

Synthesis of N-acetyllactosamine-terminated oligosaccharides — fragments of glycoprotein O-chains

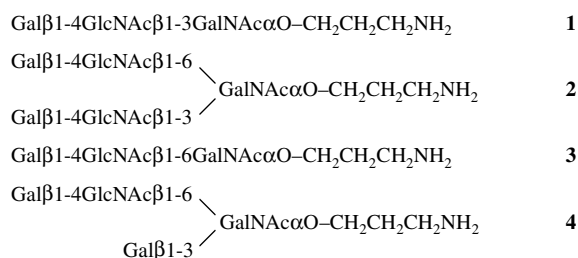
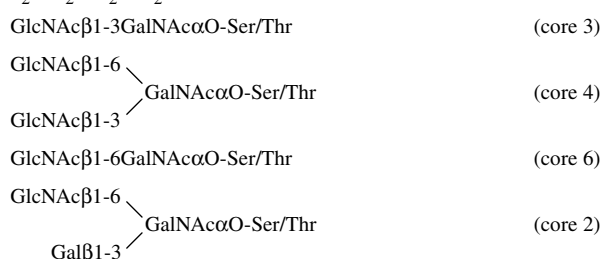
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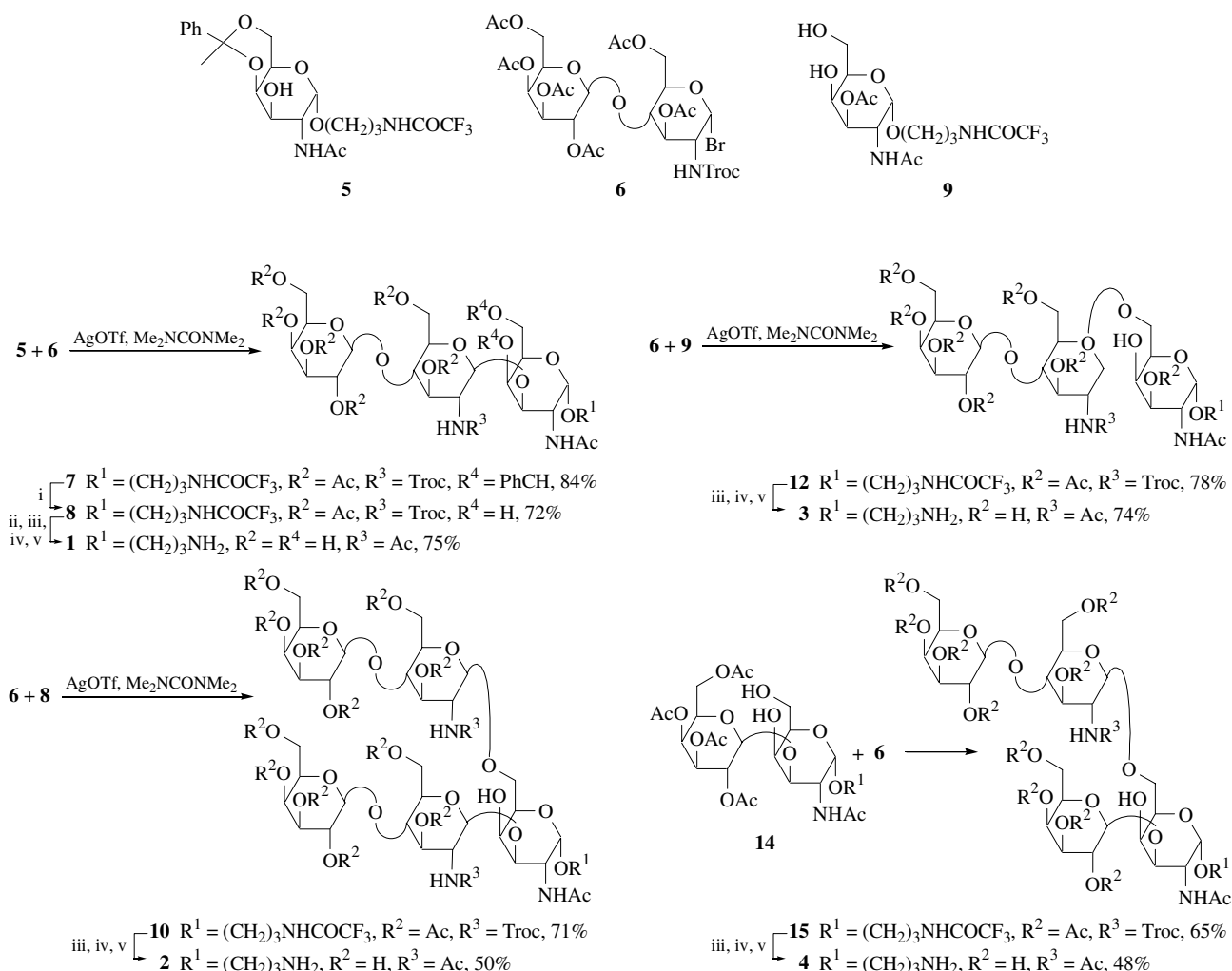
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The pentasaccharide Gal β 1-4GlcNAc β 1-6(Gal β 1-4GlcNAc β 1-3)GalNAc α OR, its partial trisaccharides Gal β 1-4GlcNAc β 1-3GalNAc α OR and Gal β 1-4GlcNAc β 1-6GalNAc α OR and the tetrasaccharide Gal β 1-4GlcNAc β 1-6(Gal β 1-3)GalNAc α OR were synthesised as spacers derivatives ($R = \text{CH}_2\text{CH}_2\text{CH}_2\text{NH}_2$) using a Troc-protected lactosamine glycosyl donor at the key stage.

Eight different cores, *i.e.*, regions connected with Ser or Thr of glycoprotein O-chains, are known.¹ Recently,² we reported the synthesis of oligosaccharides repeating four core structures, *i.e.*, GlcNAc β 1-3GalNAc α OR (core 3), GlcNAc β 1-3-(GlcNAc β 1-6)GalNAc α OR (core 4), GlcNAc β 1-6GalNAc α OR (core 6) and Gal β 1-3(GlcNAc β 1-6)GalNAc α OR (core 2); $R = \text{CH}_2\text{CH}_2\text{CH}_2\text{NH}_2$.



Here, we describe the synthesis of more complex oligosaccharides, namely, containing an additional terminal β -Gal moiety, as spacers compounds 1–4 suitable for further transformations into various glycoprobes.³ The above core glycopeptides have been identified in mucin-type glycoproteins; *e.g.*, core 6 structure is a cancer-associated antigen⁴ and core 4



Scheme 1 Reagents and conditions: i, AcOH; ii, Ac $_2$ O, Py; iii, Zn, AcOH, Ac $_2$ O; iv, MeONa, MeOH; v, NaOH, H $_2$ O.

trisaccharide is probably a crypted carcinoma-associated Tk antigen.⁵ Core 2 oligosaccharide is the specific motif, which defines P-selectin binding properties⁶ of SiaLe^x-containing glycoprotein PSGL-1. Gal-terminated oligosaccharides **1–4** are potential ligands for galectins, the family of β -galactoside binding lectins, whose natural cellular ligands are not yet clarified.

We used 3'-trifluoroacetamidopropyl 2-acetamido-4,6-O-benzylidene-2-deoxy- α -D-galactopyranoside **5**, which was described previously,⁷ as a starting building block. To synthesise trisaccharide **1**, compound **5** reacted with bromide **6** (Scheme 1). Resulting trisaccharide **7** was used as the starting material in the synthesis of pentasaccharide **2**: it was 4,6-O-debenzylidenated, and obtained diol **8** was glycosylated regioselectively at the 6-position with bromide **6**. To synthesise trisaccharide **3**, the 3-OH group in compound **5** was protected by acetylation, the benzylidene group was removed and the obtained diol **9** was 6-O-glycosylated as described above. Tetrasaccharide **4** was synthesised by 6-O-regioselective glycosylation of disaccharide **14**⁸ with bromide **6**.

In all cases, N-Troc-protected lactosaminyl bromide **6** (Troc is 1,1,1-trichloroethyloxycarbonyl),⁹ obtained by acetylation and subsequent bromine treatment of the corresponding commercial ethyl thioglycoside, was used for stereocontrolled β -glycosylation. This choice is stipulated by good yields, high β -stereoselectivity of glycosylation,^{2,9} and compatibility of this protection with other protecting groups used in the synthesis. Preparative yields in glycosylation with this reagent are noted in Scheme 1, and they are comparable with published data.⁹ β -Stereoselectivity of glycosylation with donor **6** was confirmed by the corresponding $J_{1,2}$ (8.0–9.5 Hz). The structures of synthesised oligosaccharides were confirmed by ¹H NMR data using homonuclear correlation spectroscopy and conventional analysis of coupling constants.[†]

Deprotection of sugar moieties and the spacer-arm by conventional methods (Scheme 1) gave rise to oligosaccharides **1–4** as 3-aminopropyl glycosides; the synthesis of these oligosaccharides in the convenient spacers form has not been described earlier.

Compounds **1–4** coupled with fluorescein-labelled polyacrylamide were used for the study of malignant cell galectins, the data will be published elsewhere.

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[†] *Gal β 1-4GlcNAc β 1-3GalNAc α 1-O(CH₂)₃NH₂ 1*: ¹H NMR (500 MHz, D₂O) δ : 4.778 (d, 1H, H-1 GalNAc, $J_{1,2}$ 3.9 Hz), 4.540 (d, 1H, H-1 GlcNAc, $J_{1,2}$ 7.6 Hz), 4.410 (d, 1H, H-1 Gal, $J_{1,2}$ 7.8 Hz), 4.196 (dd, 1H, H-2 GalNAc, $J_{1,2}$ 3.9 Hz, $J_{2,3}$ 11.3 Hz), 4.151 (dd, 1H, H-4 GalNAc, $J_{3,4}$ 2.7 Hz, $J_{4,5} \leq 1$ Hz), 3.85–3.92 (m, 4H, H-3 and H-5 GalNAc, H-4 Gal, H-6^a GlcNAc), 3.781 (dd, 1H, H-6^b GlcNAc, $J_{5,6}$ 4.7 Hz, $J_{6^a,6^b}$ 12.2 Hz), 3.63–3.76 (m, 9H, H-2, H-3, H-4 GlcNAc, H-5, H-6^a, H-6^b Gal, H-6^a, H-6^b GalNAc, OCH₂ sp), 3.607 (dd, 1H, H-3 Gal, $J_{2,3}$ 10.0 Hz, $J_{3,4}$ 3.5 Hz), 3.47–3.53 (m, 2H, H-5 GlcNAc, OCH₂ sp), 3.475 (dd, 1H, H-2 Gal, $J_{2,3}$ 10.0 Hz, $J_{1,2}$ 7.8 Hz), 3.055 (m, 2H, NCH₂ sp), 1.983 (s, 3H, NHCOMe), 1.953 (s, 3H, NHCOMe), 1.940 (m, 2H, CH₂ sp). MS, m/z : 666 (643 + 23) (M⁺ + Na⁺). [α]_D +49 (c1, H₂O).

Gal β 1-4GlcNAc β 1-6(Gal β 1-4GlcNAc β 1-3)GalNAc α 1-O(CH₂)₃NH₂ 2: ¹H NMR (500 MHz, D₂O) δ : 4.515 (d, 1H, H-1 GlcNAc, $J_{1,2}$ 7.8 Hz), 4.453 (d, 1H, H-1 GlcNAc, $J_{1,2}$ 8.3 Hz), 4.391 (d, 2H, 2H-1 Gal, $J_{1,2}$ 7.8 Hz), 4.172 (dd, 1H, H-2 GalNAc, $J_{1,2}$ 3.7 Hz, $J_{2,3}$ 11.0 Hz), 4.124 (dd, 1H, H-4 GalNAc, $J_{3,4}$ 2.9 Hz, $J_{4,5} \leq 1$ Hz), 4.077 (dd, 1H, H-6^a GalNAc, $J_{5,6^a}$ 2.2 Hz, $J_{6^a,6^b}$ 10.5 Hz), 3.85–3.96 (m, 6H, 2H-6^a GlcNAc, H-6^a and H-3 GalNAc, 2H-4 Gal), 3.764 (dd, 2H, 2H-6^b GlcNAc, $J_{5,6^b}$ 4.9 Hz, $J_{6^a,6^b}$ 12.2 Hz), 3.60–3.73 (m, 14H, 2H-5, 2H-6^a, 2H-6^b Gal, 2H-2, 2H-3, 2H-4 GlcNAc, H-5 GalNAc, OCH sp), 3.586 (dd, 2H, 2H-3 Gal, $J_{3,4}$ 3.4 Hz, $J_{2,3}$ 10.0 Hz), 3.532 (m, 1H, H-5 GlcNAc), 3.40–3.50 (m, 4H, H-5 GlcNAc, 2H-2 Gal, OCH sp), 3.045 (m, 2H, NCH₂ sp), 1.963 (s, 3H, NHCOMe), 1.936 (s, 3H, NHCOMe), 1.934 (s, 3H, NHCOMe), 1.85–1.98 (m, 2H, CH₂ sp). MS, m/z : 1031 (1008 + 23) (M⁺ + Na⁺). [α]_D +18 (c1, H₂O).

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Gal β 1-4GlcNAc β 1-6GalNAc α 1-O(CH₂)₃NH₂ 3: ¹H NMR (500 MHz, D₂O) δ : 4.865, (d, 1H, H-1 GalNAc, $J_{1,2}$ 3.7 Hz), 4.526 (d, 1H, H-1 GlcNAc, $J_{1,2}$ 8.1 Hz), 4.453 (d, 1H, H-1 Gal, $J_{1,2}$ 7.9 Hz), 4.139 (dd, 1H, H-2 GalNAc, $J_{1,2}$ 3.7 Hz, $J_{2,3}$ 11.0 Hz), 4.069 (dd, 1H, H-6^a GalNAc, $J_{5,6^a}$ 3.2 Hz, $J_{6^a,6^b}$ 10.8 Hz), 3.96–4.01 (m, 2H, H-6^a GlcNAc, H-6^b GalNAc), 3.953 (dd, 1H, H-4 GalNAc, $J_{3,4}$ 3.2 Hz, $J_{4,5} \leq 1$ Hz), 3.909 (dd, 1H, H-4 Gal, $J_{3,4}$ 3.4 Hz, $J_{4,5} \leq 1$ Hz), 3.884 (dd, 1H, H-3 GalNAc, $J_{3,4}$ 3.2 Hz, $J_{2,3}$ 11.0 Hz), 3.825 (dd, 1H, H-6^b GlcNAc, $J_{5,6^b}$ 4.9 Hz, $J_{6^a,6^b}$ 12.5 Hz), 3.66–3.79 (m, 8H, H-2, H-3, H-4 GlcNAc, H-5, H-6^a, H-6^b Gal, H-5 GalNAc, OCH sp), 3.648 (dd, 1H, H-3 Gal, $J_{3,4}$ 3.4 Hz, $J_{2,3}$ 9.8 Hz), 3.593 (m, 1H, H-5 GlcNAc), 3.527 (dd, 1H, H-2 Gal, $J_{1,2}$ 7.9 Hz, $J_{2,3}$ 9.8 Hz), 3.46–3.53 (m, 1H, OCH₂ sp), 3.104 (m, 2H, NCH₂ sp), 2.022 (s, 3H, NHCOMe), 2.002 (s, 3H, NHCOMe), 1.91–2.01 (m, 2H, CH₂ sp). MS, m/z : 666 (643 + 23) (M⁺ + Na⁺). [α]_D +42 (c1, H₂O).

Gal β 1-4GlcNAc β 1-6(Gal β 1-3)GalNAc α 1-O(CH₂)₃NH₂ 4: ¹H NMR (500 MHz, D₂O) δ : 4.900 (d, 1H, H-1 GalNAc, $J_{1,2}$ 3.7 Hz), 4.557 (d, 1H, H-1 GlcNAc, $J_{1,2}$ 8.2 Hz), 4.491 (d, 1H, H-1 Gal β 1-3, $J_{1,2}$ 8.1 Hz), 4.474 (d, 1H, H-1 Gal β 1-4, $J_{1,2}$ 9.0 Hz), 4.350 (dd, 1H, H-2 GalNAc, $J_{1,2}$ 3.7 Hz, $J_{2,3}$ 11.0 Hz), 4.245 (dd, 1H, H-4 GalNAc, $J_{4,5} \leq 1$ Hz, $J_{3,4}$ 2.3 Hz), 4.114 (dd, 1H, H-6^a GalNAc, $J_{5,6^a}$ 2 Hz, $J_{6^a,6^b}$ 10.5 Hz), 3.99–4.08 (m, 3H, H-3 and H-6^b GalNAc, H-6^a GlcNAc), 3.947 (dd, 1H, H-4 Gal β 1-4, $J_{3,4}$ 3.2 Hz, $J_{4,5} < 1$ Hz), 3.927 (dd, 1H, H-4 Gal β 1-3, $J_{3,4}$ 3.1 Hz, $J_{4,5} \leq 1$ Hz), 3.862 (dd, 1H, H-6^b GlcNAc, $J_{5,6^b}$ 4.9 Hz, $J_{6^a,6^b}$ 12.3 Hz), 3.70–3.83 (m, 11H, H-5 GalNAc, H-2, H-3, H-4 GlcNAc, 2H-5, 2H-6^a, 2H-6^b Gal, OCH sp), 3.685 (dd, 1H, H-3 Gal β 1-4, $J_{2,3}$ 10.0 Hz, $J_{3,4}$ 3.2 Hz), 3.634 (dd, 1H, H-3 Gal β 1-3, $J_{3,4}$ 3.2 Hz, $J_{2,3}$ 9.8 Hz), 3.60–3.66 (m, 1H, H-5 GlcNAc), 3.51–3.59 (m, 3H, 2H-2 Gal, OCH sp), 3.152 (m, 2H, NCH₂ sp), 2.040 (s, 3H, NHCOMe), 1.90–2.10 (m, 2H, CH₂ sp). MS, m/z : 828 (805 + 23) (M⁺ + Na⁺). [α]_D +40 (c1, H₂O).