

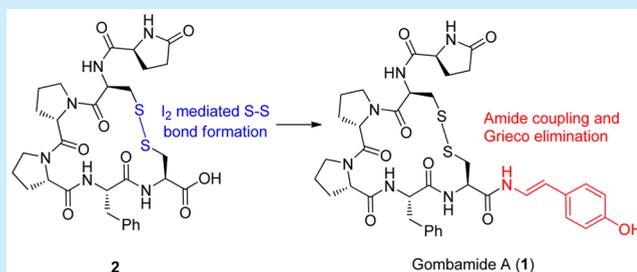
Divergent Approach for the Synthesis of Gombamide A and Derivatives

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S Supporting Information

ABSTRACT: A synthesis of gombamide A (**1**) using N-terminal peptide extension, oxidative disulfide bond formation, and late-stage 4-hydroxystyrylamide installation has been achieved. This divergent method was also utilized to synthesize several gombamide A derivatives with modification to the 4-hydroxystyrylamide via cyclic peptide **2**. The natural product and four derivatives were found to be devoid of Na⁺/K⁺-ATPase activity at 10 μM. In addition, the compounds were not cytotoxic at 10 μM against a panel of cancer cells.



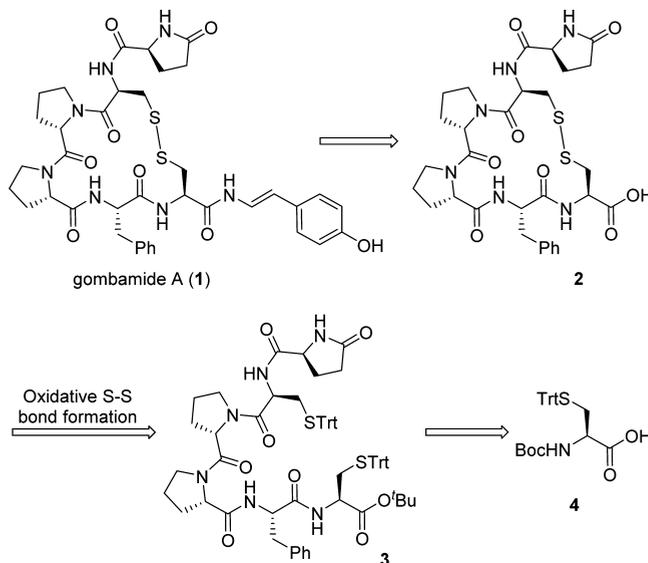
Disulfide-containing natural products possess an array of biological activities.¹ In addition, disulfide-containing cyclic peptides constitute a broad structural motif found in natural products,^{1a,2a} endogenous compounds (e.g., oxytocin and calcitonin), and pharmacological agents (e.g., linaclotide and eptifibatid). The disulfide functional group provides structural rigidity that can enhance binding affinity to biomolecular targets and potentially improve oral bioavailability.^{2b} Our laboratory has become interested in synthesizing and evaluating disulfide-containing cyclic peptide natural products and derivatives as potential pharmacological probes. For example, we recently reported the synthesis and antiproliferative activity evaluation of thiochondrilline C and several derivatives.³

Gombamide A (**1**) is a disulfide-containing cyclic peptide recently isolated from the marine sponge *Calthria gombawuensis*.⁴ In addition, one of the cysteine residues that comprise the disulfide is linked to pyroglutamic acid while the other is attached to a styrylamide moiety, an uncommon functional group among isolated natural products.⁵ Gombamide A was also reported to have moderate Na⁺/K⁺-ATPase inhibitory activity and weak cytotoxicity against human A549 nonsmall cell lung cancer and K562 leukemia cell lines.⁴

Gombamide A has attracted interest from the organic chemistry community with a synthesis using a convergent approach recently reported by Garcia-Barrantes and Lindsley.⁶ Herein, we describe a complementary synthesis of gombamide A and several derivatives that utilizes a divergent strategy that was exploited to generate several derivatives with modifications to the 4-hydroxystyrylamide. In addition, we report the evaluation of these compounds for Na⁺/K⁺-ATPase and antiproliferative activities.

A retrosynthetic analysis of **1** is outlined in Scheme 1. This divergent approach introduces the 4-hydroxystyrylamide or other amides late in the synthesis via carboxylic acid **2**,

Scheme 1. Retrosynthetic Analysis of Gombamide A (1)

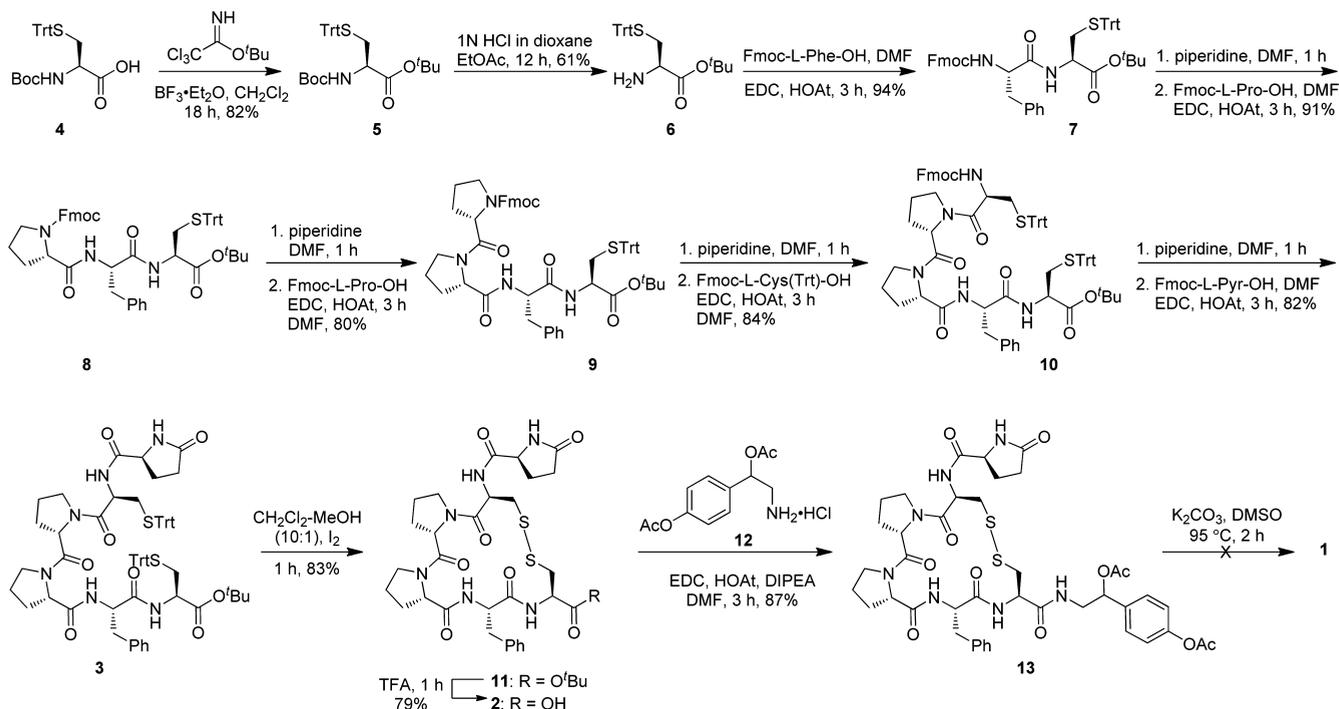


providing flexibility for modification of this exocyclic portion of the molecule. Cyclic intermediate **2** would be prepared by oxidative disulfide bond formation of **3**, which would be generated by an N-terminal peptide extension starting from **4**.

The synthesis of gombamide A and several derivatives began with esterification of N-Boc-S-tritylcysteine, **4**, in the presence of *tert*-butyl 2,2,2-trichloroacetimidate to furnish the corresponding *tert*-butyl ester **5** in 82% yield (Scheme 2). Selective removal of the Boc in the presence of the trityl group was accomplished with 1 N HCl to provide **6** in 61% yield.⁷ Amine

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Scheme 2. Synthesis of Disulfide-Containing Cyclic Peptide Intermediate 2



6 was coupled with Fmoc-L-Phe-OH using EDC and HOAt to provide dipeptide 7 in 94% yield. The dipeptide was treated with piperidine to remove the Fmoc protecting group followed by coupling with Fmoc-L-Pro-OH in the presence of EDC and HOAt to generate tripeptide 8 in 91% yield. The tripeptide was again treated with piperidine to remove the Fmoc protecting group followed by coupling with Fmoc-L-Pro-OH using EDC and HOAt to generate the tetrapeptide 9 in 80% yield. Fmoc deprotection of the tetrapeptide with piperidine followed by coupling with Fmoc-L-Cys(Trt)-OH using EDC and HOAt gave pentapeptide 10 in 84% yield. This material was treated with piperidine and then coupled with Fmoc-L-Pyr-OH using EDC and HOAt to generate 3 in 82% yield. The hexapeptide was then exposed to iodine to mediate oxidative disulfide bond formation that provided the cyclic peptide 11 in 83% yield.^{3,8} Deprotection of the *tert*-butyl ester was carried out with trifluoroacetic acid to generate the corresponding carboxylic acid 2 in 79% yield.

Our initial plan for generating the 4-hydroxystyrylamide was via an intermolecular copper-mediated coupling of *p*-(*tert*-butyldimethylsilyloxy)- β -bromostyrene with the corresponding primary amide of 2^{9a,b} since intramolecular versions of this type of process have been applied to the synthesis of cyclopeptide alkaloids.^{9c-f} However, a model reaction of this styrene derivative with *N*-acetyl-DL-phenylalaninamide was not successful.

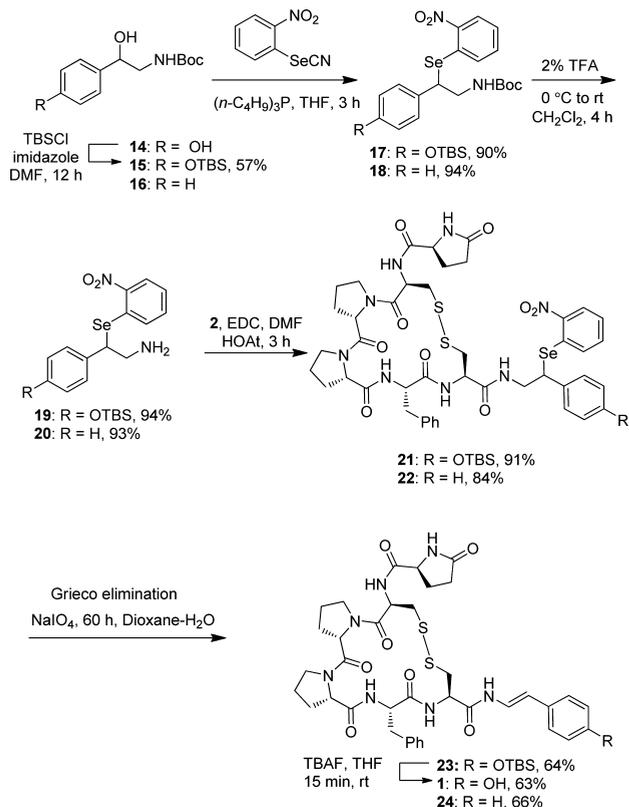
An alternative strategy for introducing the 4-hydroxystyrylamide that we selected was elimination of the corresponding β -acetoxyphenethylamide.¹⁰ In pursuit of this approach, 12 was prepared from (\pm)-octopamine hydrochloride via *N*-Boc protection followed by treatment with acetic anhydride in the presence of pyridine and subsequent removal of the Boc group using 1 N HCl. Coupling of 12 with the model substrate *N*-Boc-Cys(Trt)-OH, followed by treatment with K₂CO₃ in DMF at 95 °C, furnished the corresponding 4-hydroxystyrylamide product. Encouraged by this result, we attempted to employ

this method to introduce 4-hydroxystyrylamide in the synthesis of gombamide A. Coupling of acid 2 with amine 12 using EDC and HOAt generated 13 in 87% yield. However, treatment of 13 with K₂CO₃ in DMF at 95 °C resulted in decomposition.

Due to difficulties encountered with the acetoxy elimination strategy, Grieco's method for elimination of *O*-nitrobenzeneselenenic acid was pursued in order to introduce the styrylamide.^{11,12} *N*-Boc-(\pm)-octopamine 14 was treated with TBSCl in the presence of imidazole to furnish 15 in 57% yield (Scheme 3). Compound 15 was allowed to react with *O*-nitroselenocyanide and tributylphosphine to furnish 17 in 90% yield. This compound was treated with 2% trifluoroacetic acid in DCM to remove the Boc group, generating 19 in 94% yield. Amine 19 was coupled with 2 using EDC and HOAt to generate 21 in 91% yield. Treatment of 21 with NaIO₄ (6 equiv) under standard Grieco elimination conditions resulted in the formation of the *O*-TBS-protected gombamide A derivative 23 along with a byproduct likely due to mono-oxidation at one of the sulfurs (monitored by LCMS).^{11b,12d} In order to enhance formation of the desired product and to suppress unwanted *S*-oxidation, 21 was treated with only 1 equiv of NaIO₄, which resulted in the formation of 23 in 64% isolated yield. Removal of the TBS group was carried out using TBAF (1 equiv) providing gombamide A (1) in 63% yield. The spectral data of the synthetic material are consistent with the previously reported data.^{4,6}

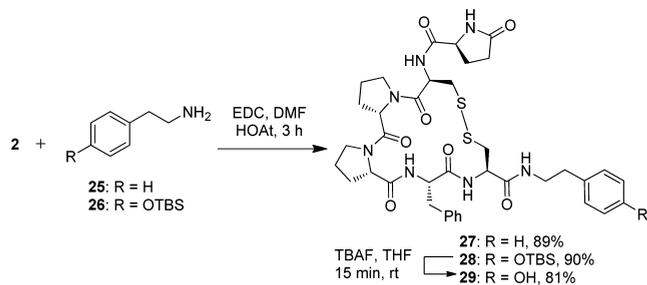
In order to highlight the versatility of this divergent approach, several gombamide A derivatives were prepared using the intermediate carboxylic acid 2. First, another styrylamide derivative was synthesized (Scheme 3). Des-hydroxyl derivative 16 was treated with *O*-nitroselenocyanide and tributylphosphine to furnish 18 in 94% yield. Again, removal of the Boc group using 2% TFA in DCM gave 20 in 93% yield. This material was coupled with 2 using EDC and HOAt to give 22 in 84% yield. Treatment of 22 with NaIO₄ (1 equiv) furnished derivative 24 in 66% yield. Next, two

Scheme 3. Synthesis of Gombamide A (1) and Derivative 24



phenethylamine derivatives were prepared (Scheme 4). Compound 27 was obtained by coupling carboxylic acid 2

Scheme 4. Synthesis of Gombamide A Derivatives 27 and 29



with 25 using EDC and HOAt in 89% yield. Similarly, 2 was coupled with 26 to furnish 28 in 90% yield. Removal of the TBS group with TBAF gave 29 in 81% yield.

Gombamide A (1) and four derivatives (2, 24, 27, and 29) were evaluated for activity against Na^+/K^+ -ATPase at 10 μM in duplicate.¹³ However, all of the compounds were found to be inactive (<2% inhibition). In addition, this set of compounds was profiled for antiproliferative activity utilizing the National Cancer Institute (NCI) 60 cancer cell line assay, which included both A549 and K562 cell lines. All of the compounds were found to be noncytotoxic at 10 μM as determined by their effect on cell growth (mean inhibition of each compound against 60 cell lines was <5%).¹⁴

In summary, a divergent approach was developed for the synthesis of gombamide A providing the natural product in 5.7% overall yield in 16 steps starting from N -Boc-L-Cys(Trt)-OH (4). Two key features of the synthesis were iodine-

mediated oxidative disulfide bond formation to generate the versatile cyclic peptide carboxylic acid 2 and introduction of the 4-hydroxystyrylamide late in the synthesis via a Grieco elimination procedure. The utility of this methodology was further illustrated with the synthesis of several gombamide A derivatives. The natural product and four analogs (2, 24, 27, and 29) were shown to be devoid of Na^+/K^+ -ATPase activity at 10 μM . Finally, these five compounds were not cytotoxic against a panel of cancer cell lines at 10 μM , which should encourage exploration of this chemotype for nononcology pharmacological applications.

■ ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.orglett.6b02379.

Description of the detailed experimental procedures and NMR spectral data (PDF)

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Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) (a) Jiang, C.-S.; Müller, W. E. G.; Schröder, H. C.; Guo, Y.-W. *Chem. Rev.* **2012**, *112*, 2179–2207. (b) Lee, S. H. *Arch. Pharmacol. Res.* **2009**, *32*, 299–315.
- (2) (a) Chung, B. K. W.; Yudin, A. K. *Org. Biomol. Chem.* **2015**, *13*, 8768–8779. (b) Góngora-Benítez, M.; Tulla-Puche, J.; Albericio, F. *Chem. Rev.* **2014**, *114*, 901–926.
- (3) Vippila, M. R.; Ly, P. K.; Cuny, G. D. *J. Nat. Prod.* **2015**, *78*, 2398–2404.
- (4) Woo, J.-K.; Jeon, J.-e.; Kim, C.-K.; Sim, C. J.; Oh, D.-C.; Oh, K.-B.; Shin, J. *J. Nat. Prod.* **2013**, *76*, 1380–1383.
- (5) (a) Lin, J.-H. *Phytochemistry* **1989**, *28*, 621–622. (b) Koguchi, Y.; Kohno, J.; Nishio, M.; Takahashi, K.; Okuda, T.; Ohnuki, T.; Komatsubara, S. *J. Antibiot.* **2000**, *53*, 105–109. (c) Palermo, J. A.; Flower, P. B.; Seldes, A. M. *Tetrahedron Lett.* **1992**, *33*, 3097–3100. (d) Jansen, R.; Washausen, P.; Kunze, B.; Reichenbach, H.; Höfle, G. *Eur. J. Org. Chem.* **1999**, *1999* (5), 1085–1089. (e) Kunze, B.; Jansen, R.; Sasse, F.; Höfle, G.; Reichenbach, H. *J. Antibiot.* **1998**, *51*, 1075–1080.
- (6) Garcia-Barrantes, P. M.; Lindsley, C. W. *Org. Lett.* **2016**, *18*, 3810–3813.
- (7) Gibson, F. S.; Bergmeier, S. C.; Rapoport, H. *J. Org. Chem.* **1994**, *59*, 3216–3218.
- (8) (a) Boger, D. L.; Ichikawa, S. *J. Am. Chem. Soc.* **2000**, *122*, 2956–2957. (b) Kamber, B.; Hartmann, A.; Eisler, K.; Riniker, B.; Rink, H.; Sieber, P.; Rittel, W. *Helv. Chim. Acta* **1980**, *63*, 899–915.
- (9) (a) Georgiades, S. N.; Clardy, J. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 3117–3121. (b) Cheung, C. W.; Buchwald, S. L. *J. Org. Chem.* **2012**, *77*, 7526–7537. (c) Toumi, M.; Couty, F.; Evano, G. *Synlett* **2008**, *2008* (1), 29–32. (d) Toumi, M.; Couty, F.; Evano, G. *J. Org. Chem.* **2007**, *72*, 9003–9009. (e) Toumi, M.; Couty, F.; Evano, G. *Angew. Chem., Int. Ed.* **2007**, *46*, 572–575. (f) Toumi, M.; Rincheval,

V.; Young, A.; Gergeres, D.; Turos, E.; Couty, F.; Mignotte, B.; Evano, G. *Eur. J. Org. Chem.* **2009**, *20*, 3368–3386.

(10) (a) Takada, K.; Imamura, N.; Gustafson, K. R.; Henrich, C. J. *Bioorg. Med. Chem. Lett.* **2010**, *20*, 1330–1333. (b) Snider, B. B.; Song, F.; Foxman, B. M. *J. Org. Chem.* **2000**, *65*, 793–800.

(11) (a) Grieco, P. A.; Miyashita, M. *J. Org. Chem.* **1974**, *39*, 120–122. (b) Grieco, P. A.; Gilman, S.; Nishizawa, M. *J. Org. Chem.* **1976**, *41*, 1485–1486.

(12) For examples of this methodology applied to peptide substrates, see: (a) Schmidt, U.; Wild, J. *Angew. Chem., Int. Ed. Engl.* **1984**, *23*, 991–993. (b) Schmidt, U.; Lieberknecht, A.; Griesser, H.; Bökens, H. *Liebigs Ann. Chem.* **1985**, *4*, 785–793. (c) Schmidt, U.; Wild, J. *Liebigs Ann. Chem.* **1985**, *9*, 1882–1894. (d) Horenstein, B. A.; Nakanishi, K. *J. Am. Chem. Soc.* **1989**, *111*, 6242–6246. (e) Kim, D.; Li, Y.; Horenstein, B. A.; Nakanishi, K. *Tetrahedron Lett.* **1990**, *31*, 7119–7122. (f) Schmidt, U.; Zäh, M.; Lieberknecht, A. *J. Chem. Soc., Chem. Commun.* **1991**, *15*, 1002–1004. (g) Laib, T.; Bois-Choussy, M.; Zhu, J. *Tetrahedron Lett.* **2000**, *41*, 7645–7649. (h) Kim, Y.-A.; Shin, H.-N.; Park, M.-S.; Cho, S.-H.; Han, S.-Y. *Tetrahedron Lett.* **2003**, *44*, 2557–2560. (i) Cristau, P.; Temal-Laib, T.; Bois-Choussy, M.; Martin, M.-T.; Vors, J.-P.; Zhu, J. *Chem. - Eur. J.* **2005**, *11*, 2668–2679.

(13) (a) Matsui, H.; Schwartz, A. *Biochim. Biophys. Acta, Enzymol. Biol. Oxid.* **1966**, *128*, 380–390. (b) Assay performed by Eurofins, Inc. (c) Source: pig heart. Substrate: 100 μ M ATP. Vehicle: 1% DMSO. Incubation buffer: 50 mM Tris-HCl, pH 7.0, 5 mM MgCl₂, 100 mM NaCl, 20 mM KCl. Pre-incubation time: 15 min at 37 °C. Incubation time: 60 min at 37 °C. Quantitation method: spectrophotometric qualification of P_i.

(14) Shoemaker, R. H. *Nat. Rev. Cancer* **2006**, *6*, 813–823.