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Introduction

Thromboxane A_2 (1a, Fig. 1), an unstable metabolite of arachidonic acid, is responsible for a wide range of pulmonary, renal, circulatory and other disorders. Thromboxane A2 causes irreversible platelet aggregation, vasoconstriction and smooth muscle cell proliferation. It is released from activated platelets, monocytes and damaged vessel walls. The activity of thromboxane A_2 is through the thromboxane-type prostanoid (TP) receptor extensively present on platelets and vascular smooth muscle. TP receptor agonists and thromboxane A2 synthase inhibitors are potential antiplatelet agents. Many of these are also TP receptor agonists and have very short half-lives.¹ Thus, there is always a requirement for a more effective, selective and long-lasting thromboxane receptor antagonist. In one of the studies to find a selective antagonist, it was observed that compounds containing a carboxylic acid and a benzenesulfonamide group separated by a spacer were found to be the best among the compounds tested. Ramatroban (1b),² (Fig. 1) a compound with these features, marketed as Baynas® by Bayer and Nippon Shinyaku Co. for allergic rhinitis, is a selective TP receptor antagonist. The pharmacophore was further explored by Lavielle et al., which led to the identification of a tetrahydronaphthalene derivative, terutroban (2) (Fig. 1) as a TP receptor antagonist. The only reported synthesis of 2 to date is achieved using Diels-Alder as the key reaction between an appropriate 2-pyrone and an acetylenic derivative.³ Terutroban is being developed by the Institut de Recherches Servier (France) and has entered Phase III clinical trials under PERFORM (prevention of cerebrovascular events of ischemic origin with terutroban in patients with a history of ischemic

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Total synthesis of a thromboxane receptor antagonist, terutroban†

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A total synthesis of terutroban is achieved using the Claisen rearrangement, Friedel–Crafts acylation and Heck coupling as key reactions, avoiding the classical Diels–Alder approach used before.



Fig. 1 Structure of ramatroban (1) and terutroban (2).

stroke or transient ischemic attack). The study was prematurely stopped, and the reasons and results have not been disclosed. Nonetheless, the skeleton attracted our attention due to our interest in molecules with important biological activities, including the synthesis of prostanoids, beraprost (antiplatelet drug)⁴ and iloprost (drug for pulmonary arterial hypertension).⁵ We wish to explore a non-Diels–Alder approach for the synthesis of this molecule which can be further extended to synthesize analogues.

Present work

In general, the tetralin units, such as that present in terutroban, are built using a Diels–Alder reaction.⁶ The reported synthesis of terutroban considered the molecule as a substituted benzene analogue which was built by a Diels–Alder reaction, as is the norm. We explored the synthesis of 2 as a tetrahydronaphthalene analogue built starting from a benzene ring. In a retrosynthetic analysis, (Fig. 2) terutroban can be synthesized

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from olefin 3 in six steps involving sulphonamidation under Mitsunobu conditions. Compound 3 can be obtained from 4 in five sequential steps using dehydration and Heck coupling as the key reactions. The tetralone 4 in turn can be synthesized starting from commercially available *o*-cresol 5 in seven steps involving o-allylation, Claisen rearrangement, oxidative olefin cleavage, Wittig olefination, olefin reduction, ester hydrolysis and a Friedel–Crafts acylation reaction.

The use of an easily available and low cost starting material, o-cresol (~15 USD Kg⁻¹), was considered to enable a facile, large scale synthesis of the target molecule 2. Thus o-cresol gave 6 following a known sequence.⁷ Lemieux–Johnson oxidation–dihydroxylation with catalytic osmium tetraoxide followed by carbon–carbon bond cleavage using sodium periodate resulted in aldehyde 7 in 75% yield over two steps (Scheme 1).

Wittig olefination with carbethoxymethylene triphenylphosphorane delivered the olefin, which upon reduction with sodium borohydride and NiCl₂·6H₂O gave 8 in 72% yield over two steps. Hydrolysis of the ester using lithium hydroxide afforded 9 (86% yield), and intramolecular Friedel-Crafts acylation with PPA resulted in tetralone 4 in 79% yield. Reduction of the ketone with sodium borohydride at 0 °C provided alcohol 10 in 90% yield, followed by dehydration of the secondary alcohol with PPA⁸ at 100 °C which gave 11 in 76% yield. Demethylation of the phenolic methyl ether using boron trichloride⁹ afforded 12 in 77% yield. Conversion of the phenolic hydroxyl group to a triflate followed by Heck coupling¹⁰ with methyl acrylate in presence of bis(triphenylphosphine)palladium(II) chloride provided unsaturated ester 13 in 60% yield over two steps. The double bond in 13 was selectively reduced with magnesium in methanol¹¹ to yield the key intermediate 3 in 84% yield. Compound 3 has both the rings and



Scheme 1 Synthesis of intermediate 7.



Scheme 2 Synthesis of key intermediate 3.

the required functional groups to synthesize the target molecule (Scheme 2).

The next logical step was to synthesize an aziridine¹² analogue, which can be selectively opened to obtain the target molecule. The reaction of sodium chloro(4-chlorophenyl)sulfonyl amide and iodine at pH 7 buffer with 3 did not yield the required product. Other different reaction conditions also did not yield the required aziridine.

As the desired aziridine was not obtained, the next effort was to synthesize an epoxide and open it to obtain an alcohol. Thus, an epoxidation reaction was tried with *m*CPBA, H_2O_2 -titanium isopropoxide, H_2O_2 *etc.*¹² The reaction always resulted in an inseparable mixture of compounds.

A stepwise approach was then followed to synthesize 2. Thus, key intermediate **3** was reacted with osmium tetroxide to get diol **14** in 78% yield. Diol **14** was converted to a carbonate derivative with triphosgene¹³ with 84% yield, which on reduction with 10% Pd/C under hydrogenation conditions¹⁴ afforded alcohol **16** in 88% yield. Mitsunobu conditions¹⁵ were employed to convert the alcohol to an azide in 83% yield. Reduction over 10% Pd/C under hydrogenation conditions¹⁶, followed by *in situ* conversion of the amine, provided the ester **17** in 78% yield. Hydrolysis of the ester with sodium hydroxide³ gave terutroban **2**, in 82% yield, which resembled the reported acid in all respects (Scheme 3).

Experimental

General

¹H NMR and ¹³C NMR spectra were recorded on a Varian Gemini 200, Bruker Avance 300 or Varian Innova 500 at 75, 125, 300 or 500 MHz in CDCl₃ or DMSO(D₆) solvent at ambient temperature. Chemical shifts δ are given in ppm, coupling constants *J* are in Hz. The chemical shifts are reported in ppm on a scale downfield from TMS as the internal standard, and signal patterns are indicated as follows: s, singlet; d, doublet;



2

Scheme 3 Synthesis of terutroban 2.

dd, doublet of doublets; dt, doublet of triplets; t, triplet; m, multiplet; bs, broad singlet, quin., quintet. FT-IR Spectra: a Perkin Elmer FT-IR was used to record spectra as KBr thin films or neat; $\nu_{\rm max}$ are given in cm⁻¹. ESI- and HR-ESI-MS were recorded using a Finnigan MAT 1020B; values are given in m/z. All the reagents and solvents were reagent grade and used without further purification unless specified otherwise. Technical grade ethyl acetate and hexanes used for column chromatography were distilled prior to use. Column chromatography was carried out using silica gel (60–120 mesh) packed in glass columns. All the reactions were performed under an atmosphere of nitrogen in oven-dried glassware with magnetic stirring.

2-(2-Methoxy-3-methylphenyl)acetaldehyde (7). To a solution of 6 (28.5 g, 175 mmol) in acetone-acetonitrile-water (1:1:1) (300 mL), were added OsO4 (44 mg, 0.18 mmol) and NMO (41.16 g, 351 mmol). The reaction was stirred at room temperature for 12 h, quenched with aqueous sodium sulphite solution and stirred for an additional 30 min. The solution was concentrated in vacuo to remove organic solvents. The aqueous phase was extracted with ethyl acetate $(3 \times 150 \text{ mL})$. The combined organic extracts were washed with brine (50 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The crude was purified by column chromatography (1:1 hexanes-EtOAc) to afford diol (29.3 g, 85%) as a colorless thick liquid. ¹H NMR (300 MHz, CDCl₃) δ 7.11-6.95 (m, 3H), 4.17-4.03 (m, 1H), 3.95-3.86 (m, 2H), 3.76 (s, 3H), 3.60 (dd, J = 11.5, 3.2 Hz, 1H), 3.46 (q, J = 5.7 Hz, 1H),2.92–2.78 (m, 2H), 2.31 (s, 3H); 13 C NMR (75 MHz, CDCl₃) δ 156.7, 131.0, 130.6, 130.0, 128.9, 124.3, 72.8, 65.8, 60.3, 31.2, 16.1; IR (KBr): ν_{max} 3382, 2942, 1246, 753 cm⁻¹; HRMS (ESI): Calcd for $C_{11}H_{16}O_3Na [M + Na]^+$: 219.0991, found: 219.0989.

To a solution of diol (29.2 g, 148 mmol) in acetone–water (4:1, 400 mL) was added sodium periodate (63.6 g, 297 mmol). The reaction mixture was stirred at 0 °C for 1 h. Upon completion of the reaction, it was filtered on celite and the filtrate was concentrated *in vacuo* to remove acetone. The aqueous phase was extracted with ethyl acetate (3 × 150 mL), washed with brine (100 mL), dried over anhydrous Na₂SO₄, fil-

tered and concentrated *in vacuo*. The crude was purified by flash column chromatography (9 : 1 hexanes–EtOAc) to afford 7 (21.6 g, 88%) as a liquid. ¹H NMR (300 MHz, CDCl₃) δ 9. 9.72 (t, J = 2.3 Hz, 1H), 7.19–7.12 (m, 1H), 7.02 (d, J = 5.3 Hz, 2H), 3.69 (s, 3H), 3.68 (dd, J = 2.3, 0.8 Hz, 2H), 2.33 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 199.6, 157.1, 131.3, 131.0, 128.9, 125.6, 124.3, 60.0, 45.3, 16.1; IR (KBr): ν_{max} 2937, 1731, 1248, 1124, 752 cm⁻¹; HRMS (ESI): Calcd for C₁₀H₁₃O₂ [M + H]⁺: 165.0910, found: 165.0911.

Ethyl 4-(2-methoxy-3-methylphenyl)butanoate (8). To a solution of 7 (21.4 g, 130 mmol) in benzene (200 mL) was added carbethoxymethylene triphenylphosphine (54.42 g, 156 mmol). The reaction mixture was stirred at rt for 12 h under a nitrogen atmosphere, after which it was concentrated *in vacuo*. The crude was purified by column chromatography (19 : 1 hexanes–EtOAc) to afford the olefinic compound (25.3 g, 83%) as a light yellow liquid. ¹H NMR (300 MHz, CDCl₃) δ 7.18–7.11 (m, 1H), 7.10–7.05 (m, 1H), 6.99–6.96 (m, 2H), 5.79 (dt, *J* = 15.7, 1.5 Hz, 1H), 4.16 (q, *J* = 7.2 Hz, 2H), 3.71 (s, 3H), 3.55 (dd, *J* = 6.6, 1.5 Hz, 2H), 2.30 (s, 3H), 1.26 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 166.5, 156.7, 147.5, 131.2, 130.0, 128.1, 124.1, 122.1, 119.6, 60.5, 60.1, 32.5, 16.1, 14.2; IR (KBr): ν_{max} 2930, 1719, 1654, 1183, 1036, 770 cm⁻¹; HRMS (ESI): Calcd for C₁₄H₁₈O₃Na [M + Na]⁺: 257.1148, found: 257.1145.

To a solution of olefin (25 g, 106 mmol) in methanol (300 mL) was added NiCl₂·6H₂O (25.3 g, 106 mmol). The mixture was cooled to 0 °C and NaBH₄ (8 g, 213 mmol) was added portion-wise. Upon completion of the reaction, as indicated by TLC, it was filtered on celite and concentrated in vacuo. The crude mixture was quenched with aq. saturated NH₄Cl (40 mL). The aqueous phase was extracted with ethyl acetate (2 \times 150 mL), washed with brine (30 mL), dried over anhydrous Na2SO4, filtered and concentrated in vacuo. The crude was purified by column chromatography (19:1 hexanes-EtOAc) to afford 8 (21.9 g, 87%) as a colorless liquid. ¹H NMR (300 MHz, CDCl₃) δ 7.05-7.0 (m, 2H), 6.99-6.92 (m, 1H), 4.13 (q, J = 6.8 Hz, 2H), 3.72 (s, 3H), 2.67 (t, J = 7.5 Hz, 2H), 2.35 (t, J = 6.8, 2H), 2.29 (s, 3H), 1.95 (quin., J = 7.5 Hz, 2H), 1.26 (t, J = 6.8 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 173.5, 156.7, 134.2, 130.9, 129.2, 127.7, 123.9, 60.3, 60.1, 33.9, 29.1, 25.9, 16.1, 14.2; IR (KBr): ν_{max} 2945, 1734, 1245, 1017, 753 cm⁻¹; HRMS (ESI): Calcd for $C_{14}H_{20}O_3Na [M + Na]^+$: 259.1304, found 259.1295.

4-(2-Methoxy-3-methylphenyl)butanoic acid (9). To a solution of 8 (20.8 g, 87.89 mmol) in THF–H₂O (1 : 1) (200 mL) was added LiOH·H₂O (18.4 g, 439 mmol). The reaction mixture was stirred at room temperature for 12 h. The reaction mixture was concentrated *in vacuo* to remove THF. Organic impurities were extracted with ethyl acetate (1 × 100 mL). The aqueous phase was cooled to 0 °C and the pH was adjusted to 2 by the addition of 2N aq. HCl. The aqueous phase was extracted with ethyl acetate (3 × 100 mL), washed with brine (50 mL), dried over anhydrous Na₂SO₄, filtered and concentrated *in vacuo* to afford 9 (15.8 g, 86%) as a colorless liquid. ¹H NMR (300 MHz, CDCl₃) δ 7.06–7.0 (m, 2H), 6.99–6.92 (m, 1H), 3.72 (s, 3H), 2.69 (t, *J* = 7.5 Hz, 2H), 2.41 (t, *J* = 7.4 2H), 2.29 (s, 3H), 1.96 (quin.

J = 7.5 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 179.9, 156.8, 134.0, 131.0, 129.3, 127.8, 123.9, 60.3, 33.6, 29.1, 25.5, 16.2; IR (KBr): ν_{max} 2942, 2673, 1710, 1244, 1013, 753 cm⁻¹; MS (ESI): m/z 207 [M - H]⁺.

5-Methoxy-6-methyl-3,4-dihydronaphthalen-1-(2H)-one (4). Compound 9 (15.7 g, 75.41 mmol) was subjected to an intramolecular Friedel-Crafts acylation with PPA (50.96 g, 150 mmol) at 90 °C for 3 min, followed by an second addition of hot PPA (38 g, 113 mmol), and the resultant mixture was stirred for 30 min. Ice (100 mL) was then placed into the reaction flask. The flask was allowed to cool and the mixture was extracted with ethyl acetate (3 × 100 mL). The organic layer was washed with 5% NaOH (2 × 100 mL), H_2O (2 × 100 mL), 3% CH₃COOH (1 × 100 mL), 5% NaHCO₃ (1 × 100 mL) and brine $(1 \times 100 \text{ mL})$. The organic layer was dried over anhydrous Na₂SO₄, filtered and concentrated. The crude was purified by column chromatography (9:1 hexanes-EtOAc) to afford 4 (11.3 g, 79%) as an orange liquid. ¹H NMR (300 MHz, CDCl₃) δ 7.75 (d, J = 7.9 Hz, 1H), 7.14 (d, J = 7.9 Hz, 1H), 3.75 (s, 3H), 2.97 (t, J = 6.0, 2H), 2.62 (t, J = 6.2 Hz, 2H), 2.34 (s, 3H), 2.11 (quin., J = 6.2 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 198.0, 155.7, 137.5, 137.1, 132.2, 129.0, 122.8, 59.8, 38.7, 23.4, 22.8, 16.5; IR (KBr): ν_{max} 2943, 1683, 1283, 1018, 767 cm⁻¹; HRMS (ESI): Calcd for $C_{12}H_{15}O_2 [M + H]^+$: 191.1066, found 191.1065.

5-Methoxy-6-methyl-1,2,3,4-tetrahydronaphthalen-1-ol (10). To a solution of 4 (11.12 g, 58.46 mmol) in CH₃OH (100 mL) was added NaBH₄ (2.65 g, 70.16 mmol) portion-wise at 0 °C. The reaction was allowed to stir for 30 min at the same temperature. After completion of the reaction, as indicated by TLC, the reaction was quenched with aq. saturated NH₄Cl solution (20 mL). The reaction mixture was concentrated under reduced pressure and aq. saturated NH₄Cl (50 mL) was added. The aqueous phase was extracted with ethyl acetate $(3 \times 60 \text{ mL})$, washed with brine (50 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The crude was purified by column chromatography (6:1 hexanes-EtOAc) to afford 10 (10.2 g, 90%) as a yellow oil. ¹H NMR (300 MHz, $CDCl_3$) δ 7.12 (d, J = 8.3 Hz, 1H), 7.04 (d, J = 7.5 Hz, 1H), 4.77-4.72 (m, 1H),3.71 (s, 3H), 2.86 (dt, J = 18.9, 5.3 Hz, 1H), 2.64 (dt, J = 17.4, 6.0 Hz, 1H), 2.27 (s, 3H), 1.98-1.85 (m, 2H), 1.84-1.72 (m, 2H); $^{13}\mathrm{C}$ NMR (75 MHz, CDCl_3) δ 155.9, 138.1, 130.4, 129.8, 128.7, 124.2, 67.9, 59.4, 31.9, 23.4, 18.3, 15.9; IR (KBr): v_{max} 3382, 2937, 1453, 1015, 820 cm⁻¹; MS (ESI): m/z 215 [M + Na]⁺.

8-Methoxy-7-methyl-1,2-dihydronaphthalene (11). Compound 10 (10.0 g, 52.18 mmol) and PPA (88 g, 260 mmol) were stirred at 100 °C for 1.5 h. After cooling to room temperature, the mixture was diluted with water and basified with ammonium hydroxide (200 mL). The aqueous phase was extracted with chloroform (3 × 100 mL), washed with brine (100 mL), dried over anhydrous Na₂SO₄, filtered and concentrated *in vacuo*. The crude was purified by column chromatography (99 : 1 hexanes–EtOAc) to afford **11** (6.9 g, 76%) as a yellow oil. ¹H NMR (300 MHz, CDCl₃) δ 6.96 (d, *J* = 7.5 Hz, 1H), 6.73 (d, *J* = 7.5 1H), 6.43 (dt, *J* = 9.8, 1.5 Hz, 1H), 6.02–5.94 (m, 1H), 3.70 (s, 3H), 2.83 (t, *J* = 8.3 Hz, 2H), 2.33–2.24 (m, 5H); ¹³C NMR (75 MHz, CDCl₃) δ 155.4, 133.5, 129.9, 128.4, 127.7,

127.6, 121.8, 59.9, 22.8, 20.5, 16.1; IR (KBr): ν 2934, 1566, 1086, 826 cm $^{-1};$ MS (ESI): m/z 197 $[\rm M + Na]^+.$

2-Methyl-7,8-dihydronaphthalen-1-ol (12). A solution of 11 (6.8 g, 39.21 mmol) in dry CH₂Cl₂ (30 mL) was cooled to 0 °C and BCl3 (1 M, 78.4 mL, 78.41 mmol) was added to it dropwise. The reaction mixture was warmed to rt and heated to reflux temperature. The reaction was quenched with water (50 mL) and the organic layer was separated. The aqueous phase was extracted with CH_2Cl_2 (2 × 50 mL). The combined organic extracts were washed with brine (30 mL), dried over anhydrous Na2SO4, filtered and concentrated in vacuo. The crude was purified by column chromatography (19:1 hexanes-EtOAc) to afford 12 as a red oil (4.8 g, 77%). ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 6.91 \text{ (d}, J = 7.4 \text{ Hz}, 1\text{H}), 6.58 \text{ (d}, J = 7.5 \text{ Hz},$ 1H), 6.41 (dt, J = 9.6, 1.9 Hz, 1H), 6.0-5.91 (m, 1H), 4.64 (s, 1H), 2.74 (t, J = 8.3 Hz, 2H), 2.37–2.28 (m, 2H), 2.23 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 150.4, 133.2, 127.9, 127.7, 127.1, 122.6, 119.9, 118.7, 22.6, 19.9, 15.9; IR (KBr): ν_{max} 3469, 2928, 1658, 823 cm⁻¹; HRMS (ESI): Calcd for $C_{11}H_{13}O [M + H]^+$: 161.0960, found: 161.0963.

(E)-Methyl 3-(2-methyl-7,8-dihydronaphthalen-1-yl)acrylate (13). To a solution of 12 (4.7 g, 29.4 mmol) in dry CH_2Cl_2 (50 mL) was added NEt₃ (12 mL, 88.2 mmol), followed by a slow addition of triflic anhydride (5.9 mL, 35.28 mmol) at 0 °C. The mixture was stirred at same temperature for 1 h. The reaction was quenched with a slow addition of aq. 1 N HCl (30 mL). The organic layer was separated, and the aqueous phase was extracted with CH_2Cl_2 (2 × 40 mL). The combined organic extracts were washed with brine (20 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The crude was purified by column chromatography (hexanes) to afford triflate (6.9 g, 81%) as a light yellow liquid. ¹H NMR (500 MHz, CDCl₃) δ 7.07 (dd, J = 7.6, 0.6 Hz, 1H), 6.94 (d, J = 7.6, 1H), 6.45 (dt, J = 9.6, 1.8 Hz, 1H), 6.09-6.05 (m, 1H), 2.87 $(t, J = 8.3 \text{ Hz}, 2\text{H}), 2.36 (s, 3\text{H}), 2.33-2.28 (m, 2\text{H}); {}^{13}\text{C} \text{ NMR}$ (75 MHz, CDCl₃) δ 145.6, 134.6, 129.8, 129.6, 129.1, 128.5, 126.8, 125.6, 22.2, 21.7, 17.0; IR (KBr): v 2930, 1556, 1410, 1215, 771 cm⁻¹; MS (ESI): m/z 293 [M + H]⁺.

PdCl₂(PPh₃)₂ (31.9 mg, 2 mol%) was added to a degassed solution of triflate (6.8 g, 23.27 mmol), methyl acrylate (10.5 mL, 116 mmol) and NEt₃ (9.8 mL, 69.81 mmol) in DMF (50 mL). The reaction mixture was stirred at 120 °C under an argon atmosphere for 48 h. Ice water (50 mL) was then added to it. The aqueous phase was extracted with ethyl acetate (3 \times 50 mL). The combined organic extracts were washed with brine (50 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The crude was purified by column chromatography (29:1 hexanes-EtOAc) to afford 13 as a red oil (3.9 g, 75%). ¹H NMR (500 MHz, CDCl₃) δ 7.84 (d, J = 16.5 Hz, 1H), 7.03 (d, J = 7.6 Hz, 1H), 6.93 (d, J = 7.6 Hz, 1H), 6.44 (dt, J = 9.5, 1.7 Hz, 1H), 6.03–5.99 (m, 2H), 3.82 (s, 3H), 2.81 (t, J = 8.2 Hz, 2H), 2.31 (s, 3H), 2.29–2.24 (m, 2H); ¹³C NMR (75 MHz, $CDCl_3$) δ 166.9, 143.4, 135.3, 133.5, 133.0, 132.3, 127.9, 127.8, 127.7, 126.3, 124.3, 51.7, 24.9, 23.1, 20.8; IR (KBr): ν_{max} 2949, 1722, 1641, 1169, 772 cm⁻¹; HRMS (ESI): Calcd for C₁₅H₁₇O₂ 229.1223 [M + H]⁺, found: 229.1224.

Methyl 3-(2-methyl-7,8-dihydronaphthalen-1-yl)propanoate (3). To a solution of 13 (3.75 g, 16.43 mmol) in CH₃OH (40 mL) were added Mg turnings (788 mg, 32.87 mmol). The reaction mixture was heated at reflux temperature for 12 h. After completion of the reaction, as indicated by TLC, the solvent was removed under reduced pressure. The reaction mixture was partitioned between aq. saturated NH₄Cl and ethyl acetate. The aqueous phase was extracted with ethyl acetate (2 \times 40 mL). The combined organic extracts were washed with brine (20 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The crude was purified by column chromatography (29:1 hexanes-EtOAc) to afford 3 (3.17 g, 84%) as a liquid. ¹H NMR (500 MHz, CDCl₃) δ 6.97 (d, J = 7.5 Hz, 1H), 6.83 (d, J = 7.6 Hz, 1H), 6.42 (dt, J = 9.5, 1.8 Hz, 1H), 6.0-5.96 (m, 1H), 3.71 (s, 3H), 3.01-2.96 (m, 2H), 2.79 (t, J = 8.2 Hz, 2H), 2.47–2.42 (m, 2H), 2.33–2.28 (m, 5H); ¹³C NMR (75 MHz, CDCl₃) δ 173.5, 135.7, 135.2, 133.4, 132.4, 128.2, 128.1, 127.2, 124.5, 51.7, 33.6, 24.4, 23.7, 23.3, 19.9; IR (KBr): $\nu_{\rm max}$ 2926, 1738, 1219, 772 cm⁻¹; HRMS (ESI): Calcd for $C_{15}H_{18}O_2Na [M + Na]^+: 253.1199$, found: 253.1204.

Methyl 3-(5,6-dihydroxy-2-methyl-5,6,7,8-tetrahydro naphthalen-1-yl)propanoate (14). To a solution of 3 (3.1 g, 13.46 mmol) in acetone-acetonitrile-water (1:1:1, 30 mL) were added OsO4 (33.7 mg, 0.1 mmol) and NMO (3.15 g, 26.92 mmol). The reaction was stirred at room temperature for 12 h. The mixture was quenched with aq. sodium sulphite solution and stirred for an additional 30 min. The solution was concentrated in vacuo to remove organic solvents and 30 mL water was added. The aqueous phase was extracted with ethyl acetate (3×30 mL). The combined organic extracts were washed with brine (30 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The crude was purified by column chromatography (1:1 hexanes-EtOAc) to afford 14 (2.5 g, 78%) as a colorless liquid. ¹H NMR (300 MHz, CDCl₃) δ 7.24 (d, J = 7.5 Hz, 1H), 7.08 (d, J = 8.3 Hz, 1H), 4.67 (d, J = 3.8 Hz, 1H), 4.05-3.95 (m, 1H), 3.72 (s, 3H), 3.04-2.91 (m, 3H), 2.78-2.63 (m, 1H), 2.48-2.39 (m, 2H), 2.32 (s, 3H), 2.14-1.90 (m, 2H); 13 C NMR (75 MHz, CDCl₃) δ 173.4, 136.5, 136.3, 134.6, 134.1, 128.8, 128.2, 70.3, 69.0, 51.7, 32.9, 26.0, 24.6, 24.3, 19.6; IR (KBr): ν_{max} 3394, 2933, 1735, 1291, 1198, 773 cm⁻¹; HRMS (ESI): Calcd for C₁₅H₂₀O₄Na: 287.1253 $[M + Na]^+$, found 287.1252.

Methyl 3-(7-methyl-2-oxo-3a,4,5,9b-tetrahydronaphtho[2,1-*d*] [1,3]dioxol-6-yl)propanoate (15). To a solution of 14 (2.35 g, 8.89 mmol) in dry CH₂Cl₂ (20 mL) was added NEt₃ (3.7 mL, 26.67 mmol). The solution was cooled to 0 °C and triphosgene (2.64 g, 8.89 mmol) was added slowly in portions. The reaction was stirred at rt for 1 h and quenched with water (20 mL). The aqueous phase was extracted with ethyl acetate (3 × 20 mL). The combined organic extracts were washed with brine (20 mL), dried over anhydrous Na₂SO₄, filtered and concentrated *in vacuo*. The crude was purified by column chromatography (6:1 hexanes–EtOAc) to afford 15 (2.17 g, 84%) as a light yellow oil. ¹H NMR (300 MHz, CDCl₃) δ 7.19 (d, *J* = 7.9 Hz, 1H), 7.12 (d, *J* = 7.7 Hz, 1H), 5.68 (d, *J* = 7.9 Hz, 1H), 5.20–5.12 (m, 1H), 3.71 (s, 3H), 3.06–2.97 (m, 2H), 2.85–2.78 (m, 2H), 2.47–2.27 (m, 6H), 1.98–1.85 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 172.9, 154.7, 138.3, 136.7, 136.5, 129.2, 129.1, 127.5, 76.3, 74.9, 51.8, 33.3, 27.2, 24.4, 20.0, 19.9; IR (KBr): $\nu_{\rm max}$ 2924, 1796, 1733, 1168, 772 cm⁻¹; HRMS (ESI): Calcd for C₁₆H₁₈O₅Na 313.1046 [M + Na]⁺, found 313.1043.

Methyl 3-(6-hydroxy-2-methyl-5,6,7,8-tetrahydronaphthalen-1-yl)propanoate (16). To a solution of 15 (2.1 g, 7.23 mmol) in CH₃OH (20 mL) was added Pd/C (45.65 mg, 10% w/w) in sulfur-free conditions. The reaction mixture was stirred under a hydrogen atmosphere at rt for 3 h. After completion, the reaction mixture was filtered through celite and the solvent was evaporated under vacuum. The crude was purified by column chromatography (6:1 hexanes-EtOAc) to afford 16 (1.59 g, 88%) as a liquid. ¹H NMR (300 MHz, CDCl₃) δ 6.97 (d, J = 7.7 Hz, 1H), 6.88 (d, J = 7.7 Hz, 1H), 4.18–4.08 (m, 1H), 3.72 (s, 3H), 3.11-2.90 (m, 4H), 2.84-2.70 (m, 2H), 2.48-2.39 (m, 2H), 2.30 (s, 3H), 2.15–2.03 (m, 1H), 1.90–1.76 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 173.5, 136.6, 133.9, 133.6, 132.3, 128.2, 127.8, 66.7, 51.7, 38.8, 33.0, 31.6, 24.5, 24.3, 19.4; IR (KBr): $\nu_{\rm max}$ 3406, 2926, 1736, 1219, 772 cm⁻¹; HRMS (ESI): Calcd for $C_{15}H_{20}O_3Na 271.1304 [M + Na]^+$, found 271.1302.

Methyl 3-(6-(4-chlorophenylsulfonamido)-2-methyl-5,6,7,8tetrahydronaphthalen-1-yl)propanoate 3-(6-(4-chlorophenyl sulfonamido)-2-methyl-5,6,7,8-tetrahydronaphthalen-1-yl) propanoate (17). Alcohol 16 (1.42 g, 5.72 mmol) and triphenylphosphine (2.24 g, 8.58 mmol) were dissolved in dry THF (20 mL). The mixture was cooled to -15 °C, and diisopropyl azodicarboxylate (DIAD, 2.84 mL, 14.29 mmol) was added to it. After stirring the solution for 10 min at -15 °C, the temperature was raised to 0 °C and diphenylphosphoryl azide (DPPA, 1.79 mL, 8.58 mmol) was added. The solution was stirred for 30 min at 0 °C and then for 12 h at room temperature. Concentration and gradient flash chromatography (99:1 hexanes-EtOAc) gave the azide (1.3 g, 83%) as a sticky yellow liquid, which was directly used for the next step.

To a solution of azide (1.3 g, 4.76 mmol) in EtOAc (15 mL) was added Pd/C (6.7 mg, 10% w/w) in sulfur-free conditions followed by 4-chlorobenzene-1-sulfonyl chloride (1.99 g, 9.51 mmol). The reaction mixture was stirred under a hydrogen atmosphere at rt for 12 h. After completion, the reaction mixture was filtered through celite and the solvent was evaporated under vacuum. The crude was purified by column chromatography (4:1 hexanes-EtOAc) to afford 17 (1.56 g, 78%) as a yellow oil. ¹H NMR (300 MHz, $CDCl_3$) δ 7.82 (d, *J* = 8.5 Hz, 2H), 7.48 (d, J = 8.5 Hz, 2H), 6.94 (d, J = 7.7 Hz, 1H), 6.75 (d, J = 7.7, 1H), 4.76 (d, J = 7.7 Hz, 1H), 3.71 (s, 3H), 3.68-3.57 (m, 1H), 2.97-2.53 (m, 6H), 2.43-2.35 (m, 2H), 2.29 (s, 3H), 2.03-1.91 (m, 1H), 1.83–1.69 (m, 1H); 13 C NMR (75 MHz, CDCl₃) δ 173.3, 139.6, 139.1, 136.9, 134.5, 132.9, 131.1, 129.4, 128.5, 128.4, 127.7, 51.8, 49.1, 36.8, 32.9, 29.6, 24.4, 24.0, 19.4; IR (KBr): $\nu_{\rm max}$ 2923, 1735, 1162, 772 cm⁻¹; HRMS (ESI): Calcd for $C_{21}H_{25}ClNO_4S$ 422.1187 [M + H]⁺, found 422.1188.

3-(6-(4-Chlorophenylsulfonamido)-2-methyl-5,6,7,8-tetrahydronaphthalen-1-yl)propanoic acid (2). Compound **17** (1.43 g, 3.39 mmol) was brought to reflux in methanol (15 mL), in the presence of 2 N sodium hydroxide (404 mg, 10.17 mmol) for

1 h. After cooling, the solvent was evaporated in vacuo, the mixture was washed with ethyl acetate (1 mL) and the aqueous phase was acidified with 1N HCl. The aqueous phase was extracted with ethyl acetate $(3 \times 10 \text{ mL})$. The combined organic extracts were dried over anhydrous Na2SO4, filtered and concentrated in vacuo to give terutroban (2) (1.12 g, 82%) as a white solid. ¹H NMR (300 MHz, DMSO-d₆) δ 7.91 (d, *J* = 6.6 Hz, 1H), 7.84 (d, J = 8.5 Hz, 2H), 7.66 (d, J = 8.5 Hz, 2H), 6.87 (d, J = 7.7 Hz, 1H), 6.69 (d, J = 7.7 Hz, 1H), 3.31(m, 1H), 2.83–2.65 (m, 4H), 2.63-2.54 (m, 2H), 2.30-2.21 (m, 2H), 2.19 (s, 3H), 1.86-1.74 (m, 1H), 1.63-1.50 (m, 1H); ¹³C NMR (75 MHz, DMSO-d₆) δ 174.2, 140.7, 137.3, 136.9, 133.5, 133.3, 131.9, 129.5, 128.5, 127.9, 127.1, 49.0, 36.4, 32.9, 29.5, 24.3, 24.2, 19.2; IR (KBr): ν_{max} 2924, 1709, 1219, 772 cm⁻¹; HRMS (ESI): Calcd for C₂₀H₂₃O₄NClS 408.1030 [M + H]⁺, found 408.1040. [Reported ¹H NMR^{3a} (DMSO-d₆) δ 12.5 (s, 1H), 7.9 (s, 1H), 7.8 (d, 2H), 7.7 (d, 2H), 6.9-6.7 (d, 2H), 3.3 (m, 1H), 3.0-2.5 (m, 6H), 2.3 (m, 2H), 2.2 (s, 3H), 2.0-1.5 (m, 2H).]

Conclusions

In conclusion, the total synthesis of terutroban is accomplished using a non-Diels–Alder approach. The Claisen rearrangement, Friedel–Crafts acylation and Heck coupling reaction are the key reactions utilized. The utility of costeffective chemicals together with the flexible route allows one to visualise the design of analogues with ease.

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Notes and references

- 1 L. A. Sorbera, N. Serradell, J. Bolos and M. Mayes, *Drugs Future*, 2006, **31**, 867.
- 2 (a) T. Ishizuka, T. Matsui, Y. Okamoto, A. Ohta and M. Shichijo, *Cardiovasc. Drug Rev.*, 2004, 22, 71;
 (b) E. Busto, V. Gotor-Fernandez and V. Gotor, *J. Org. Chem.*, 2012, 77, 4842.
- 3 (a) B. Cimetière, T. Dubuffet, O. Muller, J.-J. Descombes,
 S. Simonet, M. Laubie, T. J. Verbeuren and G. Lavielle, *Bioorg. Med. Chem. Lett.*, 1998, 8, 1375; (b) G. Lavielle,
 T. Dubuff, O. Muller, M. Laubie, T. Verbeuren, S. Simonet and J.-J. Descombes, 1,2,3,4-tetrahydronaphthalene compounds, US Pat, US005472979A, 1995, Dec. 5, 1995.
- 4 N. K. Reddy, B. V. D. Vijaykumar and S. Chandrasekhar, *Org. Lett.*, 2012, **14**, 299.
- 5 S. Chandrasekhar, C. Sridhar and P. Srihari, *Tetrahedron: Asymmetry*, 2012, 23, 388.

- 6 (a) K. C. Nicolaou, S. A. Snyder, T. Montagnon and G. Vassilkogiannakis, *Angew. Chem., Int. Ed.*, 2002, 41, 1668; (b) M. Juhl and D. Tanner, *Chem. Soc. Rev.*, 2009, 38, 2983; (c) P. Merino, E. Marques-Lopez, T. Tejero and R. P. Herrera, *Synthesis*, 2010, 0001.
- 7 (a) C. S. P. Rao and K. Srimannarayana, Synth. Commun., 1991, 21, 1455; (b) EP 1481959 (c) M.-Y. Zhou and Y.-Q. Li, J. Chem. Soc., Perkin Trans. 2, 2001, 1824; (d) J. Borgulya, R. Madeja, P. Fahrni, H. J. Hansen, H. Schmid and R. Barner, Helv. Chim. Acta, 1973, 56, 14; (e) D. McHale and J. Green, J. Chem. Soc., 1965, 5060; (f) A. Srikrishna, T. J. Reddy, P. P. Kumar and S. J. Gharpure, Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem., 2001, 40, 905; (g) Z. M. Wang, X. L. Zhang and K. B. Sharpless, Tetrahedron Lett., 1993, 34, 2267; (h) E. Taskinen, J. Chem. Soc., Perkin Trans. 2, 2001, 1824; (i) S. Katkevica, A. Zicmanis and P. Mekss, Chem. Heterocycl. Compd., 2010, 46, 158; (j) A. K. Mitra, A. De and N. Karchaudhuri, Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem., 2000, 39, 387.
- 8 (a) H. O. House and R. J. McCaully, J. Org. Chem., 1959, 24, 725; (b) A. Heesing and W. Muellers, Chem. Ber., 1980, 113, 9; (c) J. R. Grunwell, M. F. Wempe, J. Mitchell and J. R. Grunell, Tetrahedron Lett., 1993, 74, 7163; (d) C. Wright and G. V. Ullas, J. Radiolabel. Comp. Pharm., 2002, 45, 1265.
- 9 G. Liang, Y. Xu, I. B. Seiple and D. Trauner, J. Am. Chem. Soc., 2006, 128, 11022.
- 10 (a) C. Bernard, *Platinum Met. Rev.*, 2008, 52, 38;
 (b) P. Prediger, Y. Genisson and C. R. D. Correia, *Curr. Org. Chem.*, 2013, 17, 238; (c) D. MacCartney and P. J. Guiry, *Chem. Soc. Rev.*, 2011, 40, 5122.
- (a) T. Ando, D. Kano, S. Minakata, I. Ryu and M. Komatsu, *Tetrahedron*, 1998, 54, 13485; (b) S. Minakata, Y. Morino, Y. Oderaotoshi and M. Komatsu, *Chem. Commun.*, 2006, 31, 3337; (c) S. Minakata, D. Kano, R. Fukuoka, Y. Oderaotoshi and M. Komatsu, *Synthesis*, 2003, 289.

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(a) M. Imuta and H. Ziffer, J. Org. Chem., 1979, 44, 1351;
(b) K. Bhandari, V. L. Sharma, C. M. Singh, G. Shankar and H. K. Singh, Ind. J. Chem, Sec B: Org. Chem. Incl. Med. Chem., 2000, 39, 468; (c) Y. Shimada, S. Kondo, Y. Ohara, K. Matsumoto and T. Katsuki, Synlett, 2007, 2445;
(d) R. W. Draper, D. H. Radha Iyer, G. M. Lee, J. T. Liang, J. L. Mas, W. Tormos, E. J. Vater, F. Günter, I. Mergelsberg and D. Scherer, Org. Process Res. Dev., 1998, 2, 175.

- 13 (a) K. Matsumoto, S. Fuwa and H. Kitajima, *Tetrahedron Lett.*, 1995, 36, 6499; (b) M. Shimojo, K. Matsumoto and M. Hatanaka, *Tetrahedron*, 2000, 56, 9281; (c) K. Matsumoto, S. Fuwa, M. Shimojo and H. Kitajima, *Bull. Chem. Soc. Jpn.*, 1996, 69, 2977; (d) K. Matsumoto, M. Shimojo and M. Hatanaka, *Chem. Lett.*, 1997, 1151; (e) J. S. Yadav, M. A. Rahman, N. M. Reddy, A. R. Prasad and A. A. K. A. Ghamdi, *Synlett*, 2007, 661.
- 14 B. M. Trost, B. Vidal and M. Thommen, *Chem. Eur. J.*, 1999, **5**, 1055.
- 15 (*a*) M. Baeck, J. Nyhlen, I. Kvarnstroem, S. Appelgren, N. Borkakoti, K. Jansson, J. Lindberg, S. Nystroem,

A. Hallberg, A. Rosenquist and B. Samuelsson, *Bioorg. Med. Chem.*, 2008, 16, 9471; (b) C. Bjoerklund, S. Oscarson,
K. Benkestock, N. Borkakoti, K. Jansson, J. Lindberg,
L. Vrang, A. Hallberg, A. Rosenquist and B. Samuelsson, *J. Med. Chem.*, 2010, 53, 1458; (c) A. E. Gould,
S. J. Harrison, H. Mizutani, M. Shen, T. E. Smyser and
S. G. Stroud, US2010/197924, 2010; (d) V. Sandgren,
T. Agback, P. O. Johansson, J. Lindberg, I. Kvarnstroem,
B. Samuelsson, O. Belda and A. Dahlgren, *Bioorg. Med. Chem.*, 2012, 20, 4377.

16 E. J. Corey and J. O. Link, J. Am. Chem. Soc., 1992, 114, 1906.