

## Preliminary communication

---

### Synthesis and immunoadjuvant activities of muramoyl-L-alanyl-D-isoglutamine and some carbohydrate analogs\*

MAKOTO KISO, YOSHIMI KANEDA, HIROYUKI OKUMURA, AKIRA HASEGAWA\*\*,

*Department of Agricultural Chemistry, Gifu University, Kakamigahara, Gifu 504 (Japan)*

ICHIRO AZUMA,

*Institute of Immunological Science, Hokkaido University, N-15 W-7, Sapporo 060 (Japan)*

and YUICHI YAMAMURA

*Third Department of Internal Medicine, Medical School, Osaka University Hospital, Fukushima-ku, Osaka 553 (Japan)*

(Received November 6th, 1979; accepted for publication, November 28th, 1979)

Since the discovery that *N*-acetylmuramoyl-L-alanyl-D-isoglutamine (4) is the minimal structure required for the immunoadjuvant activity of bacterial, cell-wall peptidoglycans (Ellouz *et al.*<sup>1</sup>, 1974; Merseur *et al.*<sup>2</sup> and Kotani *et al.*<sup>3</sup>, 1975), a great number of analogs thereof have been synthesized by the same groups<sup>4,5</sup>, and their immunological and antitumor activities investigated<sup>6,7</sup>.

Interest in elucidating the mode of action, especially the role of the carbohydrate moiety in muramoyl dipeptides, has stimulated our recent studies on the synthesis of a variety of carbohydrate analogs<sup>8</sup>. In the course of our investigations, it has been found that not only is the sugar moiety essential for adjuvant activity<sup>6a,8c</sup>, but also that the position and configuration of the linkage between the lactoyl-dipeptide and the sugar moiety are critically important<sup>8d,9</sup>. On the other hand, as described in an accompanying communication<sup>10</sup>, the 6-hydroxyl group can be replaced by an amino or acylamino group. In this communication, the chemical modification of the C-2 substituent in the sugar moiety and the resulting variation of immunoadjuvant activity are presented.

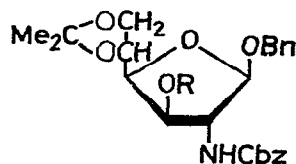
Benzyl 2-(benzyloxycarbonyl)amino-2-deoxy-5,6-*O*-isopropylidene- $\beta$ -D-glucofuranoside<sup>11</sup> (1a) was prepared by treatment of the 5,6-diol with 2,2-dibenzoyloxypropane in *N,N*-dimethylformamide containing a trace of *p*-toluenesulfonic acid. We found that, when 2,2-dimethoxypropane was used, 2-deoxy-D-*arabino*-hexose produced the methyl 2-deoxy-5,6-*O*-isopropylidene-D-*arabino*-hexofuranosides (3a),  $[\alpha]_D^{+33.2^\circ}$  (c 2.6, CHCl<sub>3</sub>), in 40% yield after column chromatography. 1,2:5,6-Di-*O*-isopropylidene- $\alpha$ -D-glucofuranose (2a) was synthesized by a conventional method, using acetone–sulfuric acid<sup>12</sup>.

The furanoid derivatives 1a, 2a, and 3a were condensed with L-2-chloropropanoic

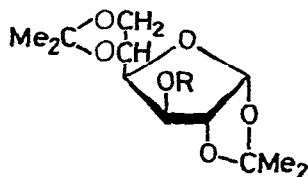
---

\*Studies on Immunoadjuvant Active Compounds, Part IV. For Part III, see ref. 8c.

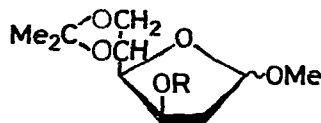
\*\*To whom enquiries should be addressed.



1a-c



2a-c



3a-c

Bn = PhCH<sub>2</sub>; Cbz = PhCH<sub>2</sub>OCO 1-3

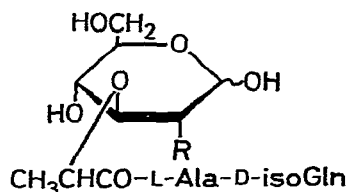
a, R = H

b, R =  $-\text{CHCO}_2\text{H}$   
 $\text{CH}_3$

c, R =  $-\text{CHCO-L-Ala-D-isogln}$   
 $\text{CH}_3$   $\text{OBn}$

acid in the presence of sodium hydride, to give the corresponding 3-*O*-(*D*-1-carboxyethyl) derivatives: **1b**,  $[\alpha]_D -28^\circ$  (*c* 0.5, CHCl<sub>3</sub>); **2b**,  $[\alpha]_D +2^\circ$  (*c* 1, CHCl<sub>3</sub>); and **3b**,  $[\alpha]_D +63.7^\circ$  (*c* 1.25, CHCl<sub>3</sub>), respectively. Couplings of **1b**–**3b** with *L*-alanyl-*D*-isoglutamine benzyl ester<sup>4a</sup> were conducted with dicyclohexylcarbodiimide and *N*-hydroxysuccinimide as activating agents, to afford the corresponding lactoyldipeptide derivatives: **1c**, m.p. 148.5°,  $[\alpha]_D -39^\circ$  (*c* 0.7, CHCl<sub>3</sub>); **2c**,  $[\alpha]_D -9.6^\circ$  (*c* 1.46, CHCl<sub>3</sub>); and **3c**,  $[\alpha]_D +34^\circ$  (*c* 0.96, CHCl<sub>3</sub>), in high yields.

Hydrolytic removal of the isopropylidene group of **1c** with 60% acetic acid, and subsequent hydrogenation with 10% palladium–carbon catalyst, followed by treatment with 0.1M hydrochloric acid, gave muramoyl-*L*-alanyl-*D*-isoglutamine hydrochloride (**5**), m.p. 159–163° (dec.),  $[\alpha]_D +55^\circ$  (*c* 0.58, H<sub>2</sub>O; equil.). Compounds **2c** and **3c** were first treated with 80–90% trifluoroacetic acid, to convert them into the pyranoid structures, and then hydrogenated, to yield the desired 2-hydroxy (**6**)  $\{[\alpha]_D +60.7^\circ$  (*c* 0.5, MeOH; equil.)} and 2-deoxy (**7**)  $\{[\alpha]_D +27.6^\circ$  (*c* 0.284, MeOH; equil.)} analogs, as amorphous materials after freeze-drying.



4 R = NHAc

5 R = NH<sub>2</sub>·HCl

6 R = OH

7 R = H

Immunoadjuvant activities (on the induction of delayed-type hypersensitivity to *N*-acetyl-L-tyrosine-3-azobenzene-4'-arsonate) of 4–7 were examined in guinea pigs<sup>13</sup>. Compound 5 showed potent activity that was even stronger than that of *N*-acetylmuramoyl-dipeptide 4. The D-glucose analog (6) had strong activity, comparable to that of 4, whereas the 2-deoxy-D-*arabino*-hexose analog (7) did not show any adjuvant activity. These results suggest that, for activity, the substituent on C-2 is not restricted to the acetamido group, but that the presence of a substituent such as an amino or a hydroxyl group at C-2 is essential for adjuvant activity.

New compounds obtained gave elemental analyses and i.r. and n.m.r. data in agreement with the structures assigned.

#### ACKNOWLEDGMENT

This work was supported, in part, by a cancer research grant (no. 401537) from the Japanese Ministry of Education.

#### REFERENCES

- 1 F. Ellouz, A. Adam, R. Ciorbaru, and E. Lederer, *Biochem. Biophys. Res. Commun.*, **59** (1974) 1317–1325.
- 2 C. Merse, P. Sinaÿ, and A. Adam, *Biochem. Biophys. Res. Commun.*, **66** (1975) 1316–1322.
- 3 S. Kotani, Y. Watanabe, F. Kinoshita, T. Shimono, I. Morisaki, T. Shiba, S. Kusumoto, Y. Tarumi, and K. Ikenaka, *Biken J.*, **18** (1975) 105–111.
- 4 (a) S. Kusumoto, Y. Tarumi, K. Ikenaka, and T. Shiba, *Bull. Chem. Soc. Jpn.*, **49** (1976) 533–539; (b) T. Shiba, S. Okada, S. Kusumoto, I. Azuma, and Y. Yamamura, *ibid.*, **51** (1978) 3307–3310.
- 5 (a) A. Adam, M. Devys, V. Souvannavong, P. Lefrancier, J. Choay, and E. Lederer, *Biochem. Biophys. Res. Commun.*, **72** (1976) 339–346; (b) P. Lefrancier, M. Derrien, I. Lederman, F. Nief, J. Choay, and E. Lederer, *Int. J. Pept. Protein Res.*, **11** (1978) 289–296.
- 6 (a) I. Azuma, K. Sugimura, T. Taniyama, M. Yamawaki, Y. Yamamura, S. Kusumoto, S. Okada, and T. Shiba, *Infect. Immun.*, **14** (1976) 18–27; (b) Y. Yamamura, I. Azuma, K. Sugimura, M. Yamawaki, M. Uemiya, S. Kusumoto, S. Okada, and T. Shiba, *Gann*, **67** (1976) 867–877; *Proc. Jpn. Acad.*, **53** (1977) 63–66.
- 7 (a) L. Chedid, F. Audibert, P. Lefrancier, J. Choay, and E. Lederer, *Proc. Natl. Acad. Sci. U.S.A.*, **73** (1976) 2472–2475; (b) C. Leclerc, L. Löwy, and L. Chedid, *Cell. Immunol.*, **38** (1978) 286–293.
- 8 (a) A. Hasegawa, K. Bito, and M. Kiso, *Gifu Daigaku Nogakubu Kenkyu Hokoku*, **40** (1977) 95–100; (b) A. Hasegawa, Y. Kaneda, M. Amano, M. Kiso, and I. Azuma, *Agric. Biol. Chem.*, **42** (1978) 2187–2189; (c) A. Hasegawa, H. Okumura, and M. Kiso, *Gifu Daigaku Nogakubu Kenkyu Hokoku*, **42** (1979) 171–177; (d) H. Okumura, Y. Kaneda, and A. Hasegawa, *Natl. Meet. Agric. Chem. Soc. Jpn. (Tokyo)*, (1979) 3L-3; Y. Kaneda, H. Okumura, Y. Gooh, M. Mitsui, and A. Hasegawa, *ibid.*, 3L-4.
- 9 M. Kiso, Y. Kaneda, A. Hasegawa, I. Azuma, and Y. Yamamura, manuscript in preparation.
- 10 A. Hasegawa, H. Okumura, M. Kiso, I. Azuma, and Y. Yamamura, *Carbohydr. Res.*, **79** (1980) C20–C23.
- 11 A. Hasegawa, N. Aritake, and M. Kiso, *Carbohydr. Res.*, **52** (1976) 137–149.
- 12 O. T. Schmidt, *Methods Carbohydr. Chem.*, **2** (1963) 318–325.
- 13 I. Azuma, K. Sugimura, Y. Yamamura, S. Kusumoto, U. Tarumi, and T. Shiba, *Jpn. J. Microbiol.*, **20** (1976) 63–66.