

**SYNTHESIS AND *IN-VITRO* STUDY OF SOME MEDICINALLY IMPORTANT
MANNICH BASES DERIVED FROM 2-AMINO-9 [(1,3 DIHYDROXY PROPANE-2YL) OXY } METHYL]
6-9 DIHYDRO-3H-PURIN-6-ONE**

SHEELA JOSHI^{A*}, PURTI BILGAIYAN^A AND ANJU PATHAK^A

^a School of Chemical Sciences, D.A.V.V., Indore 452001, M.P., India
(Received: January 4, 012 - Accepted: May 25, 2012)

ABSTRACT

A series of biologically active Mannich bases with heteroaromatic ring system have been synthesized employing Mannich reaction of 2-amino-9 [(1,3 dihydroxy propane-2yl) oxy } methyl] 6-9 dihydro-3H-purin-6-one (potent drug) with biologically active sulphonamides and secondary amines. They were analyzed by elemental analysis and characterized by spectral studies- UV, IR, ¹H NMR, Powder X-ray diffraction and Scanning Electron Microscopy. The Mannich bases were screened for their antibacterial activity against various gram + and gram - bacteria. The results had shown that the Mannich bases are quiet active against pathogens under study at varying concentrations. The toxicity of synthesized Mannich bases was ascertained by LD₅₀ test.

Key words: 2-amino-9 [(1,3 di hydroxy propane-2yl) oxy } methyl] 6-9 dihydro-3H-purin-6-one (Ganciclovir), Sulphonamides, Mannich reaction, Mannich bases, Antibacterial activity , LD₅₀ test.

1. INTRODUCTION

Heterocyclic compounds are of immense biological and industrial importance, Majority of pharmaceuticals are heterocyclic¹. 2-amino-9 [(1,3 di hydroxy propane-2yl) oxy } methyl] 6-9 dihydro-3H-purin-6-one, an antiviral agent derived from guanine (heterocyclic compound) are used in the prevention and treatment of opportunistic cytomegalovirus (CMV) infection in HIV infected patients and inhibits replication of herpes virus in-vitro. It helps in keeping CMV infection under control². The sulphonamide nucleus has well known pharmacological properties: antibacterial³, anticancer⁴, anti-inflammatory⁵, carbonic inhibitory⁶, insecticidal⁷. Even N-morpholino, piperidino, dimethylamino, diethyl amino Mannich bases show local anesthetic⁸ and hydrophilic property⁹.

These considerations provoked us to synthesize Mannich bases of 2-amino-9 [(1,3 di hydroxy propane-2yl) oxy } methyl] 6-9 dihydro-3H-purin-6-one using sulphonamides and secondary amines by Mannich reaction, which may have biological significance. This reaction offers a convenient route to introduce amino methyl chain, which alters the biological profile and physico-chemical characteristics of the drug¹⁰⁻¹².

Various drugs obtained from Mannich reaction have proved to be less toxic and more effective than their parent drug¹³. The versatile utility of Mannich bases in polymers¹⁴, dispersants¹⁵ and pharmaceutical chemistry¹⁶ prompted us to prepare aminomethyl derivatives and evaluate their biological significance. Here we reports antibacterial activity of Mannich bases of 2-amino-9 [(1,3 di hydroxyl propane-2yl) oxy } methyl] 6-9 dihydro-3H-purin-6-one including sulphonamide and secondary amines and their comparative study (Table 1).

In the present work, Mannich bases of 2-amino-9 [(1, 3 di hydroxy propane-2yl) oxy } methyl] 6-9 dihydro-3H-purin-6-one were synthesized by condensation of different primary/secondary amines with formaldehyde. The substrate first reacts with formaldehyde to form an intermediate, which then react with sulphonamides/secondary amines in the presence of acid. The synthesized Mannich bases contains amino methyl group (as justified from the spectral data), which changes the physiochemical characteristic of the drugs and makes it less toxic¹⁷⁻¹⁸. The synthesized Mannich bases were analyzed for elemental analysis and characterized by spectral studies- UV, IR, ¹H NMR, Powder X-ray diffraction and Scanning Electron Microscopy.

All the synthesized compounds were evaluated for antibacterial activity against the pathogenic strains of *S.typhi*, *K.pneumoniae*, and *P.aeruginosa* at varying concentrations 80, 160, 320 mg/ml. All the Mannich bases had shown significant activity.

2. EXPERIMENTAL

All the melting points were determined in open capillary tubes and are

uncorrected. Thin layer chromatography was used for monitoring the reaction and to check purity. UV spectra were studied on Shimadzu UV-160A, UV-visible spectrophotometer; IR spectra (KBr) were recorded as potassium bromide pellets on Shimadzu 820 IPC FTIR spectrometer and ¹HNMR spectra on Bruker DRX-300 FT NMR Spectrometer and chemical shifts were expressed as (ppm) values against tetramethylsilane (TMS) as internal reference. The XRD measurements were carried out on Bruker D8 Advance X-ray diffractometer using CuK α at a wavelength of 1.54 Å. SEM studies were performed with a Jeol JSM 5600 instrument having magnification range \times 18 to \times 300,000 and at an accelerating voltage of 0.5 to 30kV. The chemical reagents used in the synthesis were purchased from E. Merck and Aldrich.

2.1 Synthesis

2.1.1. Synthesis of Mannich bases from primary amines (3a-3e)

Synthetic pathway for the synthesis of compounds **3a-3e** is represented in **Scheme-1**. In ethanolic solution of 0.01 mol of Substrate (Comp.-1), 0.01 mol of sulfonamide and 2.5 mL of formaldehyde solution (37% v/v) were added. The mixture was kept in an efficient ice cooling for half an hour and then refluxed on water bath. The refluxed mixture was kept at 0°C for four days when crystalline product was obtained. The obtained product was recrystallized with dry distilled ethanol and DMF (1:1). The Mannich bases (**3a-3e**) were thus obtained in (\geq 85%) yield.

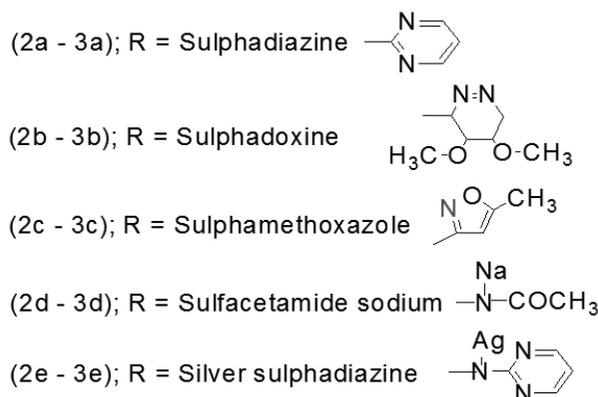
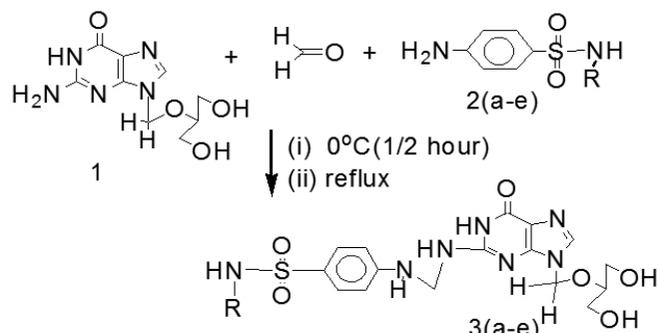
2.1.2. Synthesis of Mannich bases from secondary amines (3f-3j)

Synthetic pathway for the synthesis of compounds **3f-3j** is represented in **Scheme-2**. Secondary amine 0.01 mol was added in an ethanolic solution 50 mL of Substrate (Comp. -1) 0.01 mol in a flat bottom flask. Amount of 0.4 mL of formaldehyde solution (37%) was added slowly with constant stirring. The reaction mixture was stirred at 70-75°C for 3.0 to 8.5 hours, depending upon the secondary amine. The remaining portion of formaldehyde solution was added in two installments after 1 and 2 hours, respectively. The reaction mixture was kept over night in the refrigerator. Next day, the excess of solvent was distilled off from the reaction mixture under reduced pressure. It was again kept for crystallization in the refrigerator. The product obtained was purified by recrystallization from dry distilled ethanol and DMF (1:1). The Mannich bases (**3f-3j**) were thus obtained in (\geq 85%) yield.

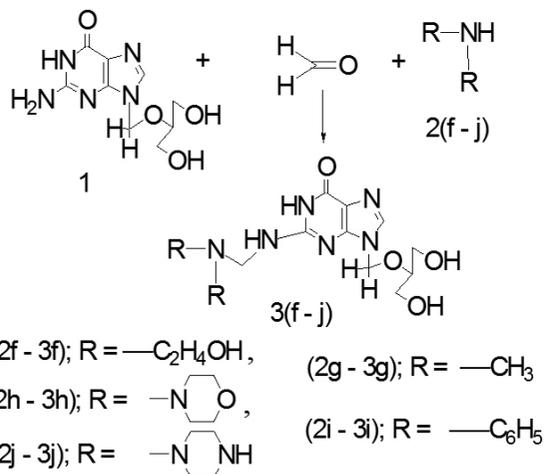
2.2 Spectral Studies

Compound 3a: C₂₀H₂₃N₉O₅S, m.p. 160-163°C. Anal. Calcd C,46.40; H,4.58; N,24.36 Found C,46.16;H,4.42; N,24.23. UV (λ max) nm: 208 (C=O), 190 (C=N), 205 (sulfonamide group), 186, 207, 251 (for benzene chromophore), 254 (sulfonamide moiety). IR (KBr) ν max in cm⁻¹: 3442 u_s N-H, 3398 u_s N-H in SO₂NH, 3302 u_s O-H, 2940 u_s CH₂, 1688 u_s C=O, 1345 u_s S=O, 1130 C-H in plane bending vibration of 1:4 disubstituted benzene. ¹H-NMR (DMSO) δ ppm: 5.04 (s, 2H, CH₂), 5.5 (s, 2H, CH₂ attached to purine

ring), 6.36 (s, 1H, NH of sulphonamide), 7.93 (s, 1H, CH of purine ring), 7.84 (s, 1H, free OH of Comp.1), 3.72(2CH₂ attached with purine ring), 8.65 (N¹H of purine ring), 9.03 (s, 1H of SO₂NH), 6.6 – 7.2 (m, ring proton of sulphonamide),



Scheme 1. Synthesis of Mannich bases from primary amines.



Scheme 2. Synthesis of Mannich bases from secondary amines.

Compound 3b: C₂₀H₂₂N₉O₈S, m.p. 105-107°C. Anal. Calcd. C, 45.74; H, 4.71; N, 21.82; Found C, 45.61; H, 4.46; N, 21.64. UV (λ max) nm: 210 (C=O), 197 (C=N), 208 S=O, 182, 205, 250 for benzene chromophore; 258 sulphonamide moiety. IR (KBr) ν max in cm⁻¹: 3440 u_s N-H, 3380 u_s N-H in SO₂NH, 3302 u_s O-H, 2910 u_s C-H in CH₂, 1653 u_s C=O, 1380 u_s S=O, 1135 C-H in plane bending vibration of 1:4 disubstituted benzene; ¹H NMR (DMSO) δ ppm: 3.43 (s, 3H of OCH₃), 5.04 (s, 2H, CH₂ attached to purine ring), 6.4 (s, 1H, NH of sulphonamide), 7.9 (s, 1H, CH of purine ring), 7.4 (s, 1H, free OH of Comp.1), 3.74(2CH₂ attached with purine ring), 8.66 (N¹H of purine ring) 9.2 (s, 1H of SO₂NH), 6.6 – 7.07 (m, ring proton of sulphonamide).

Compound 3c: C₂₀H₂₄N₈O₈S; m.p.180-181°C. Anal. Calcd. C, 46.13; H, 4.65; N, 21.53; Found C, 46.13; H, 4.25; N, 21.33. UV (λ max) nm: 201 (C=O), 192 (C=N), 208 (S=O), 186, 207, 250 for benzene chromophore; 255(sulphonamide moiety). IR (KBr) ν max in cm⁻¹: 3475 u_s N-H, 3350 u_s N-H in SO₂NH, 3318 u_s O-H, 2915 u_s C-H in CH₂, 1685 u_s C=O, 1349 u_s S=O, 1110 C-H in plane bending vibration of 1:4 disubstituted benzene. ¹H NMR (DMSO) δ ppm: 2.71(s, 3H of CH₃), 5.04 (s, 2H, CH₂ attached to purine ring), 6.3 (s, 1H, NH of sulphonamide), 7.93 (s, 1H, CH of purine ring), 7.84 (s, 1H, free OH of Comp.1), 3.73(2CH₂ attached with purine ring), 8.65 (N¹H of purine ring), 9.03 (s, 1H of SO₂NH), 6.6 – 7.2 (m, ring proton of sulphonamide).

Compound 3d: C₁₈H₂₂N₇O₇NaS; m.p. 90°C. Anal. Calcd. C, 42.93; H, 4.41; N, 19.48 Found C, 42.52; H, 4.21; N, 19.22. UV (λ max) nm: 208 (C=O), 190 (C=N), 205 S=O, 186, 207, 251 for benzene chromophore; 254 sulphonamide moiety. IR (KBr) ν max in cm⁻¹: 3482 u_s N-H, 3351 u_s N-H in SO₂NH, 3295 u_s O-H, 2950 u_s C-H in CH₂, 1647 u_s C=O, 1385 u_s S=O, 1109 disubstituted benzene. ¹H NMR (DMSO) δ ppm: 2.16 (s, 3H acetamide CH₃), 5.04 (s, 2H, CH₂), 5.5 (s, 2H, CH₂ attached to purine ring), 6.3 (s, 1H, NH of sulphonamide), 7.9 (s, 1H, CH of purine ring), 6.7 (s, 1H, OH of Comp.1), 3.72(2CH₂ attached with purine ring), 8.65 (N¹H of purine ring) 9.42 (s, 1H of SO₂NH), 6.6 – 7.07 (m, ring proton of sulphonamide).

Compound 3e: C₂₀H₂₂N₉O₆SAg; m.p. 222°C. Anal. Calcd. C, 38.46; H, 3.55; N, 20.19 Found C, 38.32; H, 3.51; N, 20.24. UV (λ max) nm: 203 (C=O), 194 (C=N), 208 S=O, 180, 203, 250 for benzene chromophore; 257 sulphonamide moiety. IR (KBr) ν max in cm⁻¹: 3496 u_s N-H, 3348 u_s N-H in SO₂NH, 3300 u_s O-H, 2952 u_s C-H in CH₂, 1638 u_s C=O, 1355 u_s S=O, 1100 C-H in plane bending vibration of 1:4 disubstituted benzene. ¹H NMR (DMSO) δ ppm: 5.04 (s, 2H, CH₂), 5.5 (s, 2H, CH₂ attached to purine ring), 6.1 (s, 1H, NH of sulphonamide), 7.9 (s, 1H, CH of purine ring), 6.72 (s, 1H, OH of Comp.1), 3.70(2CH₂ attached with purine ring), 8.66 (N¹H of purine ring), 9.03 (s, 1H of SO₂NH) 6.6 – 7.2 (m, ring proton of sulphonamide).

Compound 3f: C₁₄H₂₄N₆O₆; m.p. 160-162°C. Anal. Calcd. C, 45.14; H, 6.49; N, 22.56 Found C, 45.13; H, 6.51; N, 22.50. UV (λ max) nm: 208 (C=O), 191 (C=N), 185, 205, 250 for benzene chromophore. IR (KBr) ν max in cm⁻¹: 3482 u_s N-H, 3352 u_s O-H, 2944 u_s C-H in CH₂, 1680 u_s C=O. ¹H NMR (DMSO) δ ppm: 4.51 (s, 2H, CH₂), 5.60 (s, 2H, CH₂ attached to purine ring), 7.9 (s, 1H, CH of purine ring), 4.92 (s, 1H, OH of Comp.1), 3.72(2CH₂ attached with purine ring), 8.65 (N¹H of purine ring).

Compound 3g: C₁₂H₂₀N₆O₄; m.p. 225-227°C. Anal. Calcd. C, 46.13; H, 6.46; N, 26.90 Found C, 46.4; H, 6.51; N, 26.82. UV (λ max) nm: 210 (C=O), 195 (C=N), 185, 205, 255 for benzene chromophore. IR (KBr) ν max in cm⁻¹: 3412 u_s N-H, 3310 u_s O-H, 2909 u_s C-H in CH₂, 1655 u_s C=O. ¹H NMR (DMSO) δ ppm: 2.23(s, 3H of methyl group), 4.48 (s, 2H, CH₂), 5.5 (s, 2H, CH₂ attached to purine ring), 7.9 (s, 1H, CH of purine ring), 6.12 (s, 1H, OH of Comp.1) 3.72(2CH₂ attached with purine ring), 8.65 (N¹H of purine ring).

Compound 3h: C₁₄H₂₂N₆O₅; m.p. 215-218°C. Anal. Calcd. C, 47.43; H, 6.26; N, 23.71 Found C, 47.32; H, 6.30; N, 23.52. UV (λ max) nm: 204 (C=O), 190 (C=N), 184, 209, 254 for benzene chromophore. IR (KBr) ν max in cm⁻¹: 3477 u_s N-H, 3361 u_s O-H, 2947 u_s C-H in CH₂, 1630 u_s C=O, 1244 u_s C-O. ¹H NMR (DMSO) δ ppm: 4.4 (s, 2H, CH₂), 5.5 (s, 2H, CH₂ attached to purine ring), 7.93 (s, 1H, NH of purine ring), 6.21 (s, 1H, OH of Comp.1) 3.72(2CH₂ attached with purine ring), 8.65 (N¹H of purine ring).

Compound 3i: C₂₂H₂₄N₆O₄; m.p. 235-236°C. Anal. Calcd. C, 60.52; H, 5.55; N, 19.25 Found C, 60.61; H, 5.32; N, 19.10. UV (λ max) nm: 209 (C=O), 190 (C=N), 184, 206, 260 for benzene chromophore. IR (KBr) ν max in cm⁻¹: 3406 u_s N-H, 3318 u_s O-H, 2932 u_s C-H in CH₂, 1643 u_s C=O. ¹H NMR (DMSO) δ ppm: 4.5 (s, 2H, CH₂), 5.5 (s, 2H, CH₂ attached to purine ring), 7.9 (s, 1H, CH of purine ring), 6.17 (s, 1H, OH of Comp.1), 3.72(2CH₂ attached with purine ring), 8.60 (N¹H of purine ring).

Compound 3j: C₁₄H₂₂N₇O₃; m.p. 220°C. Anal. Calcd C, 47.57; H, 6.56; N, 27.74 Found C, 47.80; H, 6.32; N, 27.62. UV (λ max) nm: 205 (C=O), 190 (C=N), 184, 206, 260 for benzene chromophore. IR (KBr) ν max in cm⁻¹: 3458 u_s N-H, 3342 u_s O-H, 2937 u_s C-H in CH₂, 1639 u_s C=O. ¹H NMR (DMSO) δ ppm: 4.5 (s, 2H, CH₂), 5.5 (s, 2H, CH₂ attached to purine ring), 7.9 (s, 1H, CH of purine ring), 5.5 (s, 1H, OH of Comp.1) 3.72(2CH₂ attached with purine ring), 8.67 (N¹H of purine ring).

Powder X-ray diffraction studies

Powder X-ray diffraction patterns of three of the synthesized compounds namely ganciclovir methyl silver sulphadiazine (3e), ganciclovir methyl diphenylamine (3i) and ganciclovir methyl Piperazine (3j) (Fig 1,2 and 3 respectively) have been reported and important structural information have been shown in table 1,2,and 3 respectively.

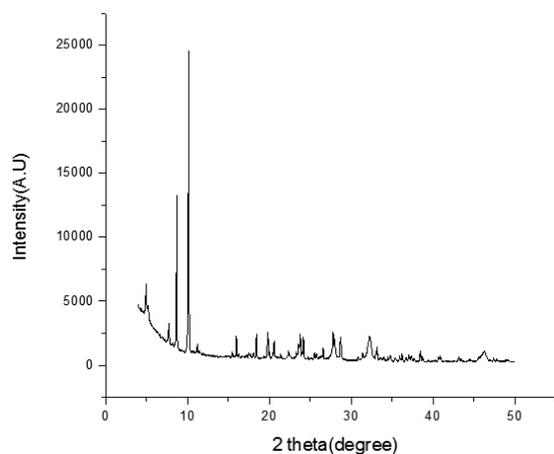


Figure 1. Powder X-ray diffraction pattern of Ganciclovir methyl Sulphadiazine (3e).

Average particle size of the synthesized compound was determined with the help of the Scherrer formula, in which particle size D is defined as $0.9\lambda / B \cos \theta$,

Where 0.9 = constant, λ = wavelength, B = angular width

And θ = diffraction angle. Average particle size of the compound determined were 94.18, 32.03 and 64.13 nm respectively. Diffraction data of the compounds are listed in table 1,2 and 3. All of our synthesized compounds had a

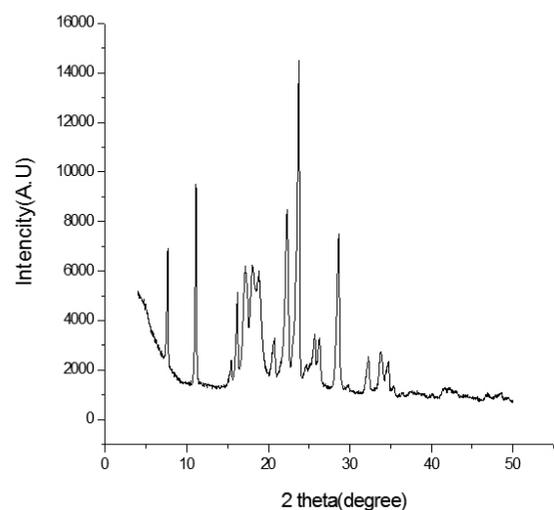


Figure 2. Powder X- Ray diffraction pattern of Ganciclovir methyl diphenylamine (3i).

Orthorhombic crystal system and XRD patterns showed that all are polycrystalline in nature. The largest relative deviation between the calculated and experimental d value was nearly equal to one, which indicates that all the synthesized compounds are multiphase compounds. Interplaner d spacing and unit cell volume of the synthesized compound were calculated by the formulae:

$$1/d^2 = h^2/a^2 + k^2/b^2 + l^2/c^2$$

$$V = abc$$

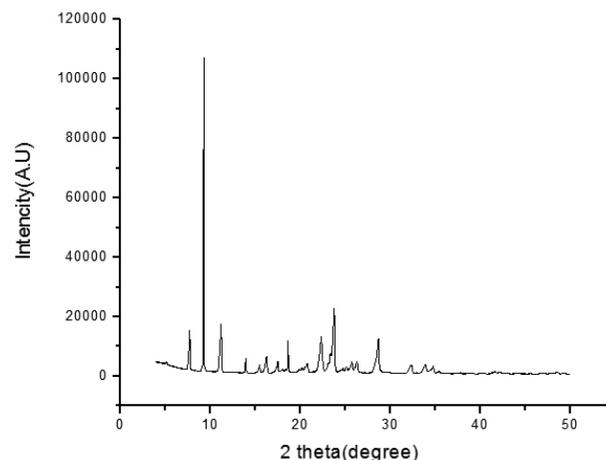


Figure 3. Powder X-ray diffraction pattern of Ganciclovir methyl Piperazine (3j).

Table 1 . The experimental data and the calculated results for powder X-ray pattern of the ganciclovir methyl Silver Sulphadiazine(3e).

2θ	d spacing(\AA°) Observed	d spacing(\AA°) Calculated	Relative intensity (%)	h k l
19.79	4.48	4.46	98.96	210
20.58	4.31	4.14	73.17	023
22.37	3.96	4.00	43.61	220
24.19	3.67	3.67	84.83	004
25.53	3.48	3.48	37.33	141
26.59	3.34	3.04	50.69	043
27.79	3.20	3.32	100	133
29.17	3.05	3.31	19.97	114
30.81	2.89	2.73	22.32	152
31.98	2.79	2.85	46.99	134
33.09	2.70	2.46	55.81	134
34.50	2.59	2.59	25.63	060
35.94	2.49	2.40	27.90	045
37.08	2.42	2.42	32.71	340
38.43	2.33	2.33	42.30	400
39.45	2.28	2.26	15.31	260
40.89	2.20	2.36	29.36	106
43.17	2.09	2.46	24.98	161
45.99	1.97	1.98	31.87	360
47.78	1.90	2.20	19.51	421
48.89	1.86	2.13	15.74	171
49.21	1.84	1.81	17.32	520

Lattice parameters and angle calculated are: $a = 9.32 \text{ \AA}^\circ$ or 0.93nm , $b = 15.54 \text{ \AA}^\circ$ or 1.55nm , $C = 14.68 \text{ \AA}^\circ$ or 1.46nm , $\beta = 94.18^\circ$. Unit cell volume of the complex: $2,126.14 \times 10^{-8} \text{ cm}^3$

Peaks having important characteristics information have been identified with the help of standard diffraction card JCPDS 31-1859, 38-1709, 49-2499 and 35-1687 and indicate formation of nanosized, polycrystalline, two phase and low symmetry compounds.

Table2. The experimental data and the calculated results for powder X-ray pattern of the ganciclovir methyl biphenyl amine(3i).

2θ	d spacing (A°) Observed	d spacing (A°) Calculated	Relative intensity (%)	h k l
10.97	8.05	8.00	27.38	110
14.87	5.94	5.94	10.07	120
20.76	4.27	4.24	22.82	102
21.99	4.03	4.00	31.25	220
23.75	3.74	4.20	100	201
24.74	3.59	3.56	15.36	140
27.97	3.18	3.18	10.12	003
29.05	3.06	3.34	10.54	202
30.46	2.14	3.01	8.14	103
32.30	2.76	3.28	17.48	212
36.46	2.46	2.61	7.52	302
37.61	2.38	2.96	7.94	301
38.40	2.34	2.34	7.63	400
40.04	2.24	2.25	7.28	260
44.00	2.05	2.40	7.03	161
45.02	2.01	1.98	6.48	360
46.81	1.93	1.93	7.01	080
47.04	1.92	2.18	6.98	421
48.54	1.87	2.10	7.49	171
49.15	1.85	1.81	6.18	520

Lattice parameters and angle calculated are: $a = 9.36 \text{ \AA}$ or 0.93 nm , $b = 15.44 \text{ \AA}$ or 1.54 nm , $C = 9.54 \text{ \AA}$ or 0.954 nm , $\beta = 32.03^\circ$. Unit cell volume of the complex: $1,378.70 \times 10^{-8} \text{ cm}$.

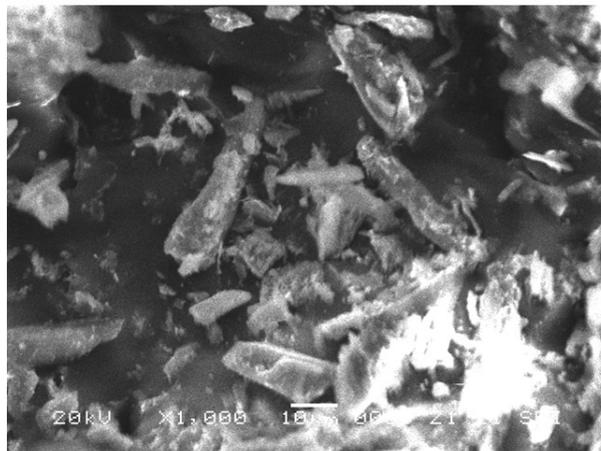
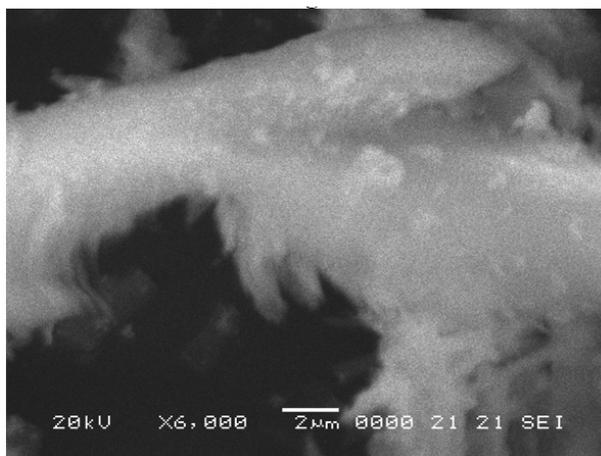
Table3. The experimental data and the calculated results for powder X-ray pattern of the ganciclovir methyl Piperazine(3j).

2θ	d spacing(A°) Observed	d spacing(A°) Calculated	Relative intensity (%)	h k l
19.85	4.53	4.53	10.43	200
22.25	4.06	4.12	46.49	113
23.78	3.81	3.81	100	040
25.71	3.54	3.51	21.29	140
27.97	3.28	3.38	6.57	042
28.67	3.20	3.27	55.90	133
30.08	3.07	3.51	4.90	140
31.98	2.90	2.80	7.92	320
34.76	2.70	2.72	14.79	025
39.16	2.43	2.43	4.59	006
40.16	2.38	2.95	4.99	301
41.68	2.31	2.83	6.86	151
43.06	2.25	2.21	4.76	260
44.26	2.20	2.52	4.55	144
46.81	2.11	2.14	4.96	206
47.89	2.07	2.41	6.18	161
49.39	2.02	1.94	4.30	360

Lattice parameters and angle calculated are: $a = 9.06 \text{ \AA}$ or 0.90 nm , $b = 15.24 \text{ \AA}$ or 1.52 nm , $C = 14.58 \text{ \AA}$ or 1.45 nm , $\beta = 64.13^\circ$. Unit cell volume of the complex: $2,013.12 \times 10^{-8} \text{ cm}$.

Scanning Electron Microscopy (SEM) Studies

Scanning Electron Microscopy Studies of two of the synthesized compound namely ganciclovir methyl silver sulphadiazine (3e), and ganciclovir methyl Piperazine (3j) have been carried out at a magnification of $\times 1,000 - 10 \mu\text{m}$ and $\times 600 - 2 \mu\text{m}$. SEM uses a focused beam of high energy electrons to generate a variety of signals from the surface of a sample, the signals reveal information about the sample, including external morphology, topography, chemical composition crystalline structure, and orientation of materials making up the sample.

**Figure 4****Figure 5**

We recorded different magnifications, as described above. Compound ganciclovir methyl silver sulphadiazine (3e) (Fig 4 and 5) has a different topography and morphology at the two different magnifications. In the images, the particles exhibit different sizes and shapes, and are present in the form of closed polygonal structure. In contrast, the compound ganciclovir methyl Piperazine (3j) (Fig 6 and 7) shows at different magnifications a finer morphology as compared to the other compound, appears in form of simple polygonal structures.

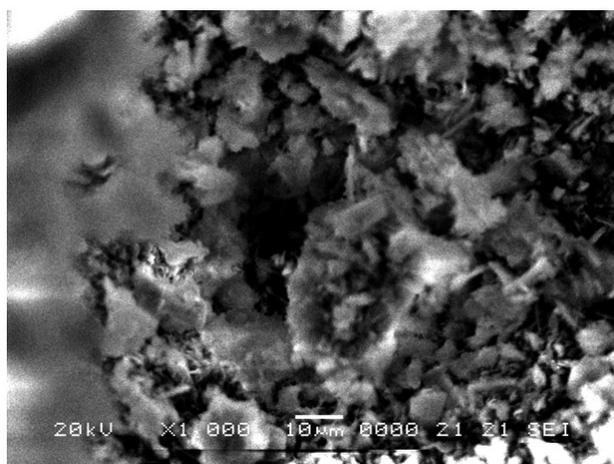


Figure 6

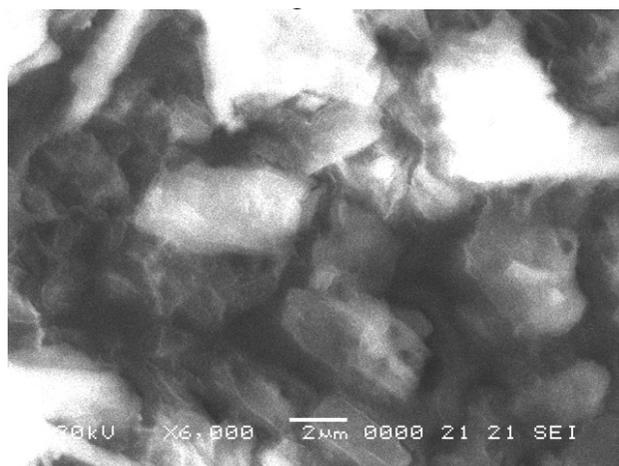


Figure 7

SEM studies show microstructure of the compounds, which mainly include surface morphology.

2.3. Antimicrobial Activity

The study was performed by the cup plate method¹⁹ on pathogenic strains of *S.typhi*, *P.aeruginosa* and *K.pneumoniae*. The media used for antibacterial screening were from hi-media. The media were prepared in triple distilled water and autoclaved at 15 lb pressure. The culture of bacterium was mixed with autoclaved media, poured in plates, and bored. The solution of Mannich bases were poured in these cups in triplicate and incubated at 37°C for 24 hours. The Mannich bases (3a-3j) were studied for their antibacterial property at concentration of 80, 160, 320 mg/mL.

2.4 LD₅₀ Test

The toxicity of synthesized Mannich bases was ascertained by LD₅₀ test. The test performed on white mice weighing 25g. Doses were given orally as well as intraperitoneally and mice were kept under observation for 72 hr for each trial. The Mannich bases showed no adverse toxic effect even of an oral dose of 1400mg/kg of the body weight of mice. However when dose was administered intraperitoneally they proved be lethal at the does level of 800mg/kg of the body weight of mice.

3. RESULTS AND DISCUSSION

The Mannich bases synthesized by Mannich reaction were obtained in good yield (≥85%). They were analyzed for elemental analysis and results were found to be in full agreement with the calculated values. The anticipated structure was in agreement with the spectral data– UV, IR and ¹HNMR. The spectral studies have shown characteristic band due to methylene group

incorporated between active hydrogen substrate and the amine component as a result of Mannich reaction at 2940-2950 cm⁻¹ and 1442-1450 cm⁻¹. This shows the presence of amino methyl linkage in the synthesized Mannich bases. The ¹H NMR also confirms aminomethyl linkage (-CH₂) between amine and active hydrogen (5.04-5.20 ppm). Some of the synthesized compounds are of nano size. The use of nano particle in medicine offers some exciting possibilities. Some techniques are only imagined, while others are at various stage of testing, or actually being used today. The Mannich bases were screened for their biological significance. They were evaluated for antibacterial activity against pathogenic strains of *S.typhi*, *P.aeruginosa* and *K.pneumoniae*. at varying concentrations–80mg/ml, 160mg/ml and 320 mg/ml. All the reported compounds exhibit remarkable *in vitro* activity against these pathogens. Their activity was also compared with their parent sulphonamides.

Table 4 Antibacterial screening of synthesized Mannich Bases and sulphonamides against *S.typhi*, *K.pneumoniae* and *P.aeruginosa* (Zone of inhibition in mm).

Comp. No	<i>S.typhi</i> (Concn. in µg/mL)				<i>K.pneumoniae</i> (Concn. in µg/mL)				<i>P.aeruginosa</i> (Concn. in µg/mL)			
	80	160	320	Av	80	160	320	Av	80	160	320	Av
3a	7.3	9.3	11.0	9.2	7	7.3	10.3	8.2	-	7.3	8.3	5.2
3b	7	7.6	9.3	7.9	9.6	13.6	18.3	13.8	-	-	7.0	2.3
3c	-	-	7.3	2.4	10.6	17.6	21.6	16.6	9.6	12.6	20.3	14.2
3d	7.3	9.3	11.6	9.4	-	-	-	-14.0	7.0	8.6	10.3	8.6
3e	-	-	8.6	2.8	18.3	20.3	17.5	-	7.3	8.3	12.3	9.3
3f	7.0	8.3	10.6	8.6	7.6	10.6	16.3	11.5	-	7.3	9.6	5.6
3g	-	-	9.3	3.1	-	-	7.0	2.3	-	-	8.6	2.8
3h	-	-	-	-	7.3	8.0	11.6	8.9	-	-	-	-
3i	7.3	8.6	10.0	8.6	8.3	8.6	10.3	9.0	7.3	9.0	11.6	9.3
3j	-	-	8.6	2.8	-	-	-	-	-	-	9.0	3.0
1	-	7.0	8.3	5.1	-	-	10.6	3.5	-	-	8.0	2.6
2a	-	-	7.3	2.4	13.6	21.0	24.3	9.5	-	7.3	13.3	6.8
2b	-	-	-	-	10.3	14.3	17.6	14.1	7	8.3	9.3	8.2
2c	-	7.3	9.3	5.5	10.3	14.3	11.6	12.1	9.6	13.3	19.6	14.1
2d	-	-	-	-	12.3	17	21.3	16.8	-	-	9.6	3.2
2e	-	-	-	-	9.3	11.6	16.6	2.3	-	9.6	13.3	7.6

Table-4 reflects that most of the compounds had shown remarkable activity only at 320 mg/ml.

Antibacterial screening of Mannich bases against *S. typhi*, showed interesting results (Table 4). That reflects, Mannich bases 3a and 3d were at par in their antibacterial activity followed by 3f and 3i were superior to other Mannich bases in case of *S. typhi*. All the Mannich bases were found to be significantly superior to their corresponding sulphonamide except 3c.

Mannich base 3e followed by 3c was found to superior in inhibitory effect against *K. pneumoniae* over other compounds. The antibacterial activity was compared with parent sulphonamides. It was found that sulphonamides were equally potential against *K. pneumoniae* as the synthesized Mannich bases. Table 1 reflects that except 3h all the synthesized Mannich bases had proven to be potent against *P.aeruginosa*, 3c are significantly superior to others in inhibiting the growth of this pathogen. Mannich bases 3e and 3i are at par their antibacterial activity followed by 3d in inhibiting the growth of this pathogen.

4. CONCLUSION

Conclusion suggest that the newly synthesized Mannich bases of 2-amino-9-[(1,3 di hydroxy propane-2yl) oxy] methyl] 6-9 dihydro-3H-purine-6-one possess a very noticeable and prolonged antibacterial activity. These derived Mannich bases are less toxic in comparison of their parent sulphonamides. This work shows that Mannich bases are a potential source of compounds for inhibition of bacteria and could be used as efficient drugs with minimum side effects.

ACKNOWLEDGEMENT

Our sincere thanks are due to CSMCRI, Bhavnagar for elemental analysis, SAIF Chandigarh and UGC-DAE CSR Indore for spectral studies. We also extend our sincere thanks to Dr. Tushar Banerjee, School of Life Sciences, DAVV, Indore for providing facilities to conduct antibacterial studies.

REFERENCES

1. Katritzky, A. R. *Chem. Rev.* **2004**, 104, 2125.
2. Sean, S. D.; Essy, M.; Eric, J.S.; Richard, W. *Clinical Therapeutics* **1996**, 18, 546.
3. Joshi, S.; Khosla, N.; Tiwari, P. *Bioorg. Med. Chem.* **2004**, 12, 571.
4. Sondhi, S. M.; Johar, M.; Singhal, N.; Dastidar, S.G.; Shukla, R.; Raghbir, R. *Monatshefte Furchem.* **2000**, 13, 511.
5. LI, J. J.; Anderson, Q. D.; Burton, E.G.; Cogburn, J. N.; Collins, J.T.; Garland, D. J.; Gregory, S. A.; Haung, H. C.; Isakson, P. C.; Koboldt, C. M.; Logush, E. W.; Norton, M. B.; Perkins, W. E.; Reinhard, E. J.; Seibert, K.; Veenhuizen, A. W.; Zang, Y.; Reitz, D.B. *J. Med. Chem.* **1995**, 38, 4570.
6. Supuran, C. T.; Scozzafava, A.; Jurca, B. C.; Ilies, M. A. *Eur. J. Med. Chem.* **1998**, 33, 83.
7. Singh, B. J. *Indian Chem. Soc.* **1976**, 56, 720.
8. Moore, M. B.; Repla, R. T. *J. Am. Chem. Soc.* **1946**, 68, 1675.
9. Tramontini, M.; Angiolini, L. *Mannich Bases: Chemistry and Uses*; CRC Press, BOCA Raton, **1994**, Ch.1, pp 07.
10. Saab, A. N.; Slowan, K. B.; Beall, H. D.; Villanueva, R. *J. Pharm. Sci.*, **1990**, 79, 1099.
11. Bonati, A.; Bombardelli, E.; Gabetto, B. *British Patent*, **1975**, 1383, 053, *Chem. Abstr.* **1975**, 83, 43342.
12. Lepetit, S. A. *Netherlands Patent Appl.*, **1965**, 6144012, *Chem. Abstr.* **1965**, 63, 13180.
13. Mandloi, D.; Joshi, S.; Khadikar, P.V.; Khosla, N. *Bioorg. Med. Chem. Lett.* **2005**, 17, 15405.
14. Tramontini, M.; Angiolini, L.; Ghedini, N. *Polymer* 1998, 29, 771.
15. Goto, M.; Minoe, T. *Jpn. Kokai Tokkyo Koho.* **1995**, JP06, 185.
16. Mitsch, A.; Wibner, P.; Sattler, I.; Schlitzer, M. *Arch. Pharm. Med. Chem.* **2001**, 334, 40.
17. Joshi, S.; Manikpuri, A.; Tiwari, P. *Bioorg. Med. Chem. Lett.* **2006**, 17, 645.
18. Manikpuri, A.; Joshi, S.; Kadikar, P.V. *J. Chil. Chem. Soc.*, 55, N° 3 2010 283.
19. *United States Pharmacopoeia*, 25th ed., Vol. II, *United States Pharmacopoeial Convention, Inc.*, Rockville, **2002**, 1882.