## Novel 5'-deoxy nucleosyl amino acid scaffolds for the synthesis of muraymycin analogues†

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Naturally occurring nucleoside antibiotics such as muraymycins represent promising lead structures for the development of novel antibacterial agents. A concise synthesis of 5'-deoxy muraymycin derivatives has been developed. The key step was the highly stereoselective asymmetric hydrogenation of suitable didehydro amino acid precursors, providing unique nucleosyl amino acid structures.

Due to emerging resistances of bacteria towards established antibiotics,<sup>1</sup> the development of novel antibacterial agents has become one of the main objectives of medicinal chemistry in recent years. Ideally, drug candidates with antimicrobial potency should display new or yet unexploited modes of action.<sup>2</sup> This is the case for the inhibition of the bacterial membrane protein translocase I (MraY), a key enzyme in the early stages of peptidoglycan biosynthesis,<sup>3,4</sup> which has recently drawn attention as a promising drug target.<sup>5</sup>

One valuable approach in drug development is to employ natural products as lead structures.<sup>6</sup> They can be of particular importance whenever structural information about the target protein is difficult to obtain, as it is often the case for membrane proteins. For MraY, naturally occurring nucleoside antibiotics

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<sup>†</sup>Electronic Supplementary Information (ESI) available: Experimental procedures for all syntheses, <sup>1</sup>H and <sup>13</sup>C NMR spectra of all compounds, details of the <sup>1</sup>H-<sup>1</sup>H NOESY NMR experiment. See DOI: 10.1039/c003092a

have been reported to act as inhibitors.<sup>7</sup> These natural products are distinctly characterised by the unique structures of their nucleoside-derived moieties. Three subclasses of nucleoside antibiotics, the liposidomycins, caprazamycins and muraymycins, as well as the muraymycin-like FR-900493, have a common (5'S,6'S)uridine-derived core structure, characterised by C–C-bond linkage of the 5'-carbon atom of the uridine unit to a glycine moiety (C-6') as well as aminoribosylation of the 5'-hydroxy group.<sup>7</sup>

Structure-activity relationship (SAR) studies have been reported for synthetic analogues of the caprazamycins,<sup>8</sup> indicating a crucial role of the aminoribosyl unit for the retention of antibacterial activity towards Mycobacterium tuberculosis.<sup>8c</sup> In case of the muraymycins, a comparison of the 19 isolated naturally occurring derivatives provides different SAR insights: the activities of aminoribosylated muraymycin A1 1 and non-aminoribosylated muraymycin A5 2 against several bacteria including Staphylococcus aureus do not differ dramatically (Fig. 1).9 In addition, (semi-)synthetic analogues of the muraymycins have been investigated.<sup>10</sup> Remarkable results were obtained for the SAR of synthetic truncated muraymycin derivatives (e.g. 3-7, Fig. 1): firstly, the absolute configuration at the 5'-position was required to be (R) in order to obtain good activities, *i.e.* only 5'-epi-analogues with respect to the (5'S)-configured natural products were active. Secondly, the presence of some synthetic protecting groups (tertbutyl ester, tert-butyldimethylsilyl (TBDMS) groups) turned out to be a prerequisite for antibacterial potency.10b

In order to obtain novel structurally simplified nucleoside scaffolds for the synthesis of muraymycin analogues, it was envisaged to prepare 5'-deoxy derivatives lacking the site for aminoribosylation. In addition, 5'-deoxy nucleosyl amino acid

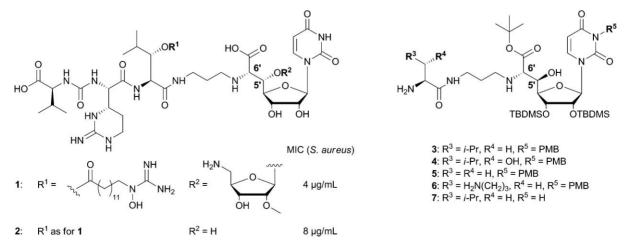


Fig. 1 Naturally occurring nucleoside antibiotics muraymycin A1 1 and muraymycin A5  $2^{\circ}$  and synthetic truncated 5'-*epi*-muraymycin analogues 3–7 displaying antibacterial activity<sup>10b</sup> (MIC = minimal inhibitory concentration, PMB = *para*-methoxybenzyl, TBDMS = *tert*-butyldimethylsilyl).

structures should provide insights into the role of the 5'-hydroxy group in case of truncated muraymycin analogues such as 3-7 (Fig. 1). Our objective therefore was to synthesise target compounds 8 and 9 with (6'S)- and (6'R)-configuration, respectively, from a common precursor in a stereocontrolled way. Thus, the influence of the configuration in 6'-position can also be elucidated. Both 8 and 9, which might also serve as suitably protected building blocks for the synthesis of more complex muraymycin analogues, were converted into their corresponding derivatives 10 and 11, respectively, which represent 5'-deoxy analogues of 3 for SAR studies on truncated muraymycin derivatives (Fig. 2).

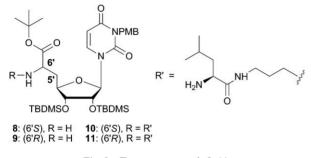
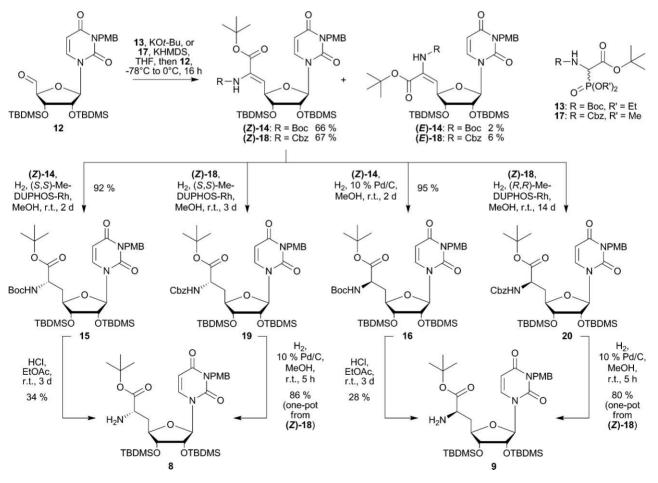


Fig. 2 Target compounds 8–11.

The synthetic route reported in this work started from protected uridine-5'-aldehyde 12 (Scheme 1), which can easily be obtained from uridine in an overall yield of 66% over 4 steps.<sup>11</sup> Wittig-Horner reaction of aldehyde 12 with the N-Boc-protected phosphonate  $13^{12}$  provided didehydro amino acid (Z)-14 in a yield of 66% and, as expected,<sup>13</sup> with high diastereoselectivity. Thus, only 2% of the diastereomer (E)-14 was found and isolated by column chromatography. The configuration of the didehydro amino acid moiety was assigned based on established <sup>1</sup>H NMR criteria for this class of compounds.<sup>14</sup> Only the (Z)-isomer was required for the subsequent asymmetric hydrogenation as it is reported that asymmetric hydrogenation using rhodium catalysts occurs more rapidly and with significantly better stereoselectivities for (Z)-didehydro amino acids than for the (E)-isomers.<sup>15</sup> Consequently, hydrogenation of (Z)-14 in the presence of the chiral rhodium catalyst (+)-1,2-bis-((2S,5S)-2,5-dimethylphospholano)benzene-(cyclooctadiene)-rhodium(I) tetrafluoroborate ((S,S)-Me-DUPHOS-Rh)<sup>16a</sup> provided product 15 in an excellent yield of 92% and with high diastereoselectivity (d.r. > 97:3) based on <sup>1</sup>H NMR). It is well established that (S,S)-Me-DUPHOS-Rh converts N-carbamate protected (Z)-didehydro amino acid esters selectively into L-amino acids.12,16b As there was clear evidence of the asymmetric homogenous hydrogenation of (Z)-14 being a catalyst-controlled reaction (vide infra), the stereochemistry at the C-6' position of 15 could therefore be assigned as (S). In contrast, when precursor (Z)-14 was hydrogenated under heterogenous conditions using palladium on charcoal, a surprising substratecontrolled selectivity (d.r. >95:5 based on <sup>1</sup>H NMR) towards



Scheme 1 Synthesis of 5'-deoxy nucleosyl amino acid building blocks 8 and 9.

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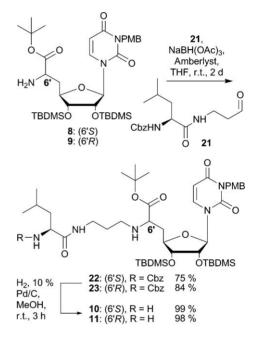
the other diastereomer with (6'R)-configuration was observed to give the according product **16** in 95% yield. For the synthesis of target bulding blocks **8** and **9**, the Boc group then had to be removed in the presence of both the *tert*-butyl ester and the silyl protecting groups. Though similar transformations employing hydrogen chloride in ethyl acetate have been previously reported,<sup>17</sup> the obtained yields of **8** and **9** (34% and 28%, respectively) were not satisfactory (Scheme 1). Most notably, prolonged reaction times in order to obtain sufficient conversion of the starting material led to unwanted side reactions such as cleavage of the silyl ether moieties.

Due to this limitation of the synthetic route utilising N-Boc protection, it was then decided to change the amino protecting group. Using phosphonate 17<sup>18</sup> for the Wittig-Horner step, N-Cbz protected didehydro amino acids (Z)-18 and (E)-18 were isolated in 67% and 6% yield, respectively, after column chromatography (Scheme 1). Surprisingly, application of the established <sup>1</sup>H NMR criteria for configurational assignment<sup>14</sup> was inconclusive in this case. Based on the results previously obtained with the N-Boc strategy, it was the most likely conclusion to propose the (Z)configuration for the major product in this case as well. This was experimentally supported by the results of the subsequent hydrogenation reactions (vide infra) as well as a <sup>1</sup>H-<sup>1</sup>H NOESY NMR experiment.<sup>19</sup> When diastereometrically pure (Z)-18 was used for the asymmetric hydrogenation in the presence of (S,S)-Me-DUPHOS-Rh, N-Cbz protected nucleosyl amino acid 19 was obtained. The N-protecting group could then be cleaved in a simple one-pot manner by further hydrogenation after the addition of palladium on charcoal to provide 8 directly in a good yield of 86% and with excellent diastereoselectivity (d.r. >97:3 based on <sup>1</sup>H NMR). In order to avoid hydrogenolysis of the Cbz group prior to reduction of the double bond, (R,R)-Me-DUPHOS-Rh was used instead of palladium on charcoal (vide supra) for the synthesis of 20, finally providing the (6'R)configured congener 9 after subsequent hydrogenolysis of the Cbz group. This one-pot sequence provided 9 in good yield (80%) and excellent diastereoselectivity (d.r. > 97:3 based on <sup>1</sup>H NMR, Scheme 1). The products 8 and 9 obtained via the N-Cbz route were identical to those furnished by the N-Boc approach, which was unambigously proven by NMR spectroscopy. Thus, nucleosyl amino acid building blocks 8 and 9 were synthesised from uridine in overall yields of 38% and 35%, respectively, over 6 steps via the N-Cbz protecting group strategy.

The reaction periods of the asymmetric hydrogenation transformations of (Z)-18 needed for sufficient conversions differed significantly for the two Me-DUPHOS-Rh catalysts. This might indicate the combination of (Z)-18 with (S,S)-Me-DUPHOS-Rh to represent the matched case, while the reaction using (R,R)-Me-DUPHOS-Rh might have suffered from a mismatched situation. As diastereoselectivities were very high in both cases, catalyst (i.e. ligand) control clearly dominated over any potential substrate control resulting from the chiral nucleoside moiety. This was at least the case for homogenous hydrogenation, while substrate control could be observed for heterogenous hydrogenation of the Boc-protected derivative (Z)-14 (vide supra). Attempts to perform the hydrogenation of (Z)-18 in the presence of the achiral Wilkinson catalyst (PPh<sub>3</sub>)<sub>3</sub>RhCl surprisingly gave no conversion. However, due to the clear evidence of the asymmetric homogenous hydrogenation reaction being a catalyst-controlled transformation

and the well-established stereoselectivity of Me-DUPHOS-Rh catalysts,<sup>12,16b</sup> assignment of the stereochemistry at C-6' was feasible.

As an efficient stereoselective synthesis of the unprecedented 5'deoxy nucleosyl amino acid scaffolds 8 and 9 could be achieved, it was desired to convert them into muraymycin derivatives 10 and 11, respectively, representing analogues of the reportedly bioactive compound 3.<sup>10b</sup> Both amines 8 and 9 were therefore reacted with aldehyde 21<sup>10b,20</sup> in reductive amination transformations, affording 22 and 23 in 75% and 84% yield, respectively. Final Cbz deprotection under hydrogenolytic conditions then gave target compounds 10 and 11 in nearly quantitative yields (Scheme 2).



Scheme 2 Synthesis of muraymycin analogues 10 and 11.

In conclusion, we report the synthesis of novel 5'-deoxy nucleosyl amino acid scaffolds derived from the nucleosidic core structure of several natural products, including muraymycin and caprazamycin antibiotics. A highly efficient and stereoselective approach using asymmetric hydrogenation of a common didehydro amino acid precursor provided both the (6'S)- and (6'R)-configured building blocks 8 and 9, which can be used for the preparation of novel non-aminoribosylated muraymycin analogues for SAR studies. The results obtained for this synthetic key step are of major general importance for the synthesis of  $\alpha$ -amino acid derivatives with highly functionalised side chains. The obtained excellent diastereoselectivities highlight the enormous versatility and broad substrate scope of the Me-DUPHOS-Rh catalysts in the hydrogenation of complex substrates. Compounds 8 and 9 were used for the concise synthesis of two analogues 10 and 11 of an established bioactive truncated muraymycin derivative. The biological evaluation of these 5'-deoxy muraymycins as part of a detailed SAR study on muraymycin analogues is currently being carried out.

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