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# Intermediates Stabilized by Tris(triazolylmethyl)amines in CuAAC Reaction

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**Abstract:** Tris(triazolylmethyl)amine ligands (**TL**) are widely used to accelerate the Cu<sup>1</sup>-catalyzed azide-alkyne cycloaddition (CuAAC) reaction, but its mechanistic role remains unclear. Using electrospray ionization mass spectrometry, for the first time we detected the trinuclear **TL**-Cu<sup>1</sup><sub>3</sub>-acetylide and the dinuclear **TL**-Cu<sup>1</sup><sub>2</sub>-acetylide complexes in aqueous solution. The apparent second-order rate constants of their reaction with an azide were 27 M<sup>-1</sup>•s<sup>-1</sup> and 783 M<sup>-1</sup>•s<sup>-1</sup> when the alkyne was tethered to **TL**. In the catalytic system without the tether, the rate constant increased to >146 M<sup>-1</sup>•s<sup>-1</sup> for the **TL**-Cu<sup>1</sup><sub>3</sub>-acetylide. The results indicated that **TL** accelerated the reaction by stabilizing the Cu<sup>1</sup><sub>2</sub>- and Cu<sup>1</sup><sub>3</sub>-acetylide and their azide adduct intermediates, but this role is largely weakened by excess alkyne and other competing ligands under catalytic conditions.

#### Introduction

The high efficiency, orthogonality and versatility of the Culcatalyzed azide-alkyne cycloaddition (CuAAC, a click reaction)<sup>[1]</sup> has rendered it a powerful tool in many fields.<sup>[2]</sup> The reaction was proposed to proceed through a rate-limiting transition state containing two Cu<sup>I,[3]</sup> However, identification of the Cu<sup>I</sup><sub>2</sub>-acetylide and -triazolide intermediates was hampered by the facile ligand exchange and multiple fast equilibria between the Cul complexes that are also air-sensitive and prone to aggregation and disproportionation.<sup>[4]</sup> To overcome these hurdles, strong Cu<sup>I</sup> ligands, such as N-heterocyclic carbenes (NHC),<sup>[5]</sup> cyclic (alkyl) (amino) carbenes (CAAC),<sup>[6]</sup> and organophosphines<sup>[7]</sup> were employed to stabilize the Cu<sup>I</sup> intermediates, allowing for isolation of single crystals of NHC-Cul-acetylide cluster.<sup>[8]</sup> (NHC-Cul)1triazolide,<sup>[5a]</sup> (CAAC-Cu<sup>1</sup>)<sub>2</sub>-acetylide and (CAAC-Cu<sup>1</sup>)<sub>2</sub>-triazolide.<sup>[6]</sup> Also, organophosphine-stabilized Cul<sub>2</sub>-acetylide and -triazolide intermediates were detected by electrospray ionization mass spectrometry (ESI-MS),<sup>[7]</sup> an important tool for studying the realtime distribution of reaction intermediates.<sup>[9]</sup> Using isotopically labeled Cul, Worrell et al. demonstrated the necessity of a second Cu<sup>I</sup> atom to activate the NHC-Cu<sup>I</sup>-acetylide for C-N bond formation with an organic azide.[5b]

The above progress was made using strong Cu<sup>I</sup> ligands. In

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comparison, it is more challenging to study the mechanism of CuAAC reaction accelerated by weaker ligands, such as the tris(triazolylmethyl)amines<sup>[10]</sup> (**TL**, Scheme 1A) that are among the most efficient and widely used ligands in CuAAC reaction.<sup>[2c, 11]</sup> Although a catalytic cycle involving di-Cu<sup>1</sup> acetylides formed from Cu<sup>1</sup><sub>2</sub>**TL** was proposed (Scheme 1A),<sup>[12]</sup> the proposed intermediates were not detected. To facilitate the interception of the weakly coordinated intermediates, we tethered the **TL** with an alkyne via an EG<sub>11</sub> chain as in **1** (Scheme 1B). This model system entropically stabilized the **TL**-Cu<sup>1</sup>-acetylide intermediates, allowing us to evaluate their reactivity using ESI-MS.



**Scheme 1.** (A) Previously proposed di-Cu<sup>1</sup> intermediates involved in **TL**-accelerated CuAAC reaction,<sup>[12]</sup> and (B) our model system to facilitate interception of the intermediates. Charges are omitted.

#### **Results and Discussion**

We first attempted to prepare the Cul<sub>2</sub>TL complex by mixing TL with [Cu(MeCN)<sub>4</sub>PF<sub>6</sub>] (2 equiv.) in MeCN under N<sub>2</sub> followed by lyophilization to completely remove MeCN. After addition of water, the Cu/TL 2:1 mixture turned into a greenish-yellow suspension with red precipitates, indicating disproportionation of the free Cu<sup>I</sup> into Cu<sup>II</sup> and Cu<sup>0</sup> (Figure S1A). In comparison, the aqueous solutions of Cul/1 1:1-3:1 remained colorless and clear for 4 months (Figure S1B). The ESI-MS spectra of Cul/1 1:1-3:1 solutions (Figure 1A) showed a mixture of Cul-alkyne and acetylide complexes. Each major signal was assigned to a Cul complex ion based on the matches with the predicted m/z value and isotopic pattern (Figure S3), and their relative abundance was estimated by the ESI-MS intensities based on the evidences that indicating these Cu<sup>I</sup> complex ions had a similar ESI-MS response factor (Figure S5 & Figure 1A). According to the following discussion, their structures were proposed in Scheme 2.

In the Cu/1 1:1 solution (Figure 1A), the predominant species was the protonated Cu<sup>1</sup>-alkyne complexes (Ia/Ib). Increasing the Cu/1 ratio to 2:1 led to a mixture of mostly Ia/Ib, the Cu<sup>1</sup><sub>2</sub>-alkyne complex IIa, and the Cu<sup>1</sup><sub>3</sub>-acetylide complex IIIa ( $[1-H+3Cu]^{2+}$ ).



**Figure 1.** (A) ESI-MS spectra for solutions of 1 (100 µM) and 1–3 equiv. of Cu<sup>1</sup> generated from CuSO<sub>4</sub> (100–300 µM) and Na ascorbate (1 mM) in deoxygenated water. (B) Averaged ESI-MS spectra of the above solutions (without NaOH) taken between 4–6 min after addition of 1 equiv. of N<sub>3</sub>EG<sub>4</sub>. All marked peaks are assigned and color-coded with cyan for mono-Cu<sup>1</sup>, blue for di-Cu<sup>1</sup> and red for tri-Cu<sup>1</sup> species (Scheme 2). The assignment is based on a good fit of the experimental (solid lines) and theoretical (dash line) isotopic patterns (Figures S3 & S4, selected examples are shown in inserts).

Only ~5% of the deprotonated  $Cu_{2}^{1}$ -acetylide IIb was present. At the ratio of Cu/1 3:1, IIIa became the predominant species (see also Table 1). Addition of NaOH produced more  $Cu_{2}^{1}$ -acetylide IIb (Figures 1A, S7 and Table 1). Overall, the stability of acetylides in aqueous solution increased as the number of  $Cu^{1}$  coordination became higher (stability: IIIa > IIb > Ic). Indeed, IIIa was present even in the Cu/1 1:1 mixture at pH 6, while Ic was not detectable (see Figure S8 for the amplified MS spectra).

The coordination mode of **III**a was proposed (Scheme 2) based on the crystal structure of a **TL**-Cu<sup>1</sup><sub>3</sub>-acetylide analogue.<sup>[13]</sup> The interaction between Cu<sup>1</sup> and alkyne or acetylide in the Cu/1 mixtures in D<sub>2</sub>O was evidenced by the large <sup>1</sup>H NMR downfield shift (0.19–0.58 ppm) of the propargylic protons (Figure S6).

Addition of 1 equiv. of the azide  $N_3EG_4$  (Scheme 1B) to the solutions of Cu/1 1:1–3:1 generated new signals (Figure 1B) assigned to the Cu<sub>2</sub>-triazolide IIc and Cu<sub>3</sub>-triazolide IIIb, and the Cu-triazole complexes 2-Cu<sub>2</sub> and 2-Cu (Scheme 2, see Figures S4 & S9–S11 for the isotopic patterns and tandem mass spectrometry analysis). Although an organophosphine-stabilized



**Scheme 2.** Proposed mechanism for the reaction of **1** and an azide in the presence of 1–3 equiv. of Cu<sup>I</sup>. Labels for the observed ions in Figures 1 & 2 are placed under the proposed mono- (I, cyan), di- (II, blue) and tri- (III, red) Cu<sup>I</sup> complexes. The proposed intermediates in a bracket were not detectable in the ESI MS. The grey curves represent the long, flexible EG<sub>11</sub> linker.



**Figure 2.** (A) Time course of ESI-MS intensities of the assigned intermediates during the reaction of 1 with N<sub>3</sub>EG<sub>4</sub>. Conditions: 100  $\mu$ M 1, 300  $\mu$ M CuSO<sub>4</sub>, 100  $\mu$ M N<sub>3</sub>EG<sub>4</sub> and 1 mM Na-Ascorbate in deoxygenated water at room temperature. For the labels, see Scheme 2. (B) Apparent second order rate constants ( $k_{obs}$ ) for reaction of 1 with N<sub>3</sub>EG<sub>4</sub> quantitatively measured by LC-MS. Conditions: 100  $\mu$ M 1, 100  $\mu$ M N<sub>3</sub>EG<sub>4</sub> and 1 mM Na-ascorbate in deoxygenated water at room temperature, with: 100  $\mu$ M CuSO<sub>4</sub> (cyan); 200  $\mu$ M CuSO<sub>4</sub> (blue); 300  $\mu$ M CuSO<sub>4</sub> (red); 600  $\mu$ M CuSO<sub>4</sub> & 500  $\mu$ M NaOH (green); 200  $\mu$ M CuSO<sub>4</sub> & 500  $\mu$ M NaOH (navy). Data from Figure S15.

azide-Cu<sup>1</sup><sub>2</sub>-acetylide adduct was reported,<sup>[7]</sup> the unstable adducts **II**c1 and **III**b1 in our system (Scheme 2) were not detectable.

The reactivity of the unprecedented tri-nuclear complex IIIa was first investigated by monitoring the reaction of the Cu/1 3:1 mixture (containing ~86% IIIa, Table 1) with 1 equiv. of  $N_3$ EG<sub>4</sub>. As shown in Figure 2A, the MS intensity of IIIa dropped

drastically during the first 10 min, accompanied by a rapid increase of the Cul<sub>3</sub>-triazolide IIIb whose signal intensity exceeded that of IIIa after 10 min, then decreased after ~15 min. Consumption of IIIa and accumulation of IIIb as the most abundant intermediate were faster than the formation of the triazole complexes 2-Cu<sub>2</sub> and 2-Cu, which was possibly via hydrolysis of IIIb to the transient 2-Cu<sub>3</sub> followed by dissociation of Cu<sup>1</sup> from 2-Cu<sub>3</sub> to 2-Cu<sub>2</sub>, or from IIIb to IIc followed by hydrolysis (Scheme 2).

The coordination mode of Cu<sub>3</sub>-triazolide IIIb is tentatively proposed in Scheme 2 and Figure S14. The three coordinating Cu<sup>1</sup> ions greatly stabilized IIIb against hydrolysis compared to the Cu<sub>2</sub>-triazolide IIc (showing a low intensity throughout the reaction, Figure S13) and the Cu-triazolide Ic (not detectable).

We next compared the reactivity of the tri-copper acetylide IIIa with the di-copper acetylide IIb as the key intermediate in the proposed mechanism.<sup>[3]</sup> To enrich the deprotonated IIb, 5 equiv. NaOH was added to the solution of Cu/1 2:1 generating the highest percentage of IIb (67%, Table 1) among all Cunalkyne/acetylide species (Figure S7), further addition of the base promoted formation of dimers and CuOH. The stoichiometric reactions of 1 with N<sub>3</sub>EG<sub>4</sub> in the presence of 1-6 equiv. of Cu<sup>1</sup> (and 5 equiv. of NaOH) were quantitatively analyzed by LC-MS and their apparent second order rate constants ( $k_{obs}$ ) were determined (Figures 2B, S15, and Table 1). It is generally agreed that the first C-N bond formation from the Cu<sub>2</sub>-acetylideazide adduct is the rate-limiting step for CuAAC reaction.[3d-f, 14] We assume that the C-N bond can be formed through the dicopper and the tri-copper pathways via the azide adducts IIc1/ IIIb1 (Scheme 2), and their barrier is much higher than those of the pre-equilibria. Under these assumptions, Eq. (1) derived from a generalized pre-equilibrium approximation method<sup>[15]</sup> and the data in Table 1 can be used to extract the apparent rate constants for converting IIb to IIc  $(k'_{II})$  and IIIa to IIIb  $(k'_{III})$ :

$$k_{\text{obs}} = x_{\text{IIb}}k'_{\text{II}} + x_{\text{IIIa}}k'_{\text{III}} \qquad \text{Eq. (1)}$$

in which  $x_{IIb}$  and  $x_{IIIa}$  are the molar fraction of IIb and IIIa derived from the ESI-MS intensities of the intermediates. Fitting the data to Eq (1) gives  $k'_{II} = 783 \text{ M}^{-1} \cdot \text{s}^{-1}$  and  $k'_{III} = 27 \text{ M}^{-1} \cdot \text{s}^{-1}$  (Table S1) with a good agreement between the calculated rate constants  $k_{cal}$  and the measured  $k_{obs}$  (Table 1). The ratio of  $k'_{II}/k'_{III} = 29:1$ agreed with result of our DFT calculation showing that the barrier for C-N bond formation via a Cu<sub>2</sub> transition state was ~ 2 kcal/mol lower than via a Cu<sub>3</sub> transition state (unpubilished result).

The apparent rate constants  $k'_{II}$  and  $k'_{III}$  depend primarily on the energy barriers for the C-N bond formation, but they can also be significantly affected by any hidden equilibria interfering the transformation. These equilibria include dissociation of the azide adduct and its association with other ligands in the system, which are minimized in the model system. Specifically, the ~50 Å EG<sub>11</sub> linker in **1** maintained a relatively high local concentration (in millimolar range) of the alkyne, which stabilized the **TL**-Cunacetylides (IIIa/IIb) and their azide adducts (IIIb1/IIc1) against dissociation, and yet should not change the barrier for the C-N bond formation. In the Cu/1 ≥ 2 system, there was no extra ligand interacting with the di- and tri-Cu acetylides and their azide adducts. Hence, the apparent rate constants  $k'_{II}$  and  $k'_{III}$ 

Table	1.	MS	Intensities	of	the	Cu <sup>I</sup> -acetylides	in	Ligand	1	System,	and
Observ	/ed	Rate	Constants	(kot	s) vs	. Calculated Ra	te C	Constants	s (ł	(cal) <sup>[a]</sup>	

Cu/ <b>1</b> /		ESI MS In	Kobs	$k_{\rm cal}{}^{[a]}$		
NaOH <sup>[b]</sup>	la/lb	llb	Illa	lla	M <sup>-1</sup> S <sup>-1</sup>	M <sup>-1</sup> S <sup>-1</sup>
1:1:0	149.0	1.5	0.8	5.3	9	8
2:1:0	45.1	5.4	43.7	31.6	41	43
3:1:0	4.3	4.7	132.0	12.3	47	47
1:1:5	47.2	45.9	0	0	386	386
2:1:5	40.2	67.2	0.8	0.7	471	483
6:1:5	1.2	6.3	185.0	5.0	38	50

[a] Rate constant calculated using Eq. (1), in which  $k'_{11} = 783 \text{ M}^{-1}\text{s}^{-1}$  and  $k'_{111} = 27 \text{ M}^{-1}\text{s}^{-1}$ . [b] The molar ratio of added NaOH.

measured with this system better represents the barriers for the C-N bond formation than with a catalytic system (see below).

Overall, this model system showed a much higher  $k_{obs}$  (9–471  $M^{-1} \cdot s^{-1}$ ) than the previously reported values ( $k_{obs} = 0.002 \ 3.5 \ M^{-1} \cdot s^{-1}$ ) for CuAAC reaction accelerated by **TL** ligands (without the linked alkyne, see Scheme 1A for structure of **TL**).<sup>[12b]</sup> Compared to the above model system, reaction of the alkyne **3** and the azide  $N_3EG_4$  under catalytic conditions (Figure 3A) showed much lower  $k_{obs}$  values of 2.1–3.6  $M^{-1} \cdot s^{-1}$  at 3/Cu = 10:6 (Table S2), which further decreased to 0.3–0.4  $M^{-1} \cdot s^{-1}$  at lower catalyst loading (**3**/Cu = 100:6, Table S3).

First of all, both the di-Cu acetylide (iia, [3 H+TL+2Cu]<sup>+</sup>) and tri-Cu acetylide (iiia, [3 H+TL+3Cu]<sup>2+</sup>) were present under catalytic conditions (Figures 3B, S17 and S22). The ratio of [iiia]/[iia] increased with higher Cu/TL ratio and catalyst loading (Tables S2 and S3). Without TL, no di- and tri-Cu acetylide was detected (Figure S17) and the reaction was 25 times slower (Figure S20), consistent with the role of TL to stabilize these active species.



Figure 3. (A) CuAAC reaction of the alkyne 3 and RN<sub>3</sub> (R = EG<sub>4</sub>). (B) ESI-MS spectra for the solutions of 3 (100  $\mu$ M), TL (20, 30, 60  $\mu$ M), Cu<sup>1</sup> (60  $\mu$ M) and Na-ascorbate (0.5 mM) in deoxygenated water. For assignment of the labeled ions, see Figure S18 & S19. (C) Effect of Cu<sup>1</sup>/TL ratio on reaction order to [Cu], with 3 (100  $\mu$ M), N<sub>3</sub>EG<sub>4</sub> (100  $\mu$ M), Cu<sup>1</sup> (30, 60, 90, 120, 180, 240, 300  $\mu$ M), Cu/L (1:1, 2:1, 3:1). See also Table S2 for the amount of iiia and iia vs  $k_{obs}$  at 3/Cu = 10:6 (boxed in C, data taken from B).

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**Scheme 3.** Proposed mechanism for the CuAAC reaction of **3** and RN<sub>3</sub> (R = EG<sub>4</sub>) in the presence of catalytic amount of Cu<sup>1</sup>/TL. For the intermediates detected by ESI-MS, see Figures 3, S17-S19 and S22; intermediates in a bracket were not detectable. L = ligands **3** and TL, etc.

The slow kinetics under catalytic conditions can be attributed mainly to the dissociation of the Cu<sub>n</sub>-acetylides **ii**a/**iii**a and their azide adducts **ii**b/**iii**b in which the Cu-acetylide bond was weakened by azide-coordination.<sup>[14]</sup> The dissociation was enhanced both by the increased entropy (compared to the EG<sub>11</sub>-tethered **1** system) and by the excess alkyne and **TL** (Scheme 3), generating inactive Cu complexes **3**-Cu, **TL**-Cu, and Cu-acetylide aggregates (Figures 3 and S17). These equilibria led to fractional rate orders in [Cu] (especially at a low Cu/TL ratio, Figure 3C), and greatly decreased the apparent rate constant for the C-N bond formation via the di-Cu pathway (*k*'<sub>11</sub>) by ~14 times to ~55 M<sup>-1</sup>•s<sup>-1</sup> (Tables S2 and S3).

In sharp contrast to the di-Cu pathway, the apparent rate constant for the C-N bond formation via the tri-Cu pathway (k'in) was increased by 5 9 times to 146 260 M<sup>-1</sup>•s<sup>-1</sup> (Tables S2 and S3). Accordingly, the  $k'_{II}/k'_{III}$  ratio drastically dropped from 29 in the model system to <0.4 in the catalytic system (Tables S1-S3), showing a greatly enhanced contribution to kobs by the tri-Cu acetylide iiib (for an example, see Figure 3B,C). To rationalize this result, we note that compared to the di-Cu acetylide iib, the tri-Cu acetylide iiib has higher affinity with the negatively charged internal nitrogen of azide.[3b, 3g, h, 10f, 16] As illustrated in Scheme 3, ligand coordination to the tri-Cu-acetylide-azide adduct iiib will facilitate the dissociation of a Cu<sup>I</sup> (internally) from the acetylide iiic to generate a di-Cu transition state TScu2 via iib or TS'<sub>Cu2</sub> (through the internal dissociation) for C-N bond formation. Furthermore, the triazolides iic and iiid (detected by ESI-MS, Figure S22) may serve as a base to deprotonate the alkyne 3, especially those formed from TS'<sub>Cu2</sub> in which the coordinated alkyne is proximal to the triazolide, to generate the product 4 and regenerate the copper acetylides iia and iiia (Scheme 3).

### Conclusions

In summary, we reported the first example of tri-Cu-acetylide as a key intermediate in CuAAC reaction. A similar intermediate was proposed by Meldal and Tornøe.<sup>[2a]</sup> Compared to the di-Cuacetylides, the tri-Cu-acetylide showed superior stability against protonation in aqueous solution at neutral pH. The **TL** ligand stabilizes the **TL**-Cu<sup>I</sup><sub>n</sub>-acetylide (n = 2, 3) complexes and their azide adducts as the reactive intermediates, but the strength of stabilization is limited, especially at a low catalyst loading where the complexes are destabilized by the large excess of alkyne and other competing ligands.

## **Experimental Section**

# ESI-MS analysis for intermediate solution equilibria in $Cu_n^1$ solutions w/ or w/o N<sub>3</sub>EG<sub>4</sub> (Figure 1 & Figure 2A)

ESI-MS spectra of Cu<sup>I</sup><sub>n</sub>1 solutions and the CuAAC reaction mixtures (with N<sub>3</sub>EG<sub>4</sub>) were acquired using a Thermo Finnigan LCQ Deca XP Plus ion trap mass spectrometer at capillary temperature of 40°C, spray voltage 4 kV, capillary voltage 28 V, multipole 1 offset 3 V, multipole 2 offset 12 V, entrance lens voltage 61 V. In an anaerobic chamber, all stock solutions were prepared in deoxygenated Milli-Q water.

To study the solution equilibria of Cu<sup>I</sup><sub>n</sub>1 complexes (Figure 1A), 100 µL 1 mM ligand 1, 100/200/300 µL 1 mM CuSO<sub>4</sub>, 100 µL 10 mM Na ascorbate, and 700/600/500 µL deoxygenated water were mixed in 3 separate Eppendorf tubes in an anaerobic chamber, vortexed, and then directly infused into the ESI source by a syringe pump at a flow rate of 10 µL/min. Each full MS spectrum was an average of 100 individual scans.

In separate experiments, to promote the deprotonation of IIb (Figures 1A, S7 and Table 1), 100  $\mu$ L 1 mM ligand 1, 100/200/600  $\mu$ L 1 mM CuSO<sub>4</sub>, 100  $\mu$ L 10 mM Na ascorbate, 100  $\mu$ L 5 mM NaOH, and 600/500/100  $\mu$ L deoxygenated water were mixed in 3 separate Eppendorf tubes and the spectra were recorded using the above procedure.

To study the solution equilibria of CuAAC reaction (Figure 1B), 100  $\mu$ L 1 mM ligand **1**, 100/200/300  $\mu$ L 1 mM CuSO<sub>4</sub>, 100  $\mu$ L 1 mM azido-tetraethylene glycol (N<sub>3</sub>EG<sub>4</sub>), 100  $\mu$ L 10 mM Na ascorbate, and 600/500/400  $\mu$ L deoxygenated water, were mixed in 3 separate Eppendorf tubes in anaerobic chamber, vortexed, and then immediately infused into ESI source by a syringe pump at a flow rate of 10  $\mu$ L/min. Reaction time was recorded on a timer. Each full MS spectrum in Figure 1B was an average of scans during the reaction time of 4–6 min. The reaction mixture was continuously infused into the ESI source during a period of 20 min. At Cu/**1** = 3:1, the variation of ESI-MS intensities of selected intermediates during the reaction was plotted on Figure 2A.

# Kinetic studies for reaction between 1 and $N_3 \text{EG}_4$ (Figure 2B)

The reactions were conducted in an anaerobic chamber at room temperature. Solutions in 900  $\mu$ L deoxygenated Milli-Q water was added into separate Eppendorf tubes, containing 100 nmol 1, 1  $\mu$ mol sodium ascorbate, and: A: 100 nmol CuSO<sub>4</sub>, B: 200 nmol CuSO<sub>4</sub>, C: 300 nmol CuSO<sub>4</sub>, D: 100 nmol CuSO<sub>4</sub> and 500 nmol NaOH, E: 200 nmol CuSO<sub>4</sub> and 500 nmol NaOH, F: 600

nmol CuSO4 and 500 nmol NaOH. 100 µL 1 mM azido tetraethylene glycol (N3EG4) aqueous solution was added into each tube to initiate the reaction. At 20 s (for A, B, C) or 10 s (for D, E, F) intervals, 20 µL aliquots of the reaction mixture were added into a 96-well plate, each well containing 180 µL airsaturated aqueous solution of 20 nmol diethylene triamine pentaacetic acid (DTPA) (pH adjusted to 7 by NaOH) as the quenching reagent and 1 nmol 5 (structure shown in Scheme S1) as internal standard. After 8 aliquots were collected, the 96well plate was moved out of anaerobic chamber. The samples were injected to a Thermo Finnigan LCQ Deca XP Plus ion trap mass spectrometer through a Thermo Finnigan Surveyor HPLC system. Reagents and product were separated by first static elution in 3% (MeCN, 0.1% formic acid) : 97% (H<sub>2</sub>O, 0.1% formic acid) for 5 min and then a linear gradient elution to 95% (MeCN, 0.1% formic acid) : 5% (H<sub>2</sub>O, 0.1% formic acid) for 5 min. A set of calibration standards of product 2, ranging from 100 nM to 10 µM, using 5 as internal standard, was injected before the unknowns. The concentration of product 2 in each sample was calculated based on the calibration curve. Each reaction under same condition was repeated three times.

# ESI-MS analysis for intermediate solution equilibria in Cu<sup>I</sup><sub>n</sub>3 solutions (Figure 3B, S17)

ESI-MS spectra of  $Cu^I_n 3$  solutions were acquired using a Thermo Finnigan LCQ Deca XP Plus ion trap mass spectrometer at capillary temperature of 40°C, spray voltage 4.5 kV, capillary voltage 32 V, multipole 1 offset 5V, multipole 2 offset 12V, entrance lens voltage 51 V. In an anaerobic chamber, all stock solutions were prepared in deoxygenated Milli-Q water.

To study the solution equilibria of 3/Cu = 10:6 solution (Figure 3B), 50 µL of 10 mM sodium ascorbate was added into 950 µL of aqueous solution containing 100 nmol 3, 60 nmol CuSO<sub>4</sub>, and 60/30/20 nmol **TL**.

To study the solution equilibria of 3/Cu = 100.6 solution (Figure S17), 50 µL of 10 mM sodium ascorbate was added into 950 µL of aqueous solution containing 1 µmol 3, 60 nmol CuSO<sub>4</sub>, and 120/60/30/20/15/0 nmol TL.

The mixtures were directly infused into ESI source by a syringe pump at a flow rate of 10  $\mu$ L/min. Each full MS spectrum was an average of about 100 individual scans. Zoom scan analysis was carried out to determine the charge state and isotopic pattern for selected Cu<sup>I</sup> adducts. Each zoom scan spectra represents an average of 50–100 individual scans (Figure S18). MS/MS analysis were carried out to determine the fragmentation pattern of selected Cu<sup>I</sup> adducts (Figure S19).

# Rate order studies for reaction between 3 and $N_3 EG_4$ with various amount of $\mbox{Cu}^1$ (Figure 3C)

In an anaerobic chamber, eight 1 mL Titertube® micro test tubes (Bio-Rad) were placed in a 96-well Titertube rack. Solutions in 900  $\mu$ L deoxygenated Milli-Q water (pH=7.0) was added into these tubes, each containing 100 nmol **3**, 100 nmol azido tetraethylene glycol and 10, 20, 30, 40, 60, 80, 100 nmol [(CuSO<sub>4</sub>)<sub>3</sub>(**TL**)] or [(CuSO<sub>4</sub>)<sub>2</sub>(**TL**)] or [(CuSO<sub>4</sub>)(**TL**)]. 100  $\mu$ L of 5 mM sodium ascorbate aqueous solution was added into each tube simultaneously by a multichannel pipette to start the reaction. At 1 min intervals, 100  $\mu$ L aliquots of the reaction

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mixture were added into a 96-well plate, each well containing 90 µL air-saturated diethylene triamine pentaacetic acid (DTPA) aqueous solution (pH adjusted to 7.0 by NaOH). After 6 sets of aliquots were collected, the 96-well plate was moved out of anaerobic chamber. 10 µL 100 µM S10 (structure shown in Scheme S1) water solution was added into each aliquot as internal standard. The samples were injected to a Thermo Finnigan LCQ Deca XP Plus ion trap mass spectrometer through a Thermo Finnigan Surveyor HPLC system. Reagents and product were separated by first static elution in 3% (MeCN, 0.1% formic acid) : 97% (H<sub>2</sub>O, 0.1% formic acid) for 2 min and then a linear gradient elution to 90% (MeCN, 0.1% formic acid) : 10% (H<sub>2</sub>O, 0.1% formic acid) for 5 min. A set of calibration standards of product 4, ranging from 500 nM to 50 µM, using S10 as internal standard, were injected before and after each set of samples. The concentration of product in each sample was calculated based on the calibration curve.

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Keywords: Click Chemistry • CuAAC Reaction • Reactive Intermediate • Mass Spectrometry • Tri-copper Acetylide

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