

A Facile Method for Preparation of Optically Active Hydantoin

Jun-ichi Yamaguchi,* Masakazu Harada, Takahito Kondo, Takeshi Noda, and Takayuki Suyama*

Department of Applied Chemistry, Faculty of Engineering, Kanagawa Institute of Technology, Shimo-ogino, Atsugi, Kanagawa 243-0292

(Received January 15, 2003; CL-030045)

Treatment of *N*-(4-nitrophenoxy-carbonyl)amino acid amide, which was prepared from amino acid amide and 4-nitrophenyl chloroformate, under weakly basic conditions proceeded to afford the corresponding hydantoin. A direct conversion of amino acid amide into hydantoin without isolation of *N*-(4-nitrophenoxy-carbonyl)amino acid amide was also accomplished.

Hydantoins and their synthesis by the action of carbonyl compounds have been known since early in the 20th century.¹ Recently, it has been found that hydantoins play an important role in medicinal activities such as fibrinogen receptor antagonist and leukocyte function-associated antigen-1 (LFA-1) antagonist.² Therefore, many synthetic methods have been reported in the past 10 years and can be classified into two types: 1) cyclization of *N*-carbamoyl amino acid ester under basic or acidic conditions,³ 2) cyclization of *N*-alkoxy or phenoxy-carbonyl amino acid amide.⁴ However, racemization check of the synthesized hydantoin has hardly been reported.

4-Nitrophenyl *N*-substituted carbamates^{5,6} as well as 4-nitrophenyl esters⁷ and carbonates⁸ are known as activated esters and react with amines to produce ureas, carboxamides, and carbamates, respectively. Among them, 4-nitrophenyl *N*-substituted carbamates generate the corresponding isocyanates on treatment with alkali or amine.⁹ From the above reports, we expected that treatment of *N*-(4-nitrophenoxy-carbonyl)amine bearing a nucleophilic group with alkaline would generate the corresponding isocyanate, which will transform into the corresponding cyclic compound by its intramolecular cyclization. In this communication, we wish to report a convenient method for preparation of (*S*)-hydantoin (**1**) from (*S*)-amino acid amide (**2**) via formation of *N*-(4-nitrophenoxy-carbonyl)amino acid amide (**3**) under weakly basic conditions (Scheme 1).

According to the method for the preparation of 4-nitrophenyl carbamate,⁵ (*S*)-**3** was prepared by reaction of (*S*)-**2** with 4-nitrophenyl chloroformate in the presence of 2.0 equivolar

amounts of NaHCO₃ in dry MeCN at room temperature for 3 h (Table 1).¹⁰ Unfortunately, because it was difficult to purify (*S*)-**3** from a small amount of 4-nitrophenol by chromatography on silica-gel, (*S*)-**3** was isolated by reprecipitation with ethyl acetate-hexane only. Therefore, the yields of (*S*)-**3** remained moderate.

Table 1. Preparation of (*S*)-**3** from (*S*)-**2**

Run	R ¹	R ²	R ³	Substrate	Product	Yield/%
1	CH ₂ Ph	H	H	2a ^a	3a	84
2	CH ₂ Ph	H	Ph	2b ^a	3b	76
3	CH ₂ Ph	H	(CH ₂) ₂ Ph	2c ^b	3c	71
4	-(CH ₂) ₃ -		Ph	2d ^a	3d	78

^aHCl salt. ^b4-Toluenesulfonic acid salt.

Next, we examined the degradation of (*S*)-**3** under basic conditions to prepare (*S*)-**1**. It was found that treatment of **3** with 1.0 equivolar amount of NaHCO₃ in H₂O-MeCN at room temperature for 3 h afforded (*S*)-**1** (Table 2). All of the reaction mixture changed from colorless into yellow instantly when an aqueous NaHCO₃ was added to a MeCN solution of (*S*)-**3**. This phenomenon shows that degradation of (*S*)-**3** and generation of 4-nitrophenol are very fast. Since formation of the corresponding isocyanate of (*S*)-**3d** derived from proline is impossible, we suspected that no transformation of (*S*)-**3d** into (*S*)-**1d** proceeded. Although the reaction rate was slow, (*S*)-**1d** was obtained. This fact indicated that (*S*)-**1d** would be given in another reaction pathway.

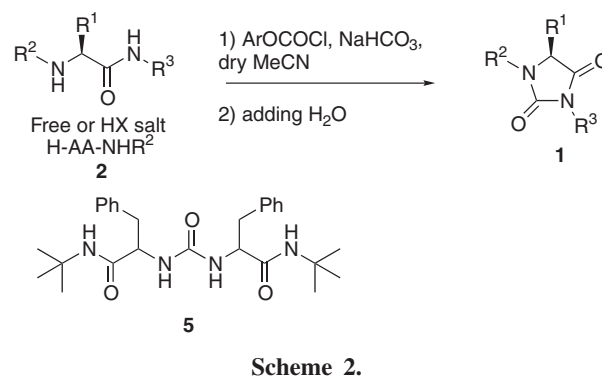
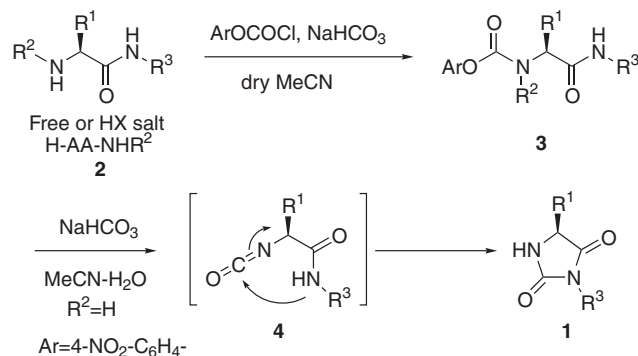


Table 2. Transformation of (*S*)-**3** into (*S*)-hydantoin (**1**)

Run	R ¹	R ²	R ³	Substrate	Product	Yield/% ^a
1	CH ₂ Ph	H	H	3a	1a	92
2	CH ₂ Ph	H	Ph	3b	1b	99
3	CH ₂ Ph	H	(CH ₂) ₂ Ph	3c	1c	93
4	-(CH ₂) ₃ -		Ph	3d	1d	80 ^b

^aThe structures of these compounds were supported by IR, Mass, and ¹H NMR spectra. ^bThe starting material (**3d**) was recovered in 14% yield.



Scheme 1.

In comparing the reaction conditions of 4-nitrophenoxycarbonylation of **2** and cyclization of **3**, the only difference is the presence or absence of water. Further, no degradation of **3** occurred under dry conditions even if excess amounts of NaHCO₃ existed. From the above results, **3** was prepared from **2** and 4-nitrophenyl chloroformate in the presence of excess amounts of NaHCO₃ in dry MeCN, and then only addition of water to the reaction mixture gave **1** (Scheme 2, Table 3).¹¹ However, in the case of Run 6, the intramolecular cyclization will be retarded by steric hindrance of the *tert*-butyl group, so that water may attack 2-isocyanatocarboxamide (**4f**) to regenerate **2f**, which reacts with **4** to give urea (**5**). The yield of **1f** was improved when 1.0 equiv. amount of triethylamine was used instead of water (Run 7).

Table 3. One-pot preparation of (S)-**1** from (S)-**2**

Run	R ¹	R ²	R ³	Product ^a	Yield/%	E.e./% ^b
1 ^c	CH ₂ Ph	H	H	(S)- 1a	96	>99
2 ^c	CH ₂ Ph	H	Ph	(S)- 1b	94	>99
3 ^d	CH ₂ Ph	H	(CH ₂) ₂ Ph	(S)- 1c	96	>99
4 ^c	-(CH ₂) ₃ -		Ph	(S)- 1d	68 ^f	>99
5 ^c	CH ₂ Ph	H	Pr	(S)- 1e	92	>99
6 ^c	CH ₂ Ph	H	<i>t</i> -Bu	(S)- 1f	28 ^g	>99
7 ^c	CH ₂ Ph	H	<i>t</i> -Bu	(S)- 1f	78 ^h	>99
8 ^d	<i>i</i> -Pr	H	Ph	(S)- 1g	96	98
9 ^d	CH ₂ -3-indolyl	H	Pr	(S)- 1h	96	>99
10 ^e	CH ₂ OBu ⁱ	H	(CH ₂) ₂ Ph	(S)- 1i	92	99
11 ^e	CH ₂ CO ₂ Bu ⁱ	H	(CH ₂) ₃ Ph	(S)- 1j	90	>99

^aThe structures of these compounds were supported by IR, Mass, and ¹H NMR spectra. ^bE.e. was determined by HPLC. Conditions: Run 1; column, Chiralpak WH (Daicel Chemical Industries, Ltd.), 1.0 mL/min of 0.25 mmol dm⁻³ CuSO₄, detection, 254 nm, other Runs; column, Sumichiral OA-4700 (Sumika Chemical Analysis Service, Ltd.), 1.0 mL/min of hexane:EtOH=9:1, detection, 254 nm. ^cHCl salt. ^d4-Toluenesulfonic acid salt. ^eFree amine. ^fThe starting material (**3d**) was recovered in 14% yield. ^gUrea (**5**) was obtained as a main product. ^hTriethylamine (1.0 equiv.) was used instead of water and the reaction mixture was stirred for 12 h after adding Et₃N. Urea (**5**) was obtained in 21% yield.

It has been reported that racemic-**1a** has a melting point of 186 °C¹² and (S)-**1a** has a melting point of 181–183 °C¹³ or 182 °C.¹⁴ Although the melting point (185–186 °C) of synthesized racemic-**1a** according to the literature was the same as that of the reported one, the melting point of (S)-**1a** obtained by our procedure was 171–172 °C, and was apparently different from that of the reported (S)-**1a**. Therefore, we undertook a racemization check of synthesized **1** by HPLC. A peak corresponding to the optically active hydantoin was identified by comparison with the standard hydantoin derived from racemic- or (R)-amino acid run under identical conditions (Table 3). It was found that the present reaction proceeded without racemization under the reaction conditions.

Similar to degradation of 4-nitrophenyl *N*-methylcarbamate under basic conditions, it is assumed that the present reaction will mainly proceed through the initial formation of **4** by treatment of **3** with NaHCO₃ in MeCN-H₂O. Isocyanate (**4**), in turn, gives the corresponding hydantoin (**1**) by its cyclization (elimination-addition process). Since formation of **4** is impossible when **2d** is used as the starting material, the reaction will proceed by an

addition-elimination process.

In conclusion, we have developed one-pot preparation of hydantoin from amino acid amide utilizing decomposition of 4-nitrophenyl *N*-substituted carbamate under weakly basic conditions without racemization.

References and Notes

- E. C. Wagner and M. Bizer, *Org. Synth.*, **1955**, 323, and references therein.
- H. U. Stilz, B. Jablonka, M. Just, J. Knolle, E. F. Paulus, and G. Zoller, *J. Med. Chem.*, **39**, 2118 (1996); T. A. Kelly, D. D. Jeanfavre, D. W. McNeil, J. R. Woska, Jr., B.-J. Bormann, and R. Rothlein, *J. Immunol.*, **163**, 5173 (1999); M. Nakajima, K. Itoi, Y. Takamatsu, T. Kinoshita, T. Okazaki, K. Kawakubo, M. Shindo, T. Honma, M. Tohigamori, and T. Haneishi, *J. Antibiot.*, **44**, 293 (1991).
- K.-H. Park, E. Abbate, S. Najdi, M. Olmstead, and M. J. Kurth, *Chem. Commun.*, **1998**, 1679; A. Boeijen, J. A. W. Kruijtzter, and R. M. J. Liskamp, *Bioorg. Med. Chem. Lett.*, **8**, 2375 (1998); L. J. Wilson, M. Li, and D. E. Portlock, *Tetrahedron Lett.*, **39**, 5135 (1998); S.-H. Lee, S.-H. Chung, and Y.-S. Lee, *Tetrahedron Lett.*, **38**, 9469 (1998); K.-H. Park and M. J. Kurth, *Tetrahedron Lett.*, **40**, 5841 (1999); J. Stadlwieser, E. P. Ellmerer-Müller, A. Takó, N. Maslough, and W. Bannwarth, *Angew. Chem., Int. Ed. Engl.*, **38**, 1402 (1998); J. Matthews and R. A. Rivero, *J. Org. Chem.*, **62**, 6090 (1997); Y. Hamuro, M. A. Scialdone, and W. F. DeGrado, *J. Am. Chem. Soc.*, **121**, 1636 (1999).
- B. A. Dressman, L. A. Spangle, and S. W. Kaldor, *Tetrahedron Lett.*, **37**, 937 (1996); X.-Y. Xiao, K. Ngu, C. Chao, and D. V. Patel, *J. Org. Chem.*, **62**, 6968 (1997).
- N. Choy, K. Y. Moon, C. Park, Y. C. Son, W. H. Jung, H.-I. Choy, C. S. Lee, C. R. Kim, and H. Yoon, *Org. Prep. Proced. Int.*, **28**, 173 (1996).
- B. Raji, J. M. Kassir, and T. P. Kogan, *Bioorg. Med. Chem. Lett.*, **8**, 3043 (1998); Q. Liu, N. W. Luedtke, and Y. Tor, *Tetrahedron Lett.*, **42**, 1445 (2001).
- M. Bodanszky, *Nature*, **175**, 685 (1955); M. Bodanszky and V. D. Vigneaud, *J. Am. Chem. Soc.*, **81**, 5688 (1959).
- K. Hoste, D. Bruneel, A. D. Marre, F. D. Schrijver, and E. Schacht, *Macromol. Rapid Commun.*, **15**, 697 (1994); D. M. Dixit and C. C. Leznoff, *J. Chem. Soc., Chem. Commun.*, **1977**, 798.
- H. L. Bender and R. B. Homer, *J. Org. Chem.*, **30**, 3975 (1965); F. M. Menger and L. E. Glass, *J. Org. Chem.*, **39**, 2469 (1974).
- 3a**: mp 132 °C (decomp.); **3b**: mp 136 °C (decomp.); **3c**: mp 127–128 °C (decomp.); **3d**: mp 137–138 °C.
- One-pot preparation of **1**: to a suspension of amino acid amide (0.3 mmol) and NaHCO₃ (0.9 mmol) in dry MeCN (5 mL) was added 4-nitrophenyl chloroformate (0.3 mmol) at room temperature. After being stirred for approximately 3 h, water (3 mL) was added and the reaction mixture was stirred for approximately 3 h. After removal of MeCN under reduced pressure, the organic materials were extracted with EtOAc. The organic layer was washed with 5% K₂CO₃ solution and brine successively, and was dried over Na₂SO₄. After removal of the solvent under reduced pressure, the residue was purified by preparative TLC on silica-gel and hydantoin **1** was isolated. (S)-**1a**: ¹H NMR (300 MHz, DMSO-*d*₆) δ 2.96 (1H, dd, *J* = 6.6 and 13.9 Hz), 3.12 (1H, dd, *J* = 4.4 and 13.9 Hz), 4.23 (1H, dd, *J* = 4.4 and 6.6 Hz), 7.20–7.30 (5H, m), 7.35 (1H, brs), 10.28 (1H, brs).
- F. Lippich, *Hoppe-Seyler's Z. Physiol. Chem.*, **90**, 124 (1914).
- H. D. Dakin and H. W. Dudley, *J. Biol. Chem.*, **17**, 29 (1914).
- T. Suzuki, K. Igarashi, K. Hase, and K. Tuzimura, *Agric. Biol. Chem.*, **37**, 411 (1973).