

Crystallographic Characterization of 12-Helical Secondary Structure in  $\beta$ -Peptides Containing Side Chain GroupsSoo Hyuk Choi,<sup>†</sup> Ilia A. Guzei, Lara C. Spencer, and Samuel H. Gellman\*

Department of Chemistry, University of Wisconsin, Madison, Wisconsin 53706

Received July 14, 2010; E-mail: gellman@chem.wisc.edu

**Abstract:** Helices are the most extensively studied secondary structures formed by  $\beta$ -peptide foldamers. Among the five known  $\beta$ -peptide helices, the 12-helix is particularly interesting because the internal hydrogen bond orientation and macrodipole are analogous to those of  $\alpha$ -peptide helices ( $\alpha$ -helix and  $3_{10}$ -helix). The  $\beta$ -peptide 12-helix is defined by  $i, i+3$  C=O $\cdots$ H–N backbone hydrogen bonds and promoted by  $\beta$ -residues with a five-membered ring constraint. The 12-helical scaffold has been used to generate  $\beta$ -peptides with specific biological functions, for which diverse side chains must be properly placed along the backbone and, upon folding, properly arranged in space. Only two crystal structures of 12-helical  $\beta$ -peptides have previously been reported, both for homooligomers of *trans*-2-aminocyclopentanecarboxylic acid (ACPC). Here we report five additional crystal structures of 12-helical  $\beta$ -peptides, all containing residues that bear side chains. Four of the crystallized  $\beta$ -peptides include *trans*-4,4-dimethyl-2-aminocyclopentanecarboxylic acid (*dm*-ACPC) residues, and the fifth contains a  $\beta^3$ -hPhe residue. These five  $\beta$ -peptides adopt fully folded 12-helical conformations in the solid state. The new crystal structures, along with previously reported data, allow a detailed characterization of the 12-helical conformation; average backbone torsion angles of  $\beta$ -residues and helical parameters are derived. These structural parameters are found to be similar to those for  $i, i+3$  C=O $\cdots$ H–N hydrogen-bonded helices formed by other peptide backbones generated from  $\alpha$ - and/or  $\beta$ -amino acids. The similarity between the conformational behavior of *dm*-ACPC and ACPC is consistent with previous NMR-based conclusions that 4,4-disubstituted ACPC derivatives are compatible with 12-helical folding. In addition, our data show how a  $\beta^3$ -residue is accommodated in the 12-helix, thus enhancing understanding of the diverse conformational behavior of this flexible class of  $\beta$ -amino acids.

## Introduction

$\beta$ -Peptides are oligomers of  $\beta$ -amino acids and constitute perhaps the most extensively studied class of unnatural foldamers.<sup>1,2</sup> Many examples of  $\beta$ -peptides that display folded conformations analogous to secondary structures of  $\alpha$ -peptides and proteins have been identified, including examples that form helices, turns, or extended conformations.  $\beta$ -Peptide helices have drawn considerable attention because they may be used to mimic helical substructures within proteins.<sup>1</sup> Five  $\beta$ -peptide helices (14-, 12-, 10/12-, 10-, and 8-helix) have been reported to date.<sup>1,2</sup> The nomenclature of these helices is based on the number of atoms in the characteristic hydrogen bonds. For example, a  $\beta$ -peptide 12-helix is defined by  $i, i+3$  C=O $\cdots$ H–N hydrogen bonds that contain 12 atoms. This particular helix is interesting from the perspective of mimicking  $\alpha$ -helices among conventional peptides and proteins. The internal hydrogen bond orientation and macrodipole of the  $\beta$ -peptide 12-helix are analogous to those for the helices most commonly observed in proteins:  $3_{10}$ -helix ( $i, i+3$  C=O $\cdots$ H–N hydrogen bonds) and

$\alpha$ -helix ( $i, i+4$  C=O $\cdots$ H–N hydrogen bonds).<sup>3</sup> In contrast, the hydrogen bond orientations of other  $\beta$ -peptide helices (e.g.,  $i, i-2$  C=O $\cdots$ H–N hydrogen bonds for the 14-helix and alternating  $i, i-1$  C=O $\cdots$ H–N and  $i, i+3$  C=O $\cdots$ H–N hydrogen bonds for the 10/12-helix) are not observed among  $\alpha$ -peptide helices.<sup>2a</sup>

Experimental and computational studies suggest that the 12-helix is not intrinsically favorable for acyclic  $\beta$ -residues, such as the commonly used  $\beta^3$ -homoamino acids (side chain adjacent to nitrogen), but that this secondary structure is promoted by cyclic  $\beta$ -residues with a five-membered ring constraint.<sup>4</sup> We have shown that  $\beta$ -peptide oligomers that contain residues derived from *trans*-2-aminocyclopentanecarboxylic acid (ACPC) and/or *trans*-4-aminopyrrolidine-3-carboxylic acid (APC) adopt 12-helical conformations.<sup>5,6a</sup> In complementary work, Martinek et al. have recently shown that 12-helical folding is promoted by  $\beta$ -amino acid residues with a bicyclo[3.3.1]heptane skeleton.<sup>6b</sup> Even more recently, Aitken et al. have shown that *trans*-2-aminocyclobutanecarboxylic acid residues favor 12-helical folding.<sup>6c</sup> The 12-helical scaffold has served as a basis

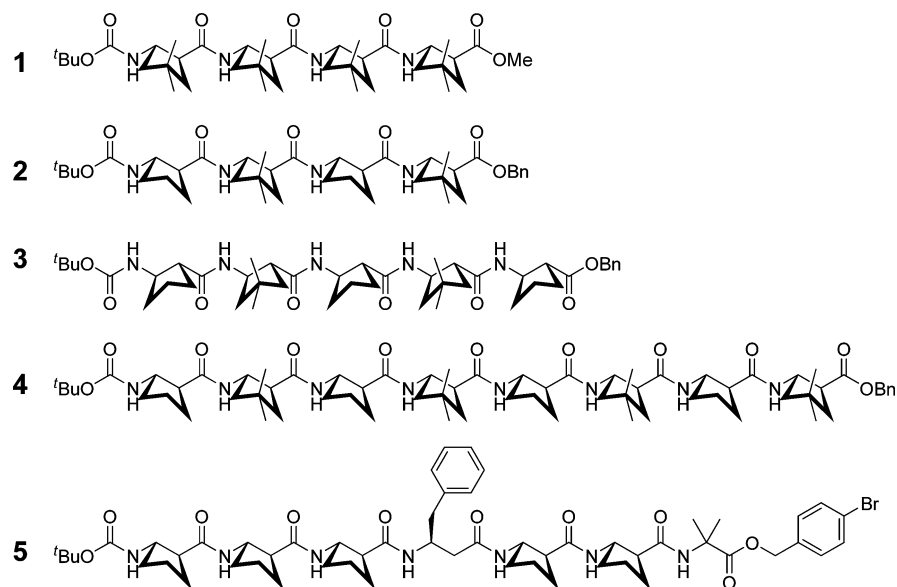
<sup>†</sup> Present address: Department of Chemistry, Northwestern University, Evanston, IL 60208.

(1) Gellman, S. H. *Acc. Chem. Res.* **1998**, *31*, 173.

(2) (a) Cheng, R. P.; Gellman, S. H.; DeGrado, W. F. *Chem. Rev.* **2001**, *101*, 3219. (b) Seebach, D.; Hook, D. F.; Glatli, A. *Biopolymers* **2006**, *84*, 23.

(3) Crisma, M.; Formaggio, F.; Moretto, A.; Toniolo, C. *Biopolymers* **2006**, *84*, 3.

(4) (a) Wu, Y. D.; Wang, D. P. *J. Am. Chem. Soc.* **1999**, *121*, 9352. (b) Gunther, R.; Hofmann, H. J.; Kuczera, K. *J. Phys. Chem. B* **2001**, *105*, 5559. (c) Beke, T.; Somlai, C.; Perczel, A. *J. Comput. Chem.* **2006**, *27*, 20.

**Chart 1.**  $\beta$ -Peptides for Which Crystal Structures Are Reported

for developing  $\beta$ -peptides that display antimicrobial activity,<sup>7</sup> inhibit protein–protein interactions,<sup>8</sup> and project carbohydrates in specific three-dimensional patterns.<sup>9</sup> For these biological applications, functional groups must be introduced at precise locations along the 12-helical backbone, which can be achieved

by incorporation of acyclic  $\beta$ -amino acid residues<sup>5g,i</sup> or cyclic  $\beta$ -amino acid residues that bear side chains.<sup>5e,f,h,j</sup> Data obtained in solution suggest that a few  $\beta^2$ - or  $\beta^3$ -residues can be incorporated into a 12-helix if most other residues have the five-membered ring constraint.<sup>5g,i</sup> Use of  $\beta$ -residues with functionalized five-membered rings, however, provides functional diversity in 12-helical scaffolds without loss of conformational stability. Several approaches to five-membered ring functionalization have been reported: sulfonylation of the pyrrolidine nitrogen atom of APC,<sup>5e,f</sup> 3-substitution,<sup>5h</sup> 4,4-disubstitution<sup>5j</sup> or 3,4-disubstitution of ACPC,<sup>10a</sup> N-functionalization of a pyrrolidinone-based  $\beta$ -amino acid,<sup>10b</sup> and generation of  $\beta$ -amino acid building blocks from AZT.<sup>10c,11</sup>

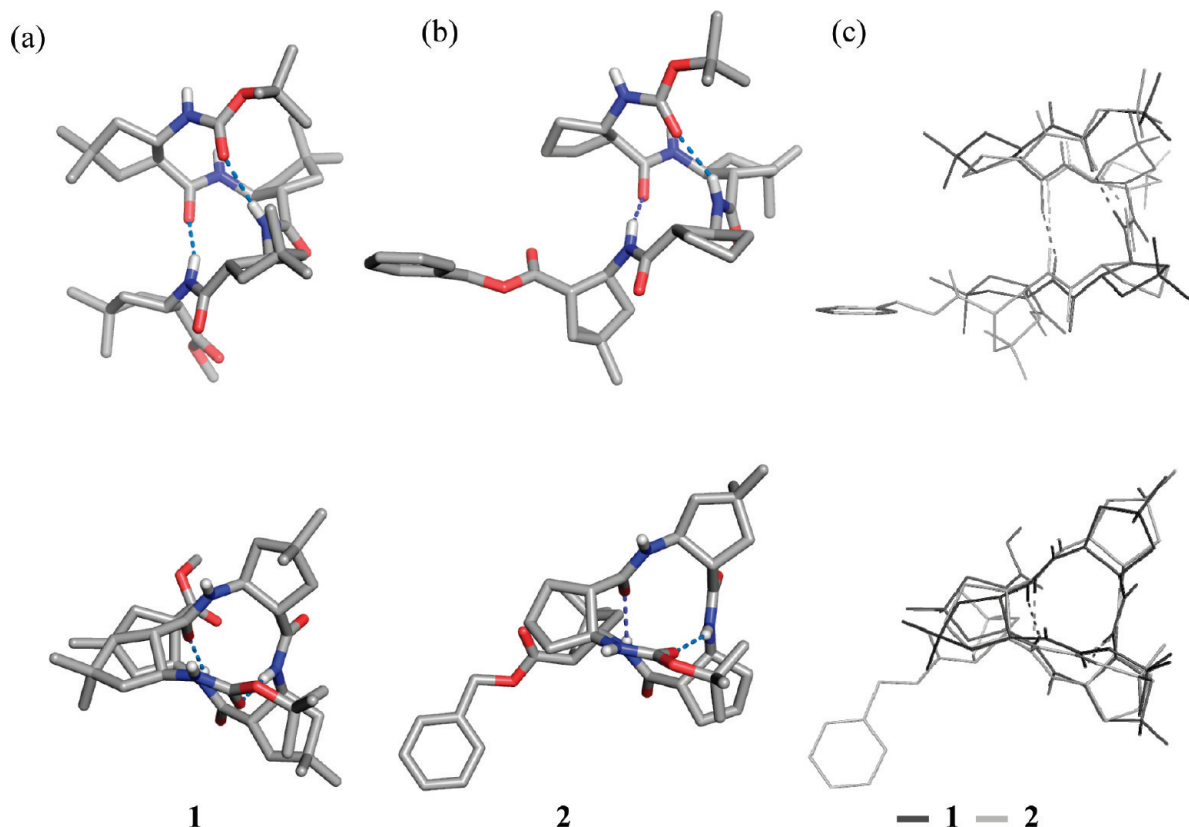
Only two crystal structures of 12-helical  $\beta$ -peptides have been reported to date, and both have been obtained with ACPC homooligomers.<sup>5c</sup> Here we provide five additional crystal structures that display 12-helical conformations. Four of the new structures contain *trans*-4,4-dimethyl-2-aminocyclopentanecarboxylic acid (*dm*-ACPC) residues, and the fifth structure contains a  $\beta^3$ -homophenylalanine ( $\beta^3$ -hPhe) residue embedded in an ACPC-rich sequence. This structural information enables us to identify characteristic structural parameters for the 12-helical secondary structure and to make comparisons with other well-characterized helices.

## Results and Discussion

**Design and Synthesis.** Chart 1 shows the five  $\beta$ -peptides for which crystal structures are reported. These molecules include a homooligomer of *dm*-ACPC (1) and three oligomers that contain both ACPC and *dm*-ACPC residues (2–4). Heptamer 5 contains a  $\beta^3$ -hPhe residue along with five ACPC residues; the crystal structure of 5 represents the first high-resolution data for a 12-helical  $\beta$ -peptide containing an acyclic residue. Oligomer 5 contains an  $\alpha$ -residue,  $\alpha$ -aminoisobutyric acid (Aib), at the C-terminus, which was included to enhance solubility and crystallization.<sup>12</sup> This  $\alpha$ -residue does not participate in the

- (5) (a) Appella, D. H.; Christianson, L. A.; Klein, D. A.; Powell, D. R.; Huang, X. L.; Barchi, J. J.; Gellman, S. H. *Nature* **1997**, 387, 381. (b) Applequist, J.; Bode, K. A.; Appella, D. H.; Christianson, L. A.; Gellman, S. H. *J. Am. Chem. Soc.* **1998**, 120, 4891. (c) Appella, D. H.; Christianson, L. A.; Klein, D. A.; Richards, M. R.; Powell, D. R.; Gellman, S. H. *J. Am. Chem. Soc.* **1999**, 121, 7574. (d) Barchi, J. J.; Huang, X. L.; Appella, D. H.; Christianson, L. A.; Durell, S. R.; Gellman, S. H. *J. Am. Chem. Soc.* **2000**, 122, 2711. (e) Wang, X. F.; Espinosa, J. F.; Gellman, S. H. *J. Am. Chem. Soc.* **2000**, 122, 4821. (f) Lee, H. S.; Syud, F. A.; Wang, X. F.; Gellman, S. H. *J. Am. Chem. Soc.* **2001**, 123, 7721. (g) LePlae, P. R.; Fisk, J. D.; Porter, E. A.; Weisblum, B.; Gellman, S. H. *J. Am. Chem. Soc.* **2002**, 124, 6820. (h) Woll, M. G.; Fisk, J. D.; LePlae, P. R.; Gellman, S. H. *J. Am. Chem. Soc.* **2002**, 124, 12447. (i) Park, J. S.; Lee, H. S.; Lai, J. R.; Kim, B. M.; Gellman, S. H. *J. Am. Chem. Soc.* **2003**, 125, 8539. (j) Peelen, T. J.; Chi, Y. G.; English, E. P.; Gellman, S. H. *Org. Lett.* **2004**, 6, 4411.
- (6) (a) Hetényi, A.; Tóth, G. K.; Somlai, C.; Vass, E.; Martinek, T. A.; Fülöp, F. *Chem.—Eur. J.* **2009**, 15, 10736. (b) Hetényi, A.; Szakonyi, Z.; Mándity, I. M.; Szolnoki, É.; Tóth, G. K.; Martinek, T. A.; Fülöp, F. *Chem. Commun.* **2009**, 177. (c) While our paper was under review, a very thorough characterization of 12-helix formation by oligomers of *trans*-2-aminocyclobutanecarboxylic acid appeared: Fernandes, C.; Faure, S.; Pereira, E.; Théry, V.; Declerck, V.; Guillot, R.; Aitken, D. J. *Org. Lett.* **2010**, 12, 3606–3609. The crystal structure reported by Aitken et al. is not included in our structural analysis. (d) See also: Winkler, J. D.; Piatnitski, E. L.; Mehlmann, J.; Kaspárec, J.; Axelsen, P. H. *Angew. Chem. Int. Ed.* **2001**, 40, 743.
- (7) (a) Porter, E. A.; Wang, X. F.; Lee, H. S.; Weisblum, B.; Gellman, S. H. *Nature* **2000**, 404, 565. (b) Porter, E. A.; Weisblum, B.; Gellman, S. H. *J. Am. Chem. Soc.* **2002**, 124, 7324.
- (8) (a) English, E. P.; Chumanov, R. S.; Gellman, S. H.; Compton, T. J. *Biol. Chem.* **2006**, 281, 2661. (b) Imamura, Y.; Watanabe, N.; Umezawa, N.; Iwatsubo, T.; Kato, N.; Tomita, T.; Higuchi, T. *J. Am. Chem. Soc.* **2009**, 131, 7353.
- (9) Simpson, G. L.; Gordon, A. H.; Lindsay, D. M.; Promsawan, N.; Crump, M. P.; Mulholland, K.; Hayter, B. R.; Gallagher, T. J. *Am. Chem. Soc.* **2006**, 128, 10638.
- (10) (a) Kiss, L.; Kazi, B.; Forró, E.; Fülöp, F. *Tetrahedron Lett.* **2008**, 49, 339. (b) Menegazzo, I.; Fries, A.; Mammi, S.; Galeazzi, R.; Martelli, G.; Orena, M.; Rinaldi, S. *Chem. Commun.* **2006**, 4915. (c) Chandrasekhar, S.; Reddy, G. P. K.; Kiran, M. U.; Nagesh, C.; Jagadeesh, B. *Tetrahedron Lett.* **2008**, 49, 2969.

- (11) An alternative secondary structure has been observed for an AZT-derived subunits in a different solvent: Threlfall, R.; Davies, A.; Howarth, N. M.; Fisher, J.; Cosstick, R. *Chem. Commun.* **2008**, 585.

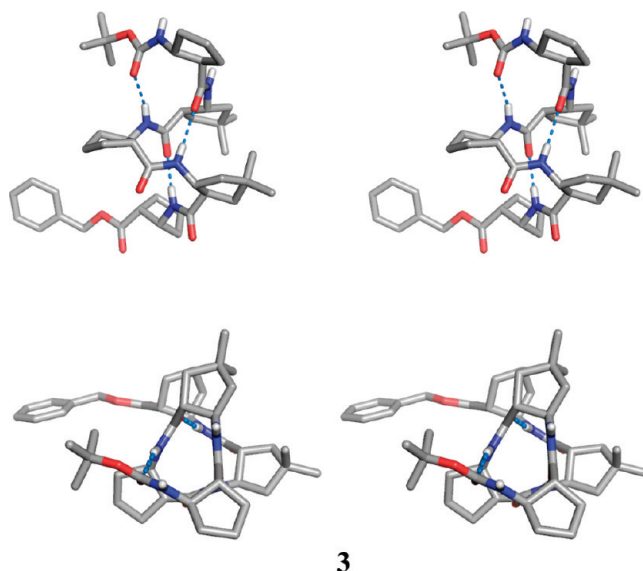


**Figure 1.** Crystal structures of tetramers: (a) **1**, (b) **2**, (c) overlay of the two structures. The top views are perpendicular to the helical axis, and the bottom views are along the helical axis. Dashed lines indicate hydrogen bonds. A cocrystallized chloroform molecule for **2** and most hydrogen atoms for both **1** and **2** are omitted for clarity.

helical hydrogen bonding pattern because there is no hydrogen bond donor at the C-terminus.

$\beta$ -Peptides **1–5** were synthesized via conventional solution-phase methods involving carbodiimide-mediated coupling.<sup>12b–d</sup> ACPC<sup>13</sup> and *dm*-ACPC<sup>5j</sup> building blocks were prepared by reported procedures. Each of the three shortest  $\beta$ -peptides (**1–3**) was crystallized by solvent diffusion of *n*-pentane into a diethyl ether/chloroform solution of the oligomer. Crystals of octamer **4** were grown by solvent diffusion of *n*-heptane into a 1,2-dichloroethane solution of the oligomer. Heptamer **5** was crystallized in two ways, by slow evaporation of either a methanol–water mixture or an isopropyl alcohol solution.

**Crystal Structures of  $\beta$ -Peptides Containing *dm*-ACPC Residues.** All of the four *dm*-ACPC-containing  $\beta$ -peptides display fully folded 12-helical conformations with the maximum number of *i*, *i*+3 C=O $\cdots$ H–N hydrogen bonds in the crystal-line state (Figures 1–3). Pentamer **3** displays a left-handed 12-helical conformation in the solid state because **3** contains (1*R*,2*R*)-forms of ACPC and *dm*-ACPC residues (Figure 2). The other  $\beta$ -peptides discussed here contain (1*S*,2*S*)-forms of ACPC and *dm*-ACPC residues and therefore display right-handed helical conformations. In the crystal structure of octamer **4**

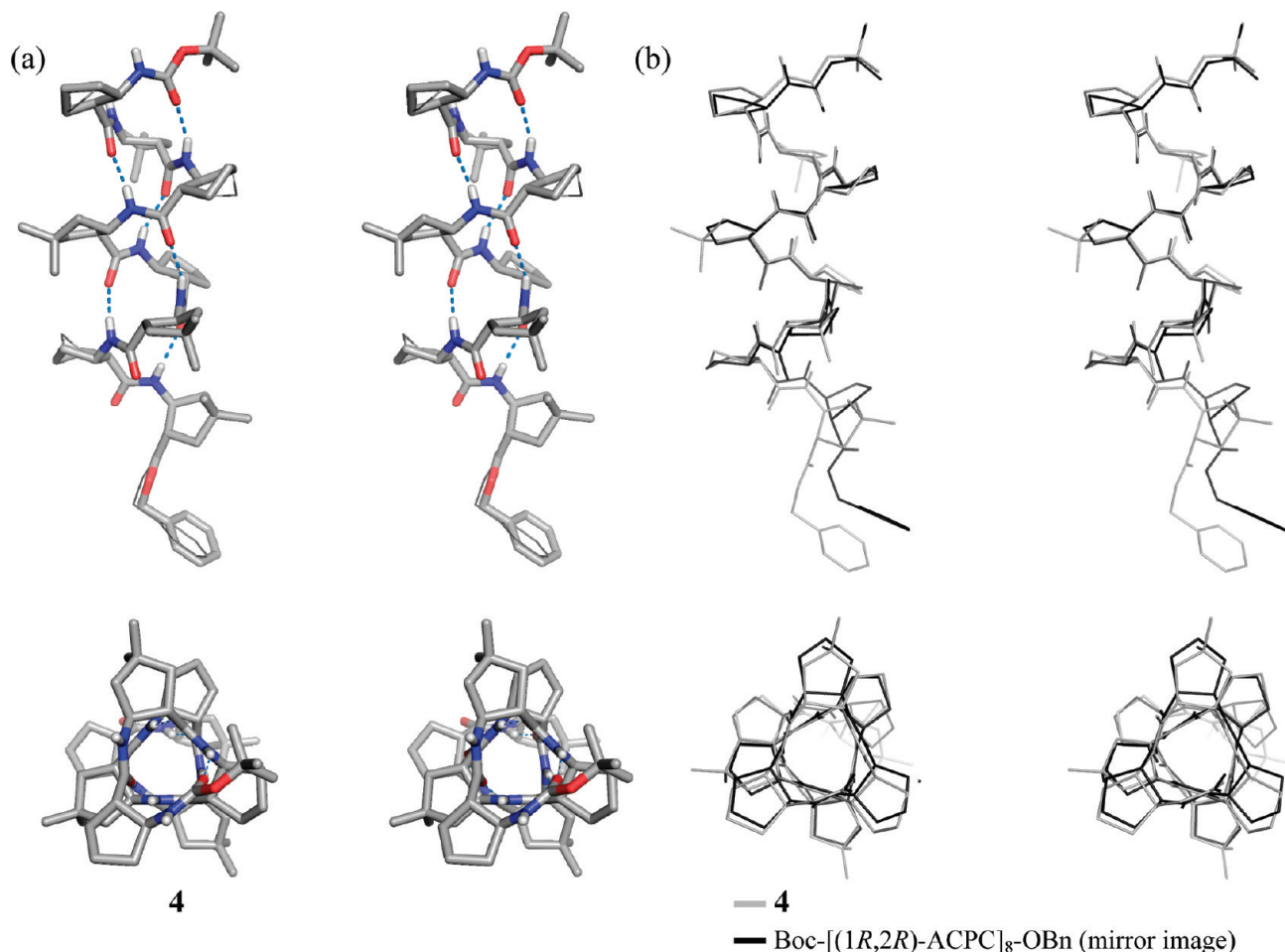


**Figure 2.** Crystal structure of pentamer **3** (stereoview). The top view is perpendicular to the helical axis, and the bottom view is along the helical axis. Dashed lines indicate hydrogen bonds. A cocrystallized ether molecule, a disordered carbon atom in ACPC(1), and most hydrogen atoms are omitted for clarity.

(Figure 3), the C-terminal benzyl group and some cyclopentane ring puckers are disordered over two positions (9:1 to 6:4 ratio). Interestingly, ring pucker disorder is limited to ACPC residues; no disordered carbons are found among the four *dm*-ACPC residues. The sequence of **4** is analogous to that of a previously reported homooligomer, Boc-[(1*R*,2*R*)-ACPC]<sub>8</sub>-OBn.<sup>5c</sup> The

- (12) (a) Moretto, V.; Crisma, M.; Bonora, G. M.; Toniolo, C.; Balaram, H.; Balaram, P. *Macromolecules* **1989**, *22*, 2939. (b) Choi, S. H.; Guzei, I. A.; Gellman, S. H. *J. Am. Chem. Soc.* **2007**, *129*, 13780. (c) Choi, S. H.; Guzei, I. A.; Spencer, L. C.; Gellman, S. H. *J. Am. Chem. Soc.* **2008**, *130*, 6544. (d) Choi, S. H.; Guzei, I. A.; Spencer, L. C.; Gellman, S. H. *J. Am. Chem. Soc.* **2009**, *131*, 2917.
- (13) (a) Lee, H. S.; LePlae, P. R.; Porter, E. A.; Gellman, S. H. *J. Org. Chem.* **2001**, *66*, 3597. (b) LePlae, P. R.; Umezawa, N.; Lee, H.-S.; Gellman, S. H. *J. Org. Chem.* **2001**, *66*, 5629.





**Figure 3.** (a) Crystal structure of octamer **4**. Alternative positions of disordered atoms are indicated with thin lines (stereoview). (b) Overlay of the structure of **4** (gray lines) and the mirror image of the structure of Boc-[(1*R*,2*R*)-ACPC]<sub>8</sub>-OBn (black lines; ref 5c) (stereoview). The top view is perpendicular to the helical axis, and the bottom view is along the helical axis. Dashed lines indicate hydrogen bonds. Most hydrogen atoms are omitted for clarity.

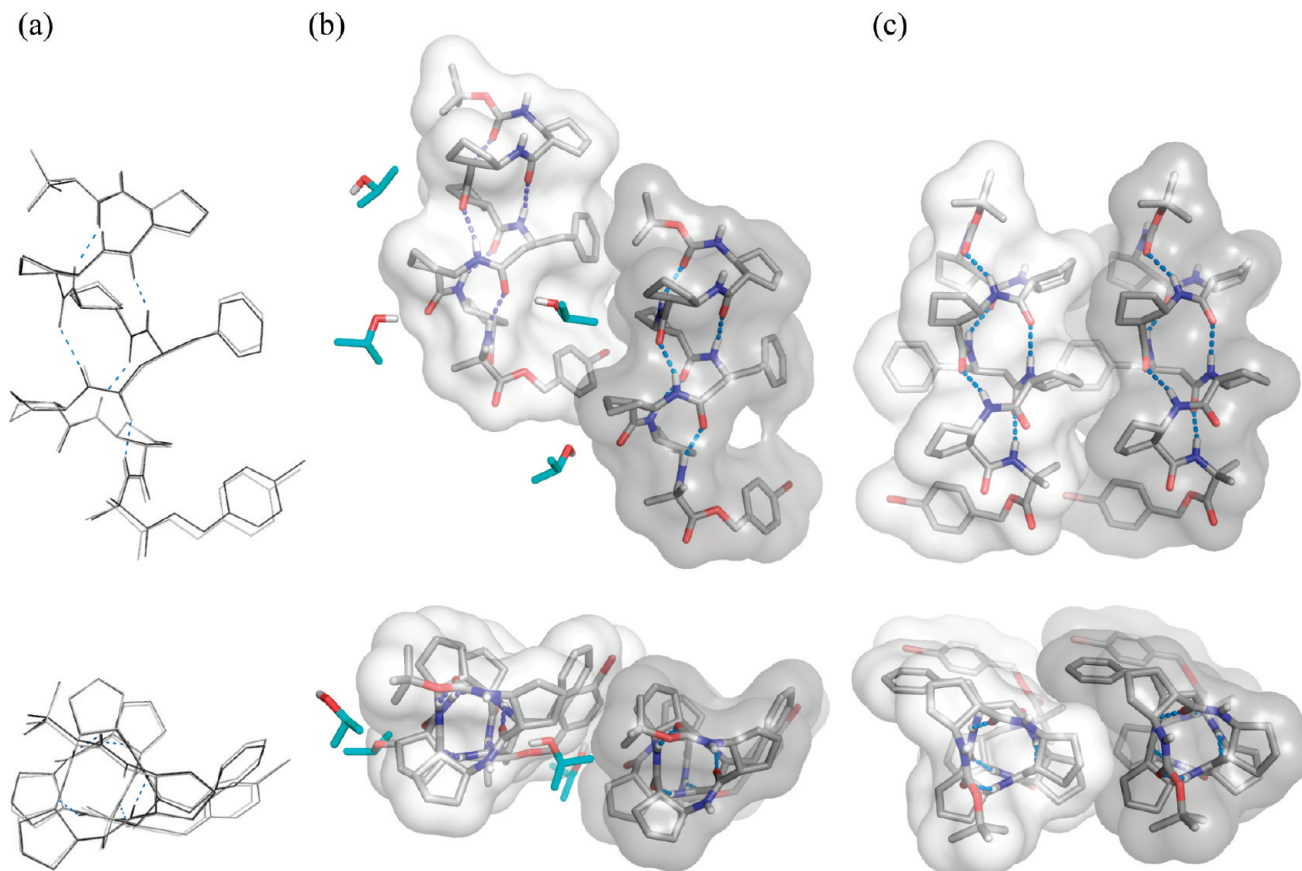
principal differences between **4** and Boc-[(1*R*,2*R*)-ACPC]<sub>8</sub>-OBn are the *gem*-dimethyl substituents of the *dm*-ACPC residues in **4** and the opposite handedness of the two helices. In Figure 3b, the right-handed 12-helical conformation observed for **4** in the solid state is superimposed on the mirror image of the left-handed 12-helical conformation observed for Boc-[(1*R*,2*R*)-ACPC]<sub>8</sub>-OBn. The backbones of these two 12-helices are virtually identical except for the C-terminal residue in each  $\beta$ -peptide, the ester of which cannot serve as a hydrogen bond donor. These results suggest that the *gem*-dimethyl substituents of the *dm*-ACPC residues do not cause any distortion of the 12-helical conformation, a conclusion that is consistent with the results of two-dimensional NMR studies of analogous  $\beta$ -peptides in solution.<sup>5j</sup>

**Crystal Structure of a  $\beta$ -Peptide Containing a  $\beta^3$ -hPhe Residue.** Figure 4 shows the two symmetry-independent conformations in the crystal of heptamer **5** grown from isopropyl alcohol solution. These two independent conformations are very similar to one another (Figure 4a); they are fully 12-helical, without any significant distortion around the  $\beta^3$ -hPhe residue. The C-terminal Aib residue curls away from the helical axis in each molecule and does not affect the 12-helical conformation. There are two types of lateral interactions between adjacent molecules: close packing between two symmetry-independent conformations (Figure 4b) and close packing between two identical molecules (Figure 4c). In both cases, the interacting molecules are parallel to one another. These crystal packing

interactions may provide guidance for future efforts to design 12-helical segments that participate in discrete tertiary or quaternary structures.<sup>5c</sup> There is space near the  $\alpha$ -carbon of the  $\beta^3$ -hPhe residue at the interface shown in Figure 4b because no substituent projects outward from this carbon (in contrast to ACPC and *dm*-ACPC residues). This space is occupied by an isopropyl alcohol molecule, which minimizes lateral contact between these two  $\beta$ -peptide molecules. In contrast, the interface highlighted in Figure 4c involves extensive intermolecular contact between cyclopentane rings of ACPC residues. This interaction is very similar to a lateral packing interaction previously observed in the crystal of an ACPC hexamer.<sup>5c</sup>

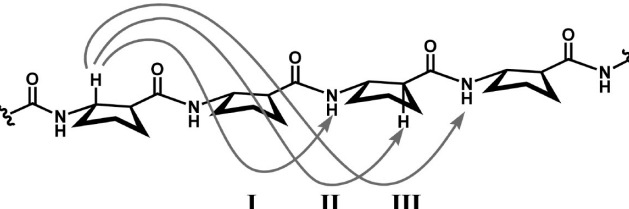
A second crystal form of heptamer **5** was obtained from methanol–water solution (see Supporting Information). These crystals did not provide diffraction data of high quality; nevertheless, it was possible to determine that this crystal form contained two symmetry-independent molecules, both of which displayed 12-helical conformations similar to those seen in the crystal form from isopropyl alcohol, though the crystal packing is different from the packing seen in Figure 4b,c. Overall, these crystallization results suggest that adoption of the 12-helical conformation by **5** is not determined by crystal packing forces but is instead intrinsically favorable for the  $\beta$ -peptide portion of this oligomer, despite the presence of a flexible  $\beta^3$ -residue.

**Relationship between Interproton Distances and NOE Patterns.** Medium-range nuclear Overhauser enhancements (NOEs) between protons on residues that are not adjacent in a



**Figure 4.** Crystal structure of heptamer **5**, based on a crystal grown from isopropyl alcohol: (a) overlay of two symmetry-independent molecules in the crystal, viewed perpendicular to the helical axis (top) and along the helical axis (bottom); (b) close packing between the two symmetry-independent molecules, viewed along the crystallographic *a* axis (top) and *b* axis (bottom); (c) close packing between two identical molecules, viewed along the crystallographic *c* axis (top) and *b* axis (bottom). Solvent molecules (isopropyl alcohol) are highlighted in cyan. Dashed lines indicate hydrogen bonds. Disordered atoms and most hydrogen atoms are omitted for clarity. Molecular surfaces for the two helical conformations are rendered as shaded areas.

**Table 1.** Average Interproton Distances (Å) Corresponding to Medium-Range NOE Patterns Observed for 12-Helical  $\beta$ -Peptides



	NOE type	distance <sup>a</sup>
<b>I</b>	$C_{\beta}H(i)-NH(i+2)$	3.5(4) [25]
<b>II</b>	$C_{\beta}H(i)-C_{\alpha}H(i+2)$	2.9(4) [21]
<b>III</b>	$C_{\beta}H(i)-NH(i+3)$	3.6(5) [21]

<sup>a</sup> The number in brackets indicates the number of measured and averaged distances.

peptide sequence can develop if the corresponding interproton distance is within 5 Å. Table 1 lists three types of medium-range NOEs between backbone protons that have been associated with the  $\beta$ -peptide 12-helix in solution<sup>5a,j</sup> and corresponding average interproton distances derived from the seven available 12-helical  $\beta$ -peptide crystal structures (the five structures in this paper and the two previously reported ACPC homooligomer structures<sup>5c</sup>). The three average distances for each of these types of proton pairing is <4 Å. This correlation confirms that the three NOE patterns in Table 1 are excellent indicators of 12-helical folding in solution. Inspection of the crystal structures

does not reveal any other type of medium-range NOE that would be expected between backbone proton pairs from nonadjacent  $\beta$ -amino acid residues within a 12-helical conformation.

We have previously reported that an unexpected NOE pattern,  $C_{\beta}H(i)-C_{\alpha}H(i+3)$ , was observed for a *dm*-ACPC-containing  $\beta$ -peptide hexamer, in addition to the three NOE types in Table 1.<sup>5j</sup> This unexpected NOE pattern is not consistent with a 12-helical conformation because the corresponding interproton distances are >5 Å in the available crystal structures. Interestingly, comparable NOE patterns have been observed for oligomers containing both  $\alpha$ - and  $\beta$ -amino acid residues (“ $\alpha/\beta$ -peptides”) in either a 2:1 and a 1:2 ratio, in which all  $\beta$ -residues are ACPC.<sup>12d</sup> Numerous crystal structures for these types of  $\alpha/\beta$ -peptides have revealed *i*, *i*+3  $C=O\cdots H-N$  hydrogen-bonded helical conformations, which are not consistent with the  $C_{\beta}H(i)-C_{\alpha}H(i+3)$  NOE pattern. We have proposed that the  $C_{\beta}H(i)-C_{\alpha}H(i+3)$  NOEs observed for the 2:1 and 1:2  $\alpha/\beta$ -peptides can be explained by alternative helical conformations involving *i*, *i*+4  $C=O\cdots H-N$  hydrogen bonds. Indeed, this alternative helix has been documented crystallographically for a 33-residue 2:1  $\alpha/\beta$ -peptide.<sup>14</sup> By analogy, the  $C_{\beta}H(i)-C_{\alpha}H(i+3)$  NOE pattern observed for the *dm*-ACPC-containing  $\beta$ -peptide hexamer could arise from an *i*, *i*+4  $C=O\cdots H-N$  hydrogen-bonded helical conformation, which would be designated a “16-helix”.<sup>5j</sup> Although this alternative

(14) Horne, W. S.; Price, J. L.; Gellman, S. H. *Proc. Natl. Acad. Sci. U.S.A.* **2008**, *105*, 9151.

**Table 2.** Average  $\beta$ -Residue Torsion Angles (deg) for the 12- and the 14-Helices

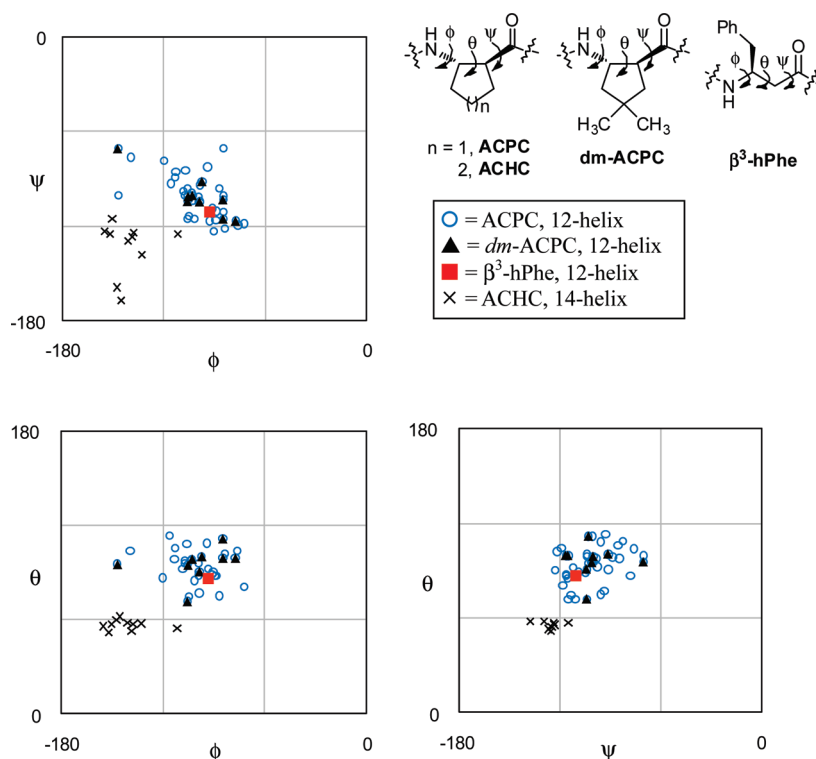
	12-helix <sup>a,e</sup>				14-helix <sup>a,e</sup>
	ACPC	<i>dm</i> -ACPC	$\beta^3$ -hPhe <sup>b</sup>	overall <sup>c</sup>	ACHC <sup>d</sup>
$\phi$	-100(15) [25]	-101(20) [9]	-93	-100(16) [35]	-141(13) [10]
$\psi$	-102(14) [25]	-101(14) [9]	-111	-102(14) [35]	-126(6) [9]
$\theta$	94(11) [25]	96(11) [9]	87	94(10) [35]	57(3) [10]

<sup>a</sup> The number in brackets indicates the number of measured and averaged distances. <sup>b</sup> Only one set of torsion angles was used. <sup>c</sup> Torsion angle averages identified from ACPC, *dm*-ACPC, and  $\beta^3$ -hPhe residues. <sup>d</sup> From ref 18; ACHC = (1*S*,2*S*)-2-aminocyclohexanecarboxylic acid. <sup>e</sup> For heptamer **5** and the two ACHC homooligomers, only one independent molecule was used.

helical conformation is not observed in the crystal structures of *dm*-ACPC-containing oligomers **1–4**, which are fully 12-helical, it is possible that both the 12- and 16-helical conformations are populated in solution. Previous 2D NMR analysis of 1:1  $\alpha/\beta$ -peptides led us to conclude that the analogous pair of helical conformations (designated the 11- and 14/15-helices, respectively) is populated in solution for examples containing <10 residues,<sup>15</sup> and subsequent crystallographic studies have verified the occurrence of distinct helical conformations of 1:1  $\alpha/\beta$ -peptides that contain either *i*, *i*+3 C=O...H-N or *i*, *i*+4 C=O...H-N hydrogen bonds.<sup>12b,c</sup> Comparable behavior is well-known among  $\alpha$ -peptides, involving the  $3_{10}$ -helix (*i*, *i*+3 C=O...H-N hydrogen bonds) and the  $\alpha$ -helix (*i*, *i*+4 C=O...H-N hydrogen bonds).<sup>16</sup> Among  $\alpha$ - and  $\alpha/\beta$ -peptides, the available data suggest that increasing the number of amino acid residues causes a preference for *i*, *i*+4 C=O...H-N hydrogen-bonded helices relative to *i*, *i*+3 C=O...H-N hydrogen-bonded helices. If this trend holds for  $\beta$ -peptides that are rich in residues with a five-membered ring constraint, then

it is possible that the 16-helical conformation will become dominant when the length of an ACPC-rich  $\beta$ -peptide is >10 residues.

**Backbone Torsion Angle Analysis.** The set of available crystal structures (five from this study and two from ref 5c) enables the identification of characteristic backbone torsion angle averages for the  $\beta$ -peptide 12-helix (Table 2). Figure 5 presents the crystallographically determined torsion angles for 12-helical  $\beta$ -peptides in Ramachandran-type plots.<sup>17</sup> For comparison, these plots contain also backbone torsion angles derived from the two 14-helical crystal structures of homooligomers of 2-aminocyclohexanecarboxylic acid (ACHC).<sup>18</sup> Torsion angles for left-handed helical structures (pentamer **3**, the two ACPC homooligomers,<sup>5c</sup> and one of the 14-helical structures<sup>18</sup>) were inverted for this comparison. For heptamer **5** and the two 14-helical ACHC homooligomers, torsion angles of only one independent molecule in the crystal are included. In all three Ramachandran-type plots, the ACPC and *dm*-ACPC residues fall in the same region, which is distinct from the region that is characteristic of the 14-helix. The  $\beta^3$ -hPhe residue in the structure of **5** falls into the region for the 12-helix instead of the region for the 14-helix, even though acyclic  $\beta^3$ -amino acid residues are generally understood to prefer 14-helical secondary structure relative to 12-helical secondary structure.<sup>2</sup> Table 2 shows that the local conformation of  $\beta^3$ -hPhe in **5** is adjusted to the 12-helical environment; for example, the  $\theta$  angle is 87°, which is substantially larger than  $\theta$  angles common for  $\beta^3$ -amino acid residues in a 14-helical environment ( $\sim 60^\circ$ )<sup>2a</sup> and close to the average  $\theta$  observed for residues with a five-membered ring constraint in a 12-helical environment (94°). The ability of the lone  $\beta^3$ -hPhe in **5** to conform to the 12-helical environment is consistent with the view that this type of  $\beta$ -amino acid



**Figure 5.** Ramachandran-type plots for right-handed helical  $\beta$ -peptides. Torsion angles for the 14-helix were measured from the crystal structures in ref 18. Torsion angles for left-handed helical structures have been inverted. For heptamer **5** and the two 14-helical ACHC homooligomers, torsion angles of only one independent molecule in the crystal are shown. ACHC = (1*S*,2*S*)-2-aminocyclohexanecarboxylic acid.



**Table 3.** Average Parameters for  $i, i+3$  C=O $\cdots$ H–N Hydrogen-Bonded Helices for  $\alpha$ -,  $\beta$ -, and  $\alpha/\beta$ -Peptides

peptide backbone (helix type)	res/turn $n$	rise/turn $p$ (Å)	rise/res $d$ (Å)	radius $r$ (Å)	$\beta$ - residue %
$\beta$ -peptide (12-helix)	2.7(1)	5.4(1)	2.0(1)	2.1(1)	100
1:2 $\alpha/\beta$ - peptide <sup>a</sup>	2.9	5.5	1.9	2.1	67
1:1 $\alpha/\beta$ -peptide (11-helix) <sup>b</sup>	2.8	5.6	2.0	2.1	50
2:1 $\alpha/\beta$ - peptide <sup>a</sup>	2.9	5.8	2.0	2.0	33
$\alpha$ -peptide (3 <sub>10</sub> -helix) <sup>c</sup>	3.2	5.8	1.8	2.0	0

<sup>a</sup> From ref 12d. <sup>b</sup> From ref 12c. <sup>c</sup> From ref 20.

residue is intrinsically flexible and can adapt to a variety of conformational contexts.

**Helical Parameter Analysis.** Average structural parameters for the  $\beta$ -peptide 12-helix were derived from five of the available crystal structures (oligomers containing five or more  $\beta$ -residues). Each helical parameter was calculated from sets of four consecutive  $\alpha$ -carbons by reported methods.<sup>19</sup> Nonhelical residues at C-termini were excluded from these calculations. This approach prevented the use of structures for tetramers **1** and **2** in the analysis. For heptamer **5**, only one independent conformation was used because all four independent conformations of **5** in the two crystal structures are quite similar.

The calculated parameters reveal the characteristic features of the  $\beta$ -peptide 12-helix. The rise-per-turn of the  $\beta$ -peptide 12-helix is 5.4 Å, which is the same as the pitch of the  $\alpha$ -helix and a bit smaller than that of the 3<sub>10</sub>-helix (5.8 Å). The radius of  $\beta$ -peptide 12-helix is 2.1 Å, which is slightly smaller than that of the  $\alpha$ -helix (2.3 Å) and comparable to that of the 3<sub>10</sub>-helix (2.0 Å).<sup>20</sup> Table 3 lists the parameters for five  $i, i+3$  C=O $\cdots$ H–N hydrogen-bonded helices adopted by  $\beta$ -peptides (the 12-helix),  $\alpha$ -peptides (the 3<sub>10</sub>-helix),<sup>20</sup> or  $\alpha/\beta$ -peptides.<sup>12c,d</sup>

The radii and the rise-per-residue values for these five helices are quite similar to one another. The rise-per-turn ( $p$ ) becomes slightly smaller as the proportion of  $\beta$ -residues increases, suggesting that fine-tuning of the helical pitch can be achieved by changing the proportion of  $\beta$ -residues. This correlation should be useful in design efforts that require specific arrangements of side chain groups along the helical surface.

## Conclusions

The five crystal structures disclosed here, along with two previously reported structures of ACPC homooligomers, have allowed us to identify canonical structural characteristics of the  $\beta$ -peptide 12-helix. The conformational behavior of *dm*-ACPC is seen to be very similar to that of ACPC, implying that functional groups could be placed at C4 of the five-membered ring without interfering with 12-helical propensity. Analysis of torsion angles and helical parameters suggests that the  $\beta$ -peptide 12-helix is comparable to the  $i, i+3$  C=O $\cdots$ H–N hydrogen-bonded helices formed by  $\alpha$ -peptides and by 1:1, 1:2, and 2:1  $\alpha/\beta$ -peptides. The structural insight that emerges from this study should facilitate the design of 12-helical  $\beta$ -peptides intended to perform specific functions.

**Acknowledgment.** We thank Dr. T. Peelen for providing synthetic materials. This work was supported in part by NSF Grant CHE-0848847 and NIH Grant GM-56414. S.H.C. was supported in part by a fellowship from the Samsung Scholarship Foundation. X-ray equipment purchase was supported in part by grants from the NSF.

**Note Added after ASAP Publication.** Chart 1 contained an error in the version published asap September 9, 2010; the correct version and an updated SI file were reposted September 15, 2010.

**Supporting Information Available:** Experimental details, characterization data, crystallographic information, full table of helical parameters, and alternative crystal structure of heptamer **5**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

**Note Added in Proof.** Very recently, Lee et al. have reported extraordinary self-assembly behavior of an ACPC heptamer: Kwon, S.; Jeon, A.; Yoo, S. H.; Chung, I. S.; Lee, H.-S. *Angew. Chem. Int. Ed.* **2010** *Early View* (DOI: 10.1002/anie.201003302).

JA1062532

- (15) Hayen, A.; Schmitt, M. A.; Ngassa, F. N.; Thomasson, K. A.; Gellman, S. H. *Angew. Chem., Int. Ed.* **2004**, *43*, 505.
- (16) (a) Karle, I. L.; Balaram, P. *Biochemistry* **1990**, *29*, 6747. (b) Toniolo, C.; Benedetti, E. *Trends Biochem. Sci.* **1991**, *16*, 350.
- (17) Ramachandran, G. N.; Sasisekharan, V. *Adv. Protein Chem.* **1968**, *23*, 283.
- (18) Appella, D. H.; Christianson, L. A.; Karle, I. L.; Powell, D. R.; Gellman, S. H. *J. Am. Chem. Soc.* **1999**, *121*, 6206.
- (19) (a) Sugeta, H.; Miyazawa, T. *Biopolymers* **1967**, *5*, 673. (b) Kahn, P. C. *Comput. Chem.* **1989**, *13*, 185.
- (20) Barlow, D. J.; Thornton, J. M. *J. Mol. Biol.* **1988**, *201*, 601.