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Investigation on the ZBG-functionality of phenyl-4-ylacrylohydroxamic acid derivatives as histone deacetylase inhibitors

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ABSTRACT

A series of alternative Zn-binding groups were explored in the design of phenyl-4-yl-acrylohydroxamic acid derivatives as histone deacetylase (HDAC) inhibitors. Most of the synthesized compounds were less effective than the parent hydroxamic acid. However, the profile of activity shown by the analog bearing a hydroxyurea head group, makes this derivative promising for further investigation.

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Over the last 10 years the reversible acetylation of histones and other proteins has emerged as an important regulatory mechanism implicated in cell proliferation and has been identified as a valuable target for anticancer drug design.

Acetylation is executed by histone acetyltransferases and is reversed by histone deacetylases (HDACs).¹ Eighteen mammalian HDAC enzymes have been identified so far, which can be subdivided into two families according to a cofactor (Zn²⁺ for Classes I, II and IV or NAD⁺ for Class III) for activity.²

A number of HDAC inhibitors reached clinical trials, and five of them were approved by authorities like the US FDA: Vorinostat (Zolinza; 2006), Romidepsin (Istodax; 2009) both for use in peripheral T-cell lymphoma (CTCL) with the latter also for use in peripheral T-cell lymphoma (CTCL) (2011). Belinostat (Beleodaq; 2014) and Chidamide (Epidaza; 2015) both for use in CTCL and, more recently, Panobinostat (Farydak; 2015) for use in multiple myeloma.³ They all fit in the widely accepted HDAC pharmacophore model for HDAC inhibitors characterized by a cap group, a linker chain and a zinc binding group (ZBG).⁴ Based on molecular

modelling studies involving the histone deacetylase-like protein (HDLP), these molecules appear to bind in a pocket of the HDAC active site in such a way that the metal binding moiety (a hydrox-amic acid for SAHA, Belinostat and Panobinostat, a thiol derivative for Romidepsin and a benzamide for Chidamide), coordinates the catalytic zinc ion at the end of the pocket. Binding of the Zn²⁺ ion has proven to be crucial for the activity.

Since the discovery of SAHA, structural studies to identify novel HDAC inhibitors have mainly focused on modifications of the linker or the cap group, while less attention has been paid to the metal binding group.

Hydroxamic acid derivatives represent the most studied and successful class of HDAC inhibitor drug candidates. However, although very powerful, they suffer from poor metabolic stability and a rapid excretion.^{5,6} The different clinical applications of this class of drugs, such as CNS, inflammatory and HIV diseases, require a renewed effort in the search for new ZBG.

Several efforts have been made to replace the hydroxamic acid with alternative chemical moieties that could also further improve the isoform selectivity of HDAC inhibitors.^{7–11}

As part of a previous research program aimed at studying new HDAC inhibitors, we have developed a series of hydroxamic acidbased compounds, characterized by a cinnamic spacer capped with a substituted phenyl group (**1**, Fig. 1).¹² Extensive research has been conducted on the derivatization of the cap group, to optimize drug-target interactions.¹²⁻¹⁴

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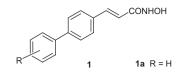
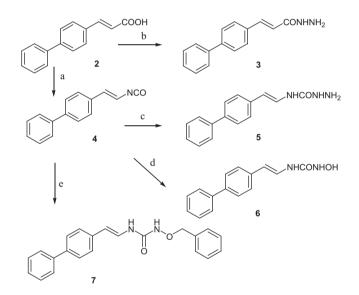


Figure 1. General structure of compounds 1.

Recently, we have initiated the search for moieties that would retain the zinc-binding properties and enzyme inhibitory activity of the hydroxamic acid group. In this Letter we report the attempts made to modify the ZBG-functionality of the 4-vinylbiphenyl scaffold.

A series of analogs incorporating non-hydroxamic groups was designed to probe the potential of various ZBGs. Indeed, very stringent constraints should be satisfied in order to have zinc binding (as with the hydroxamic acid group), and also interactions with amino acid residues of the enzymes to have a strong binding affinity.¹⁵ The *N*-hydroxyurea group appears to be an interesting possible replacement for the hydroxamate group commonly found in HDAC inhibitors, because both groups display an identical binding mode.^{15–17} SAHA *N*-hydroxyurea derivative was found to show anti-HDAC activity even though it was much less effective than



Scheme 1. Synthesis of compounds **2–7**. Reagents and conditions: (a) DPPA, TEA, CH₂Cl₂, reflux, 7 h, 100%; (b) HOBT, EDC, NH₂NH₂·H₂O, cyclohexene, acetonitrile, 0–10 °C, 10 min, 77%; (c) NH₂NH₂·H₂O, toluene, 80 °C, 1 h, 63%; (d) NH₂OH, toluene, reflux, 1 h, 53%; (e) *O*-benzylhydroxylamine, toluene, reflux, 3.5 h, 23%.

SAHA.¹⁶ To investigate the effect of the hydroxyurea group on the 4-vinylbiphenyl scaffold, compound **6** was synthesized starting from acid **2** (weak Zn^{2+} coordinator), which was tested for its HDAC inhibitory activity as well. The acid was quantitatively converted into the corresponding isocyanate **4** by treatment with diphenylphosphorylazide (DPPA) in dry CH₂Cl₂. This latter was reacted with hydroxylamine in toluene to give the hydroxyurea **6**. The same strategy was used to prepare the *O*-benzyl derivative **7** (Scheme 1).

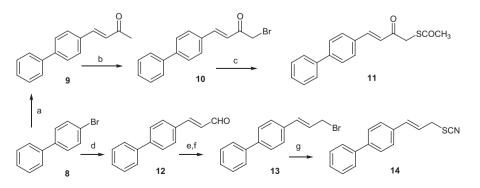
Isocyanate **4** was also used as an intermediate to prepare semicarbazide **5**, based on the consideration that a SAHA semicarbazide derivative was found to be active as HDAC inhibitor.¹⁶ As other derivatives with a hydrazide group showed HDAC inhibitory activity,¹⁸ we also replaced the hydroxamic acid moiety of reference compound **1a** with a hydrazide counterpart. Accordingly, compound **3** was prepared by condensation of acid **2** with hydrazine in the presence of HOBt and EDC (Scheme 1).¹⁹

Other groups were selected based on the concept that it is possible for them to bind to the zinc ion. Sulfur ligands are well known to bind tightly to Zn containing enzymes. In particular, HDAC inhibitors with α -thioacetate²⁰ as well as α -thioacetoxyketone moiety in place of the hydroxamic acid showed higher inhibition of the enzyme than SAHA.^{21,22} Based on these results, the synthesis of analogs harboring an α -thioacetoxyketone as a ZBG on the phenyl-4-yl-acryloyl scaffold was performed. Heck condensation between 4-bromobiphenyl **8** and but-3-en-2-one afforded ketone **9** that was brominated with pyrrolidone hydrotribromide to obtain **10**. Treatment of this latter with potassium thioacetate gave the desired thioacetoxyketone **11** in good yield (Scheme 2).

We also speculated that another possible ZBG could be the thiocyanate group, which is known to bind the Zn ion through the nitrogen atom.²³ Thus, compound **14** was prepared from 4-bromobiphenyl **8**, which was coupled with 3,3-diethoxypropene to obtain aldehyde **12** in 78% yield. Reduction by NaBH₄, followed by treatment with PBr₃ afforded the bromide **13** in 88% overall yield. Reaction with silica gel-supported potassium thiocyanate²⁴ afforded compound **14** in good yield (Scheme 2).

During the past years, Cohen's group has investigated several new cyclic ketones and lactones as ZBGs for Zn-dependant enzymes. Among them, they proposed some 6-membered cyclic hydroxamic and thiohydroxamic acids.²⁵ The purpose to investigate the effect of a five-membered cyclic hydroxamic acid, which should be the closest structural analog of hydroxamic acid, prompted us to synthesize compound **17** (Scheme 3). When this work was already in an advanced stage, the same group was reported as a novel ZBGs for HDAC inhibitors.²⁶

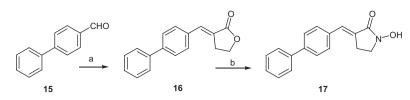
The synthesized compounds were tested for their inhibitory activity towards HDAC2 isoform (Table 1), and for cell growth



Scheme 2. Synthesis of compounds 11 and 14. Reagents and conditions: (a) but-3-en-2-one, palladium acetate, tri(*o*-tolyl)phosphine, TEA, 100–110 °C, 12 h, 42%; (b) pyrrolidone hydrotribromide, THF, rt, 24 h, 41%; (c) potassium thioacetate, DMF, rt, 2.5 h, 56%; (d) 3,3-diethoxypropene, Bu₄NOAc, K₂CO₃, KCl, Pd(OAc)₂, DMF, 100 °C, 5 h, 78%; (e) NaBH₄, EtOH, rt, 30 min, 98%; (f) PBr₃, Et₂O, rt, 1 h, 90%; (g) potassium thiocyanate–SiO₂, hexane, 50 °C, 3.5 h, 78%.

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Scheme 3. Synthesis of compound 17. Reagents and conditions: (a) butyrolactone, NaOMe, toluene, -10 °C for 5 min, then rt for 1.5 h, 89%; (b) NH₂OH·HCl, EtONa, reflux, 3.5 h, 20%.

Table 1HDAC2 inhibitory activity of synthesized compounds

Compd	HDAC2 ^a (IC ₅₀ μM)		
SAHA	0.15 ± 0.02		
1a	0.82 ± 0.14		
2	>5		
3	>5		
5	>5		
6	1.05 ± 0.62		
7	>5		
11	>5		
14	>5		
17	>5		

^a HDAC activity was determined using HDAC2 from HeLa cells as indicated in SI.

Table 2 Cell growth inhibition (IC₅₀, μ M) of synthesized compounds and reference HDAC inhibitor (SAHA) against a panel of tumor cell lines

$\begin{array}{c c c c c c c c c c c c c c c c c c c $						
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Compd	IGROV-1	IGOV/Pt1	HCT116	NB4	H460
1a 3.5 ± 0.6 4.0 ± 0.4 1.6 ± 0.3 0.9 ± 0.1 5.4 ± 0.1 2>20>10>203 7.6 ± 0.2 25.1 ± 5.9 11.9 ± 1.2 7.1 ± 0.7 >205 14.5 ± 0.3 26.7 ± 0.3 16.4 ± 3.0 9.8 ± 0.9 >206 3.5 ± 0.3 6.7 ± 0.2 4.0 ± 0.7 1.96 ± 0.1 9.5 ± 0.4 7 40 ± 1 >50 40 ± 3 >20>2011 26.4 ± 0.8 18.7 ± 0.6 14.0 ± 1.5 >20>2014 3.8 ± 0.6 58.1 ± 2.6 12.6 ± 3.8 >20 9.4 ± 1.6				$I{C_{50}}^a(\mu M)$		
2>20>10>203 7.6 ± 0.2 25.1 ± 5.9 11.9 ± 1.2 7.1 ± 0.7 >205 14.5 ± 0.3 26.7 ± 0.3 16.4 ± 3.0 9.8 ± 0.9 >206 3.5 ± 0.3 6.7 ± 0.2 4.0 ± 0.7 1.96 ± 0.1 9.5 ± 0.4 7 40 ± 1 $\gg 50$ 40 ± 3 >20>2011 26.4 ± 0.8 18.7 ± 0.6 14.0 ± 1.5 >20>2014 3.8 ± 0.6 58.1 ± 2.6 12.6 ± 3.8 >20 9.4 ± 1.6	SAHA	2.2 ± 0.3	2.2 ± 0.2	1.2 ± 0.1	0.70 ± 0.03	3.4 ± 0.8
3 7.6 ± 0.2 25.1 ± 5.9 11.9 ± 1.2 7.1 ± 0.7 >205 14.5 ± 0.3 26.7 ± 0.3 16.4 ± 3.0 9.8 ± 0.9 >206 3.5 ± 0.3 6.7 ± 0.2 4.0 ± 0.7 1.96 ± 0.1 9.5 ± 0.4 7 40 ± 1 $\gg 50$ 40 ± 3 >20>2011 26.4 ± 0.8 18.7 ± 0.6 14.0 ± 1.5 >20>2014 3.8 ± 0.6 58.1 ± 2.6 12.6 ± 3.8 >209.4 \pm 1.6	1a	3.5 ± 0.6	4.0 ± 0.4	1.6 ± 0.3	0.9 ± 0.1	5.4 ± 0.1
5 14.5 ± 0.3 26.7 ± 0.3 16.4 ± 3.0 9.8 ± 0.9 >206 3.5 ± 0.3 6.7 ± 0.2 4.0 ± 0.7 1.96 ± 0.1 9.5 ± 0.4 7 40 ± 1 $\gg 50$ 40 ± 3 >20>2011 26.4 ± 0.8 18.7 ± 0.6 14.0 ± 1.5 >20>2014 3.8 ± 0.6 58.1 ± 2.6 12.6 ± 3.8 >20 9.4 ± 1.6	2	_	-	>20	>10	>20
6 3.5 ± 0.3 6.7 ± 0.2 4.0 ± 0.7 1.96 ± 0.1 9.5 ± 0.4 7 40 ± 1 $\gg 50$ 40 ± 3 >20 >20 11 26.4 ± 0.8 18.7 ± 0.6 14.0 ± 1.5 >20 >20 14 3.8 ± 0.6 58.1 ± 2.6 12.6 ± 3.8 >20 9.4 ± 1.6	3	7.6 ± 0.2	25.1 ± 5.9	11.9 ± 1.2	7.1 ± 0.7	>20
7 40 ± 1 $\gg 50$ 40 ± 3 > 20 > 20 11 26.4 ± 0.8 18.7 ± 0.6 14.0 ± 1.5 > 20 > 20 14 3.8 ± 0.6 58.1 ± 2.6 12.6 ± 3.8 > 20 9.4 ± 1.6	5	14.5 ± 0.3	26.7 ± 0.3	16.4 ± 3.0	9.8 ± 0.9	>20
11 26.4 ± 0.8 18.7 ± 0.6 14.0 ± 1.5 >20 >20 14 3.8 ± 0.6 58.1 ± 2.6 12.6 ± 3.8 >20 9.4 ± 1.6	6	3.5 ± 0.3	6.7 ± 0.2	4.0 ± 0.7	1.96 ± 0.1	9.5 ± 0.4
14 3.8 ± 0.6 58.1 ± 2.6 12.6 ± 3.8 >20 9.4 ± 1.6	7	40 ± 1	≫50	40 ± 3	>20	>20
	11	26.4 ± 0.8	18.7 ± 0.6	14.0 ± 1.5	>20	>20
17 ≫60 ≫60 >20 >20 >20	14	3.8 ± 0.6	58.1 ± 2.6	12.6 ± 3.8	>20	9.4 ± 1.6
	17	≫60	≫60	>20	>20	>20

^a Cells were exposed for 72 h. Cell growth inhibitory activity was measured by cell counting and expressed as concentration required for 50% inhibition of cell growth (IC₅₀). IC₅₀ values were determined by dose–response curves. Results are the means \pm SD of three independent experiments.

inhibition against a panel of tumor cell lines of different tissue origin: NB4 (human acute promyelocytic leukemia), H460 (human lung carcinoma), HCT116 (human colon cancer), IGROV-1 and its subline resistant to cisplatinum IGROV-1/Pt1 (human ovarian carcinoma) (Table 2).

Most of the synthesized compounds were characterized by a low activity (compared to SAHA) in terms of both HDAC inhibition and cell growth inhibition. In spite of a lack of enzymatic activity, compounds **3**, **5**, **11**, and **14** showed a moderate cell growth inhibition, most likely related to the inhibition of multiple targets. In fact, there appears to be a common opinion that inhibitors containing metal binding groups can be prone to widespread off-target enzymatic inhibition, due to non-specific metal binding.²⁷

The introduction of an O-benzyl hydroxyurea moiety (compound **7**) and of a five-membered cyclic hydroxamic acid (compound **17**) led to a loss of activity Only compound **6**, bearing a hydroxyurea as ZBG, exhibited activity comparable to the parent compound **1a**, both in HDAC2 and cell growth inhibitions.

Thus, the profiling of **6** was carried out also against HDAC1 and HDAC6 isoforms, using SAHA as a reference compound. The results (HDAC1: compound **6**, IC₅₀ 1.31 ± 0.21 μ M. SAHA, IC₅₀ 0.12 ± 0.01 μ M; HDAC6: compound **6**, IC₅₀ 1.05 ± 0.09 μ M. SAHA, IC₅₀ 0.07 ± 0.01 μ M) suggested that the compound was a multi-HDAC inhibitor, its activity against HDAC1 and 6 being comparable to the activity against HDAC2 (see SI for details).

A structural rationalization study was conducted in order to understand the main interaction features of compounds **1a** and **6** with the enzyme. The molecular docking protocol adopted here has been validated in the past through the re-docking of the cocrystallized SAHA within the HDAC2 protein structure.²⁸ Both the hydroxamate and *N*-hydroxyurea groups of compound **1a** (ChemPLP score of the best ranked pose: 69.8; 100% convergent docking solutions) and **6** (ChemPLP score of the best ranked pose:

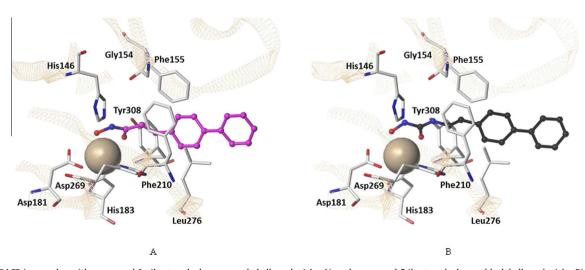


Figure 2. HDAC2 in complex with compound 1a (best-ranked pose, purple balls and sticks, A), and compound 6 (best-ranked pose, black balls and sticks, B). The protein is reported as gray ribbons. Zinc ion is indicated as a gray ball. Key residues for ligands stabilization are labeled.

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75.4; 100% convergent docking solutions), respectively, were involved in zinc chelation. Polar interactions of these chelating groups with neighbor residues (His146, Tyr308) were also observed. The additional nitrogen atom present in compound **6** made a hydrogen bond with Gly154. The zinc binding groups of both molecules were connected to two phenyl rings, the first one stabilized by hydrophobic interactions with Phe105, Phe210, and Leu276, the second one solvent exposed in both cases (Fig. 2).

In conclusion, we explored alternative ZBGs in the design of phenyl-4-yl-acrylohydroxamic acid derivatives as HDAC inhibitors. Most of the synthesized compounds were less effective than the parent hydroxamic acid **1a**. However, the profile of activity shown by compound **6**, bearing a hydroxyurea head group, makes this derivative a promising candidate for further investigation, also taking into account the potential improvement in terms of bioavail-ability/metabolism of hydroxyurea over hydroxamate.^{29,30}

Acknowledgments

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.bmcl.2015.09. 006.

References and notes

- 1. Grunstein, M. Nature 1997, 389, 349.
- 2. Mottamal, M.; Zheng, S.; Huang, T. L.; Wang, G. Molecules 2015, 20, 3898.
- 3. Falkenberg, K. J.; Johnstone, R. W. Nat. Rev. Drug Disc. 2014, 13, 673.
- Paris, M.; Porcelloni, M.; Binaschi, M.; Fattori, D. J. Med. Chem. 2008, 51, 1505.
 Summers, J. B.; Gunn, B. P.; Mazdiyasni, H.; Goetze, A. M.; Young, P. R.; Bouska,
- J. B.; Dyer, R. D.; Brooks, D. W.; Carter, G. W. J. Med. Chem. **1987**, 30, 2121. 6. Kelly, W. K.; Richon, V. M.; O'Connor, O.; Curley, T.; MacGregor-Curtelli, B.;
- Tong, W.; Klang, M.; Schwartz, L.; Richardson, S.; Rosa, E.; Drobnjak, M.; Cordon-Cordo, C.; Chiao, J. H.; Rifkind, R.; Marks, P. A.; Scher, H. *Clin. Cancer Res.* 2003, 9, 3578.
- Sodji, Q. H.; Patil, V.; Kornacki, J. R.; Mrksich, M.; Oyelere, A. K. J. Med. Chem. 2013, 56, 9969.

- Ononye, S. N.; VanHeyst, M. D.; Oblak, E. Z.; Zhou, W.; Ammar, M.; Anderson, A. C.; Wright, D. L. ACS Med. Chem. Lett. 2013, 4, 757.
- 9. Dallavalle, S.; Pisano, C.; Zunino, F. Biochem. Pharmacol. 2012, 84, 756.
- 10. Madsen, A. S.; Kristensen, H. M. E.; Lanz, G.; Olsen, C. A. *ChemMedChem* 2014, 9, 614.
- 11. Suzuki, T.; Miyata, N. Curr. Med. Chem. 2005, 12, 2867.
- Dallavalle, S.; Cincinelli, R.; Nannei, R.; Merlini, L.; Morini, G.; Penco, S.; Pisano, C.; Vesci, L.; Barbarino, M.; Zuco, V.; De Cesare, M.; Zunino, F. *Eur. J. Med. Chem.* 2009, 44, 1900.
- Cincinelli, R.; Musso, L.; Giannini, G.; Zuco, V.; De Cesare, M.; Zunino, F.; Dallavalle, S. Eur. J. Med. Chem. 2014, 79, 251.
- Giannini, G.; Brunetti, T.; Pezzi, R.; Battistuzzi, G.; Di Marzo, M.; Marzi, M.; Cabri, W.; Vesci, L.; Pisano, C. Bioorg. Med. Chem. Lett. 2009, 19, 2346.
- Vanommeslaeghe, K.; Loverix, S.; Geerlings, P.; Tourwé, D. Bioorg. Med. Chem. 2005, 13, 6070.
- Suzuki, T.; Nagano, Y.; Kouketsu, A.; Matsuura, A.; Maruyama, S.; Kurotaki, M.; Nakagawa, H.; Miyata, N. J. Med. Chem. 2005, 48, 1019.
- Hanessian, S.; Vinci, V.; Auzzas, L.; Marzi, M.; Giannini, G. Bioorg. Med. Chem. Lett. 2006, 16, 4784.
- Hou, X.; Du, J.; Liu, R.; Zhou, Y.; Li, M.; Xu, W.; Fang, H. J. Chem. Inf. Model. 2015, 55, 861.
- Zhang, X.; Breslav, M.; Grimm, J.; Guan, K.; Huang, A.; Liu, F.; Maryanoff, C. A.; Palmer, D.; Patel, M.; Qian, Y.; Shaw, C.; Sorgi, K.; Stefanick, S.; Xu, D. J. Org. *Chem.* 2002, 67, 9471.
- Giannini, G.; Vesci, L.; Battistuzzi, G.; Vignola, D.; Milazzo, F. M.; Berardino Guglielmi, M.; Barbarino, M.; Santaniello, M.; Fanto, N.; Mor, M.; Rivara, S.; Pala, D.; Taddei, M.; Pisano, C.; Cabri, W. J. Med. Chem. 2014, 57, 8358.
- Wash, P. L.; Hoffman, T. Z.; Wiley, B. M.; Bonnefous, C.; Smith, N. D.; Sertic, M. S.; Lawrence, C. M.; Symons, K. T.; Nguyen, P.-M.; Lustig, K. D.; Guo, X.; Annable, T.; Noble, S. A.; Hager, J. H.; Hassig, C. A.; Malech, J. W. Bioorg. Med. Chem. Lett. 2008, 18, 6482.
- Gu, W.; Nusinzon, I.; Smith, R. D., Jr.; Horvath, C. M.; Silverman, R. B. Bioorg. Med. Chem. 2006, 14, 3320.
- Diaz, A.; Massacesi, M.; Ponticelli, G.; Paschina, G. J. Inorg. Nucl. Chem. 1975, 37, 2469.
- 24. Ando, T.; Clark, J. H.; Cork, D. G.; Fujita, M.; Kimura, T. J. Org. Chem. 1987, 52, 681.
- Jacobsen, J. A.; Fullagar, J. L.; Miller, M. T.; Cohen, S. M. J. Med. Chem. 2011, 54, 591.
- Mutule, I.; Borovika, D.; Rozenberga, E.; Romanchikova, N.; Zalubovskis, R.; Shestakova, I.; Trapencieris, P. J. Enzyme Inhib. Med. Chem. 2015, 30, 216.
- 27. Day, J. A.; Cohen, S. M. J. Med. Chem. 2013, 56, 7997.
- Zwick, V.; Chatzivasileiou, A.-O.; Deschamps, N.; Roussaki, M.; Simoes-Pires, C.; Nurisso, A.; Denis, I.; Blanquart, C.; Martinet, N.; Carrupt, P.-A.; Detsi, A.; Cuendet, M. Bioorg. Med. Chem. Lett. 2014, 24, 5497.
- Campestre, C.; Agamennone, M.; Tortorella, P.; Preziuso, S.; Biasone, A.; Gavuzzo, E.; Pochetti, G.; Mazza, F.; Hiller, O.; Tschesche, H.; Consalvi, V.; Gallina, C. Bioorg. Med. Chem. Lett. 2006, 16, 20.
- 30. Stewart, A. O.; Bhatia, P. A.; Martin, J. G.; Summers, J. B.; Rodriques, K. E.; Martin, M. B.; Holms, J. H.; Moore, J. L.; Craig, R. A.; Kolasa, T.; Ratajczyk, J. D.; Mazdiyasni, H.; Kerdesky, F. A. J.; DeNinno, S. L.; Maki, R. G.; Bouska, J. B.; Young, P. R.; Lanni, C.; Bell, R. L.; Carter, G. W.; Brooks, C. D. W. J. Med. Chem. 1997, 40, 1955.