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Discovery of Selective, Orally Bioavailable, N-linked Arylsulfonamide Na_v1.7 Inhibitors with Pain Efficacy in Mice

Anthony J. Roecker,^a Melissa Egbertson,^a Kristen L. G. Jones,^a Robert Gomez,^a Richard

L. Kraus,^b Yuxing Li,^b Amy Jo Koser,^b Mark O. Urban,^b Rebecca Klein,^c Michelle Clements,^c Jacqueline Panigel,^c Christopher Daley,^b Jixin Wang,^c Eleftheria N. Finger,^b John Majercak,^b Vincent Santarelli,^c Irene Gregan,^b Matthew Cato,^b Tracey Filzen,^b Aneta Jovanovska,^b Ying-Hong Wang,^d Deping Wang,^e Leo A. Joyce,^f Edward C. Sherer,^f Xuanjia Peng,^g Xiu Wang,^g Haiyan Sun,^g Paul J. Coleman,^a Andrea K. Houghton,^b Mark E. Layton,^a

Department of ^aDiscovery Chemistry, ^bPharmacology, ^cNeuroscience, ^dDrug Metabolism, ^eChemistry Modeling & Informatics, ^fAnalytical Chemistry Merck & Co., Inc., West Point, PA 19486 USA; ^gWuXi AppTec Co., Ltd, Shanghai, China

* Corresponding author. Tel.: + 1-215-652-8257; fax: + 1-215-652-3971; e-mail: anthony_roecker@merck.com



Abstract

The voltage-gated sodium channel $Na_v 1.7$ is a genetically validated target for the treatment of pain with gain-of-function mutations in man eliciting a variety of painful disorders and loss-of-function mutations affording insensitivity to pain. Unfortunately, drugs thought to garner efficacy via $Na_v 1$ inhibition have undesirable side effect profiles due to their lack of selectivity over channel isoforms. Herein we report the discovery of a novel series of orally bioavailable arylsulfonamide $Na_v 1.7$ inhibitors with high levels of selectivity over $Na_v 1.5$, the Na_v isoform responsible for cardiovascular side effects, through judicious use of parallel medicinal chemistry and physicochemical property optimization. This effort produced inhibitors such as compound **5** with excellent potency, selectivity, behavioral efficacy in a rodent pain model, and efficacy in a mouse itch model suggestive of target modulation.



Keywords: Nav1.7, ion channel, pain, arylsulfonamide, selectivity



It is estimated that 15% of the US population currently suffers from chronic pain conditions. In addition to the debilitating nature of these ailments, this spectrum of diseases confers a significant economic burden of approximately \$600 billion per year in the US due to increased medical costs and lost productivity in the workforce.¹ A range of pharmaceutical agents are utilized to treat chronic pain. First line treatments include anticonvulsants (pregabalin and gabapentin),² antidepressants (i.e. duloxetine and desipramine),³ and non-steroidal anti-inflammatory drugs (NSAIDs, i.e. ibuprofen and naproxen).⁴ All current first-line treatment options have significant limitations including incomplete efficacy and undesirable side effect profiles. Opioids (i.e. morphine, oxycodone, and hydrocodone) are effective agents for chronic pain management; however, these are not considered first line options due to the potential for tolerance, potential for addiction and abuse, and narrow therapeutic indices with adverse effects consisting of constipation, decreased respiration, sedation, and nausea.^{5,6} For these reasons, novel, safe, and effective chronic pain treatments remain an area of significant unmet medical need and a focus of pharmaceutical research.⁷

A more recent option for chronic pain management is the topical application of lidocaine in the form of a patch (Lidoderm[®]) approved for the treatment of postherpetic neuralgia, a painful skin condition resulting from shingles.⁸ The mechanism of action of lidocaine is thought to arise from non-selective blockade of Na_v1 voltage-gated sodium channels (VGSCs); key regulators of electrical signaling in a variety of tissues through the generation and propagation of action potentials via sodium ion flux in response to changes in membrane voltage potential.⁹ There are nine VGSCs currently known, Na_v1.1 – 1.9, that possess differential tissue distribution patterns and associated pharmacology.¹⁰



 $Na_v 1.1, 1.2, and 1.3$ are expressed in the central nervous system (CNS) while $Na_v 1.4$ and 1.5 are expressed in skeletal tissue and cardiac muscle, respectively. $Na_v 1.6$ is expressed widely throughout the CNS and peripheral nervous system (PNS), and $Na_v 1.7$, 1.8, and 1.9 are restricted mainly to the PNS. Recent human genetic studies have revealed that loss-of-function (LOF) mutations in Na_v1.7 afford channelopathy-associated insensitivity to pain suggesting that blockade of this VGSC might be a contributing factor to efficacy for agents such as lidocaine.¹¹ Additional investigations revealed that gain-of-function (GOF) mutations in Nav1.7 elicit painful conditions such as erythromelalgia and paroxysmal extreme pain disorder further linking $Na_v 1.7$ and pain.¹² Unfortunately, systemic exposure to local anesthetics such as lidocaine can engender severe cardiovascular and CNS side effects through blockade of Nav1.5 in the heart and Nav channels in the CNS (Nav1.1 / Nav1.2), respectively, thus limiting their therapeutic utility.¹³ The combination of human genetic validation for Na_v1.7 and marketed therapeutics which target Nav1 channels has drawn extremely high levels of interest and investment from the scientific community with the goal of developing a safe and effective therapeutic for patients suffering from chronic pain.¹⁴

Historically, identifying selective inhibitors of $Na_v 1.7$ has been extremely challenging due to high levels of homology among the $Na_v 1$ channels. Indeed, the binding site for lidocaine resides in the pore region of the channels which is almost completely conserved; consistent with the non-selective nature of the drug.¹⁵ Pioneering work by Pfizer and Icagen described the discovery of arylsulfonamides such as PF-04856264 (**1a**, Figure 1) that potently inhibited $Na_v 1.7$ through a binding site distinct from the pore region (DIV = domain four: S2-S3 = between segments 2 and 3) with



unprecedented selectivity over Na_v1.5, ¹⁶ potentially ameliorating the risk of cardiotoxicity.¹⁷ This important finding has inspired further chemistry efforts from Pfizer affording the clinical candidate PF-05089771 (**1b**, Figure 1),¹⁸ an X-ray crystal structure of GX-936 (**1c**, Figure 1) bound to DIV of human Na_v1.7 from scientists at Genentech and Xenon,¹⁹ and several arylsulfonamide subseries efforts reported by Amgen (**1d**, Figure 1),²⁰ Genentech / Xenon (**1e**, Figure1),²¹ and MSD (**1f**, Figure 1).²² One hallmark feature of the arylsulfonamide series referenced above is the use of aromatic motifs in Western SAR. This observation suggested that Western aromatic motifs might be necessary to achieve adequate target potency. Herein we describe initial parallel chemistry efforts to identify novel Western SAR devoid of aromatic motifs and further medicinal chemistry efforts to optimize potency, selectivity, physicochemical properties, oral bioavailability, and behavioral pain efficacy in mice.





Figure 1. Known arylsulfonamide (1a - 1f) Na_v1.7 inhibitors from Pfizer / Icagen, Genentech, Amgen, Xenon, and MSD.

The generic aryl sulfonamide scaffold (2) shown in Figure 2 was utilized as a template for design of nucleophilic aromatic substitution (S_NAr) libraries taking advantage of the MSD collection of alcohol- and amine-containing building blocks lacking aromatic motifs. Initial library exploration afforded several interesting leads including compound **3** containing a novel (tetrahydro-1*H*-pyrrolizin-7a(5*H*)-yl)methanamine (3.3.0 amine hereafter) Western substituent.²³ This compound possessed moderate potency against Na_v1.7 (inactivated state potency)²⁴ of 112 nM, no activity against Na_v1.5 (> 30 μ M), and good binding affinity to the same binding site as PF-04856264.²⁵ Encouraged by this finding, compound **3** was profiled further and was found to be very polar (LogD = 0.0) with low permeability ($P_{app} = 2$; Table 1). Arylsulfonamide **3** had limited oral bioavailability (%F = 4), consistent with its permeability and lipophilicity profile, and low intrinsic clearance (CL_{int}) in rat. Hence, additional optimization focused on increasing lipophilicity without compromising pharmacokinetics.



Figure 2. Generic arylsulfonamide scaffold (2) utlized in library design via S_NAr chemistry yielding novel Na_v1.7 inhibitor leads such as **3**.

Modification of the 5-position fluorine to chlorine on the central aromatic ring (4, Table 1) increased lipophilicity (LogD = 0.4), improved Na_v1.7 potency by an order of



magnitude, and maintained selectivity over Na_v1.5. Unfortunately, rat CL_{int} increased significantly and permeability was not improved. Exploration of substituted thiazole sulfonamides based upon compound **4** afforded compounds **5** – **11** which all possessed moderate to excellent Na_v1.7 potency, high levels of selectivity over Na_v1.5, and increased LogD (0.7 – 1.3; Table 1). Surprisingly, compounds **5** -**7** and **9** still displayed low permeability (P_{app} = 2 - 6) but were orally bioavailable in rat, indicating that high levels of permeability were not a prerequisite for oral bioavailability.²⁶ A selection of 6-membered ring sulfonamides was also synthesized to probe SAR, and representative examples **12** and **13** showed decreased Na_v1.7 potency compared to **5** suggestive of tight sulfonamide SAR. Although **12** significantly increased permeability, the increase in rat CL_{int} deterred further analog design using this core template. Arylsulfonamide **5** possessed the best balance of potency, selectivity, rat CL_{int}, and oral bioavailability and was utilized for further compound design.

Table 1. Arylsulfonamide core SAR.



Compound	Ar	Х	$Na_v 1.7 IC_{50}$ (nM) ^a	$\frac{\text{Na}_{\text{v}}1.5 \text{ IC}_{50}}{(\text{nM})^{\text{a}}}$	LogD ^b / PSA (Å)	$\frac{P_{app}}{(10^{-6} \text{ cm/s})^c}$	Rat CL _{int} ^d / %F
3	N N N S	F	112	>30000	0.0 / 96	2	62 / 4
4	N N N S	Cl	10	7200	0.4 / 95	2	660 / ND



5	N S	Cl	8	>30000	0.7 / 77	2	210 / 40
6	N S F	Cl	5	1900	1.0 / 77	2	230 / 55
7	N S CI	Cl	12	17600	1.3 / 77	3	780 / 45
8	M Me S Me	Cl	142	24200	1.1 / 78	9	560 / ND
9	Me N S	Cl	44	22500	1.1 / 78	6	71 / 36
10	Me N F	Cl	30	>30000	1.3 / 78	5	630 / ND
11	N= S	Cl	91	>30000	0.7 / 78	20 ^e	ND / ND
12	♦ N F	Cl	45	19000	1.0 / 77	23	610 / 26
13		Cl	215	>30000	-0.2 / 92	2	61 / ND

(a) Estimated inactivated state potency as measured by PatchXpress[®] in HEK293 cells stably expressing human Nav1.7 or Nav1.5; IC₅₀ values are estimated from \geq 3 cellular measurements at varying compound concentrations; (b) MSD HPLC LogD assay; (c) Monolayer assay in MDCK cells; (d) Rat CL_{int} = (84*rat CL) / [rat f_u * (84-rat CL)]; units mL/min/kg; IV: 0.05 mpk cassette dosing or 2 mpk single dosing in DMSO/PEG400/water (20/60/20); PO: 10 mpk in PEG400/Tween90/water (40/10/50); (e) permeability determined in LLCPK cell line.

Western amine SAR utilizing the core template of arylsulfonamide **5** is displayed in Table 2. Addition of one carbon atom to the linker of **5** afforded **14** which decreased Na_v1.7 potency by over an order of magnitude, increased rat CL_{int} substantially, and had no impact on permeability. Somewhat surprisingly, the (octahydro-9a*H*-quinolizin-9ayl)methanamine (4.4.0) substituent in **15**²⁷ dramatically increased permeability (P_{app} =



32) compared to **5**, albeit at the expense of potency and pharmacokinetics. Modifying the basicity of the 3.3.0 amine via the *beta*-fluorine effect improved permeability for compounds **16** and **18**,²⁸ however, potency was diminished on Na_v1.7. A substitution pattern which was tolerated with regard to Na_v1.7 potency was dimethyl-3.3.0 ring system in **19** and **20**. This pair of enantiomers had excellent selectivity over Na_v1.5, moderate permeability (9 – 11), lipophilicity (LogD = 1.7), rat CL_{int} and oral bioavailability. Compound **19** possessed a very comparable profile to **5** (Table 1) with slight differentiation in permeability (P_{app} = 9), hence both compounds were moved forward for additional characterization.





Compound	R	Na _v 1.7 IC ₅₀ (nM)	Na _v 1.5 IC ₅₀ (nM)	LogD / PSA (Å)	$\frac{P_{app}}{(10^{-6} \text{ cm/s})}$	Rat CL _{int} / %F
14		129	24200	0.6 / 80	2	1600 / 30
15		136	>30000	2.2 / 76	32	3000 / ND
16 (racemic)	F N	375	>30000	1.7 / 77	23	NA / NA
17 (enant. A)	F	160	>30000	1.5 / 77	ND	ND / ND



18 (enant. B)	F,	210	5700	1.5 / 77	11	810 / 57
19 (<i>R</i>)	N N	12	>30000	1.7 / 78	9	460 / 61
20 (<i>S</i>)	N	27	>30000	1.7 / 78	13	850 / 45

(a) See Table 1 legend for definition of terms.

Representative synthetic routes for the compounds in Tables 1 and 2 are exemplified in Schemes 1 and 2. Compound 5 was synthesized from commercially available (tetrahydro-1*H*-pyrrolizin-7a(5*H*)-yl)methanamine 21^{29} and known 2,4-dimethyoxybenzyl-protected sulfonamide scaffold 22^{30} under nucleophilic substitution reaction conditions, deprotected under acidic conditions, and transformed to the HCl salt to afford 5 in moderate overall yield. This synthetic approach was utilized for all compounds in the preceding tables.



Scheme 1. Synthesis of compound 5. Reagents and reaction conditions: (a) Et_3N , DMF, 40 °C, 18 h, 62%; (b) TFA, DCM, 25 °C, 1 h, then 2.0 M HCl / MeOH, 80%: DMF = *N*,*N*-dimethylformamide, DCM = dichloromethane.

Substituted or homologated 3.3.0 amines were assembled using several methods (Scheme 2). Known intermediate 23^{31} was ozonized and subsequently difluorinated to afford 24 which was globally reduced to afford 25 in moderate yield (Scheme 2A).



Alcohol 25 was converted to primary amine 26 via standard mesylate formation, azide displacement, and hydrogenation sequence in modest yield. Alternatively, the ozonolysis product of intermediate 23 could be stereoselectively reduced with NaBH₄ to afford alcohol 27 which can be inverted through a DAST fluorination to afford 28 (Scheme 2B). This intermediate underwent the same alcohol to primary amine interconversion sequence as before to afford 29. Finally, methyl benzylprolinate 30 was alkylated with ethyl-3-bromopropionate, de-benzylated and cyclized to afford 31 which was selectively reduced (NaBH₄) and TBS protected (32). Dimethylation of the lactam was achieved under LDA / MeI conditions to afford 33 in modest yield after TBS deprotection. The resulting alcohol was taken through a phthalimide Mitsunobu / hydrazine deprotection sequence to afford amines 36 and 37, respectively, with a chiral resolution occurring at the phthalimide stage (Scheme 2C).³²



Scheme 2. Representative synthesis of amine monomers. Reagents and reaction conditions: A) (a) ozone, DCM, 25 °C, then Me₂S, 67%; (b) DAST, THF, 0 °C, 18 h, 57%; (c) LiAlH₄, THF, 0 °C to reflux, 4 h, 80%; (d) MsCl Et₃N, DCM, 0 °C, 1 h, aqueous work up, then NaN₃, DMF, 80 °C, 18 h, 83%; (e) 10% Pd/C, MeOH, H₂ (1 atm), 0 °C, 2 h, ~ 100%; B) (a) see A-a; 67%; (f) NaBH₄, MeOH, 0 °C, 5 min, 86%; (b) DAST, DCM, 0 °C to 25 °C, 18 h; 74%; (c); see A-c; 99%; (d) see A-d; 71%; (e); see A-e; ~ 100%; C) (g)



LDA, HMPA, 3-bromopropionate, THF, -78 °C to 25 °C, 16 h, 22%; (h) 10% Pd/C, cat. formic acid, MeOH, H₂ (50 psi), 25 °C, 10 h, 92%; (i) toluene, reflux, 16 h, 81%; (j) NaBH₄, MeOH, 0 °C, 30 min, 75%; (k) TBSCl, imidazole, DCM, 0 °C to 25 °C, 5 h, 100%; (l) LDA, MeI, THF, -78 °C, (reaction performed twice for bis-addition), 3 h, 36% overall; (m) 2.0 M HCl, MeOH, 1 h, 76%; (n) phthalimide, PPh₃, DEAD, THF, 0 °C to 25 °C, 18 h, 71%; (o) SFC separation; IC column, 5 to 40% EtOH in CO₂; (p) hydrazine hydrate, EtOH, reflux, 4 h, 54%; (q) BH₃•DMS, THF, 25 °C ~ quant. DAST = diethylaminosulfurtrifluoride; MsCl = methanesulfonyl chloride; LDA = lithium diisopropylamide; HMPA = hexamethylphosphoramide; TBSCl = *tert*-butyldimethylsilyl chloride; DEAD = diethylazodicarboxylate; LiHMDS = lithium bis(trimethylsilyl)amide.

Compound	$ \begin{array}{c} F & O & N \\ F & O & N \\ S & N \\ H \\ Cl \\ Compound 5 \end{array} $	F O, O N S N H Cl Compound 19
Molecular Weight	431 g/mol	459 g/mol
pK _a (sulfonamide / amine)	6.7, 9.8	7.0, 9.3
Solubility (μ M, pH 2 and 7) ^a	159, 43	143, 28
HPLC $LogD^{b} / P_{app}^{c}$	0.7 / 2	1.7 / 9
Mouse $Na_v 1.7 IC_{50} (nM)^d$	15 nM	13 nM
Mouse PPB	59%	82%
CYP IC ₅₀ (3A4/2C9/2D6, μM)	> 50, > 50, 32	> 50, > 50, 3.1
PXR EC ₅₀ (µM)	16	5.6

Table 3.	Additional	profiling	of com	pounds 5	and 19.
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(a) MSD HPLC kinetic solubility assay; (b) MSD HPLC logD assay; (c) Monolayer assay in MDCK cells; (d) Estimated inactivated state inhibition potency as measured by PatchXpress[®] in HEK293 cells stably expressing mouse Nav1.7; IC₅₀ values are estimated from \geq 3 cellular measurements at varying compound concentrations.

Compounds **5** and **19** possessed moderate molecular weight, zwitterionic character, and moderate kinetic solubility at neutral and acidic pH (Table 3). As stated above, the two compounds represented a 1 log unit range of lipophilicity (0.7 versus 1.7) and apparent differentiation in permeability (2 versus 9 x 10^{-6} cm/s), and the impact of these properties on *in vivo* performance was of high interest to our team. Na_v1.7 potency shifts in preclinical species, especially rodents, were recently identified in the literature.³³ Hence, we were delighted to observe that compounds **5** and **19** possessed a mouse Na_v1.7 IC₅₀ of 15 and 13 nM, respectively; sufficient to support *in vivo* efficacy evaluation. Both compounds had adequate unbound fraction in mouse plasma to interpret our *in vivo*



experiments, and compound **5** displayed excellent functional selectivity in a general offtarget screening panel with no hits of less than 400-fold selectivity.³⁴ Both compounds were not potent CYP inhibitors with the exception of moderate activity at CYP2D6,³⁵ and did elicit moderate PXR activation. Compound **5** demonstrated minimal CNS penetration when dosed orally to mice with less than 1% and 6% of the plasma exposure existing in the CSF and brain compartments (30 mpk oral dose), respectively. While this same experiment was not performed on **19**, this compound was a Pgp substrate in the rat (LLCPK cell line) with a BA/AB ratio of 17.³⁶ These two experiments partially alleviated concerns around CNS penetration of these molecules which could elicit known toxicity from blockade of Na_v1 isoforms (Na_v1.1 and 1.2) expressed in the CNS (*vide supra*). On the basis of the data above, both compounds **5** and **19** were advanced into behavioral pain models in mice.

A common assay to evaluate antinociceptive effects of potential pain therapeutic agents is the mouse formalin paw test assay.³⁷ When a dilute solution of formalin is injected into the hind paw of a mouse, characteristic biting and licking of the affected area is elicited. The efficacy assessment is composed of two phases; an acute phase (or Phase I) generally reflective of direct activation of nociceptors and a tonic phase (or Phase II) related to inflammatory responses. Compounds **5** and **19** were administered orally two hours prior to formalin challenge, and both compounds demonstrated statistically significant, dose-dependent reversal of these effects in the acute phase of the experiment (0 - 5 minute period post formalin injection) and the tonic phase of the experiment (20 - 35 minute period post formalin injection) with full reversal of formalin effects in the tonic phase (Figure 3). Plasma concentrations were evaluated at the end of



the experiment with an unbound plasma concentration IC_{50} of 170 nM ($C_p = 420$ nM) and 45 nM ($C_p = 250$ nM) for **5** and **19**, respectfully, during the tonic phase. These exposures were 11-fold (**5**) and 3.5-fold (**19**) over the mouse Na_v1.7 IC₅₀ value, respectively.

Recently, GOF mutations in Na_v1.7 have been linked to paroxysmal itch in humans.³⁸ Hence, a model of histamine-induced itch was developed to compare the pain efficacy observed for the compounds characterized above to effects that our team viewed as suggestive of Na_v1.7 target modulation. Briefly, intradermal treatment of histamine to mice engendered a significant increase in the number of scratching events over the course of 15 minutes (Figure 4). Oral administration of compound **5** two hours prior to histamine challenge demonstrated dose-dependent blockade of these scratching events with statistically significant, complete blockade occurring at the same oral dose as that used in the mouse formalin paw test (30 mpk). This suggested that the efficacy observed in the mouse formalin paw test was mediated through the inhibition of Na_v1.7.

In conclusion, parallel synthetic efforts utilizing an arylsulfonamide core template led to the discovery of the novel Western 3.3.0-amine class of arylsulfonamide $Na_v1.7$ inhibitors represented by compound **3**. Medicinal chemistry targeting the balance of potency, selectivity over $Na_v1.5$, PSA, and lipophilicity afforded advanced compounds **5** and **19**: potent, selective, and orally bioavailable inhibitors of $Na_v1.7$ with limited CNS penetration. Arylsulfonamides **5** and **19** were effective at reversal of formalin-induced nociceptive events in mice, and the efficacy of **5** was consistent with $Na_v1.7$ target modulation as evidenced by efficacy in the mouse itch assay. Additional research in the $Na_v1.7$ inhibitor arena will be published in due course.





Figure 3. Oral efficacy of compounds 5 and 19 in mouse formalin paw test.^a

(a) Measurement of formalin-induced nociceptive behaviors in mice (C57BL/6 mice) following administration of vehicle (40% PEG400 / 10% Tween 80 / 50% water) or rising doses of compounds **5** (Panel A) and **19** (Panel B) PO. Compounds were administered 2 hours prior to formalin injection to coincide with T_{max} from separate pharmacokinetic studies (3 hours). Pharmacokinetics were evaluated 3 hours post administration. Data were analyzed using within-subject ANOVA to determine main effects and one sample t-test to compare to vehicle (N = 8 / group); * P > 0.01, ** > 0.001.

Figure 4. Oral efficacy of compound 5 in mouse itch assay.^a





(a) Measurement of histamine-induced nociceptive behaviors in mice (C57BL/6 mice) following administration of vehicle (40% PEG400 / 10% Tween 80 / 50% water), 3, 10, and 30 mg/kg of compound 5 PO. Compound 5 was administered 2 hours prior to histamine injection to coincide with T_{max} from separate pharmacokinetic studies (3 hours). Data were analyzed using within-subject ANOVA to determine main effects and one sample t-test to compare to vehicle (N = 8 or 9 / group); * P > 0.01.

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²⁵ The "arylsulfonamide" binding site potency for **3** was measured by a radioligand displacement assay with a $K_i = 42$ nM using a proprietary Merck radioligand residing in the structural class from reference 22. No binding was detected for compound **3** in a standard local anesthetic binding site displacement assay with $K_i > 30 \mu$ M.

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³⁴ General off-target screening (Ricerca panel, 145 assays) of compound **5** revealed several initial binding hits including adrenergic activity (α 1D, α 2A, α 2B), serotonin (5HT_{2B}), and muscarinic (M1) activites. Second messenger assays for adrenergic activities resulted in functional antagonist activity with an IC₅₀ ~ 8 μ M (all 3 isoforms listed), M1 IC₅₀ and EC₅₀ > 30 μ M, and 5HT_{2B} IC₅₀ = 3.8 μ M. Compound **19** was only tested in selected off-target assays and demonstrated a similar profile to **5**.

 35 CYP inhibition has been identified as an issue for the arylsulfonamide series of Na_v1.7 inhibitors (see reference 21.

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