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Mechanistic Insights into the Origin of Stereoselectivity in an Asymmetric Chlorolactonization Catalyzed by (DHQD)₂PHAL

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ABSTRACT: Electrophilic halofunctionalization reactions have undergone a resurgence sparked by recent discoveries in the field of catalytic asymmetric halocyclizations. To build mechanistic understanding of these asymmetric transformations, a toolbox of analytical methods has been deployed, addressing the roles of catalyst, electrophile (halenium donor), and nucleophile in determining rates and stereopreferences. The test reaction, (DHQD)₂PHAL-catalyzed chlorocyclization of 4-aryl-4-pentenoic acids with 1,3-dichloro-5,5-dimethylhydantoin (DCDMH), is revealed to be first order in catalyst and chlorenium ion donor and zero order in alkenoic acid substrate under synthetically relevant conditions. The simplest interpretation is that rapid substrate-catalyst binding precedes rate-limiting chlorenium attack, controlling the face selectivity of both chlorine attack and lactone closure. ROESY and DFT studies, aided by crystal structures of carboxylic acids bound by the catalyst, point to a plausible resting state of the catalyst-substrate complex predisposed for asymmetric chlorolactonization,. As revealed by our earlier labeling studies, these findings suggest modes of binding in the (DHQD)₂PHAL chiral pocket that explain the system's remarkable control over rate- and enantioselection-determining events. Though a comprehensive modeling analysis is beyond the scope of the present work, quantum chemical analysis of the fragments' interactions and candidate reaction paths point to a one-step concerted process with the nucleophile playing a critical role in activating the olefin for concomitant electrophilic attack.

Introduction

Electrophilic olefin halocyclizations (Figure 1a) are long-known workhorse organic transformations,¹ now returning to prominence as their catalytic asymmetric variants offer a powerful strategy for enlarging the chiral pool.²⁻⁵⁰ In early work on 4-aryl-4-pentenoic acids, we found Sharpless's ligand, (DHQD)₂PHAL, to be an efficient mediator for asymmetric chlorolactonization (Figure 1b).^{40,51} Since then, reports from our own and other labs have confirmed the efficacy of cinchona alkaloid dimers in a range of asymmetric halofunctionalizations.^{17,19,22,41-43,52-55} With the growing list of examples, mechanistic insight into the catalytic activation and stereoinduction in these processes is necessary to support the design of new reactions and catalyst scaffolds. Taking (DHQD)₂PHAL-catalyzed asymmetric chlorolactonization of 4-aryl-4-propenoic acid as a test reaction, we present here kinetic, spectroscopic, structural, and computational studies leading to a mechanistic model.

Our recent labeling and spectroscopic analyses⁵⁶⁻⁵⁷ revealed the absolute and relative face selectivities of Cl (electrophile) and O (nucleophile) addition across the double bond in **1** (Figure 1c) as well as the rate and stereochemical effects of varying the chlorohydantoins (chlorenium ion donors).⁵⁶⁻⁵⁷ The cumulative findings of this work led to the following conclusions:

 Catalyst templated addition across the olefin 1 shows both a strong pro-*R* preference (>20:1) for chlorenium ion attachment at C-6, and closure favoring the 5*R* over the 5*S* lactone by a factor of >10:1. The net result is predominant *syn* Cl, O addition across the olefinic π -bond in **1-D**, obviating the potential intermediacy of a 3-membered cyclic chloronium ion (Figure 1c).

- In principle, the two new bonds in the major product stereoisomer 2-D could be formed stepwise, *via* a carbocation intermediate, or concertedly (Figure 1c).
- 3. In either mechanistic case, the catalyst templates the ring closure. Catalyst binding, mainly *via* hydrogen bonding and van der Waals interactions, presumably limits the conformational choices of substrate **1**, guiding the enantioselective cyclization.
- 4. As depicted in Figure 2, we have probed the effects of tuning the chlorenium source in our asymmetric chlorolactonization protocol. By preparing and studying a series of previously unknown *N*-aroylated *N*-chlorohydantoins, we have shown that *N*1 substituents inductively activate delivery of the *N*3 chlorine to the substrate during the course of the chlorolactonization. The reaction rates of these electronically perturbed chlorinating agents vary as predicted by their *HalA* values (halenium affinity).⁵⁸ The linearity and low slope of the $\Delta\Delta G^{\dagger}$ [=RTln(k_{Ar}/k_{Ph})] vs *HalA* plot suggests an early transition state for chlorine transfer. Furthermore, chiral chlorohydantoins in these reactions display classic match-mismatch behaviors.⁵⁷ These results indicate direct involvement of the

chlorohydantoins in the rate and stereoselectivity-determining events in the asymmetric chlorocyclization.

These preliminary mechanistic findings call for a full investigation of the reaction mechanism to identify the modes of interaction among the participants, and to map out the catalytic cycle of this $(DHQD)_2PHAL$ catalyzed asymmetric chlorolactonization.





c. Stereochemical Outcomes of the Chlorolactonization of 1-D



Figure 1. a. Generalized depiction of halocyclization reactions; b. Catalytic asymmetric chlorolactonization of 4-aryl-4-pentenoic acid 1 mediated by (DHQD)₂PHAL; c. Summary of observed stereoselectivities with deuterated substrate analog **1-D**, highlighting the *syn* addition as the major product of the catalytic reaction.

Results and Discussion

1. Kinetic Studies

Kinetic investigation of multistep organic reactions can provide key insights into reaction mechanisms by revealing the order of individual components in the rate-determining step (RDS). As articulated in recent years by Blackmond and co-workers,⁵⁹⁻⁶² Reaction Progress Kinetic Analysis (RPKA) exploits modern reaction monitoring methods and easily accessible fitting and graphing software to improve the ease and accuracy of analyses over classical approximation methods. A great advantage of RPKA is that it can probe practical reactions such as halofunctionalization under their native conditions, unlike classical kinetic treatments that require e.g. the unbalanced concentrations used in pseudo first order studies. In this work, RPKA results place critical boundary conditions on the mechanisms proposed for this asymmetric chlorolactonization. In the original optimized reaction, 4-phenyl-4-pentenoic acid **1** at 0.051 M concentration was cyclized in the presence of 0.056 M 1,3-dichloro-5,5-diphenylhydantoin (DCDPH) as the chlorine source, along with 10 mol% (DHQD)₂PHAL as the chiral catalyst. The highest enantioinduction was observed in a 1:1 chloroform:hexane mixture as solvent with benzoic acid as an additive at 0.051 M (Figure 1b).⁴⁰

To follow the kinetics of this reaction, NMR analysis proved optimal as the reagent and products displayed well-resolved peaks under the reaction conditions. The following modifications to the standard protocol were introduced to simplify the mixture for kinetic studies: (i) deuterated chloroform was employed instead of 1:1 chloroform:hexane; (ii) 4 mol% of the catalyst was used to slow the reaction, enabling capture of the important early data points. This approach allowed time for temperature equilibration in the probe and shimming, while retaining $\geq 85\%$ of the starting material at the initial point of measurement; (iii) 1,3-dichloro-5,5-dimethylhydantoin (DCDMH) was employed since the originally used DCDPH results in the insoluble byproduct 1-chloro-5,5-diphenylhydantoin, adversely affecting the spectra of the evolving reaction mixture. (iv) to reduce the complexity of the initial kinetic studies, benzoic acid was omitted, although, as described in the SI, and later in the manuscript, including benzoic acid was illuminating with respect to the order of the substrate. These adjustments for the kinetic studies led to only a 5% drop in ee and presumably do not qualitatively change the reaction mechanism.

A detailed RPKA analysis of the chlorolactonization reaction revealed the following results (see SI for detailed experimental analysis and a brief theoretical description of RPKA in the context of asymmetric halogenation):



(a) The asymmetric chlorolactonization is *zero order* in alkene carboxylic acid **1**.

Figure 2. Electronic perturbation at the *N*1 substituent of chlorohydantoins affecting the rate of chlorolactonization. The plot represents the linear variation of $\Delta\Delta G^{\ddagger} = -RTln(k_{Ar}/k_{Ph})$, relating the experimentally determined rate constants (k_{Ar}) with the theoretically calculated absolute *HalA* (CI) values.

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- (b) The reaction shows *first order* dependence with respect to the catalyst (DHQD)₂PHAL and the chlorenium ion donor-DCDMH.
- (c) The reaction does not suffer from any catalyst deactivation or product inhibition, as demonstrated by 'the same excess' experiments.
- (d) Addition of an external carboxylic acid such as benzoic acid or an inert alkenoic acid, that can potentially compete with the substrate's binding to the catalyst's active site, retards the overall rate of the reaction.

The above observations can be summarized as follows:

 $Rate = k \left[(DHQD)_2 PHAL \right]^1 \left[DCDMH \right]^1 [\mathbf{1}]^0$

The fact that the reaction is zeroth order in substrate **1** (4-phenylpent-4-enoic acid) suggests that the catalyst is saturated, binding **1** rapidly to the basic quinuclidine moiety to form the strongly hydrogen-bonded acid-base adduct. As detailed further below, this **resting state** species is a 1:2 complex of (DHQD)₂PHAL and **1**.

Quantum chemical modeling of the reaction components and their interactions finds that quinuclidine has a substantially stronger affinity for alkenoic acid 1 (-15.8 kcal/mol) than for DCDMH (-10.6 kcal/mol; see inset Table in Figure 3). It is thus not surprising that alkene 1 outcompetes DCDMH for binding to the catalyst. NMR evidence further supports this conclusion: 1,3-dichlorohydantoin with (DHQD)₂PHAL shows splitting of the CH₂ hydrogens into a diastereotopic pair, implying a complex formed with the chiral catalyst. However, addition of 1 completely reverses this complexation, returning the hydantoin spectrum to that of uncomplexed reagent.⁴⁰

Collision of suitable conformations of the **resting state** complex $(DHQD)_2PHAL-1$ with DCDMH results in chlorenium transfer to the alkene in a rate determining step (RDS, see SI for the detailed kinetic model that leads to the rate law and proposed catalytic cycle). This process may follow either of two paths: (a) formation of a β -chloromethyl carbenium ion **intermediate A** (Path A, Figure 3), or (b) a concerted addition *via* **transition state B**, to directly access the products (Path B, Figure 3). This latter Ad_E3-type process is an example of nucleophile assisted alkene activation (*NAAA*).⁶³

In the absence of (DHQD)₂PHAL or quinuclidine catalysts, the DCDMH and alkene **1** are essentially unreactive at -40 °C. Therefore, apart from controlling the enantioselectivity by providing a chiral pocket, the catalyst must also activate the alkene or the DCDMH (or both). To promote the stepwise pathway leading to **intermediate A**, the catalyst must activate the DCDMH to form the proposed chlorocarbenium ion intermediate. The potential viability of forming the ionic species depicted in **intermediate A** is supported by NMR evidence that quinuclidine can deprotonate 3-chloro-5,5-dimethyl hydantoin (MCDMH) (shaded box in Figure 3). Rapid cyclization, guided by the catalyst, would then afford the product. On the other hand, the concerted pathway via **transition state B**



Figure 3. Putative catalytic cycle for (DHQD)₂PHAL catalyzed chlorolactonization of alkenoic acid 1 (in the absence of benzoic acid additive). *Path A* depicts a stepwise addition via a carbocationic intermediate **A**. The shaded box on the right confirms the possibility of generating the ion-pair depicted in intermediate **A** ('H NMR studies show that quinuclidine deprotonates 3-chloro-5,5-dimethyl hydantoin in CDCI₃). *Path B* depicts a possible one step concerted addition to the alkene 1. NMR studies and computational modeling of the proposed intermediates and transition states are detailed in the following section. The left table insert displays calculated binding enthalpies of alkene 1, DCDMH and quinuclidine (a truncated model for the catalyst). These pairwise association energies argue for the validity of the resting state. For clarity, only one alkenoic acid is shown bound to the catalyst, although experimental results indicate two molecules are bound at one time (*vida infra*).

hinges on the activation of the alkene, irrespective of additional activation of the chlorenium source.

In the case of stepwise cyclization, the cation closure (step three in Figure 3) could not be rate limiting as this would predict a buildup of cation intermediate, which is not observed. Also, reversible formation of the carbocation should scramble the stereochemistry of the alkene =CHD site in deuterium labeled substrate **1-D** (see Scheme 1), a process ruled out by the finding that recovered starting materials retain their stereochemical integrity.^{4a} In the alternative event of a concerted addition *via NAAA* (**transition state B**), the concerted chlorenium ion attack and ring closure would directly form the chlorolactone product. In either case (Path A or B), the chlorenium ion delivery to the olefin (step 2 or 2') must be part of the rate-determining step.

The kinetic studies also helped clarify the role of the benzoic acid additive, which marginally increases the enantioselectivity. We surmise that its presence aids in maintaining the rigidity of the C2-symmetric (DHQD)₂PHAL catalyst (see next section for validation of this hypothesis by NMR and X-ray studies), and perhaps also aids in shuttling the protons required to neutralize the byproduct hydantoin anions, especially toward the end of the reaction when the concentration of 1 is low. Consistent with this idea, benzoic acid addition at an equimolar ratio to substrate lowers the rate of chlorolactonization (see SI for experimental details); presumably benzoic acid competes with 4-phenylpent-4-enoic acid 1 for binding in the catalyst's active site (vida infra). This interpretation gained support from RPKA studies with benzoic acid added, which raised the measured order of the alkene from zero to 0.5 in the rate equation. This change is expected due to the fact that alkene 1 has to compete with the benzoic acid for binding in the catalyst, making its concentration relevant with respect to the rate. The following is the observed rate law in presence of benzoic acid:

$Rate = k \left[(DHQD)_2 PHAL \right]^1 \left[DCDMH \right]^1 [\mathbf{1}]^{0.5}$

To further explore the proposed catalytic cycle, we resorted to competition studies (Figure 4). Prior analysis of the catalytic asymmetric chlorolactonization methodology revealed that 4-(4-(trifluoromethyl)phenyl)pent-4-enoic acid **3** is one tenth as reactive as **1**.⁴⁰ Nonetheless, the trifluoromethyl substituent should not interfere with binding, as other sterically comparable substrates behaved well under the same reaction conditions. If substrate **3** binds to the catalyst to form an unreactive analogue of the **resting state**, it should serve as a competitive inhibitor, decreasing the concentration of the reactive complex with substrate **1**. Indeed, as shown in Figure 4, the chlorolactonization of **1** was slowed dramatically (decreased by 58%) upon addition of one equivalent of **3** to a 1:1 mixture of **1** and DCDMH with 1.5 mol% catalyst.

A similar competition study was performed between 4-phenylpent-4-enoic acid 1 and 4-(3-nitrophenyl)pent-4-enoic acid 4, known to be a slow substrate. With a 1:1:1 mixture of substrates 1, 4 and DCDMH under 1.5 mol% catalyst loading, the rate of the chlorolactonization of 1 was decreased by 71% (Figure 4). At a 1:1 ratio of inhibitor 4 to substrate 1, a 50% decrease in rate would suggest similar binding affinity of 4 and 1 with (DHQD)₂PHAL; the larger observed inhibition suggests a stronger complexation with 4, consistent with the formation of a putatively stronger π -acid-base complex.

Further evidence supporting the idea that chlorine delivery from the hydantoin to **1** is rate-determining may be found in the effects of structural variations in the hydantoin chlorenium donors. As noted



Figure 4. Rate of formation of product 2 in presence of alkenoic acids 3 and 4. The reduced rates imply 3 and 4 are competitive inhibitors of the catalyst, presumably binding to the active site and retarding the binding of substrate 1.

earlier (Figure 2), the *N*3 chlorine is activated when the *N*1 substituent of DCDMH is an electron withdrawing group (EWG). As quantified by the *HalA* values, stronger EWGs on *N*1 accelerate the reaction, consistent with chlorine delivery to **1** in the RDS. A related result is that chiral chlorohydantoins show match/mismatch effects in reaction, pointing to direct involvement of the chlorenium delivery reagent in the chlorocyclizations.⁵⁷

To probe the involvement of the carboxylic acid moiety in the rate determining step, chlorocyclization of the deuterated 4-phenylpent-4-enoic acid **1-OD** (RCO₂D) was studied and compared to the same reaction with unlabeled substrate **1**. An inverse isotope effect ($k_{\rm H}/k_{\rm D}$ = 0.82, see SI) was measured, suggesting that the carboxylate plays a direct role in the reaction. Considering *Path A*, the change from H to D should not lead to the observed KIE, as the electrophilic transfer of the chlorenium ion is ostensibly insensitive to the nature of the hydrogen-bonded nucleophile. On the other hand, the concerted *Path B* via **transition state B** would presumably be accelerated by the H to D change, as the carboxylate oxygen's, and therefore the alkene's nucleophilicity is more activated as a consequence of the increased acidity of **1-OD** vs **1-OH**.

Overall, reaction progress kinetic analysis (RPKA), competition, reagent, and isotope studies find that (a) rate is first order in catalyst and in DCDMH, but zero order in substrate; (b) chlorenium donor ability and asymmetry characteristics affect reaction rates; and (c) the carboxylic acid's hydrogen isotope also modulates reaction rate. The implication is that the rate-determining step involves chlorenium transfer and the carboxylic acid's degree of deprotonation.

2. Resting state of the catalytic cycle

With the elementary steps of the catalytic cycle deduced from the kinetic findings, we undertook NMR and computational studies to gain structural insight into intermediates and transition states along the reaction paths. Our first target in this process was the **resting state** (catalyst-substrate complex) of the catalytic cycle.

A comprehensive conformational search of the free catalyst using molecular mechanics and density functional simulations revealed three low energy conformational orientations for the two sidechains on the phthalazine (see SI for computational details). These showed H_aCCH_b dihedral angles of roughly 80° and 170°, consistent with prior NMR and theoretical work.⁶⁴⁶⁶ Figure 5 (left) shows a DFT-B3LYP/6-31G^{*} (gas) minimized structure of

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Figure 5. Conformational changes observed upon binding of 1 to (DHQD)₂PHAL. The left model represents the minimized structure of free (DHQD)₂PHAL with H_aCCH_b dihedral angle ~170°, supported by the 9 Hz coupling of H_b with H_a. H NMR titration induces a change in the H_b/H_a coupling from 9 to <3 Hz with the addition of the alkenoic acid 1. The model on the right depicts a minimized structure of the catalyst bound to two alkenoic acid 1, where the two H_aCCH_b dihedral angle are 73.8° and 80.7°. The dihedral angles of the bound catalyst fit well with the observed smaller coupling constant for H_a/H_b. Computed binding free energies for complexation of the first and second alkenoic acid 1, and for DCDMH with (DHQD)₂PHAL are shown in the shaded box.

 $(DHQD)_2PHAL$ with a H_aCCH_b dihedral of 169.4°. This is consistent with the NMR of free $(DHQD)_2PHAL$, where the H_b resonance exhibits a *ddd* with three equivalent 9 Hz coupling constants. Dihedral angles between H_b and the methylene protons in the quinuclidine ring lead to the anticipated 9 Hz coupling. Thus, the third 9 Hz coupling of H_b is ascribed to its interaction with H_a, resulting from the *anti* orientation of the two protons.

Table 1. ^1H NMR analysis of alkenoic acid 5 and its complexes with (DHQD)_2PHAL and quinuclidine at -40 $^\circ\text{C}$ in CDCl_3.

H _I H _m H _t	proton	δ (ppm)	∆δ with (DHQD)₂PHAL	∆δ with quin.
СО2Н	Ht	2.52	-0.52	-0.19
F H _k H _j 5	Hr	2.80	-0.43	-0.06
	Hm	5.06	-0.46	-0.04
quinuclidine	H	5.25	-0.54	-0.04
	Hj	7.36	-0.23	+0.04
	Hĸ	7.01	-0.29	-0.06

Titration of (DHQD)₂PHAL with substrate 1 lowers the catalyst's H_a-H_b coupling constant from 9 to <3 Hz, suggesting a conformation with an averaged H_aCCH_b dihedral angle close to 90°. This geometry, as depicted in Figure 5, also orients the quinuclidine N toward the chiral cleft of the catalyst. Furthermore, the 1:1 catalyst complex exhibits broad peaks which sharpen on addition of another equivalent of alkenoic acid suggesting that in the resting state, the catalyst binds two alkenes. Finally, conformational searching using the MMFF94 force field, followed up by B3LYP-D3/6-31G*/postSMD(CHCl₃) structural optimization of the (DHQD)₂PHAL bound with two alkenoic acids found calculated binding free energies for the first and second molecules of 1 in (DHQD)₂PHAL of -14.6 and -13.3 kcal/mol, respectively. Among the lowest energy complex minima, the structure shown at right in Figure 5 is used here as the resting state catalyst model; this structure has H_aCCH_b dihedral angles of 73.8° and 80.7°, consistent with the <3 Hz coupling of Ha with Hb in the NMR of the fully saturated complex.

Given the desymmetrization observed in the NMR of dichlorohydantoin interacting with the catalyst, one might expect that DCDMH binding would compete for the catalyst active site. However, for both the empty and singly occupied forms of the catalyst, the -14.6 and -13.3 kcal/mol binding energies of an additional acid substantially outcompete the -5.7 kcal/mol binding free energy of a molecule of DCDMH. Interacting with the doubly occupied **resting state**, the association of DCDMH is even weaker, as shown by the free energy values in Figure 5.

Further structural data on the resting state complex were obtained from NMR studies of a 2:1 mixture of 4-(4-fluorophenyl)pent-4-enoic acid 5 and (DHQD)₂PHAL in CDCl₃ carried out at -40 °C. Carboxylic acid 5 was used as a substitute for 1 since its aromatic protons (ortho to fluorine) do not overlap with the aromatic protons of the catalyst. Table 1 compares the ¹H NMR chemical shifts of the substrate with and without (DHQD)₂PHAL present. It is notable that the vinylic protons and also two methylene groups of the alkenoic acid 5 are shielded by ~0.5 ppm upon complexation with catalyst. We interpret this shielding as a combination of two effects: (a) deprotonation of the carboxylic acid and (b) binding in the cleft of the catalyst. To distinguish these contributions, ¹H NMR spectra of 5 were taken with quinuclidine in CDCl₃. There, it was the methylene group adjacent to the carboxylate moiety that showed the highest upfield shift (0.19 ppm). The other protons' shifts showed at most slight changes (~0.04 ppm). Thus, acid-base complex formation does not account fully for the large shielding seen with the catalyst-bound substrate.

To further probe the interactions between the substrate and the catalyst, ROESY experiments using the methods described by Bodenhausen *et al.*^{67.69} found correlations between protons of **5** and (DHQD)₂PHAL offering further insights into binding. As depicted in Figure 6, ROESY correlations, leading to average distances are grouped into intramolecular and intermolecular correlations. The <u>ROESY Correlations of (DHQD)₂PHAL + 5</u>



Figure 6. Intramolecular and intermolecular ROESY correlations of **5** bound to (DHQD)₂PHAL. The intramolecular correlations place the ethyl group near the phthalazine ring, while the methoxy shows close intermolecular relations with the substrate methylene units. These interactions are present in the lowest energy conformation of (DHQD)₂PHAL (see Figure 5, bound structure).



Figure 7. a. Prior work illustrating experimental evidences that support concerted addition to an alkene b. Transition states leading to syn and anti-addition for quinuclidine catalyzed chloro-lactonization. Calculated using density functionals B3LYP-D3/6-31G* SM8 (CHCl₃)

distances derived for the intramolecular interactions fit well with the minimized structure of the catalyst, illustrated in Figure 5. Probing intermolecular interactions, the OMe sidechains of the quinoline group (H_n) showed contacts with the substrate's methylene (H_r and H_t) and phenyl protons (H_k). This also matches well with the calculated **resting state** model, depicted in Figure 5, where the calculated distances fall well within the ROESY measured values (H_n/H_t 2.6 Å calc'd, 3.0 Å expt, H_n/H_r 2.5 calc'd, 3.2 Å expt, H_n/H_k 2.9 Å calc'd, 3.8 Å expt). The ROESY verified model for the **resting state** shown in Figure 5 illustrates carboxylate binding to the protonated quinuclidine moiety, positioning the vinylic protons in the shielding regions of the quinoline rings, which leads to their observed upfield shifts.

As a C₂ symmetric cinchona alkaloid, (DHQD)₂PHAL can potentially bind to two substrates at the same time (as depicted in Figure 5). Consistent with this idea, when (DHQD)₂PHAL is treated with only one equivalent of the alkenoic acid 1, the ¹H NMR peak broadening at room temperature clearly suggests loss of structural symmetry due to partial occupancy of the substrate. The spectra sharpen up in the presence of two equivalents of 1. These results support a picture in which (DHQD)₂PHAL requires two substrates (or other carboxylic acids) to maintain its conformational rigidity in the C2-symmetric anti-open form. On the other hand, addition of four equivalents of alkenoic acid to (DHQD)₂PHAL resulted in an averaged spectrum for the substrates, indicating fast equilibrium (on the NMR time scale) between bound and free forms of the alkenoic acid, while the catalyst's spectra remained sharp. Similar behavior is seen with benzoic acid (see SI for NMRs of (DHQD)₂PHAL with benzoic acid).

To further explore the stoichiometry of complexation, (DHQD)₂PHAL was treated with two equivalents of a 1:1 mixture of alkenoic acids 1 and 4. This NMR experiment showed shielding behavior for both 1 and 4 similar to that seen in 2:1 substrate:catalyst mixtures with the individual substrates. If catalyst had only a single binding site, such mixtures should show more shielding of resonances of the more tightly bound alkenoic acid. Since the same degree of chemical shift is observed in the above case with the spectra being well-resolved, it appears that the catalyst binds both substrates in a 1:1:1 complex without competition. This 2:1 binding model is in accord with the reactivity results involving 1, 3, and 4 previously shown in Figure 4, and with the calculated binding energetics in Figure 5. As discussed further below, the 2:1 complex may undergo attack by the chlorenium ion donor on either of the two essentially independently bound substrate molecules; it is thus the substrate for attack by the chlorenium ion donor.

An alternative scenario might involve dissociation to an active 1:1 complex, with the catalytically incompetent 2:1 complex serving simply as an off-cycle reservoir. In this scenario, however, the initial rate would not be the same for two reactions with different starting concentrations, such as the different excess experiments carried out in this study. We would anticipate the reaction with a higher concentration of the starting alkene to have a smaller initial rate. This behavior is not observed (see Figure S1 and the description of kinetic models in the S1).

Although we were unable to obtain crystals of the **resting state** itself, we were able to co-crystallize the catalyst (DHQD)₂PHAL with benzoic acid. This crystal structure displayed essentially the conformation deduced from the NMR studies (see SI for crystal structure), with the two H_aCCH_b dihedral angle being 73.7° and 77.6°.

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3. Structural insights into transition state B

To understand the asymmetric induction achieved in the (DHQD)₂PHAL-catalyzed chlorocyclization of **1** with DCDMH, a model of the interaction of the chlorenium ion donor with the **rest-ing state** is needed, which must entail both activation and asymmetric specificity. The combination of substrate **1** with DCDMH alone is essentially unreactive (at -30 °C); a basic catalyst, such as quinuclidine or (DHQD)₂PHAL is needed to promote the reaction. Also, as noted earlier, both rate and stereoselectivity are affected by electronic and stereochemical variations in the dichlorohydantoins used. Thus, the ring-forming reaction involves both the state of carboxylate deprotonation, and the identity of the chlorine donor.

One path that might be envisioned is indirect chlorine transfer to substrate via a chlorinated (DHQD)₂PHAL. This can be ruled out as it predicts that the enantioselectivity should be independent of the chlorenium ion source (hydantoin) as the *N*-chloroquinuclidinium form of (DHQD)₂PHAL would now serve as the active *in situ* generated halogenating reagent. Also, as assessed in terms of *HalA* (Cl) values, chlorenium ion transfer to the quinuclidine nitrogen in the catalyst would be strongly endothermic (Δ *HalA* (Cl) 24.0 kcal/mol in CHCl₃), even when the stabilizing effects of tight ion pairing and solvation are included in the calculation.

Evidence for concerted addition (NAAA) pathway

Without assistance, chlorenium transfer from DCDMH to the alkene would be unfavorable (Δ *HalA* (Cl) = 22.3 kcal/mol in CHCl₃) (see *HalA* values in Figure 7). Despite its low *HalA*, alkene **1** does slowly react with DCDMH. However, the rate is accelerated greatly in the presence of a basic catalyst such as quinuclidine. This strongly suggests that the catalyzed halocyclization of **1** proceeds via a nucleophilic activated alkene addition (*NAAA*) mechanism. Previous reports on chlorolactonization (summarized in Figure 7, dashed box) have noted that a low energy barrier pathway via concerted addition can be triggered when the substrate adopts conformations with the anionic carboxylate oxygen atom near the alkene moiety. This close proximity of the nucleophile sharply enhances the alkene halenium affinity, enabling capture of the chlorenium ion from its donor, as the nucleophile approach now stabilizes for the developing charge in the alkene sp² carbon. This view is verified further by the contrasting isotope effects seen in the uncatalyzed reactions of DCDMH with alkenes 1 and 6 (Figure 7, dashed box). Here, in 1, replacement of the protons vicinal to the putative carbenium site with deuterium showed no observable kinetic isotopic effect $(k_{\rm H}/k_{\rm D} = 1.0)$.⁶³ As depicted in Figure 7, TS-I illustrates the concerted transition state with bond distances that indicate a one-step, asynchronous addition of the chlorenium and the carboxylate to the olefin. In the case of 6, the 4-methoxy analog of 1, the alkene's nucleophilicity is inherently enhanced enough that the nucleophile-assisted pathway is unimportant; uncatalyzed reaction of 6 with DCDMH is much faster as compared to 1. However, as expected for a stepwise pathway via a carbocationic intermediate, reaction of alkene 6 displayed a significant secondary kinetic isotopic effect $(k_H/k_D = 1.2)$. In contrast to the concerted addition calculated for alkene 1, TS-II depicts the attack to form the carbocation in the stepwise addition of the chlorenium ion to substrate 6. The HalA (Cl) values of 1 and 6 (158.8 and 169.4 kcal/mol) differ by >10 kcal/mol, supporting the formation of a carbocation intermediate. The HalA (Cl) value of **6** closely matches that of the hydrogen-bonded MCDMH anion (~173 kcal/mol), consistent with unassisted chlorenium capture by 6 to form the carbocation, an action too endothermic for olefin 1 without further activation.

The *m*-NO₂ analogue of **1**, alkene **3**, is intrinsically deactivated and unreactive towards DCDMH, hinting that *NAAA* can only offer so much activation if there are strong inherent factors retarding the intrinsic nucleophilicity of the alkene. Though it appears quite general for otherwise nonpolarized alkenes, the concerted *NAAA* mechanism evidently strikes a fine balance. With the experimental evidence firmly indicating concerted addition to the alkene **1**, transition states with quinuclidine as well as (DHQD)₂PHAL as a catalyst were computationally modeled and analyzed in the following section.



Figure 8. Transition states for (DHQD)₂PHAL catalyzed chloro-lactonization using DCDMH and the chiral chlorenium reagent. The non-degeneracy of the two transition states conforms with the previously reported matched-mismatched selectivity. Structures were calculated using the dispersion-corrected density functional method B3LYP-D3, with all optimizations carried out in SMD simulated CHCl₃. This scheme is denoted B3LYP-D3/6-31G*/SMD(CHCl₃).

Quinuclidine-catalyzed TS for syn and anti chlorocyclization

For the quinuclidine catalyzed reaction, transition structures for the lowest energy syn and anti concerted addition pathways were identified and optimized. (Figure 7, bottom). In search of optimized structures, conformational analyses using the MMFF94 force field provided poses, which were further refined via B3LYP-D3/6-31G*/postSMD (CHCl₃) calculations. Though the developing negative charge on the hydantoin ring in the syn TS benefits from proximity to the partially positive protonated quinuclidine, that advantage is outweighed by the steric freedom of approach from the far side (anti approach). In the CHCl₃ "solvent" as represented by the SMD dielectric continuum simulation, the anti-addition pathway is found to be 0.7 kcal/mol lower in energy than its synpartner, a result consistent with experimental findings. Both structures show similar parameters, with the developing C-Cl and C-O bond lengths slightly longer in the anti (2.2, 2.6 Å) than in the syn case (2.1, 2.4 Å). All attempts to initiate chlorenium atom transfer from DCDMH to 1 were monotonically endothermic in conformations that did not place the nucleophilic carboxylate moiety nearby (Figure 7, bottom). This implies that a stable β -chloromethyl carbenium ion intermediate is not a minimum along the reaction coordinate as would be expected for the stepwise mechanism.

The experimentally observed *syn: anti* ratio for quinuclidine catalyzed chlorocyclization of **1-D** is 1:5.⁵⁶ This agrees with the discussion above, and also with the calculated difference in activation energies computed for the *syn* and *anti* TS structures shown in Figure 7 ($\Delta\Delta G^{\dagger} = 0.7$ kcal/mol).

Modeling of the *syn*-addition TS for (DHQD)₂PHAL catalyzed reactions

Having identified the lowest energy TS structures for quinuclidine-catalyzed chlorocyclization of 1, we proceeded to model the syn structure into the optimized geometry of (DHQD)₂PHAL, reoptimizing at the B3LYP-D3/6-31G* level of theory. The reader is reminded that the asymmetric catalyzed chlorolactonization of 1-D favors the syn adduct 2-D (syn: anti ratio ~9:1).56 The energy of activation (10.5 kcal/mol) and partial bond lengths (2.4 Å for C-O and 2.1 Å for C-Cl) for the lowest energy (DHQD)₂PHAL catalyzed transition state were comparable to the quinuclidine catalyzed addition (Figure 8a). The acute H_aCCH_b dihedral angle of 61.7° in the TS effectively oriented the quinuclidine towards the center of the catalyst allowing for the alkenoic acid to sit inside the chiral pocket as it further enjoys π - π -stacking with the phthalazine linker. This dihedral angle also fits well with the NMR studies depicted in Figure 5 for (DHQD)₂PHAL bound to **1**. As illustrated in Figure 8a, the *Re* face of the alkene is now readily accessible by both the chlorenium ion donor and the quinuclidine-activated carboxylate nucleophile, the simultaneous addition of which leads to the experimentally observed major syn enantiomer. Computational searches for NAAAtype TS structures leading to the minor products were much less successful; those products may well arise via more complex stepwise paths, and further studies in this direction are deferred to a future detailed computational analysis.

Prior studies have demonstrated that chlorolactonization using chiral chlorohydantoins exhibits matched-mismatched behavior when used with (DHQD)₂PHAL (Figure 8b, table in dashed box). This provides further support for the hydantoin's direct involvement in the rate determining step as the chlorenium source. To connect our computational results to these experimental findings, we reoptimized the (DHQD)₂PHAL catalyzed transition state with the two enantiomers of the chiral hydantoin **8** and alkene **1**. Consistent with the experimental findings, the TS calculated with the (*R*)-**8** (mismatch) shows steric crowding of the isopropyl group with the quinuclidine moiety of $(DHQD)_2PHAL$, presumably magnified as the hydantoin is approaching the **resting state** prior to reaching the TS. The (*S*)-congener (match), however, has a more relaxed approach, having the isopropyl group pointed in the opposite direction. This is also represented in the activation energies for each transition state, with the match TS having a slightly lower barrier ($\Delta\Delta G^{\dagger} = 0.5$ kcal/mol). Owing to less steric conflict with the catalyst, the approach of (*S*)-**8** to the olefin is also more linear, avoiding the DCDMH distortion seen in the TS from the (*R*) isomer.

Summary

Reaction progress kinetic analysis (RPKA) and competition studies revealed the delivery of chlorenium ion to the olefin as a key feature of the rate determining step (RDS). These kinetic studies also define the order of each reagent and consequently the rate equation of this catalytic reaction both in the presence and absence of the benzoic acid: rate = $k[(DHQD)_2PHAL][DCDMH]$. Moreover, these studies were conducted at concentrations relevant to the optimized standard protocol, obviating the need for extrapolations outside the range studied. The observed inverse isotope effect (acid catalysis) suggests that the alkene-COOD is more activated for DCDMH attack in transition state B than alkene-COOH. Between the kinetic results, ROESY NMR studies, X-ray structures, and DFT analysis, we propose a resting state model in which two molecules of substrate 1 occupy the binding pockets of (DHQD)₂PHAL. In the proposed catalytic cycle (Figure 3), the alkene face that is ultimately chlorinated is exposed to solution, allowing collision with DCDMH. Ring closure is achieved when the backbone chain achieves a conformation placing C-4 near the partially negatively charged carboxylate oxygen, activating the olefin for the concerted NAAA closure. The syn preference reflects the electrostatic attraction between the developing negative charge on the hydantoin and the positively charged quinuclidinium ion. These mechanistic investigations, coupled with our previously reported labeling studies, suggest that the rate-determining and enantioselectivity-determining events occur together in the predominant pathway. Taken together, the experimental results were combined with an initial DFT analysis that provided structural insight into the resting state model and the corresponding transition state B. Overall, using the asymmetric chlorolactonization of 1 as a proof of principle, a toolbox of analytical techniques has been successfully optimized and applied to probe the nuances underlying the (DHQD)₂PHAL catalyzed halofunctionalization of olefins. These mechanistic studies have established an optimal range of alkene HalA values over which concerted reactions dominate, firmly defining both relative and absolute stereochemical relationships. This conceptual framework and set of tools for reaction assessment will serve as a guide, in a broad sense, to the emerging field of catalytic stereoselective halofunctionalizations of olefins. More specifically, the localized interactions identified in exploring these chlorolactonization reaction paths point the way to key features needed in the design of simplified organocatalysts.

ASSOCIATED CONTENT

Supporting Information. Experimental details, *HalA* calculations, characterization data, DFT computational data and MS

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office excel template for RPKA analysis. This material is available free of charge *via* the Internet at http://pubs.acs.org.

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