 3H), 1.26–1.39 (m, 14H), 1.42-1.54 (m, 6H), 2.25 (t, J = 7.0 Hz, 2H), 2.50 (br s, 1H), 2.71 (br s, 2H), 3.51-3.56 (m, 1H), 4.01-4.17 (m, 4H), 5.56-5.68 (m, 2H); ¹³C NMR: δ 14.2, 18.9, 24.8, 26.2, 26.6, 31.2, 31.5, 34.2, 36.5, 37.2, 73.0, 74.2, 75.0, 78.3, 131.2, 137.1, 173.7. Anal. Calcd for C₂₀H₃₈O₅: C, 67.00; H, 10.68. Found: C, 67.26; H, 10.88.

3.1.15. (6S,9R,10S)-6,9,10-Trihydroxyoctadeca-7E-enoic acid 1d

A solution of **17** (0.1 g, 0.28 mmol) in aqueous-ethanolic KOH (2 M, 5 mL) was stirred at room temperature for 4 h. Most of the solvent was removed in vacuo. The residue was taken in CHCl₃ (10 mL), and the extract washed with water (2 × 5 mL) and brine (1 × 5 mL), and dried. Removal of solvent followed by preparative thin layer chromatography of the residue (silica gel, 5% MeOH/CHCl₃) furnished pure **1d**. Yield: 0.087 g (95%); semi-solid; $[\alpha]_D^{24} = +5.7$ (*c*, 0.831, CHCl₃), $[\alpha]_D^{24} = +6.3$ (*c* 0.980, MeOH) {lit.² $[\alpha]_D^{25} = -10.7$ (*c* 0.15, MeOH) for natural **1d**}; IR: 3630–3550, 3488, 1729 cm⁻¹; ¹H NMR (500 MHz): δ 0.86 (t, *J* = 6.2 Hz, 3H), 1.22–1.39 (m, 14H), 1.40–1.61 (m, 6H), 2.25 (t, *J* = 6.8 Hz, 2H), 1.83 (br s, 1H), 2.41 (br s, 2H), 3.52–3.55 (m, 1H), 4.04–4.10 (m, 2H), 5.56 (dd, *J* = 15.4, 5.4 Hz, 1H); 5.67 (dd, *J* = 15.4, 5.4 Hz, 1H); ¹³C NMR: δ 14.2, 23.8, 26.4, 26.9, 30.7, 30.8, 31.2, 33.9, 34.2, 37.7, 73.2, 75.0, 76.8, 130.8, 137.0, 177.2.

References

- (a) Brack Egg, A. Diccionario Enciclopedico de Plantas utiles del Perú; Centro de Estudios Regionales Andinos Bartolome de las Casas: Cuzco, Peru, 1999. p 187;
 (b) Miyake, M.; Sasaki, K.; Ide, K.; Matsukura, Y.; Shijima, K.; Fujiwara, D. J. Immunol. 2006, 176, 5797–5804; (c) Lovera, A.; Bonilla, C.; Hidalgo, J. Rev. Perú Med. Exp. Salud Publica 2006, 23, 177–181.
- Benavides, A.; Napolitano, A.; Bassarello, C.; Carbone, V.; Gazzerro, P.; Malfitano, AM.; Saggese, P.; Bifulco, M.; Piacente, S.; Pizza, C. J. Nat. Prod. 2009, 72, 813–817.
- (a) Vyavahare, V. P.; Chakraborty, C.; Maity, B.; Chattopadhyay, S.; Puranik, V. G.; Dhavale, D. D. J. Med. Chem. 2007, 50, 5519–5523; (b) Maity, B.; Chattopadhyay, S. Curr. Bioact. Compat. 2008, 4, 225–244; (c) Guha, P.; Dey, A.; Sarkar, B.; Dhyani, M. V.; Chattopadhyay, S.; Bandyopadhyay, S. K. J. Exp. Pharmacol. Ther. 2009, 328, 1–10; (d) Maity, B.; Banerjee, D.; Bandyopadhyay, S. K.; Chattopadhyay, S. Int. Immunopharmacol. 2009, 9, 491–498; (e) Yadav, S. K.; Adhikary, B.; Maity, B.; Bandyopadhyay, S. K.; Chattopadhyay, S. K.; Chattopadhyay, S. Int. Immunopharmacol. 2009, 9, 491–498; (e) Yadav, S. K.; Adhikary, B.; Maity, B.; Bandyopadhyay, S. K.; Chattopadhyay, S. Eur. J. Pharmacol. 2009, 614, 106–113; (f) Guha, P.; Dey, A.; Chatterjee, A.; Chattopadhyay, S.; Bandyopadhyay, S. K. Br. J. Pharmacol. 2010, 159, 726–734; (g) Patro, B. S.; Maity, B.; Chattopadhyay, S. Antioxid. Redox. Signal. 2010, 12, 945–960.
- (a) Turner, W. B.; Aldridge, D. C. Fungal Metabolites II; Academic Press: London, 1983; (b) Gardner, H. W. J. Lipid Res. **1970**, *11*, 311–321; (c) Kawagishi, H.; Ando, M.; Micno, T.; Yokota, H.; Konishi, S. Agric. Biol. Chem. **1990**, *54*, 1329– 1331; (d) Kuga, H.; Ejima, A.; Mitui, I.; Sato, K.; Ishihara, N.; Fukunda, K.; Saito, F.; Uenaki, K. Biosci. Biotech. Biochem. **1993**, *57*, 1020–1021; (e) Simon, B.; Anke, T.; Sterner, O. Phytochemistry **1994**, *36*, 815–816; (f) Nagai, T.; Kiyohara, H.; Munakata, K.; Shirahata, T.; Sunazuka, T.; Harigaya, Y.; Yamada, H. Int. Immunopharmacol. **2002**, *2*, 1183–1193.
- (a) Salaskar, A.; Mayekar, N. V.; Sharma, A.; Chattopadhyay, A.; Nayak, S. K.; Chattopadhyay, S. Synthesis 2005, 2777–2781; (b) Salaskar, A.; Sharma, A.; Chattopadhyay, S. Tetrahedron: Asymmetry 2006, 17, 325–329; (c) Roy, S.; Sharma, A.; Chattopadhyay, N.; Chattopadhyay, S. Tetrahedron Lett. 2006, 47, 7067–7069; (d) Sharma, A.; Gamre, S.; Chattopadhyay, S. Tetrahedron Lett.

2007, *48*, 633–634; (e) Sharma, A.; Gamre, S.; Chattopadhyay, S. *Tetrahedron Lett.* **2007**, *48*, 3705–3707; (f) Sharma, A.; Gamre, S.; Roy, S.; Goswami, D.; Chattopadhyay, A.; Chattopadhyay, S. *Tetrahedron Lett.* **2008**, *49*, 3902–3905; (g) Sharma, A.; Das, P.; Chattopadhyay, S. *Tetrahedron: Asymmetry* **2008**, *19*, 2167–2170.

- (a) Roy, S.; Sharma, A.; Mula, S.; Chattopadhyay, S. Chem. A Eur. J. 2009, 15, 1713–1722; (b) Roy, S.; Sharma, A.; Dhotare, B.; Vichare, P.; Chattopadhyay, A.; Chattopadhyay, S. Synthesis 2007, 1082–1090.
- For reviews on olefin metathesis, see: (a) Armstrong, S. K. J. Chem. Soc., Perkin Trans. 1 1998, 371–388; (b) Schrock, R. R. Tetrahedron 1999, 55, 8141–8153; (c) Fürstner, A. Angew. Chem., Int. Ed. 2000, 39, 3012–3043; (d) Trnka, T. M.;

Grubbs, R. H. Acc. Chem. Res. 2001, 34, 18–29; (e) Blackwell, H. E.; O'Leary, D. J.; Chatterjee, A. K.; Washenfelder, R. A.; Bussmann, D. A.; Grubbs, R. H. J. Am. Chem. Soc. 2000, 122, 58–71; (f) Chatterjee, A. K.; Choi, T.-L.; Sanders, D. P.; Grubbs, R. H. J. Am. Chem. Soc. 2003, 125, 11360–11370.

- (a) Sharma, A.; Mahato, S.; Chattopadhyay, S. Tetrahedron: Lett. 2009, 50, 4986– 4988; (b) Sharma, A.; Gamre, S.; Chattopadhyay, S. Tetrahedron: Asymmetry 2009, 20, 1164–1167; (c) Biswas, S.; Chattopadhyay, S.; Sharma, A. Tetrahedron: Asymmetry 2010, 21, 27–32.
- 9. Goswami, D.; Chattopadhyay, A.; Chattopadhyay, S. Tetrahedron: Asymmetry 2009, 20, 1957–1961.
- 10. Chattopadhyay, A.; Mamdapur, V. R. J. Org. Chem. 1995, 60, 585-587.