$\begin{array}{l} Synthesis and biological evaluation of sodium \\ 2\alpha-(1,2,3-triazol-1-yl)methyl-2\beta-methyl-6,6-dihydropenam-3\alpha-carboxylate-1,1-dioxide\end{array}$

SN Maiti¹, P Spevak¹, AVN Reddy¹, RG Micetich¹, N Ishida², Y Miyake², K Ogawa²

¹SynPhar Laboratories Inc, #2, 4290-91A Street, Edmonton, Alberta, T6E 5V2 Canada;

²Tokushima Research Institute, Taiho Pharmaceutical Co Ltd, 224-2 Ebisuno Hiraishi, Kawauchi-cho, Tokushima, 771-01 Japan

(Received 17 March 1994; accepted 20 June 1994)

Summary — The synthesis and β -lactamase inhibitory activity of sodium $2\alpha - (1,2,3-\text{triazol-1-yl})$ methyl- 2β -methyl-6,6-dihydropenam- 3α -carboxylate-1,1-dioxide **2** (R = Na) is described. Its activity was compared with tazobactam **1** (R = Na). Compound **2** showed excellent synergism in combination with piperacillin (PIPC) and the results were either comparable or superior to tazobactam.

 β -lactam / β -lactamase inhibitor / penam sulfone

Introduction

Though the principal direction of recent B-lactam antibiotic research has been towards the synthesis of cephalosporins with enhanced antibacterial activity and superior stability to the bacterial β -lactamases, the current approach to overcoming the bacterial resistance to β -lactam antibiotics caused by β -lactamase production is to develop agents that can inhibit the action of the β -lactamase. The success of clavulanic acid [1] stimulated extensive research leading to the discovery of other β -lactamase inhibitors such as sulbactam [2] and YTR-830 [3, 4]. A number of 6-(substituted methylene)penams have been reported in the literature [5-8] as potent inhibitors of cell-free β -lactamases, but were ineffective in synergistic antibacterial tests probably because of poor penetration through the bacterial cell wall. More recently, 6-(substituted methylene)penems [9-13] have been shown to be very potent inhibitors of most bacterial β -lactamases including the class I β -lactamase, which is resistant to commercially available B-lactamase inhibitors. Although the penem derivative BRL-42715 [14-16] was discovered several years ago, there is little information about the further development of this compound. In our previous reports [3, 4] we have demonstrated that substitution of the 2B-methyl group of penicillanic acid sulfone molecule by a heterocyclic moiety (particularly by the triazole ring) provides a new class of very active β -lactamase inhibitors. One

member of this family of β -lactamase inhibitors, tazobactam 1 is being developed for clinical use. In a continuation of these studies with series 1, we pursued the closely related structure 2 since its derivatives should follow a similar mechanistic pathway and should also act as good suicide inhibitors. Here, we report a brief synthesis of 2 (scheme 1) and report on its synergy and β -lactamase inhibition data.

Chemistry

 2α -(Chloromethyl)penam 1 β -oxide **3** [17] was deoxygenated smoothly with phosphorus pentasulfide and pyridine [18] to give the 2α -(chloromethyl)penam **4**. The azido group at the 2α -methyl carbon was introduced by heating the corresponding 2α -(chloromethyl)penam **4** with excess sodium azide in dimethylformamide at 50–60°C for 4 h. Notably, no detectable amount of C(3)-substituted cepham was





Scheme 1.

isolated; this fact is in sharp contrast to the substitution reaction of 2β -(chloromethyl)penam with sodium azide affording a mixture of 2β -(azidomethyl)penam and 3β -azido- 3α -methyl cepham [19]. Oxidation of 2α -(azidomethyl)penam **5** was accomplished in 19% yield employing potassium permanganate in glacial acetic acid to provide **6**. Heating of **6** with acetylene in acetone gave the 2α -(triazolyl methyl)penam sulfone **7** as the sole product. Removal of the benzhydryl ester by hydrogenation over Pd/C afforded the 2α -(triazolylmethyl)penam- 3α -carboxylic acid-1,1-dioxide **2** (R = H) which was converted to the sodium salt and purified over a HP-20 column.

Results and discussion

The β -lactamase inhibitory activity of sodium 2α -(triazolylmethyl)penam- 3α -carboxylate-1,1-dioxide **2** (R = Na) and tazobactam **1** (R = Na) was determined by spectrophotometrically measuring the hydrolysis of the substrate (penicillin G) in the presence and absence of the β -lactamase inhibitors using penicillinase as β -lactamase. These results are summarized in table I.

The minimum inhibitory concentrations (MIC) of piperacillin (PIPC) in combination with the triazoles (10 μ g/ml) were determined against a series of β -lactamase-producing microorganisms (table II). The bacteria were cultivated in Mueller Hinton Broth

Table I. Inhibition of β -lactamase by inhibitor^a.

	I(R = Na)	2 (R = Na)
IC ₅₀ (µM)	0.57	5.09

^aSubstrate: PCG (200 μ M, sigma); enzyme: penicillinase from *B cereus* (5000 units, 30 μ l, Tokyo Kasei); preincubation 30°C, 5 min; incubation 30°C, 3 min.

(Difco) and diluted to 10⁷ cfu/ml and were then inoculated into the same medium containing piperacillin and the prepared triazoles in a specific concentration and incubated at 37°C for 20 h. The growth of the microorganisms was observed to determine the minimal inhibitory concentration (MIC) for rendering the inoculated medium free from turbidity.

Conclusion

The synergistic data (table II) indicate that the 2α -triazole (2, R = Na) exhibits excellent β -lactamase inhibitory activity and the results are either comparable or superior to tazobactam (1, R = Na) except against TEM-2 producing *E coli* and *Acinetobacter* 450 L, in which case tazobactam showed better synergism.

Experimental protocols

General procedures

Melting points were determined on a Thomas–Hoover melting point apparatus and are uncorrected. The ¹H-NMR spectra (δ ppm) were obtained in CDCl₃ with tetramethylsilane as an internal standard on a Varian EM-360 spectrometer. Infrared spectra were recorded with a Nicolet DX FTIR. Only significant maxima are listed. Analytical results for compounds followed by elemental symbols were \pm 0.4% of calculated values and were determined by the Department of Chemistry, University of Alberta. All column chromatographic purifications were accomplished on silica gel 60 (E Merck, 230–400 mesh) with the appropriate solvent gradients.

Benzhydryl 2α -chloromethyl- 2β -methyl-6,6-dihydropenam- 3α -carboxylate **4**

To a solution of benzhydryl 2α -chloromethyl- 2β -methyl-6,6dihydropenam- 3α -carboxylate 1β -oxide [17] (**3**, 0.208 g, 0.0005 mol) in methylene chloride (10 ml), were added pyridine (0.16 ml) and phosphorus pentasulfide (0.111 g), and the mixture was stirred at room temperature. After 3 h, an additional portion of phosphorus pentasulfide (0.08 g) was added and the reaction mixture was stirred at room temperature for a total period of 18 h. The mixture was diluted with water, extracted with methylene chloride, washed successively with water, 0.1 N HCl, water, NaHCO₃ solution and finally with brine, and dried over anhydrous Na₂SO₄. The organic layer was

Test organism	β -Lactamase	<i>ΜIC</i> (μg/ml)		
		PIPC	+ Tazobactam (R = Na)	+ Compound $2 (R = Na)$
E coli	SHV 1	12.5	0.78	0.78
E coli	TEM 1	50	0.78	0.78
E coli	TEM 2	> 200	0.78	100
E coli	OXA 1	25	1.78	0.78
E coli	OXA 3	3.13	0.78	0.78
K pneumoniae 366 L	SHV 1	100	6.25	3.13
K pneumoniae 101 L	TEM 1	> 200	3.13	3.13
P mirabilis 60		3.13	0.20	0.20
S marcescens 200 L	TEM + C	50	6.25	6.25
<i>C freundii</i> 2046 E	С	25	0.78	0.39
C freundii 962 L	TEM 2	100	3.13	1.56
E cloaceae P 99	С	200	6.25	6.25
Acinetobacter 450 L	TEM + C	> 200	0.78	12.5
P aeruginosa	PSE 2	25	12.5	12.5
P aeruginosa	PSE 3	50	6.25	3.13
P aeruginosa	PSE 4	100	12.5	12.5
S aureus 54 K		12.5	0.78	0.78
S aureus 123 K		0.78	0.39	0.78

Table II. In vitro synergy of compound 2 (R = Na) with piperacillin (PIPC) against β -lactamase-producing isolates.

filtered and evaporated *in vacuo* to give a brown viscous oil (0.1364 g, 67.92%). This product was directly used for the next step without further purification. ¹H-NMR (CDCl₃): 1.78 (s, 3H, CH₃); 3.08 (dd, J = 2 and 16 Hz, 1H, 6-H); 3.59 (dd, J = 4 and 16 Hz, 1H, 6-H); 3.58 (s, 2H, CH₂Cl); 4.78 (s, 1H, 3-H); 5.33 (dd, J = 2 and 4 Hz, 1H, 5-H); 7.0 (s, 1H, COOCHPh₂); 7.4 (broad s, 10H, aromatic).

Benzhydryl 2α -azidomethyl- 2β -methyl-6,6-dihydropenam- 3α carboxylate 1,1-dioxide **6**

A mixture of 0.4063 g (0.001 mol) of benzhydryl 2 α -chloromethyl-2 β -methyl-6,6-dihydropenam-3 α -carboxylate 4 sodium azide (0.394 g, 0.006 mol) in dimethyl formamide (8.1 ml) and water (2.6 ml) was heated at 60–65°C for 4 h. The mixture was diluted with water, extracted with chloroform, the chloroform extract was washed successively with small portions of icewater, dried over anhydrous Na₂SO₄ and concentrated to give a brown viscous oil (0.3443 g, 83.34%) of benzhydryl 2 α -azidomethyl-2 β -methyl-6,6-dihydropenam-3 α -carboxylate 5. This crude product was directly used for the next step.

Benzhydryl 2α -azidomethyl- 2β -methyl-6,6-dihydropenam- 3α -carboxylate **5** (0.4971 g, 0.0012 mol) from the previous step was dissolved in a mixture of glacial acetic acid (22 ml) and water (3 ml). Potassium permanganate (0.423 g, 0.0026 mol) was added portionwise, the mixture was stirred at room temperature for 3 h. Excess potassium permanganate was decomposed by adding 30% hydrogen peroxide, the mixture was diluted with ice-cold water, the precipitated solid was filtered off and dissolved in methylene chloride, washed with water, followed by NaHCO₃ solution and brine. The methylene chloride layer was dried (Na₂SO₄) and evaporated *in vacuo* to yield a viscous residue (0.3735 g). The product was purified by column chromatography on silica gel (3:1, hexane/ethyl acetate) to yield 0.1016 g of **6** (18.95% yield). IR (CHCl₃) cm⁻¹ 2114, 1802, 1745; ¹H-NMR (CDCl₃): 1.52 (s, 3H, CH₃); 3.4 (broad d, 2H, 6-H); 3.65 (broad d, 2H, CH₂N₃); 4.5 (s, 1H, 3-H); 4.68 (broad t, 1H, 5-H); 7.0 (s, 1H, COOCHPh₂); 7.4 (broad s, 10H, aromatic). Anal $C_{21}H_{20}N_4O_5S$ (C, H, N).

Benzhydryl 2α -(1,2,3-triazol-1-yl)methyl- 2β -methyl-6,6-dihydropenam- 3α -carboxylate-1,1-dioxide 7

A solution of benzhydryl 2 α -azidomethyl-2 β -methyl-6,6-dihydropenam-3 α -carboxylate 1,1-dioxide **6** (0.05 g) in acetone (4 ml) was heated in a sealed steel vessel with excess of acetylene (4.0 g) at 85°C for 24 h, cooled to room temperature, evaporation of the solvent gave a pale-yellow semisolid (0.0499 g) which on crystallization from ethyl acetate/hexane gave the pure triazole **7** (0.042 g, 79.72%); mp 156–159°C; ¹H-NMR (CDCl₃): 1.58 (s, 3H, CH₃); 3.43 (broad d, 2H, 6-H); 3.68 (broad t, 1H, 5-H); 4.8 (ABq, *J* = 13.4 Hz, 2H, CH₂-triazole); 4.72 (s, 1H, 3-H); 7.08 (s, 1H, COOCHPh₂); 7.4 (broad s, 10H, ar); 7.7 (s, 2H, triazole). Anal C₂₃H₂₂N₄O₅S (C, H, N).

Sodium $2\alpha \cdot (1,2,3-triazol-1-yl)$ methyl- 2β -methyl-6,6-dihydropenam- 3α -carboxylate-1,1-dioxide (2, R = Na)

A mixture of benzhydryl 2α -(1,2,3-triazol-1-yl)methyl-2\beta-methyl-6,6-dihydropenam-3 α -carboxylate-1,1-dioxide (7, 0.012 g), sodium bicarbonate (2.3 mg), 10% Pd/C (50 mg) in ethyl acetate (10 ml) and water (5 ml) was hydrogenated at a low pressure $(2-3 \text{ kg/cm}^2)$ at room temperature. The catalyst was removed by filtration and the aqueous layer was separated, washed with ethyl acetate, and then lyophilized to afford a white powder. The crude product was purified over MCI gel (CHP-20P, Mitsubishi Kasei Co) using water as eluent to afford a white powder of **2** (R = Na, 0.005 g, 60%); mp 235°C (decomp). ¹H-NMR (D₂O): 1.54 (s, 3H, CH₃); 3.40 (dd, J = 2 and 16 Hz, 1H, 6-H); 3.67 (dd, J = 4 and 16 Hz, 1H, 6-H); 4.97 (dd, J = 2 and 4 Hz, 1H, 5-H); 4.98 (s, 2H, CH₂ triazole); 5.09 (s, 1H, 3-H); 7.82 (s, 1H, triazole); 8.09 (s, 1H, triazole).

Acknowledgment

The authors wish to thank Taiho Pharmaceutical Co Ltd, Japan for their generous support of this work.

References

- 1 Howarth TT, Brown AG, King TJ (1976) J Chem Soc Chem Commun 266–267
- 2 English AR, Retsema JA, Girard AE, Lynch JE, Barth WE (1978) Antimicrob Agents Chemother 14, 414-419

- 3 Hall TW, Maiti SN, Micetich RG et al (1985) In: Recent Advances in the Chemistry of β-Lactam Antibiotics (Roberts SM, Brown AG, eds) Royal Society of Chemistry, London, UK, 242–254
- 4 Micetich RG, Maiti SN, Spevak P et al (1987) J Med Chem 30, 1469–1474
- 5 Chen YL, Chang CW, Hedberg K (1986) Tetrahedron Lett 27, 3449–3452
- 6 Chen YL, Chang CW, Hedberg K et al (1987) J Antibiot 40, 803-822
- 7 Angehrn P, Arisawa M (1982) J Antibiot 35, 1584-1589
- 8 Arisawa M, Then RL (1982) J Antibiot 35,1578–1583
- 9 Basker MJ, Osborne NF (1990) J Antibiot 43, 70–75
- 10 Broom NJP, Coleman K, Hunter PA, Osborne NF (1990) J Antibiot 43, 76–82
- 11 Bennett IS, Broom NJP, Bruton G et al (1991) J Antibiot 44, 331–337
- 12 Bennett IS, Broom NJP, Coleman K et al (1991) J Antibiot 44, 338–343
- 13 Bennett IS, Brooks G, Broom NJP, Calvert SH, Coleman K, Francois I (1991) J Antibiot 44, 969–978
- 14 Coleman K, Griffin DRJ, Page JWJ, Upshon PA (1989) Antimicrob Agents Chemother 33, 1580–1587
- 15 Woodnutt G, Berry V, Mizen L (1992) Antimicrob Agents Chemother 36, 1427–1431
- 16 Coleman K, Griffin DRJ, Upshon PA (1991) Antimicrob Agents Chemother 35, 1748–1752
- 17 Maiti SN, Spevak P, Ogawa K, Micetich RG (1988) J Org Chem 53, 3803-3807
- 18 Micetich RG (1976) Tetrahedron Lett 39, 971–974
- 19 Micetich RG, Maiti SN, Spevak P, Tanaka M, Yamazaki T, Ogawa K (1986) Synthesis 292–296