

## REVISED STRUCTURE OF CERIFERIC ACID

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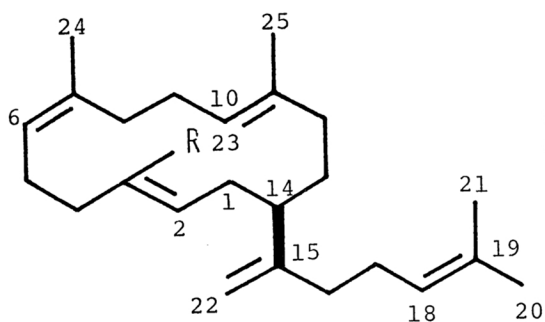
The structure of ceriferic acid, a scale insect sester-  
terpene secretion, has been revised from 10 to 5. Naturally  
occurring ceriferol has been correlated with ceriferic acid.

We recently reported the structures of a series of 14-membered monocyclic  
sesterterpenoids isolated from the secretion of a Japanese scale insect  
*Ceroplastes ceriferus* Anderson (Coccidae).<sup>1,2</sup> Eight of these possess the 2-t/6-  
c/10-t skeletal framework as exemplified by cericerol-I (1),  $[\alpha]_D^{27} -84.1^\circ$ , for which  
the absolute configuration at C-14 was established by chemical correlations;<sup>1</sup>  
two minor congeners possibly have 2-t/6-c/10-c and 2-t/6-t/10-t skeletons.<sup>2</sup>

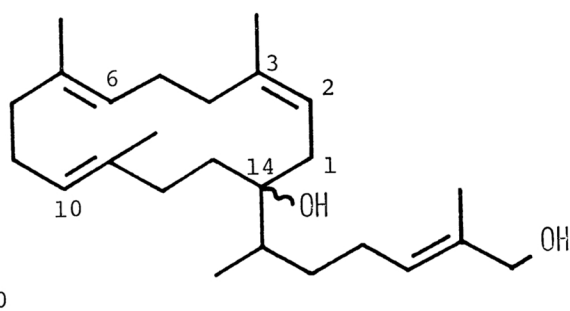
The arrangement of annular double bonds in these compounds is in contrast to  
that of albocerol (2) which was isolated from the Mexican species *C. albolineatus*  
by Veloz et al.<sup>3</sup> It should be noted that the skeletons of 1 and 2 are derived,  
respectively, from two opposing cyclization modes of the biogenetic precursor  
geranylfarnesyl pyrophosphate (3) and (4).<sup>2,3</sup> It was thus quite remarkable that  
spectral data of ceriferic acid isolated from the Japanese species *C. ceriferus*  
(collected in Tokyo area) led to the albocerol- or Mexican-type structure (10);<sup>4</sup>  
it even implied necessity for a taxonomic reinvestigation. In the following,  
however, we show that ceriferic acid should after all be represented by the  
cericerol-I type structure (5).

The evidence leading to the revised structure is as follows:

1) Ceriferic acid showed no CD Cotton effect. Therefore, the  $\alpha, \beta$ -unsaturated  
carboxylic acid group, which from <sup>1</sup>H- and <sup>13</sup>C-NMR data is clearly part of a *cis*

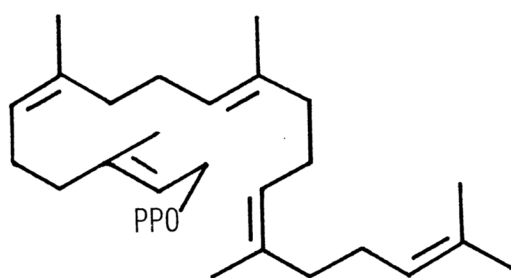


(1):  $R=CH_2OH$   
(cericerol-I)

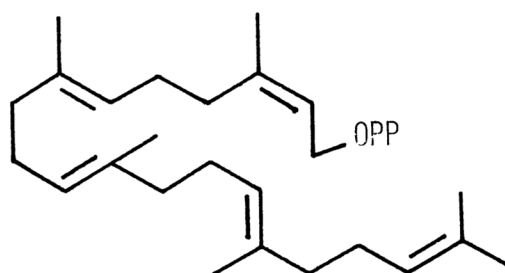


(2)  
(albocerol)

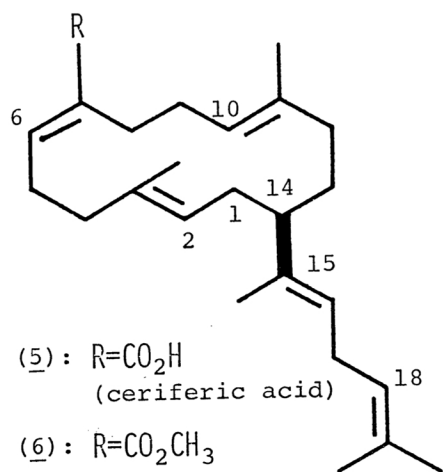
(11):  $R=CH_3$



(3)



(4)



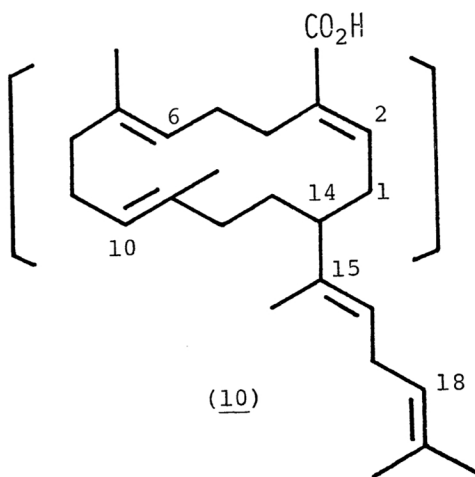
(5):  $R=CO_2H$   
(ceriferic acid)

(6):  $R=CO_2CH_3$

(7):  $R=CH_2OH$   
(ceriferol)

(8):  $R=CH_2OTs$

(9):  $R=CH_3$



(10)

Table 1.  $^{13}\text{C}$ -NMR chemical shifts of methyl ceriferate (6), ceriferol (7), " $\alpha$ -cericerene" (9), and cericerene (11) in  $\text{CDCl}_3$ . Peak assignments are based on measurements of  $T_1$  and  $^{13}\text{C}$ -NOE<sup>1</sup> in addition to the more conventional techniques.

| C-atom              | <u>6</u>           | <u>7</u>           | <u>9</u>           | <u>11</u>          |
|---------------------|--------------------|--------------------|--------------------|--------------------|
| C-1                 | 29.8 <sup>a</sup>  | 29.6 <sup>a</sup>  | 29.4 <sup>a</sup>  | 30.5 <sup>a</sup>  |
| C-2                 | 125.3 <sup>b</sup> | 125.0 <sup>b</sup> | 125.2 <sup>b</sup> | 125.1 <sup>b</sup> |
| C-3                 | 133.7 <sup>c</sup> | 133.2 <sup>c</sup> | 133.8 <sup>c</sup> | 134.1 <sup>c</sup> |
| C-4                 | 31.2 <sup>a</sup>  | 29.9 <sup>a</sup>  | 30.4 <sup>a</sup>  | 31.1 <sup>a</sup>  |
| C-5                 | 26.9               | 27.2               | 31.0 <sup>a</sup>  | 31.4 <sup>a</sup>  |
| C-6                 | 142.4              | 127.6              | 124.9 <sup>b</sup> | 125.1 <sup>b</sup> |
| C-7                 | 131.3 <sup>d</sup> | 137.6              | 133.1 <sup>c</sup> | 132.9 <sup>c</sup> |
| C-8                 | 36.0               | 35.8               | 35.8               | 36.2               |
| C-9                 | 26.0               | 24.6 <sup>d</sup>  | 24.5               | 24.6               |
| C-10                | 125.4 <sup>b</sup> | 125.0 <sup>b</sup> | 124.9 <sup>b</sup> | 125.0 <sup>b</sup> |
| C-11                | 132.9 <sup>c</sup> | 133.1 <sup>c</sup> | 132.9 <sup>c</sup> | 133.0 <sup>c</sup> |
| C-12                | 40.3               | 30.2               | 40.2               | 40.3               |
| C-13                | 24.5               | 24.5 <sup>d</sup>  | 24.5               | 24.6               |
| C-14                | 49.9               | 46.4               | 46.6               | 44.6               |
| C-15                | 136.4              | 136.8              | 137.1              | 153.0              |
| C-16                | 123.4              | 123.5              | 123.7              | 33.7               |
| C-17                | 26.9               | 26.9               | 26.8               | 26.6               |
| C-18                | 124.6              | 124.8 <sup>b</sup> | 124.5              | 124.6 <sup>b</sup> |
| C-19                | 131.1 <sup>d</sup> | 131.2              | 131.0              | 131.3              |
| C-20                | 25.7               | 25.6               | 25.6               | 25.7               |
| C-21                | 17.7               | 17.7               | 17.7               | 17.8               |
| C-22                | 12.3               | 12.0               | 12.0               | 108.9              |
| C-23                | 15.6 <sup>e</sup>  | 15.6 <sup>e</sup>  | 15.6 <sup>d</sup>  | 15.6 <sup>d</sup>  |
| C-24                | 168.4              | 66.6               | 22.4               | 22.5               |
| C-25                | 15.3 <sup>e</sup>  | 15.4 <sup>e</sup>  | 15.4 <sup>d</sup>  | 15.5 <sup>d</sup>  |
| -COOCH <sub>3</sub> | 51.3               |                    |                    |                    |

a-e) Assignments denoted by same alphabet are interchangeable.

skeleton framework,<sup>4</sup> must be remote from the sole chiral center at C-14; if it were as in 10, the CD should have shown a Cotton effect at the  $\lambda_{\max}$  of ceriferic acid. Of the two 14-membered skeletal possibilities, 5 (or 1) and 10, it is only structure 5 which satisfies the two criteria underscored above.

2) Reduction of methyl ceriferate (6) with  $\text{LiAlH}_4$  gave "ceriferol" (7), which was further converted into the tosylate (8) by careful treatment with  $\text{TsCl/py}$  at 0 °C. Tosylate 8 with its strongly absorbing chromophore also lacked a CD Cotton effect, thus confirming the deductions described above.

3) Acid treatment of tosylate 8 simply resulted in hydrolysis to alcohol 7. In contrast, acid treatment of the tosylate derived from cericerol-I 1 underwent transannular cyclization to a bicyclocericerene.<sup>5</sup>

4) The neutral fraction of *C. ceriferus* collected around Kyoto in 1979 has given, in addition to cericerol-I 1, an alcohol,  $[\alpha]_{\text{D}}^{24} -83.5^\circ$  (c, 1.33), the physical constants (MS, NMR, IR, and rotation) of which were identical with those of allyl alcohol 7 (ceriferol). Furthermore, the acidic fraction<sup>1</sup> gave ceriferic acid,  $[\alpha]_{\text{D}}^{26} -97.6^\circ$  (c, 1.02); these findings are in line with the taxonomical identify of the Tokyo and Kyoto species.

5) The  $^{13}\text{C}$ -NMR data of ceriferic acid 5 derivatives are in good agreement with cericerol-I 1 derivatives, a further support for the skeletal identity. This is exemplified by a comparison of the data for hydrocarbons (9),  $[\alpha]_{\text{D}}^{19} -28.8^\circ$  (c, 0.24) (derived from tosylate 8 by  $\text{LiAlH}_4$  treatment) and (11),<sup>1</sup>  $[\alpha]_{\text{D}}^{27} -48.3^\circ$  (c, 0.48) (derived from cericerol-I) (Table 1). The data for methyl ceriferate 6 and ceriferol 7 given in Table 1 are in accord with the structural variations.

Finally, in analogy with other cericerol-I 1 derivatives, we assign an R-configuration (or " $\beta$ " as depicted) to the C-14 of ceriferic acid 5.

#### REFERENCES

- 1) F. Miyamoto, H. Naoki, T. Takemoto, and Y. Naya, *Tetrahedron*, **35**, 1913 (1979).
- 2) F. Miyamoto, H. Naoki, Y. Naya, and K. Nakanishi, *Tetrahedron* in press.
- 3) R. Veloz, L. Quijano, J.S. Calderon, and T. Rios, *J.C.S. Chem. Commun.*, **1975**, 191.
- 4) T. Kusumi, T. Kinoshita, K. Fujita, and H. Kakisawa, *Chem. Lett.*, **1979**, 1129.
- 5) To be published elsewhere.

\*\* Previous name: The Institute of Food Chemistry

(Received May 6, 1980)