Carbohydrate Research 345 (2010) 780-786

Contents lists available at ScienceDirect

Carbohydrate Research

journal homepage: www.elsevier.com/locate/carres

# Synthesis of N-substituted iminosugar derivatives and their immunosuppressive activities

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### ARTICLE INFO

Article history: Received 15 December 2009 Received in revised form 26 January 2010 Accepted 29 January 2010 Available online 4 February 2010

Keywords: Iminosugar Synthesis Alkylation Immunosuppressive agent MTT assay

### ABSTRACT

Several N-alkyl and hydroxyethyl-substituted iminosugar derivatives, including L-altro and D-galacto epimers, were synthesized and the effects of these synthetic iminosugars on proliferation of mouse splenocytes were evaluated in the MTT assay. The experimental data demonstrated that the N-alkylated D-galacto iminosugars showed better inhibitory activities than L-altro analogues, which might hold potential as immunosuppressive agents.

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### 1. Introduction

The currently clinically used small-molecule immunosuppressive agents, such as cyclosporin A (CyA), sirolimus (Rapamycin), tacrolimus (FK506), and mycophenolate mofetil (MMF), have serious side effects including nephrotoxicity, neurotoxicity, infection, new onset post-transplant diabetes mellitus, hyperlipidemia, and hypertension. Furthermore, so far there is no antidote for the toxicity of these organ transplantation drugs. To improve transplant survival and reduce the toxicity of current agents, there is a great need to find highly efficacious immunosuppressants with low toxicity.<sup>1–4</sup>

Iminosugars, also known as the 'sugar-shaped alkaloids', have been found to be potent inhibitors of many carbohydrate-processing enzymes (i.e., glycosidases and glycosyltransferases).<sup>5–8</sup> Because these enzymes are frequently involved in essential biological processes,<sup>9,10</sup> iminosugars have great potential as therapeutic agents in many diseases such as cancer,<sup>11</sup> diabetes,<sup>12</sup> viral infections,<sup>13</sup> and lysosomal storage disorders.<sup>14,15</sup> However, the inhibition effects of iminosugars on immune system responses and their application as immunosuppressants have been less explored. To date only castanospermine, a naturally occurring indolizidine alkaloid, has been found to exhibit immunosuppressive activity.<sup>16</sup> Our preliminary results disclosed that some synthetic iminosugars (e.g., **1** and **2**, Fig. 1) displayed immunosuppressive



Figure 1. Structures of iminosugars 1 and 2, and their derivatives 3-10.

activity.<sup>17,18</sup> Encouraged by these experiments, to further understand the structure–activity relationships (SARs) of this type of compounds and to find iminosugars with better activities, we report herein the synthesis of several N-alkylated iminosugar derivatives and the evaluation of their immunosuppressive activities by the MTT assay.

### 2. Results and discussion

### 2.1. Design

Nojirimycin (NJ) and 1-deoxynojirimycin (DNJ), isolated from microorganism cultures and/or plants, are competitive inhibitors of *N*-glycan-processing enzymes (Fig. 2).<sup>19,20</sup> N-Alkylated





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Figure 2. Some emblematic iminosugars.

deoxynojirimycin and deoxygalactonojirimycin (DGJ) span a wide range of biological activities. For instance, *N*-butyl-1-deoxynojirimycin (NB-DNJ) has been approved as medicine for type I Gaucher disease, the most common lysosomal storage disorder. In addition, NB-DNJ was evaluated in Phase II clinical trials as an anti-HIV agent.<sup>21,22</sup> *N*-Hydroxyethyl-DNJ (miglitol) served as an oral drug for the treatment of type II diabetes taken either alone or in combination with other anti-diabetic medicines.<sup>23–25</sup> In addition, the galactose analogue *N*-nonyl-1-deoxygalactonojirimycin (NN-DGJ) was in clinical trials as an anti-HBV and an anti-HCV agent.<sup>26,27</sup> Based on our previous report,<sup>17</sup> to investigate whether the N-alkylated side chain influences the immunosuppressive activity or not, we designed some N-alkylated iminosugar derivatives **3–10** with side chains consisting of four-, six- or nine-carbons, or a hydroxyethyl group (Fig. 1).

### 2.2. Chemistry

HOAc = 4:2:1.

The preparation of iminosugars **3–8** is shown in Scheme 1. The key materials, cyclized isomeric alcohols **11** and **17**, were obtained

from 2,3,4,6-tetra-O-benzyl-D-galactopyranose according to a procedure described previously.<sup>17</sup> The alcohol **11** was treated with sodium methoxide to provide 12 smoothly, which was further treated with 50% aqueous potassium hydroxide to cleave the protecting group on the amino functionality, affording **13**.<sup>28</sup> The N-alkylated side chains were successfully introduced to the iminosugar moiety by reductive amination<sup>29,30</sup> leading to compounds **14**, 15. and 16 in 70%. 89%. and 65% vield, respectively, based on the recovered starting materials. Debenzylation of 14-16 under a hydrogen atmosphere over Pd-C provided iminosugars 3-5 in quantitative yield. In a similar way, iminosugar derivatives 6-8 were prepared from the corresponding p-galacto alcohol **17**. Interestingly, when the above-mentioned reductive amination conditions were applied to the preparation of compounds **20**, **21**, and **22** from **19**. the reaction gave a low yield (less than 40%). However, when using ethanol instead of methanol as the solvent, satisfactory yields (80-86%) were obtained. Finally, it is noteworthy that it was not easy to obtain pure samples of iminosugars 6 and 7. The crude product must be successively subjected to silica gel column chromatography (CH<sub>2</sub>Cl<sub>2</sub>-MeOH as the eluent), C-18 reverse-phase



n=2 (80%) 20 : 6 n=2 (90%) **21** : n=4 (86%) 7 n=4 (92%) 22 : 8 n=7 (100%) n=7 (80%) Scheme 1. Reagents and conditions: (a) NaOMe, MeOH, rt; (b) 50% KOH, MeOH, 80 °C; (c) CH<sub>3</sub>(CH<sub>2</sub>)<sub>n</sub>CHO, MeOH or EtOH, HOAc then NaBH<sub>3</sub>CN, 80 °C; (d) Pd/C, H<sub>2</sub>, H<sub>2</sub>O/THF/

column chromatography ( $H_2O$  as eluent), and ion-exchange resin column chromatography.

Starting from compound **13** and its isomer **19**, the miglitol derivatives 9 and 10 were obtained (Scheme 2). Initially, compound 13 was treated with BrCH<sub>2</sub>CH<sub>2</sub>OH and K<sub>2</sub>CO<sub>3</sub> in DMF in the hope of getting 24, but no product was obtained. Although many other bases and solvent systems such as THF/K<sub>2</sub>CO<sub>3</sub>, THF/ NaOH/tetra-n-butylammonium iodide (TBAI), and THF/Et<sub>3</sub>N/TBAI were evaluated, all these efforts failed to give the desired product. Eventually, treatment of 13 with 40% glyoxal aqueous solution afforded the lactone **23** in 74% isolated yield.<sup>31</sup> The carbonyl group of 23 was subsequently reduced by LiAlH<sub>4</sub>, providing tetra-O-benzyl-protected iminosugar derivative 24. It was noteworthy that Dgalacto iminosugar intermediates 25 and 26 were less stable than their corresponding L-altro isomers. Final deprotection of 24 by catalvtic hydrogenolysis vielded the target compound 9, which was purified by silica gel column chromatography. C-18 reverse-phase column chromatography, and ion-exchange resin. Iminosugar 10 was prepared in the same manner as described for the preparation of 9.

#### 2.3. Biology

With all the synthetic compounds in hand, the effect of the N-substituted iminosugars **3–10** and the N-unsubstituted iminosugars **1–2**<sup>17</sup> on concanavalin A (Con A)-induced proliferation of splenocytes in mice were assessed by the MTT assay.<sup>32</sup> The splenocytes were induced by 2.5 µg/mL of concanavalin A with 30 µM concentration of the iminosugars at 37 °C, 5% CO<sub>2</sub> for 48 h, using the Con A-treated splenocytes as the experimental control. Compared with the control, the levels of cell proliferation were reduced 25.4%, 16.5%, 21.6%, 17.9%, 26.6%, 35.8%, 38.3%, 27.7%, 18.4%, and 18.2% when including 30 µM of compounds **1–10**, respectively.

As shown in Figure 3, iminosugars **6** and **7** displayed the strongest inhibition effects. The N-alkylation of p-galacto iminosugar **2** was important for the improvement of the inhibitory activity and the immunosuppressive effects were increased by 1.7-2.3-fold (compared compounds **6–8** to **2**). The six-carbon side chain showed the best inhibitory effect among the three N-substituted iminosugars. However, in the case of the L-altro iminosugar epimers, N-alkylation did not improve the activity (compared compounds **3–5** to **1**). Comparing compounds **6–8** with **10**, it seemed that iminosugars with N-substituted non-polar side chains showed better inhibitory activity than that with polar chains. This suggests that the hydrophobic character may be important to the immunosuppressive activity. By comparison of the inhibition of the p-epimer **7** (38.3%) with L-epimer **4** (17.9%), it was clear that the



**Figure 3.** Effects of compounds **1–10** on concanavalin A-induced mouse splenocytes proliferation were assessed by the MTT assay. Values are mean  $\pm$  SEM. \*\*\*p <0.001 versus control.

configuration of the hydroxymethyl group was important to improve the inhibitory activity. However, the mechanisms of these differences are not understood.

### 3. Conclusions

N-Alkylated iminosugars **3–8** and miglitol derivatives **9** and **10** were designed and synthesized, and their immunosuppressive activities were evaluated. It was clear that N-alkylation of the D-galacto iminosugar gave rise to better inhibitory activities than that of the L-altro iminosugar epimer, and the D-galacto iminosugar derivative with N-substituted six-carbon side chain showed the strongest inhibitory activity. With this information, our future work will focus on the preparation and evaluation of more N-substituted D-galacto iminosugar derivatives.

### 4. Experimental

#### 4.1. General methods

All chemicals were purchased and used without further purification. Tetrahydrofuran (THF) was distilled over sodium/



Scheme 2. Reagents and conditions: (a) glyoxal, MeOH/HOAc (v/v) = 200:1, NaBH<sub>4</sub>, 80 °C; (b) LiAlH<sub>4</sub>, THF, rt; (c) Pd/C, H<sub>2</sub>, H<sub>2</sub>O/THF/HOAc = 4:2:1.

benzophenone and methylene chloride (CH<sub>2</sub>Cl<sub>2</sub>) over calcium hydride. The reactions were monitored with analytical TLC on Silica Gel 60- $F_{254}$ -precoated aluminum plates and visualized under UV (254 nm) and/or by staining with acidic ceric ammonium molybdate. Column chromatography was performed on silica gel (35–75 µm). NMR spectra were recorded on a Varian VXR-300M or Varian INOVA-500M spectrometer. Mass spectra were recorded using a PE SCLEX QSTAR spectrometer. Elemental analysis data were recorded on PE-2400C elemental analyzer. Concanavalin A was purchased from Sigma. Other reagents were from commercial sources.

### 4.2. 1,3,4,5-Tetra-O-benzyl-2,6-dideoxy-2,6-imino-D-glycero-Laltro-heptitol O (7), N-cyclic carbamate (12)

To a solution of  $11^{17}$  (210 mg, 0.31 mmol) in MeOH (15 mL) was added sodium methoxide (30% solution in MeOH) (0.5 mL. 0.009 mmol). The reaction mixture was stirred for 1 h at room temperature. MeOH was removed under reduced pressure, and the residue was extracted with EtOAc ( $3 \times 20$  mL). The organic phases were combined, dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether-EtOAc 4:1) to provide 12 (159 mg, 90%) as a yellow oil. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  3.42 (d, 1H, *J* = 3.0 Hz), 3.80–3.81 (m, 1H), 3.87–3.91 (m, 1H), 3.98 (dd, 1H, J = 2.5, 9.5 Hz), 4.11–4.18 (m, 3H), 4.27 (dd, 1H, J = 3.5, 10.0 Hz), 4.33-4.37 (m, 2H), 4.41 (d, 1H, J = 12.0 Hz,), 4.43 (d, 1H, J = 12.5 Hz), 4.47 (d, 1H, J = 12.0 Hz), 4.51 (d, 1H, J = 11.5 Hz), 4.66 (s, 2H), 4.74 (d, 1H, J = 12.0 Hz)  $(8 \times -CH_2Ph)$ , 7.19–7.43 (m, 20H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$ 54.53, 55.05, 62.06, 64.65, 72.14, 72.37, 73.00, 73.07, 73.15, 73.51  $(4 \times -CH_2Ph)$ , 74.58, 127.37, 127.75, 128.03, 128.09, 128.16, 128.36, 128.47, 137.30, 137.67, 137.92, 138.42 (Ph), 156.14 (CO). HRMS (ESI, positive) Calcd for C<sub>36</sub>H<sub>38</sub>NO<sub>6</sub>: 580.2694 [M+H]<sup>+</sup>. Found: 580.2705.

# 4.3. 3,4,5,7-Tetra-O-benzyl-2,6-dideoxy-2,6-imino-*D-glycero-L-altro*-heptitol (13)

To a solution of **12** (220 mg, 0.38 mmol) in MeOH (15 mL) was added 50% aqueous KOH (15 mL). The reaction mixture was stirred for 20 h at 80 °C. MeOH was removed under reduced pressure, and the residue was extracted with EtOAc ( $3 \times 50$  mL). The organic phases were combined, dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether-EtOAc 1:1) to provide 13 (190 mg, 90%) as a white solid. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  2.03 (br s, 2H, –OH and -NH), 3.15-3.21 (m, 2H, H-2, H-6), 3.50 (dd, 1H, J = 1.5, 3.5 Hz, H-5), 3.53 (dd, 1H, J = 5.5, 11.0 Hz, H-7), 3.61–3.66 (m, 2H, H-7, H-1), 3.77 (dd, 1H, J = 3.0, 10.0 Hz, H-3), 3.78-3.82 (m, 2H, H-1, H-4), 4.30 (d, 1H, J = 12.0 Hz), 4.37 (d, 1H, J = 12.5 Hz), 4.39 (br s, 2H), 4.48 (d, 1H, J = 12.0 Hz), 4.51 (d, 1H, J = 12.0 Hz), 4.56 (d, 1H, J = 12.0 Hz, 4.70 (d, 1H, J = 12.0 Hz), 7.16–7.35 (m, 20H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  54.17 (C-2), 54.79 (C-6), 63.59 (C-7), 69.91 (C-1), 71.79 (CH<sub>2</sub>Ph), 71.90 (CH<sub>2</sub>Ph), 72.26 (C-4), 72.93 (CH<sub>2</sub>Ph), 73.26 (CH<sub>2</sub>Ph), 74.93 (C-3), 76.75 (C-5), 127.50, 127.69, 127.81, 127.93, 127.98, 128.12, 128.30, 128.35, 137.90, 138.37, 138.42, 138.53 (Ph). HRMS (ESI, positive) Calcd for  $C_{35}H_{40}NO_5$ : 554.2901 [M+H]<sup>+</sup>. Found: 554.2900.

### 4.4. 3,4,5,7-Tetra-O-benzyl-N-butyl-2,6-dideoxy-2,6-imino-Dglycero-L-altro-heptitol(14)

To a solution of **13** (140 mg, 0.25 mmol) in MeOH and acetic acid (v/v 200:1, 10 mL) was added CH<sub>3</sub>(CH<sub>2</sub>)<sub>2</sub>CHO (160  $\mu$ L, 1.75 mmol). After stirring for 1 h at 60 °C, a portion of NaBH<sub>3</sub>CN (80 mg, 1.25 mmol) was added. The reaction mixture was stirred overnight at 80 °C and the reaction was quenched with 1 N HCl aqueous solu-

tion (0.2 mL). The mixture was extracted with EtOAc  $(3 \times 30 \text{ mL})$ and washed with aqueous NaHCO<sub>3</sub> (10 mL) and aqueous NaCl (10 mL). The organic phases were combined. dried (Na<sub>2</sub>SO<sub>4</sub>), filtered. and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether-EtOAc 4:1) to provide 14 (59 mg, 38%) as a yellow oil. The material 13 (63 mg) was recovered and the conversion was 70%. <sup>1</sup>H NMR (500 MHz,  $CDCl_3$ ):  $\delta$  0.85 (t, 3H, J = 7.5 Hz, -CH<sub>3</sub>), 1.15-1.45 (m, 4H, -CH<sub>2</sub>-), 2.81-2.93 (m, 2H, -NCH<sub>2</sub>-), 3.10 (q, 1H, J = 5.0, 11.0 Hz, H-2), 3.20 (q, 1H, J = 5.5, 10.5 Hz, H-6), 3.41–3.46 (m, 2H, H-1, H-7), 3.53 (dd, 1H, J = 4.5, 10.0 Hz, H-7), 3.65 (dd, 1H, J = 2.5, 7.0 Hz, H-4), 3.75 (dd, 1H, *J* = 6.5, 11.5 Hz, 1H), 3.88 (dd, 1H, *J* = 2.5, 5.5 Hz, H-5), 3.98 (dd, 1H, J = 4.5, 7.0 Hz, H-3), 4.41 (d, 1H, J = 12.0 Hz), 4.47-4.53 (m, 5H), 4.56 (d, 1H, J = 12.5 Hz), 4.61 (d, 1H, J = 12.0 Hz), 7.22-7.32 (m, 20H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  13.95 (-CH<sub>3</sub>), 20.21 (-CH<sub>2</sub>-), 28.05 (-CH<sub>2</sub>-), 53.08 (-NCH<sub>2</sub>-), 59.05 (C-2 and C-6), 61.26 (C-1), 68.94 (C-7), 71.92, 72.33, 72.91, 73.11 (4 × -CH<sub>2</sub>Ph), 74.09 (C-4), 75.30 (C-5), 77.00 (C-3), 127.46, 127.58, 127.63, 127.79, 127.86, 127.91, 128.03, 128.22, 128.25, 128.39, 137.98, 138.07, 138.48, 138.58. MS-ESI: 610 [M+H]<sup>+</sup>. Anal. Calcd for C<sub>39</sub>H<sub>47</sub>NO<sub>5</sub>: C, 76.82; H, 7.77; N, 2.30. Found: C, 77.06; H, 7.56; N, 2.36.

### 4.5. 3,4,5,7-Tetra-O-benzyl-N-hexyl-2,6-dideoxy-2,6-imino-Dglycero-L-altro-heptitol (15)

Compound 15 was prepared from 13 (500 mg, 0.9 mmol) as described in the preparation of 14, providing 15 (340 mg, 59%) as a yellow oil. The material 13 (170 mg) was recovered and the conversion was 89%. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  0.86 (t, 3H, J = 7.5 Hz, -CH<sub>3</sub>), 1.14-1.42 (m, 8H, -CH<sub>2</sub>-), 2.80-2.92 (m, 3H, -OH, -NCH<sub>2</sub>-), 3.10 (q, 1H, J=6.0, 10.5 Hz, H-2), 3.20 (q, 1H, J = 5.0, 10.5 Hz, H-6), 3.43 (dd, 1H, J = 5.5, 9.5 Hz, H-7), 3.40–3.47 (m, 1H, H-1), 3.54 (dd, 1H, J = 5.0, 10.0 Hz, H-7), 3.64 (dd, 1H, J = 2.5, 6.5 Hz, H-4), 3.74 (dd, 1H, J = 6.5, 11.0 Hz, 1H), 3.87 (dd, 1H, J = 3.0, 6.0 Hz, H-5), 3.97 (dd, 1H, J = 4.5, 7.0 Hz, H-3), 4.41 (d, 1H, / = 12.0 Hz), 4.47 (d, 1H, / = 11.5 Hz), 4.49–4.52 (m, 4H), 4.55 (d, 1H, / = 12.0 Hz), 4.59 (d, 1H, / = 11.5 Hz), 7.17-7.32 (m, 20H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): δ 14.00 (-CH<sub>3</sub>), 22.62, 25.77, 26.75, 31.68  $(4 \times -CH_2)$ , 53.27  $(-NCH_2)$ , 58.99 (C-6), 59.05 (C-2), 61.32 (C-1), 68.93 (C-7), 71.96, 72.38, 72.92, 73.15 (4 × -CH<sub>2</sub>Ph), 74.05 (C-4), 75.31 (C-5), 77.00 (C-3), 127.50, 127.62, 127.67, 127.84, 127.91, 127.94, 128.07, 128.26, 128.42, 138.00, 138.07, 138.49, 138.60 (Ph). HRMS (ESI, positive) Calcd for C<sub>41</sub>H<sub>52</sub>NO<sub>5</sub>: 638.3840 [M+H]<sup>+</sup>. Found: 638.3843.

### 4.6. 3,4,5,7-Tetra-O-benzyl-N-nonyl-2,6-dideoxy-2,6-imino-Dglycero-L-altro-heptitol (16)

Compound 16 was prepared from 13 (390 mg, 0.71 mmol) as described in the preparation of 14, providing 16 (190 mg, 40%) as a yellow oil. The material 13 (150 mg) was recovered and the conversion was 65%. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  0.88 (t, 3H, J = 7.2 Hz, -CH<sub>3</sub>), 1.23-1.27 (m, 14H, -CH<sub>2</sub>-), 2.82-2.89 (m, 2H, -NCH<sub>2</sub>-), 3.10 (q, 1H, J = 5.1 Hz, H-2), 3.19 (q, 1H, J = 5.4 Hz, H-6), 3.40-3.47 (m, 2H, H-7, H-1), 3.54 (dd, 1H, J = 4.8, 9.9 Hz, H-7), 3.65 (dd, 1H, J = 2.7, 6.9 Hz, H-4), 3.74 (dd, 1H, J = 6.6, 11.4 Hz, H-1), 3.87 (dd, 1H, J = 2.7, 6.0 Hz, H-5), 3.96 (dd, 1H, J = 4.5, 6.9 Hz, H-3), 4.42 (d, 1H, J = 12.0 Hz), 4.44–4.55 (m, 6H), 4.60 (d, 1H, I = 11.4 Hz), 7.23–7.30 (m, 20H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$ 14.09 (-CH<sub>3</sub>), 22.64, 25.70, 27.11, 29.24, 29.52, 29.59, 31.85  $(7 \times -CH_2-)$ , 53.14 (-NCH<sub>2</sub>-), 58.91 (C-6), 58.98 (C-2), 61.35 (C-1), 68.85 (C-7), 71.94, 72.38, 72.89, 73.14 (4 × -CH<sub>2</sub>Ph), 73.98 (C-4), 75.29 (C-5), 77.09 (C-3), 127.51, 127.68, 127.92, 128.09, 128.26, 128.43, 138.00, 138.05, 138.48, 138.59 (Ph). MS-ESI: 680 [M+H]<sup>+</sup>. Anal Calcd for C<sub>44</sub>H<sub>57</sub>NO<sub>5</sub>: C, 77.72; H, 8.45; N, 2.06. Found: C, 77.88; H, 8.67; N, 1.97.

#### 4.7. N-Butyl-2,6-dideoxy-2,6-imino-p-glycero-L-altro-heptitol (3)

A mixture of **14** (43 mg, 0.07 mmol) and 10% Pd-C (10.0 mg) in acetic acid (1.0 mL), H<sub>2</sub>O (4.0 mL), and THF (2.0 mL) was stirred for 48 h under a H<sub>2</sub> atmosphere. The solid was removed by filtration through Celite and the filtrate was concentrated. The residue was passed through a C-18 reverse-phase column chromatography (H<sub>2</sub>O as eluent) and ion-exchange resin (Dowex  $1 \times 8$ , OH<sup>-</sup> form) to give 3 (17.5 mg, 100%) as white solids after lyophilization.  $[\alpha]_{D}^{26}$  -21.3 (c 0.6, H<sub>2</sub>O). <sup>1</sup>H NMR (500 MHz, D<sub>2</sub>O):  $\delta$  0.89 (t, 3H, J = 7.5 Hz, -CH<sub>3</sub>), 1.20-1.28 (m, 2H, -CH<sub>2</sub>-), 1.40-1.46 (m, 2H, -CH<sub>2</sub>-), 2.64-2.70 (m, 1H, -NCH<sub>2</sub>-), 2.74 (td, 1H, J = 2.5, 9.0 Hz, H-2), 2.78-2.85 (m, 1H, -NCH2-), 2.95-2.98 (m, 1H, H-6), 3.75 (dd, 1H, J = 7.0, 12.5 Hz, H-7), 3.80–3.85 (m, 3H, 2H-1, H-7), 3.87–3.90 (m, 1H, H-3), 3.92 (dd, 1H, J = 3.5, 6.5 Hz, H-4), 3.98 (dd, 1H, I = 2.5, 4.5 Hz, H-5). <sup>13</sup>C NMR (125 MHz, D<sub>2</sub>O):  $\delta$  13.61 (-CH<sub>3</sub>), 19.96 (-CH<sub>2</sub>-), 24.02 (-CH<sub>2</sub>-), 48.82 (-NCH<sub>2</sub>-), 55.75 (C-6), 59.59 (C-1), 60.14 (C-2), 61.03 (C-7), 63.86 (C-3), 69.57 (C-5), 70.02 (C-4). HRMS (ESI, positive) Calcd for C<sub>11</sub>H<sub>23</sub>NO<sub>5</sub>Na: 272.1468 [M+Na]<sup>+</sup>. Found: 272.1472.

#### 4.8. N-Hexyl-2,6-dideoxy-2,6-imino-D-glycero-L-altro-heptitol (4)

Compound **4** was prepared from **15** (63 mg, 0.1 mmol) as described in the preparation of **3**, providing **4** (27 mg, 100%) as white solids.  $[\alpha]_D^{26} - 22.6$  (*c* 0.8, H<sub>2</sub>O). <sup>1</sup>H NMR (500 MHz, D<sub>2</sub>O):  $\delta$  0.86 (t, 3H, *J* = 7.0 Hz, -CH<sub>3</sub>), 1.22–1.30 (m, 6H, -CH<sub>2</sub>–), 1.43–1.48 (m, 2H, -CH<sub>2</sub>–), 2.64–2.70 (m, 1H, -NCH<sub>2</sub>–), 2.74 (td, 1H, *J* = 3.0, 8.5 Hz, H-2), 2.79–2.85 (m, 1H, -NCH<sub>2</sub>–), 2.96–2.99 (m, 1H, H-6), 3.75 (dd, 1H, *J* = 7.0, 11.5 Hz, H-7), 3.80–3.88 (m, 3H, 2H-1, H-7), 3.89 (dd, 1H, *J* = 3.0, 8.5 Hz, H-3), 3.91–3.94 (m, 1H, H-4), 3.98 (dd, 1H, *J* = 2.0, 4.0 Hz, H-5). <sup>13</sup>C NMR (125 MHz, D<sub>2</sub>O):  $\delta$  14.04 (-CH<sub>2</sub>–), 21.68, 22.72, 26.86, 31.64 (4 × -CH<sub>2</sub>–), 48.21 (-NCH<sub>2</sub>–), 57.59 (C-6), 58.86 (C-1), 60.16 (C-2), 61.16 (C-7), 66.51 (C-3), 70.66 (C-5), 70.94 (C-4). HRMS (ESI, positive) Calcd for C<sub>13</sub>H<sub>27</sub>NO<sub>5</sub>Na: 300.1781 [M+Na]<sup>+</sup>. Found: 300.1786.

#### 4.9. N-Nonyl-2,6-dideoxy-2,6-imino-D-glycero-L-altro-heptitol (5)

Compound **5** was prepared from **16** (77 mg, 0.11 mmol) as described in the preparation of **3**, providing **5** (36 mg, 100%) as white solids.  $[\alpha]_D^{26} - 22.8$  (*c* 0.8, H<sub>2</sub>O). <sup>1</sup>H NMR (500 MHz, D<sub>2</sub>O):  $\delta$  0.86 (t, 3H, *J* = 7.0 Hz, -CH<sub>3</sub>), 1.27-1.37 (m, 12H, -CH<sub>2</sub>-), 1.71-1.80 (m, 2H, -CH<sub>2</sub>-), 3.30-3.47 (m, 3H, 2 × -NCH<sub>2</sub>-, H-2), 3.72-3.74 (m, 1H, H-6), 3.97 (dd, 1H, *J* = 4.5, 13.0 Hz, H-7), 4.02-4.09 (m, 4H, H-7, H-3, 2 × H-1), 4.19-4.23 (m, 2H, H-4, H-5). <sup>13</sup>C NMR (125 MHz, D<sub>2</sub>O):  $\delta$  14.12 (-CH<sub>3</sub>), 20.69, 22.75, 26.14, 28.84, 28.99, 29.11, 31.82 (7 × -CH<sub>2</sub>-), 49.24 (-NCH<sub>2</sub>-), 55.09 (C-6), 59.94 (C-1), 60.08 (C-2), 61.13 (C-7), 63.38 (C-3), 69.28 (C-5), 69.95 (C-4). HRMS (ESI, positive) Calcd for C<sub>16</sub>H<sub>34</sub>NO<sub>5</sub>: 320.2431 [M+H]<sup>+</sup>. Found: 320.2430.

### 4.10. 1,3,4,5-Tetra-O-benzyl-2,6-dideoxy-2,6-imino-*D*-*glycero*-*D*-*galacto*-heptitol *O* (7), *N*-cyclic carbamate (18)

Compound **18** was prepared from **17** (210 mg, 0.71 mmol) as described in the preparation of **12**, providing **18** (159 mg, 90%) as a yellow oil. The data were in good agreement with those reported by Martin et al.<sup>28</sup> <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  3.34 (dd, 1H, *J* = 2.5, 4.0 Hz), 3.82–3.83 (m, 1H), 3.88 (dd, 1H, *J* = 3.0, 10.5 Hz), 3.94 (dd, 2H, *J* = 2.5, 6.5 Hz), 4.02 (t, 1H, *J* = 10.5 Hz), 4.09 (dd, 1H, *J* = 4.0, 8.5 Hz), 4.13 (ABq, 1H, *J* = 7.5 Hz, H-1), 4.32 (d, 1H, *J* = 12.0 Hz), 4.40 (d, 1H, *J* = 13.0 Hz), 4.43–4.46 (m, 3H), 4.55–4.57 (m, 1H), 4.59 (d, 1H, *J* = 12.0 Hz), 4.62 (d, 1H, *J* = 12.0 Hz), 4.70 (d, 1H, *J* = 12.0 Hz), 7.12–7.35 (m, 20H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  49.42, 51.51, 62.69, 65.41, 71.42, 72.15, 72.19, 73.11, 73.62, 75.09, 127.38, 127.52, 127.70, 127.81, 127.98, 128.13, 128.28,

128.30, 128.44, 137.00, 137.67, 137.85, 138.29, 157.60 (CO). MS-ESI: 580 (M+H<sup>+</sup>). Anal Calcd for  $C_{36}H_{37}NO_6$ : C, 74.59; H, 6.43; N, 2.42. Found: C, 74.39; H, 6.58; N, 2.26.

### 4.11. 3,4,5,7-Tetra-O-benzyl-2,6-dideoxy-2,6-imino-*D*-*glycero*-*D*-*galacto*-heptitol (19)

Compound 19 was prepared from 18 (220 mg, 0.38 mmol) as described in the preparation of 13, providing 19 (178 mg, 85%) as a yellow oil. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  2.07 (br s, 2H, –OH, – NH-), 3.06 (br s, 1H, H-6), 3.27-3.31 (m, 1H, H-2), 3.45 (t, J = 8.5 Hz, 1H, H-7), 3.57–3.61 (m, 2H, H-7, H-1), 3.64 (dd, 1H, *J* = 2.5, 9.0 Hz, H-3), 3.75 (dd, 1H, *J* = 5.5, 11.0 Hz, H-1), 3.99–4.01 (m, 2H, H-5, H-4), 4.47 (s, 2H), 4.54 (d, 1H, J = 11.5 Hz), 4.58 (d, 1H, J = 12.0 Hz), 4.66 (d, 1H, J = 12.5 Hz), 4.68 (d, 1H, J = 12.5 Hz), 4.73 (d, 1H, J = 11.5 Hz), 4.84 (d, 1H, J = 11.0 Hz) (8 × -CH<sub>2</sub>Ph), 7.24-7.33 (m, 20H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): δ 52.25 (C-6), 53.85 (C-2), 58.84 (C-1), 68.91 (C-7), 72.83, 73.12, 73.33, 73.78  $(4 \times -CH_2Ph)$ , 75.20 (C-5), 77.61 (C-4), 79.36 (C-3), 127.40, 127.46, 127.61, 127.73, 127.84, 127.86, 127.95, 128.27, 128.31, 128.38, 128.42, 138.06, 138.32, 138.71, 138.74. MS-ESI: 554  $[M+H]^{+}$ . Anal Calcd for C<sub>35</sub>H<sub>39</sub>NO<sub>5</sub>: C, 75.92; H, 7.10; N, 2.53. Found: C, 75.67; H, 7.08; N, 2.45.

### 4.12. 3,4,5,7-Tetra-O-benzyl-*N*-butyl-2,6-dideoxy-2,6-imino-Dglycero-D-galacto-heptitol (20)

Compound 20 was prepared from 19 (100 mg, 0.18 mmol) as described in the preparation of 14 with the exception that ethanol was used in the reaction mixture instead of MeOH, providing **20** (88 mg, 80%) as a yellow oil. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$ 0.85 (t, 3H, J = 7.0 Hz, -CH<sub>3</sub>), 1.14-1.29 (m, 4H, -CH<sub>2</sub>-), 2.56-2.67 (m, 2H, -NCH2-), 2.82 (br s, 1H, -OH), 2.93 (br s, 1H, H-6), 3.22-3.26 (m, 1H, H-2), 3.52-3.56 (m, 2H, H-1, H-7), 3.59 (dd, 1H, J = 2.5, 9.5 Hz, H-4), 3.66 (t, 1H, J = 8.0 Hz, H-7), 3.72 (dd, 1H, J = 5.5, 10.5 Hz, H-1), 4.07 (br s, 1H, H-5), 4.18 (br s, 1H, H-3), 4.35 (d, 1H, *J* = 12.0 Hz), 4.41 (d, 1H, *J* = 11.5 Hz), 4.46 (d, 1H, J = 12.0 Hz), 4.62 (d, 1H, J = 11.5 Hz), 4.71–4.76 (m, 3H), 4.96 (d, 1H, I = 11.5 Hz) (8 × -CH<sub>2</sub>Ph), 7.22-7.36 (m, 20H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  13.97 (-CH<sub>3</sub>), 20.19 (-CH<sub>2</sub>-), 32.55 (-CH<sub>2</sub>-), 49.96 (-NCH<sub>2</sub>-), 54.97 (C-6), 56.27 (C-1), 59.43 (C-2), 68.64 (C-7), 72.78, 73.26, 73.38  $(3 \times -CH_2Ph)$ , 74.34 (C-3), 74.58 (-CH<sub>2</sub>Ph), 76.74 (C-5), 80.74 (C-4), 127.23, 127.30, 127.34, 127.40, 127.59, 127.75, 128.16, 128.30, 128.38, 137.96, 138.49, 138.79, 139.28. MS-ESI: 610 [M+H]<sup>+</sup>. Anal. Calcd for C<sub>39</sub>H<sub>47</sub>NO<sub>5</sub>: C, 76.82; H, 7.77; N, 2.30. Found: C, 76.64; H, 7.91; N, 2.20.

### 4.13. 3,4,5,7-Tetra-O-benzyl-*N*-hexyl-2,6-dideoxy-2,6-imino-*D*-glycero-*D*-galacto-heptitol (21)

Compound **21** was prepared from **19** (31 mg, 0.06 mmol) as described in the preparation of **20**, providing **21** (31 mg, 86%) as a yellow oil. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  0.87 (t, 3H, *J* = 7.5 Hz, -CH<sub>3</sub>), 1.19–1.28 (m, 8H, -CH<sub>2</sub>–), 2.55–2.66 (m, 2H, -CH<sub>2</sub>–), 2.93 (br s, 1H, H-6), 3.21–3.26 (m, 1H, H-2), 3.50–3.56 (m, 2H, H-1, H-7), 3.59 (dd, 1H, *J* = 3.0, 9.5 Hz, H-4), 3.65 (t, 1H, *J* = 8.5 Hz, H-7), 3.71 (dd, 1H, *J* = 5.5, 11.0 Hz, H-1), 4.06–4.07 (m, 1H, H-5), 4.17 (br s, 1H, H-3), 4.36 (d, 1H, *J* = 11.5 Hz), 4.41 (d, 1H, *J* = 12.0 Hz), 4.46 (d, 1H, *J* = 11.5 Hz), 4.62 (d, 1H, *J* = 11.5 Hz), 4.71–4.76 (m, 3H), 4.96 (d, 1H, *J* = 12.0 Hz) (8 × -CH<sub>2</sub>Ph), 7.22–7.36 (m, 20H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  14.05 (-CH<sub>3</sub>), 22.59, 26.70, 30.32, 31.70 (4 × -CH<sub>2</sub>–), 50.24 (-NCH<sub>2</sub>–), 54.98 (C-6), 56.28 (C-1), 59.47 (C-2), 68.63 (C-7), 72.77, 73.25, 73.36 (3 × -CH<sub>2</sub>Ph), 74.40 (C-3), 74.57 (-CH<sub>2</sub>Ph), 76.75 (C-5), 80.75 (C-4), 127.22, 127.28, 127.33, 127.38, 127.58, 127.73, 128.14, 128.28, 128.37, 137.95, 138.49,

138.78, 139.28. MS-ESI: 638  $[M+H]^+$ . Anal. Calcd for  $C_{41}H_{51}NO_5$ : C, 77.20; H, 8.06; N, 2.20. Found: C, 76.97; H, 8.06; N, 2.10.

### 4.14. 3,4,5,7-Tetra-O-benzyl-*N*-nonyl-2,6-dideoxy-2,6-imino-Dglycero-D-galacto-heptitol (22)

Compound 22 was prepared from 19 (80 mg, 0.14 mmol) as described in the preparation of 20, providing 22 (78 mg, 80%) as a yellow oil. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  0.89 (t, 3H, J = 7.0 Hz, -CH<sub>3</sub>), 1.13-1.31 (m, 14H, -CH2-), 2.55-2.66 (m, 2H, -NCH2-), 2.93 (br s, 1H, H-6), 3.21-3.26 (m, 1H, H-2), 3.52-3.56 (m, 2H, H-1, H-7), 3.59 (dd, 1H, J = 3.0, 10.0 Hz, H-4), 3.65 (t, 1H, J = 8.5 Hz, H-7), 3.71 (dd, 1H, J = 5.5, 11.0 Hz, H-1), 4.06 (br s, 1H, H-5), 4.18 (br s, 1H, H-3), 4.36 (d, 1H, J = 12.0 Hz), 4.41 (d, 1H, J = 11.5 Hz), 4.46 (d, 1H, J = 12.0 Hz), 4.62 (d, 1H, J = 11.5 Hz), 4.72–4.76 (m, 3H), 4.96 (d, 1H, J = 11.5 Hz) (8 × -CH<sub>2</sub>Ph), 7.23-7.30 (m, 20H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  14.09 (-CH<sub>3</sub>), 22.65, 27.07, 29.28, 29.54, 29.57, 30.38, 31.85 (7 × -CH<sub>2</sub>-), 50.25 (-NCH<sub>2</sub>-), 54.97 (C-6), 56.31 (C-1), 59.49 (C-2), 68.65 (C-7), 72.79, 73.25, 73.37 (3 × -CH<sub>2</sub>Ph), 74.45 (C-3), 74.58 (-CH<sub>2</sub>Ph), 76.75 (C-5), 80.77 (C-4), 127.23, 127.30, 127.34, 127.40, 127.59, 127.75, 128.15, 128.30, 128.38, 137.96, 138.49, 138.79, 139.29. HRMS (ESI, positive) Calcd for C<sub>44</sub>H<sub>58</sub>NO<sub>5</sub>: 680.4310 [M+H]<sup>+</sup>. Found: 680.4302.

### 4.15. *N*-Butyl-2,6-dideoxy-2,6-imino-*D*-glycero-*D*-galacto-heptitol (6)

Compound **6** was prepared from **20** (73 mg, 0.12 mmol) as described in the preparation of **3**, providing **6** (26.8 mg, 90%) as a white solid.  $[\alpha]_{26}^{26}$  +26.8 (*c* 0.8, H<sub>2</sub>O). <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O):  $\delta$  0.73 (t, 3H, *J* = 7.2 Hz, -CH<sub>3</sub>), 1.06–1.18 (m, 2H, -CH<sub>2</sub>–), 1.22–1.28 (m, 2H, -CH<sub>2</sub>–), 2.46–2.64 (m, 2H, -NCH<sub>2</sub>–), 2.73–2.77 (m, 1H, H-2), 3.09–3.18 (m, 1H, H-6), 3.49 (dd, 1H, *J* = 3.6, 9.9 Hz, H-4), 3.62–3.74 (m, 4H, H-7, H-1), 3.86 (br s, 1H, H-5), 3.93 (dd, 1H, *J* = 5.7, 9.9 Hz, H-3). <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O):  $\delta$  15.92 (-CH<sub>3</sub>), 22.58, 33.28 (2 × -CH<sub>2</sub>–), 51.51 (-NCH<sub>2</sub>–), 58.32 (C-6), 60.40 (C-2), 62.71 (C-7), 63.24 (C-1), 69.60 (C-3), 72.75 (C-5), 73.73 (C-4). HRMS (ESI, positive) Calcd for C<sub>11</sub>H<sub>23</sub>NO<sub>5</sub>Na: 272.1468 [M+Na]<sup>+</sup>. Found: 272.1466.

### 4.16. *N*-Hexyl-2,6-dideoxy-2,6-imino-*D*-glycero-*D*-galacto-heptitol (7)

Compound **7** was prepared from **21** (30 mg, 0.05 mmol) as described in the preparation of **3**, providing **7** (12 mg, 92%) as white solids.  $[\alpha]_D^{26}$  +30.5 (*c* 0.8, H<sub>2</sub>O). <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O):  $\delta$  0.70 (t, 1H, *J* = 6.3 Hz, -CH<sub>3</sub>), 1.13–1.14 (m, 6H, -CH<sub>2</sub>–), 1.39 (br s, 2H, -CH<sub>2</sub>–), 2.76–2.78 (m, 2H, -NCH<sub>2</sub>–), 3.02 (br s, 1H, H-2), 3.31–3.33 (m, 1H, H-6), 3.56 (dd, 1H, *J* = 3.0, 9.6 Hz, H-4), 3.65–3.74 (m, 3H, H-7, H-1), 3.78 (dd, 1H, *J* = 6.6, 11.7 Hz, H-1), 3.94 (br s, 1H, H-5), 3.98 (dd, 1H, *J* = 5.7, 9.9 Hz, H-3). <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O):  $\delta$  15.92 (-CH<sub>3</sub>), 24.54, 28.62, 30.14, 33.49 (4 × -CH<sub>2</sub>–), 52.20 (-NCH<sub>2</sub>–), 58.05 (C-6), 61.18 (C-2), 62.01 (C-7), 63.56 (C-1), 69.25 (C-3), 72.25 (C-5), 73.26 (C-4). HRMS (ESI, positive) Calcd for C<sub>13</sub>H<sub>28</sub>NO<sub>5</sub>: 278.1962 [M+H]<sup>+</sup>. Found: 278.1962.

### 4.17. *N*-Nonyl-2,6-dideoxy-2,6-imino-*D*-glycero-*D*-galacto-heptitol (8)

Compound **8** was prepared from **22** (43 mg, 0.06 mmol) as described in the preparation of **3**, providing **8** (20 mg, 100%) as a white solid.  $[\alpha]_D^{26}$  -10.2 (*c* 0.4, H<sub>2</sub>O). <sup>1</sup>H NMR (500 MHz, D<sub>2</sub>O):  $\delta$  0.86 (t, 3H, *J* = 6.0 Hz, -CH<sub>3</sub>), 1.23-1.32 (m, 12H, -CH<sub>2</sub>-), 1.60 (br s, 2H, -CH<sub>2</sub>-), 3.07 (d, 2H, *J* = 7.5 Hz, -NCH<sub>2</sub>-), 3.33-3.34 (m, 1H, H-2), 3.56-3.57 (m, 1H, H-6), 3.78 (d, 1H, *J* = 7.0 Hz, H-4), 3.84-3.94 (m, 4H, H-1, H-7), 4.16-4.20 (m, 2H, H-3, H-5). <sup>13</sup>C NMR

 $\begin{array}{l} (125 \text{ MHz}, \text{ D}_2\text{O}): \delta \ 14.38 \ (-\text{CH}_3), 23.09, 27.16, 28.02, 29.62, 29.71, \\ 29.89, 32.29 \ (7 \times -\text{CH}_2-), 51.01 \ (-\text{NCH}_2-), 55.92 \ (\text{C-6}), 59.28 \ (\text{C-2}), 60.18 \ (\text{C-7}), 66.79 \ (\text{C-1}), 68.86 \ (\text{C-3}), 67.00 \ (\text{C-5}), 70.96 \ (\text{C-4}). \\ \text{HRMS} \ (\text{ESI, positive}) \ \text{Calcd} \ \text{for} \ C_{16}\text{H}_{34}\text{NO}_5: \ 320.2431 \ [\text{M+H}]^{+}. \\ \text{Found: } 320.2427. \end{array}$ 

### 4.18. 1,3,4,5-Tetra-O-benzyl-2,6-dideoxy-2,6-imino-D-glycero-Laltro-heptitol-lactone (23)

To a solution of 13 (278 mg, 0.50 mmol) in MeOH and acetic acid (v/v 200:1, 10 mL) was added 40% glyoxal aqueous solution (86 μL, 1.88 mmol). After stirring for 1 h at 60 °C, a portion of NaBH<sub>4</sub> (38 mg, 1.0 mmol) was added. The reaction mixture was stirred overnight at 80 °C and the reaction was guenched by the addition of 1 N HCl aqueous solution (0.2 mL). The mixture was extracted with EtOAc  $(3 \times 50 \text{ mL})$  and washed with aqueous NaHCO<sub>3</sub> (10 mL) and aqueous NaCl (10 mL). The organic phases were combined, dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether-EtOAc 4:1) to provide 23 (221 mg, 74%) as a yellow oil. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  2.69 (td, 1H, I = 2.5, 10.5 Hz), 2.91 (td, 1H, / = 2.5, 10.5 Hz), 3.07 (d, 1H, / = 18.0 Hz), 3.36 (dd, 1H, / = 2.5, 4.0 Hz), 3.63–3.71 (m, 3H), 3.84 (dd, 1H, J = 2.5, 10.0 Hz), 3.93 (dd, 1H, /= 3.0, 11.0 Hz), 4.05 (d, 1H, /= 17.5 Hz), 4.19 (d, 1H, J = 12.5 Hz), 4.31 (d, 1H, J = 11.5 Hz), 4.33 (d, 1H, J = 12.5 Hz), 4.36 (d, 1H, J = 11.5 Hz), 4.44 (d, 1H, J = 12.5 Hz), 4.48 (d, 1H, J = 10.5 Hz), 4.49 (d, 1H, J = 12.5 Hz), 4.57 (d, 1H, J = 12.0 Hz), 4.67 (d, 1H, J = 12.5 Hz), 7.10-7.33 (m, 20H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): δ 53.56, 53.65, 61.72, 65.98, 70.04, 70.82, 72.18, 72.40, 73.09, 73.23, 73.42, 73.63, 127.79, 127.84, 127.87, 128.20, 128.32, 128.40, 128.43, 128.46, 128.49, 128.57, 137.11, 137.83, 137.89, 138.16, 167.41 (CO). MS-ESI: 594 [M+H]<sup>+</sup>. Anal. Calcd for C<sub>37</sub>H<sub>39</sub>NO<sub>6</sub>: C, 74.85; H, 6.62; N, 2.36. Found: C, 74.62; H, 6.55; N, 2.21.

### 4.19. 3,4,5,7-Tetra-O-benzyl-*N*-hydroxyethyl-2,6-dideoxy-2,6imino-*D*-glycero- *L*-altro-heptitol (24)

To a solution of **23** (133 mg, 0.22 mmol) in dry THF (8 mL) was added LiAlH<sub>4</sub> (70 mg, 1.76 mmol) at -15 °C under argon and the mixture was stirred for 5 h at room temperature. The reaction mixture was slowly added EtOAc (3 mL) and 1 N NaOH (1 mL) at 0 °C and was extracted with EtOAc ( $3 \times 50$  mL) and washed with aqueous NaHCO<sub>3</sub> (10 mL) and aqueous NaCl (10 mL). The organic phases were combined, dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether-EtOAc 2:1) to provide 24 (76 mg, 57%) as a yellow oil. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$ : 2.96 (td, 1H, J = 5.5, 14.5 Hz, -NCH<sub>2</sub>-), 3.09 (td, 1H, J = 4.0, 15.0 Hz, -NCH<sub>2</sub>-), 3.26 (q, 1H, J = 6.5 Hz, H-6), 3.33–3.36 (m, 1H, H-2), 3.43 (dd, 1H, J = 5.0, 10.0 Hz, H-7), 3.47-3.56 (m, 4H, 2H-1, 2 × CH<sub>2</sub>OH), 3.68 (dd, 1H, J = 2.5, 7.5 Hz, H-4), 3.72 (dd, 1H, J = 6.5, 11.5 Hz, H-7), 3.79 (dd, 1H, J = 3.0, 5.0 Hz, H-3), 4.03 (dd, 1H, J = 5.0, 7.5 Hz, H-5), 4.44 (d, 1H, J = 12.0 Hz), 4.48–4.60 (m, 6H), 4.62 (d, 1H, J = 11.5 Hz) (8 × – CH<sub>2</sub>Ph), 7.24–7.39 (m, 20H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  55.77 (-NCH2-), 60.46 (-CH2OH), 60.98 (C-2), 61.07 (C-6), 61.96 (C-7), 69.83 (C-1), 72.26, 72.63, 73.22, 73.35 (4 × -CH<sub>2</sub>Ph), 74.56 (C-4), 74.92 (C-3), 77.37 (C-5), 127.60, 127.64, 127.69, 127.77, 127.82, 127.88, 127.94, 128.33, 128.34, 128.45, 137.43, 138.06, 138.11, 138.43. HRMS (ESI, positive) Calcd for C<sub>37</sub>H<sub>44</sub>NO<sub>6</sub>: 598.3163 [M+H]<sup>+</sup>. Found: 598.3170.

## 4.20. *N*-Hydroxyethyl-2,6-dideoxy-2,6-imino-*D*-*glycero*-*L*-*altro*-heptitol (9)

Compound **9** was prepared from **24** (43 mg, 0.07 mmol) as described in the preparation of **3**, providing **9** (15.3 mg, 90%) as a

white solid.  $[\alpha]_D^{26}$  –9.0 (*c* 0.8, H<sub>2</sub>O). <sup>1</sup>H NMR (500 MHz, D<sub>2</sub>O)  $\delta$ : 2.84–2.87 (m, 1H, H-2), 2.98 (t, 2H, *J* = 6.5 Hz, -NCH<sub>2</sub>–), 3.06 (dt, 1H, *J* = 3.0, 6.5 Hz, H-6), 3.66 (t, 2H, *J* = 6.0 Hz, 2 × CH<sub>2</sub>OH), 3.74– 3.83 (m, 4H, 2H-7 and 2H-1), 3.90 (dd, 1H, *J* = 3.5, 6.0 Hz, H-3), 3.93 (dd, 1H, *J* = 3.0, 7.5 Hz, H-4), 4.01 (dd, 1H, *J* = 3.0, 5.5 Hz, H-5). <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O)  $\delta$ : 53.54 (–NCH<sub>2</sub>–), 61.38 (–CH<sub>2</sub>OH), 61.76 (C-6), 62.07 (C-1), 63.20 (C-2), 64.58 (C-7), 68.85 (C-3), 72.16 (C-5), 72.37 (C-4). HRMS (ESI, positive) Calcd for C<sub>9</sub>H<sub>19</sub>NO<sub>6</sub>-Na: 260.1105 [M+Na]<sup>+</sup>. Found: 260.1102.

# 4.21. *N*-Hydroxyethyl-2,6-dideoxy-2,6-imino-*D*-*glycero*-*D*-*galacto*-heptitol (10)

Compound **10** was prepared from **19** (90 mg, 0.16 mmol) as described in the preparation of **23**, providing yellow crude product (64 mg) after column chromatography. The product was treated under the conditions as described in the preparation of **24**, yielding crude product (53 mg). Finally, catalytic hydrogenolysis of the crude product using the method in the preparation of **9** provided **10** (21 mg, 55%, three steps) as white solids.  $[\alpha]_D^{26}$  +33.8 (*c* 0.8, H<sub>2</sub>O). <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O):  $\delta$  2.54–2.62 (m, 1H, –NCH<sub>2</sub>–), 2.73–2.89 (m, 2H, H-2, –NCH<sub>2</sub>–), 2.99–3.05 (m, 1H, H-6), 3.32–3.44 (m, 3H, H-4, –CH<sub>2</sub>OH), 3.49–3.55 (m, 3H, H-7, H-1), 3.64 (dd, 1H, *J* = 7.8, 10.8 Hz, H-1), 3.77 (br s, 1H, H-5), 3.84 (dd, 1H, *J* = 5.4, 9.3 Hz, H-3). <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O):  $\delta$  53.15 (–NCH<sub>2</sub>–), 58.82 (C-6), 59.74 (C-2), 62.17 (C-7), 62.77 (CH<sub>2</sub>OH), 64.05 (C-1), 69.56 (C-3), 73.10 (C-5), 74.12 (C-4). HRMS (ESI, positive) Calcd for C<sub>9</sub>H<sub>19</sub>NO<sub>6</sub>Na: 260.1105 [M+Na]<sup>+</sup>. Found: 260.1101.

### 4.22. Cell proliferation assay

Male BALB/c mouse splenocytes  $(4 \times 10^5$  cells per well), retreated with Con A alone (2.5 µg/mL concentration) or along with 30 µM concentration of synthetic iminosugar compounds, were incubated at 37 °C for 48 h under 5% CO<sub>2</sub> in a RPMI-1640 medium containing 10% fetal bovine serum (FBS). The proliferation of the mouse splenocytes was assayed using the MTT reduction method. MTT (5 mg/mL, 20 µL) dissolved in phosphate buffer solution (PBS) was added to each well and the plates were incubated for 4 h at 37 °C. The 96-well microplates were centrifuged at 1500g for 6 min and the media were aspirated. The resultant crystals were dissolved in 150 µL of DMSO for 10 min. Optical density was measured using a Microplate Reader at 570 nm. All data were presented as mean ± SEM. One-way ANOVA followed by Bonferroni was used to test the difference between groups. Statistical significance was indicated by p < 0.05.

### Acknowledgment

This work was financially supported by the National Natural Science Foundation of China.

### Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.carres.2010.01.021.

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