

1.90 ~ 3.32 (5 H, m, 14-H, 16-H₂, and NCH₂), 3.90 (2 H, s, 10-H₂), 4.92 (2 H, s, PhCH₂), 6.60–7.50 ppm (8 H, m, Ar H). *Anal.* (C₂₆H₃₂N₂O·HCl) C, H, N.

Evaporation of the subsequent CHCl₃-EtOH (10:1) eluate gave 1.0 g (70%) of 3-hydroxy-*N*-cyclopropylmethyl-9-azamorphinan (2) as a pale yellow oil, which was triturated with Et₂O to give a solid, which was recrystallized from EtOH to give colorless prisms, mp 172–174°. The spectral data of this compound were superimposable with that of an authentic sample.^{3c}

3-Hydroxy-*N*-cyclopropylmethyl-9-azamorphinan (2). A mixture of 200 mg of 3-benzyloxy-*N*-cyclopropylmethyl-9-azamorphinan (11) hydrochloride, 10 ml of EtOH, and 10 ml of concentrated HCl was heated under reflux for 2 hr. The aqueous layer, obtained by removal of EtOH, was basified with aqueous NH₄OH and extracted with CHCl₃. The CHCl₃ layer was washed (H₂O), dried (K₂CO₃), and evaporated to give a pale yellow oil, which was crystallized from EtOH to give 110 ml (78.6%) of 2 as colorless prisms, mp 172–174° (from EtOH). This was identical with an authentic sample^{3c} by comparison of spectroscopic data and mixture melting point test.

Acknowledgment. We thank Misses C. Yoshida, A. Kawakami, T. Yoshida, R. Kato, and F. Yishinaka for microanalyses and we also thank President A. Yanagisawa and Director O. Takagi of Grelan Pharmaceutical Co., Ltd., for their encouragement. We also thank colleagues Messrs. S. Hayashida, O. Koyama, and K. Kawasaki.

References

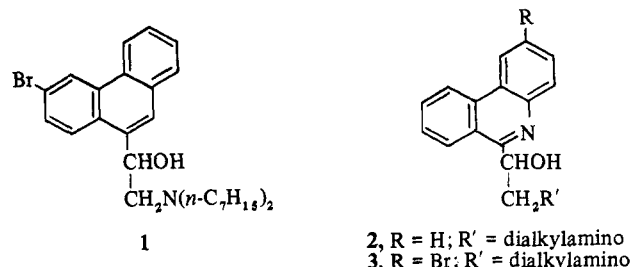
- (1) T. Kametani, K. Kigasawa, M. Hiiragi, T. Aoyama, K. Araki, and S. Saito, *Chem. Pharm. Bull.*, **20**, 2483 (1972) (paper 33).
- (2) T. Kametani, H. Takeda, F. Satoh, and S. Takano, *J. Heterocycl. Chem.*, in press (paper 508).
- (3) (a) T. Kametani, K. Kigasawa, M. Hiiragi, and N. Wagatsuma, *Chem. Pharm. Bull.*, **16**, 296 (1968); (b) T. Kametani, K. Kigasawa, M. Hiiragi, K. Wakisaka, and N. Wagatsuma, *ibid.*, **17**, 1096 (1969); (c) T. Kametani, K. Kigasawa, M. Hiiragi, K. Wakisaka, N. Wagatsuma, F. Satoh, and S. Saito, *J. Med. Chem.*, **13**, 1064 (1970).
- (4) T. Kametani, S. Noguchi, I. Agata, K. Kigasawa, M. Hiiragi, T. Hayasaka, and O. Kusama, *J. Chem. Soc. C*, 1047 (1971).
- (5) E. C. Horning, "Organic Syntheses," Collect. Vol. III, Wiley, New York, N. Y., 1955, p 223; J. D. Roberts and R. H. Mazur, *J. Amer. Chem. Soc.*, **73**, 2509 (1951).
- (6) H. Gilman, "Organic Syntheses," Collect. Vol. I, Wiley, New York, N. Y., 1948, p 156; J. Meck and J. W. Rowe, *J. Amer. Chem. Soc.*, **77**, 6675 (1955).
- (7) R. Kosta, *Fed. Proc., Fed. Amer. Soc. Exp. Biol.*, **18**, 412 (1959).
- (8) J. T. Lichfield and F. Wilcoxone, *J. Pharmacol.*, **96**, 99 (1949).
- (9) F. Haffner, *Deut. Med. Wochenschr.*, **55**, 731 (1929).

Synthesis of 6-(α -Hydroxy- β -*N,N*-dialkylaminoethyl)phenanthridines as Potential Antimalarials†

Chester W. Muth,* Bhabatosh Bhattacharya,¹
Robert L. Mahaffey,² and Howard L. Minigh

Department of Chemistry, West Virginia University,
Morgantown, West Virginia 26506. Received September 5, 1972

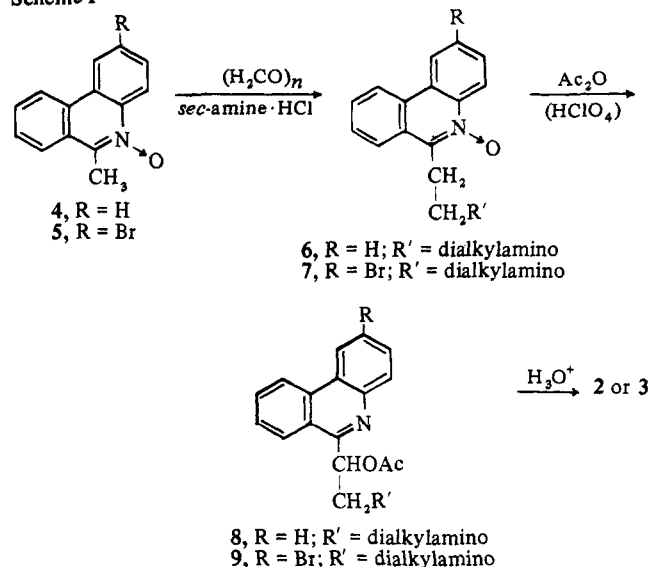
The objective of this work has been the synthesis of 6-(α -hydroxy- β -*N,N*-dialkylaminoethyl)phenanthridines (2 and 3) and water-soluble salts of these compounds for antimalarial testing. These compounds were chosen for synthesis and testing because of their similarity to the 9-phenanthrene-methanols, one of which (1) has been found to be curative



for chicks with *Plasmodium gallinaceum* and not be photo-toxic.^{3,4}

Scheme I outlines the principal method which has been studied. Mannich bases (6 and 7) were made in 51–96% yields (Table I) from 6-methylphenanthridine 5-oxides (4 and 5) in the first successful application of the Mannich reaction (in experiments modeled after a Mannich reaction of 6-methylphenanthridine⁵) to the side chain of azine *N*-oxides. The Mannich base *N*-oxides when treated with Ac₂O and HClO₄ yielded acetates 8 and 9 in about 40% yields (Table II) which were hydrolyzed to 6-(α -hydroxy- β -*N,N*-dialkylaminoethyl)phenanthridines (2 and 3) (Table III).

Scheme I



Of the Mannich base *N*-oxides (6 and 7) which were allowed to react with Ac₂O⁶ alone, only 6-(β -*N*-morpholinoethyl)-phenanthridine 5-oxide (6c) yielded an acetate which could be purified. However, by reacting 1 mol of HClO₄ and 1 mol of Mannich base *N*-oxide with Ac₂O the hydrogen perchlorates of the acetates could be precipitated and in some cases purified (Table II). In comparison with the foregoing method, the addition of HClO₄ after the Ac₂O rearrangement reaction gave an inferior product (8a·HClO₄).

The acetates and the corresponding alcohols were readily characterized by the nmr absorptions (two doublets or an irregular triplet) of their benzylic hydrogens at τ 3.0–3.25 and 4.2–4.6, respectively.

Infrared and nmr spectral analyses indicated that acetate 8e was produced but we were unable to purify 8e or its hydrolysis product, alcohol 2e.

When the acetates were hydrolyzed (Table III) by using refluxing concentrated HCl, the reaction mixtures showed more evidence of decomposition (darkening) than by using 3 *N* HCl at room temperature. With the more vigorous conditions hydramine cleavage may have occurred. The latter phenomenon is believed to have occurred during the melting point determinations of 2b·2HCl and 3a·HCl (Table IV)

†This research was supported by a contract with the U. S. Army Medical Research and Development Command (DA-17-68-C-8099) and is Contribution No. 970 to the Army Research Program on Malaria. Presented before the West Virginia Academy of Sciences, Bluefield, W. Va., April 7, 1972, and the Fourth Central Regional Meeting of the American Chemical Society, Pittsburgh, Pa., May 4, 1972.

Table I. Mannich Base *N*-Oxides from Scheme 1

Mannich base <i>N</i> -oxide	R	R'	Reflux time, hr	% yield	Mp, °C	Recrystn solvent	Formula	Analyses
6a·HCl	H	N(Me) ₂	8	95	189 dec	EtOH	C ₁₇ H ₁₈ N ₂ O·HCl	C, H, N, O
6a·0.5H ₂ O	H	N(Me) ₂			77–77.5 ^a	Petroleum ether ^g	C ₁₇ H ₁₈ N ₂ O·0.5H ₂ O	C, H, N, O
6b·HCl ^b	H	N(<i>n</i> -Bu) ₂	12	73	294–297 dec	Me ₂ CO	C ₂₃ H ₃₀ N ₂ O·HCl	C, H, Cl, N
6b·2HCl	H	N(<i>n</i> -Bu) ₂	12	73	250–253 dec	EtOH	C ₂₃ H ₃₀ N ₂ O·2HCl	C, H, Cl, N
6b	H	N(<i>n</i> -Bu) ₂			58–59 ^c	Hexane		
6c·HCl	H	<i>N</i> -Morpholino	8	96	214–215 dec	80% EtOH	C ₁₉ H ₂₀ N ₂ O ₂ ·HCl	C, H, Cl, N, O
6c·H ₂ O	H	<i>N</i> -Morpholino			82–83	Petroleum ether ^g	C ₁₉ H ₂₀ N ₂ O ₂ ·H ₂ O	C, H, N, O
6d·HCl·H ₂ O	H	<i>N</i> -Piperidino	8	77	205 dec	EtOH	C ₂₀ H ₂₂ N ₂ O·HCl·H ₂ O	C, H, Cl, N
6d·2H ₂ O	H	<i>N</i> -Piperidino			76–78 ^d	Petroleum ether ^g		
6e·HCl ^{e,f}	H	N(<i>n</i> -C ₄ H ₉) ₂	16		195–198 dec	Me ₂ CO	C ₂₉ H ₄₂ N ₂ O·HCl	C, H, Cl, N
7a·HCl	Br	N(Me) ₂	2	73	216–217 dec	EtOH	C ₁₇ H ₁₇ BrN ₂ O·HCl	C, H, Br, Cl, N
7a	Br	N(Me) ₂			139–140	Heptane–C ₆ H ₆		
7b	Br	N(<i>n</i> -C ₄ H ₉) ₂		63	119–120	Hexane	C ₂₃ H ₂₉ BrN ₂ O	C, H, Br, N
7c·HCl	Br	<i>N</i> -Morpholino	16	51	222–223 dec	EtOH		
7c	Br	<i>N</i> -Morpholino			143–144	Heptane–C ₆ H ₆	C ₁₉ H ₁₉ BrN ₂ O ₂	C, H, N

^aPicrate from EtOH, mp 165–166°. *Anal.* (C₁₇H₁₈N₂O·C₆H₃N₃O₇) C, H, N. ^bFrom 6b and 11.7% w/v HCl. ^cPicrate from EtOH, mp 130–131°. *Anal.* (C₂₃H₃₀N₂O·C₆H₃N₃O₇) N. ^dPicrate from EtOH, mp 159–160°. *Anal.* (C₂₀H₂₂N₂O·C₆H₃N₃O₇) C, H, N. ^eFrom adding dry HCl to 6e in absolute EtOH. ^fPicrate from EtOH, mp 81–83°. *Anal.* (C₂₉H₄₂N₂O·C₆H₃N₃O₇) N. ^gBp 60–80°.

Table II. Preparation of Acetates

Acetate	Method	% yield	Mp, °C	Recrystn solvent	Formula	Analyses
8a·HClO ₄	B	54	200–201 dec	MeCN	C ₁₉ H ₂₀ N ₂ O ₂ ·HClO ₄	C, H, N
8a·HCl	C	32	189 dec ^a	EtOH–Me ₂ CO	C ₁₉ H ₂₀ N ₂ O ₂ ·HCl	Cl, N
8b·HClO ₄	B	43	151–152	AcOH	C ₂₅ H ₃₂ N ₂ O ₂ ·HClO ₄	C, H, N
8c ^b	A	36	115–116	80% Me ₂ CO	C ₂₁ H ₂₂ N ₂ O ₃	C, H, N
8d	A	Not purified				
8e	A, B	Not purified				
9a·HClO ₄	C	Not purified				
9b·HClO ₄	B	Not purified				

^a8a·PcOH, mp 158–159° dec. *Anal.* (C₁₉H₂₀N₂O₂·2C₆H₃N₃O₇) N. ^b8b·PcOH, from EtOH, mp 182–183°. *Anal.* (C₂₁H₂₂N₂O₃·C₆H₃N₃O₇) C, H, N. ^cPurified by distillation.

Table III. Preparation of Alcohols from Acetates

Alcohol	Hydrolysis time, hr	% yield	Mp, °C	Recrystn solvent	Formula	Analyses
2a ^a	5 ^b	87	124–125	Et ₂ O	C ₁₇ H ₁₈ N ₂ O	C, H, N
2b	93 ^c	64	74–76	EtOH	C ₂₃ H ₃₀ N ₂ O	C, H, N
2c ^d	4 ^b	76	114–115	Me ₂ CO	C ₁₉ H ₂₀ N ₂ O ₂	C, H, N
2d·2HCl·H ₂ O	4 ^b	37 ^e	160–162	95% EtOH–Me ₂ CO	C ₂₀ H ₂₂ N ₂ O·2HCl·H ₂ O	C, H, Cl, N
2f ^f	4 ^b	90	115–118	EtOH	C ₁₄ H ₁₁ NO	C, H, N
3a	96 ^c	40 ^g	127–128	MeCN	C ₁₇ H ₁₇ BrN ₂ O	C, H, Br, N
3b	48 ^c	40 ^h	65–66	Heptane	C ₂₃ H ₂₉ BrN ₂ O	C, H, Br, N

^a2a·PcOH from Me₂CO, mp 165° dec. *Anal.* (C₁₇H₁₈N₂O·2C₆H₃N₃O₇) N. ^b12 *N* HCl, reflux. ^c3 *N* HCl, room temperature. ^d2c·PcOH from EtOH, mp 190–191° dec. *Anal.* (C₁₉H₂₀N₂O₂·C₆H₃N₃O₇) N. ^eBased on 6d. ^fFrom 6-acetoxymethylphenanthridine 5-oxide: C. W. Muth, J. C. Patton, B. Bhattacharya, D. L. Giberson, and C. A. Ferguson, *J. Heterocycl. Chem.*, in press. ^gBased on 7a. ^hBased on 7b.

because these compounds melted and resolidified before decomposition. This behavior parallels that reported by Lutz, *et al.*,⁷ for similar compounds which were shown to have undergone hydramine cleavage as they were heated.

None of the phenanthridinemethanol hydrochlorides of Table IV, 2a, nor any of the intermediates for these compounds have shown any significant antimalarial activity in mice (infected with *Plasmodium berghei*) or chicks (infected with *Plasmodium gallinaceum*).[‡]

Experimental Section

All melting points were done in a Mel-Temp apparatus and are uncorrected. The infrared spectra (ir) were recorded by using either a Beckman IR-8 or Perkin-Elmer 137 spectrometer. Nuclear magnetic resonance spectra (nmr) were recorded using either a Varian HA-60 nmr or a Varian T-60 nmr spectrometer. Elemental analyses were performed by Galbraith Microanalytical Laboratories, Knoxville, Tenn. The analyses for all new compounds are within 0.4% of the calculated values and the elements analyzed for are indicated.

‡All test results were obtained from the U. S. Army Walter Reed Institute of Research, Washington, D. C.

The analytical results are on file with the editor of this journal.

2-Bromo-6-methylphenanthridine 5-Oxide (5). 2-Bromo-6-methylphenanthridine⁸ was treated with *m*-chloroperoxybenzoic acid⁹ with the modification of extracting the reaction mixture with 5% NaOH before column chromatography. The product (5) after recrystallization from EtOH melted at 194–197° dec and the yield was 94%. The analytical sample, recrystallized from EtOH, melted at 197–198° dec. *Anal.* (C₁₄H₁₀BrNO) C, H, N.

Mannich Base *N*-Oxides and/or Hydrochlorides (6a–e, 7a–c). A mixture of 0.046 mol of *N*-oxide (4¹⁰ or 5), 0.05 mol of formaldehyde (as paraformaldehyde), and 0.051 mol of *sec*-amine·HCl was dissolved in EtOH (125 ml of 95% EtOH for 4 and 535 ml of absolute EtOH for 5) and heated under reflux for 4 hr when an additional 0.033 mol of formaldehyde was added and refluxing was continued until the total reflux time indicated in Table I had elapsed. From subsequent experiments it was learned that all of the paraformaldehyde could be added initially without any significant change in yields. In some cases when the product precipitated during the reaction or after the reaction mixture cooled, the product was recrystallized from the solvent indicated in Table I. If the product did not precipitate, the solvent was distilled and the residue was dissolved in 10% HCl and filtered (charcoal). The filtrate, on concentration *in vacuo*, yielded a residue which was crystallized from EtOH.

The Mannich base *N*-oxides were obtained from their hydrochlorides by treatment with 10% NaOH. The solvents used for re-

Table IV. Alcohol Hydrochlorides

Alcohol·HCl	Method ^a	Mp, °C	Formula	Analyses
2b·2HCl	A	93–100, resolidification, 115–165 dec	C ₂₃ H ₃₀ N ₂ O·2HCl	C, H, Cl, N, O
2c·2HCl·H ₂ O	B	165–166 ^b	C ₁₉ H ₂₀ N ₂ O ₂ ·2HCl·H ₂ O	Cl
2d·2HCl·H ₂ O	B	160–162 ^c	^d	
3a·HCl	A	124–130, resolidification, melts again 140–150 dec	C ₁₇ H ₁₇ BrN ₂ O·HCl	Cl, N
3b·2HCl	C	142 dec	C ₂₃ H ₂₉ BrNO·2HCl	C, H, Cl

^aMethod A, addition of dry HCl to ethereal solution of alcohol; method B, addition of dry HCl to EtOH containing alcohol; method C, addition of ethanolic HCl to 3b in ethanol followed by cooling to 0° and dilution with ether. ^b95% EtOH. ^c95% EtOH-Me₂CO. ^dSee Table III.

crystallization and melting points of the Mannich base *N*-oxides are listed in Table I.

Acetates from Mannich Base *N*-Oxides and Acetic Anhydride.

Method A. Acetic anhydride (306 g) was stirred with 37.0 g (0.113 mol) of 6-(*β*-*N*-morpholinoethyl)phenanthridine 5-oxide monohydrate (6c·H₂O) for 1 day. The reaction mixture was filtered and the filtrate was concentrated *in vacuo*. Water was added to the residue and the mixture was made basic by adding NaHCO₃ solution. The mixture was extracted with ether and the ethereal solution was washed with saturated brine solution and concentrated to yield a dark red viscous oil. This was purified by columnar chromatography using neutral alumina with 5:1 EtOAc-C₆H₆ as the developer. The eluent was concentrated to a residue which was triturated with acetone to yield a solid which was recrystallized from 80% acetone to give 14 g (36%) of 8c as colorless needles, mp 115–116°.

Method B. HClO₄ (70%, 2.15 g, 0.0150 mol) was added dropwise to 40 ml of Ac₂O at such a rate that the temperature did not rise above 25°. To this solution was added 5.05 g (0.0144 mol) of 6-(*β*-*N*,*N*-di-*n*-butylaminoethyl)phenanthridine 5-oxide (6b) with stirring at 23–25°. After 6 hr of stirring at 23–25°, 0.35 g of yellow solid, mp 213–215°, was collected by filtration and the reddish brown filtrate was diluted with a large volume of ether to yield an ether-insoluble brown oil. The ether was decanted and the oil was triturated with a small amount of AcOH to yield 3.05 g (43%) of 8b·HClO₄, mp 151–153°. The analytical sample was recrystallized from AcOH as fine yellow crystals, mp 151–152°.

8a·HClO₄ precipitated from its reaction mixture without the addition of ether.

Method C. This was like method B except that MeCN was added to the reaction mixture to cause dissolution of the Mannich base *N*-oxide hydrogen perchlorate or hydrochloride.

Acknowledgments. The authors wish to thank Mr. Robert Smith for recording many of the nmr spectra and Drs. Richard E. Strube and Edward Steck both of the Walter Reed Army Institute of Research for their helpful suggestions.

References

- (1) B. Bhattacharya, M.S. Thesis, West Virginia University, Morgantown, W. Va., 1970.
- (2) R. L. Mahaffey, M.S. Thesis, West Virginia University, Morgantown, W. Va., 1971.
- (3) E. L. May and E. Mosettig, *J. Org. Chem.*, **11**, 627 (1946).
- (4) G. R. Coatney, W. C. Cooper, N. B. Eddy, and J. Greenberg, "Survey of Antimalarial Agents," Public Health Monograph, No. 9, 1953, p 7.
- (5) J. Finkelstein and S. M. Linder, *J. Amer. Chem. Soc.*, **73**, 302 (1951).
- (6) V. Boekelheide and W. J. Linn, *ibid.*, **76**, 1286 (1954).
- (7) R. E. Lutz, P. S. Bailey, M. T. Clark, J. F. Codington, A. J. Deinet, J. A. Freek, G. H. Harnest, N. H. Leake, T. A. Martin, R. J. Rowlett, Jr., J. M. Salsbury, N. H. Shearer, Jr., J. D. Smith, and J. W. Wilson, III, *ibid.*, **68**, 1813 (1946).
- (8) B. L. Hollingsworth and V. Petrow, *J. Chem. Soc.*, 3771 (1961).
- (9) J. C. Craig and K. K. Purushothaman, *J. Org. Chem.*, **35**, 1721 (1970).
- (10) C. W. Muth, J. C. Patton, B. Bhattacharya, D. L. Giberson, and C. A. Ferguson, *J. Heterocycl. Chem.*, in press.

Communications to the Editor

Metabolism of Acetylmethadol. A Sensitive Assay for Noracetylmethadol and the Identification of a New Active Metabolite

Sir:

α-*l*-Acetylmethadol (1) is an orally effective analgesic in both laboratory animals and in man. Early pharmacology and metabolism studies led to the conclusion that 1 exerted its activity, at least in part, through an active metabolite (*cf.* Way and Adler¹ for a comprehensive review of the literature up to 1962). Metabolism studies on *α*-*dl*-acetylmethadol reported in 1965² strongly suggested that the active metabolite was noracetylmethadol (2). Noracetylmethadol had been synthesized in 1959 by Pohland, *et al.*,³ and shown to be an effective analgesic in animals and man.⁴

The current interest in the use of *α*-*l*-acetylmethadol as an alternative to methadone in the maintenance of heroin addicts has prompted us to reinvestigate the metabolism of this drug. In this communication we report a sensitive method for the assay of the active metabolite, noracetylmeth-

adol, in body fluids. We also wish to report the identification of a second active metabolite of acetylmethadol in the rat.

Noracetylmethadol Assay. The method is similar to that used by Änggård, *et al.*, in 1970⁵ for the assay of amphetamine. It is based on formation of the trichloroacetamide derivative which is suitable for gas-liquid chromatography with the sensitive electron-capture detector.

In an initial experiment rats were dosed with *α*-*dl*-acetylmethadol (20 mg/kg ip). The animals were sacrificed at intervals, blood was collected, and the liver, lung, and brain were removed. Plasma samples (2–3 ml) were made alkaline to pH 9.5 with 1 *N* NaOH and were extracted twice with 7 ml of butyl chloride (C₄H₉Cl). The C₄H₉Cl extracts were evaporated to dryness *in vacuo*. The residue was dissolved in 0.5 ml of dry toluene in a stoppered centrifuge tube, and 50 μl of 1% trichloroacetyl chloride in dry toluene was added. The reaction mixture was heated at 70–80° for 15 min and then evaporated to dryness *in vacuo*. The residue was taken up in hexane and 1–2 μl was injected onto the gas chromatograph (gc). Analyses were accomplished with a Hewlett-Pac-