# Total Synthesis of (S,S)-Tetramethylmagnolamine via Aerobic Desymmetrization

Zheng Huang, Xiang Ji,<sup>†</sup> and Jean-Philip Lumb\*®

Department of Chemistry, McGill University, 801 Sherbrooke Street West, Montreal, QC H3A 0B8, Canada

Supporting Information

ABSTRACT: We describe a concise synthesis of the pseudodimeric tetrahydroisoqunoline alkaloid (S,S)-tetramethylmagnolamine by a catalytic aerobic desymmetrization of phenols. Desymmetrization reactions increase molecular complexity with high levels of efficiency, but those that do so by aerobic oxidation are uncommon. Our conditions employ molecular oxygen as an oxygen atom transfer agent and a formal acceptor of hydrogen, enabling two mechanistically distinct aromatic C-H oxygenation reactions with high degrees of selectivity.



olecular oxygen  $(O_2)$  is an attractive reagent for chemical synthesis (Scheme 1A).<sup>1</sup> It is renewable and nontoxic, and its reduction to water  $(H_2O)$  provides a strong thermodynamic driving force that can offset otherwise challenging bond forming reactions. Despite these advantages, O2 can be difficult to use when oxidizing C-H bonds in complex molecules, where issues of chemo- and regioselectivity outweigh synthetic efficiency.<sup>2</sup>  $O_2$  has a small steric profile, a noncanonical 4 e<sup>-1</sup>4 H<sup>+</sup> redox stoichiometry, and a triplet electronic structure.<sup>3</sup> These factors often lead to radical chain processes that can be difficult to control when potential sites of oxidation are not electronically differentiated.<sup>4</sup>

To overcome these challenges, our group has adopted a bioinspired strategy of utilizing O<sub>2</sub> that is inspired by metalloenzymes (Scheme 1A).<sup>5,6</sup> By coordinating O<sub>2</sub> to transition metals within their active sites, metalloenzymes can exert control over aerobic oxidations by defining the coordination environment surrounding the metal center.<sup>1a,b,7</sup> Our group has been particularly interested in the type III Cu enzyme tyrosinase (Scheme 1B), which catalyzes the aerobic oxygenation of L-tyrosine in the first and rate-limiting step of melanogenesis (not shown).<sup>8</sup> Building upon bioinorganic studies of Stack and others,<sup>9,10</sup> our group showed that the enzyme's mechanism of O<sub>2</sub> activation and oxygen atom transfer (OAT) could be recreated by a simple and commercially available catalytic system, composed of  $Cu(CH_3CN)_4(PF_6)$  (abbreviated CuPF<sub>6</sub>) and  $N_{N}$ '-di-tert-butylethylenediamine (DBED).<sup>11</sup> When applied to the oxidation of simple para-substituted phenols, this system catalyzes ortho-C-H oxygenation through a series of discrete steps consisting of the following: (step 1) activation of O<sub>2</sub> as its di-Cu(II)  $\mu$ - $\eta^2$ , $\eta^2$  peroxo complex (**P**), (step 2) coordination of a phenolate, and (step 3) electrophilic aromatic substitution onto the Cu<sub>2</sub>O<sub>2</sub> core. Although these discrete steps replicate those of the enzyme, the fate of the ensuing bridged di-Cu(II)-catecholate (Cu<sub>2</sub>-Cat) differs between the two systems. Whereas tyrosinase releases a free

ortho-quinone (Q1) following (step 4) protonation of the hydroxide bridge and (step 5a) redox between the ligand and the metal centers, the synthetic system releases a Cu(II)-semiquinone radical complex (SQ) (step 5b). The ensuing oxidative C-Ocoupling with the starting phenol is rate determining (step 6) and provides the corresponding substituted ortho-quinone product (Q2) while releasing DBED-Cu(I) to close the catalytic cycle.<sup>12</sup> O<sub>2</sub> serves multiple roles over the course of this reaction. The first is as a reagent for OAT that reduces the  $C_{2\nu}$  symmetry of the phenol to the  $C_s$  symmetry of either quinone (enzymatic) or SQ (biomimetic system). This distinction is important as the retention of Cu in the SQ forces the biomimetic system to engage another equivalent of the starting phenol in a C-O coupling. This step is distinct from the first OAT and ensures that the starting material is consumed in two distinct processes over the course of the reaction. C-O bond formation is driven by the favorable reduction of  $1/2 O_{2}$ , highlighting its second role as a formal acceptor of  $H_2$ .

Herein, we describe the application of this catalytic aerobic process to the synthesis of the pseudosymmetric tetrahydroisoquinoline (THIQ) alkaloid (S,S)-tetramethylmagnolamine (1) (Scheme 2A). 1 is a member of the diaryl ether subfamily of THIQs, which encompass a number of structurally unique and biologically active members.<sup>13</sup> Among them are the well-known macrocyclic diaryl ether tubocurarine (4),<sup>14</sup> which is a potent muscle relaxant, and the chemotherapeutic (S,S)-thalicarpine (3).<sup>15</sup> The family members share defining diaryl ethers, typically linking two electron-rich aromatic rings. They also possess two basic amine functionalities separated by  $\sim 14$  Å, which are believed to be protonated at physiological pH. This has become a defining structural

Received: October 8, 2019

Scheme 1. (A) Bioinspired Strategies for the Utilization of O<sub>2</sub> and (B) Biomimetic Aerobic Oxidation of Phenol



feature of many simple neuromuscular relaxants, which contain quaternized ammonium cations separated by  ${\sim}10$  carbon atoms.  $^{16}$ 

Although this subfamily of THIQs has been extensively studied, questions remain about their biosynthesis and how the associated oxidases create the defining diaryl ethers (Scheme 2B).<sup>13a</sup> Free radical reactions of phenoxyl radicals favor C-C instead of C-O bond formation, reflecting the favorable localization of spin density on the less electro-negative carbons of the aromatic ring.<sup>1c,17</sup> Little is known about how these enzymes change selectivity to promote C-O bond formation and to what extent the metal center is involved in the bond forming event. This underscores a more general challenge of using phenols in oxidative C-O bond forming reactions as it is easy to trigger competitive free-radical pathways that erode selectivity. Instead, more commonly practiced aryl ether syntheses are isohypsic, including Ullman or Buchwald-Hartwig cross-coupling reactions that employ an oxidized aromatic halide as the coupling partner (Scheme 2C).<sup>18</sup> Cross-coupling reactions of this family have been extensively studied, but their persistent use of stoichiometric base and increased reaction

temperatures makes them capricious in complex molecule settings. This includes the formation of the diaryl ether in (S,S)-tetramethylmagnolamine (1),<sup>19</sup> which was synthesized by Opatz, in 50% yield from phenol 9 and aryl bromide 10 under Ma's optimized conditions<sup>20</sup> for catalytic Ullman-type coupling.

Recognizing that 1 possesses an element of hidden symmetry, we questioned whether our conditions for diaryl ether formation could be used for its direct synthesis from 11 (Scheme 2C). This would feature a catalytic aerobic desymmetrization that could shorten the overall step count by requiring the preparation of only a single coupling partner. It would also offer a rare example of aerobic desymmetrization, in which an atom of  $O_2$  is incorporated into the product.<sup>21</sup> Herein, we describe the successful execution of this plan and report a concise and high-yielding synthesis of 1 along these lines.<sup>22</sup>

To investigate our proposed key step, we developed an asymmetric synthesis of Boc-protected THIQ (17), inspired by the precedent of Noyori (Scheme 3).<sup>23</sup> Namely, amidation of 12 and 13 at increased temperatures without solvent, followed by a Bischler-Napieralski cyclization, set the stage for Noyori's asymmetric hydrogenation.<sup>23</sup> This introduced the benzylic stereocenter in 94% ee over a three-step sequence that could be run on a multigram scale in 70% yield. Despite our efforts to use free amine 16 or its methyl-substituted derivative (not shown) in the subsequent aerobic catalysis, these substrates afforded intractable mixtures at complete consumption of the starting material. Therefore, we advanced the synthesis by Boc protection to afford 17 before conducting the key step. Under our previously optimized conditions,<sup>6a</sup> aerobic oxidative coupling is complete within 4 h at room temperature using an applied pressure of O<sub>2</sub> (1 atm). The catalyst system is composed of 4 mol % of CuPF<sub>6</sub> and 5 mol % of DBED in CH<sub>2</sub>Cl<sub>2</sub>. Whereas ortho-quinone 19 is the immediate product of coupling, we use a reductive workup, consisting of a saturated aqueous solution of sodium dithionite  $(Na_2S_2O_4)$  to effect in situ reduction. This leads to the corresponding coupled catechol 20 in a 60% isolated yield on a 400 mg scale. Notably, the yield of this process improves to 75% when conducted in the presence of 4 Å MS, highlighting the beneficial effects of running these reactions in the presence of a desiccant.<sup>17b</sup> Subsequent methylation of the catechol and reduction of the N-Boc groups under standard conditions furnished (S,S)-tetramethylmagnolamine (1) in a longest linear sequence (LLS) of 7 steps that proceeded in 21% overall vield. This compares favorably to the only previous synthesis of 1 by Opatz (Scheme 2C), which required 16 total steps and a LLS of 8 steps that proceeded in 14% overall yield.<sup>19</sup> Our route takes advantage of the target's pseudosymmetry and forms the key aryl ether linkage at room temperature by selectively oxidizing two aromatic C-H bonds of 1 equiv of the starting material and the phenol O-H on another. Because O<sub>2</sub> is the terminal reductant, H<sub>2</sub>O is the only stoichiometric byproduct of the reaction.

In conclusion, we have described an asymmetric total synthesis of (S,S)-tetramethylmagnolamine (1) that features a catalytic aerobic desymmetrization of phenols. Despite many of its attributes,  $O_2$  remains underutilized in complex molecule synthesis due, in part, to persistent challenges of controlling selectivity. Our work highlights the beneficial effects of bioinspired metal coordination in overcoming this challenge and also highlights the unique reactivity of Cu in catalyzing

# Scheme 2. (A) Tetrahydroisoquinoline Alkaloids with Diaryl Ether Linkages, (B) Proposed Biosynthesis of the Diaryl Ether Linkages, and (C) Previous Synthesis of (S,S)-Tetramethylmagnolamine and This Work







<sup>a</sup>Reagents and conditions: (a) neat, 200 °C, 2 h, 91%; (b) POCl<sub>3</sub> (6 equiv), MeCN, reflux, 1 h; (c) **15** (0.2 mol %), HCO<sub>2</sub>H/NEt<sub>3</sub> (5:2), DMF, rt, 12 h, 78% over two steps, 94% ee; (d) Boc<sub>2</sub>O (1.1 equiv), MeOH, rt, 12 h, 89%; (e)  $[Cu(MeCN)_4](PF_6)$  (4 mol %), DBED (5 mol %), 4 Å MS, CH<sub>2</sub>Cl<sub>2</sub>, O<sub>2</sub> (1 atm), rt, 4 h, then Na<sub>2</sub>S<sub>2</sub>O<sub>4</sub> workup, 75%; (f) MeI (3.0 equiv), Cs<sub>2</sub>CO<sub>3</sub> (3.0 equiv), DMSO, rt, 12 h; (g) LiAlH<sub>4</sub> (10.0 equiv), THF, reflux, 12 h, 44% over two steps.

C–O bond formation during phenolic oxidation. We see many opportunities to extend this chemistry to other diaryl ether targets, including additional members of the THIQ-ether subfamily,<sup>15b</sup> and work toward this goal will be reported in due course.

### ASSOCIATED CONTENT

#### **Supporting Information**

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.orglett.9b03559.

#### **Organic Letters**

Experimental procedures and characterization data (PDF)

Copies of NMR spectra (PDF)

# AUTHOR INFORMATION

#### Corresponding Author

\*E-mail: jean-philip.lumb@mcgill.ca.

# ORCID ®

Jean-Philip Lumb: 0000-0002-9283-1199

#### Present Address

<sup>†</sup>Department of Chemistry, New York University, Silver Center for Arts and Science, 100 Washington Square East, New York, NY 10003, United States.

#### Notes

The authors declare no competing financial interest.

#### ACKNOWLEDGMENTS

Financial support was provided by the Natural Sciences and Engineering Council (NSERC) of Canada (Discovery Grant to J.-P.L.); the Fonds de Recherche Quebecois Nature et Technologies (FRQNT) (Team Grant to J.-P.L.); McGill University Faculty of Science (Milton Leong Fellowship in Science to Z.H.), and the FRQNT Center for Green Chemistry and Catalysis (fellowship to X.J.).

#### REFERENCES

(1) (a) Que, L.; Tolman, W. B. Nature 2008, 455, 333-340.
(b) Trammell, R.; Rajabimoghadam, K.; Garcia-Bosch, I. Chem. Rev. 2019, 119, 2954-3031.
(c) Allen, S. E.; Walvoord, R. R.; Padilla-Salinas, R.; Kozlowski, M. C. Chem. Rev. 2013, 113, 6234-6458.
(d) Campbell, A. N.; Stahl, S. S. Acc. Chem. Res. 2012, 45, 851-863.
(e) Punniyamurthy, T.; Velusamy, S.; Iqbal, J. Chem. Rev. 2005, 105, 2329-2364.

(2) Stahl, S. S.; Alsters, P. L. Liquid Phase Aerobic Oxidation Catalysis: Industrial Applications and Academic Perspectives; Wiley-VCH: Weinheim, 2016.

(3) Borden, W. T.; Hoffmann, R.; Stuyver, T.; Chen, B. J. Am. Chem. Soc. 2017, 139, 9010–9018.

(4) (a) Esguerra, K. V. N.; Xu, W.; Lumb, J.-P. Chem. 2017, 2, 533–549. (b) Esguerra, K. V. N.; Lumb, J.-P. Angew. Chem., Int. Ed. 2018, 57, 1514–1518.

(5) For review articles, see: (a) Esguerra, K. V. N.; Lumb, J.-P. *Synthesis* **2019**, *51*, 334–358. (b) Huang, Z.; Lumb, J.-P. *ACS Catal.* **2019**, *9*, 521–555.

(6) For related examples, see: (a) Esguerra, K. V. N.; Fall, Y.; Lumb, J.-P. Angew. Chem. 2014, 126, 5987–5991. (b) Huang, Z.; Kwon, O.; Huang, H.; Fadli, A.; Marat, X.; Moreau, M.; Lumb, J.-P. Angew. Chem., Int. Ed. 2018, 57, 11963–11967.

(7) (a) Elwell, C. E.; Gagnon, N. L.; Neisen, B. D.; Dhar, D.; Spaeth, A. D.; Yee, G. M.; Tolman, W. B. *Chem. Rev.* 2017, *117*, 2059–2107.
(b) Solomon, E. I.; Heppner, D. E.; Johnston, E. M.; Ginsbach, J. W.; Cirera, J.; Qayyum, M.; Kieber-Emmons, M. T.; Kjaergaard, C. H.; Hadt, R. G.; Tian, L. *Chem. Rev.* 2014, *114*, 3659–3853.

(8) (a) Borovansky, J.; Riley, P. A. *Melanins and Melanosomes*; Wiley-VCH: Weinheim, 2011. (b) Esguerra, K. V. N.; Lumb, J.-P. *Synlett* 2015, 26, 2731–2738.

(9) Mirica, L. M.; Vance, M.; Rudd, D. J.; Hedman, B.; Hodgson, K. O.; Solomon, E. I.; Stack, T. D. P. *Science* **2005**, *308*, 1890–1892.

(10) For review articles, see: (a) Rolff, M.; Schottenheim, J.; Decker, H.; Tuczek, F. *Chem. Soc. Rev.* **2011**, 40, 4077–4098. (b) Mirica, L. M.; Ottenwaelder, X.; Stack, T. D. P. *Chem. Rev.* **2004**, *104*, 1013–1046.

(11) Askari, M. S.; Esguerra, K. V. N.; Lumb, J.-P.; Ottenwaelder, X. Inorg. Chem. **2015**, *54*, 8665–8672. (12) For a more detailed study of the C–O coupling step, see: Huang, Z.; Lumb, J.-P. Angew. Chem., Int. Ed. 2016, 55, 11543–11547.

(13) (a) Weber, C.; Opatz, T. Bisbenzylisoquinoline Alkaloids. In *The Alkaloids: Chemistry and Biology*; Knölker, H.-J., Ed.; Academic Press, 2019; Vol. 81, pp 1–114, Chapter 1. (b) Shamma, M. *The Isoquinoline Alkaloids Chemistry and Pharmacology*; Elsevier, 2012; Vol. 25.

(14) Otto, N.; Ferenc, D.; Opatz, T. J. Org. Chem. 2017, 82, 1205–1217.

(15) (a) Kupchan, S. M.; Liepa, A. J.; Kameswaran, V.; Sempuku, K. J. Am. Chem. Soc. **1973**, 95, 2995–3000. (b) Xu, W.; Huang, Z.; Ji, X.; Lumb, J.-P. ACS Catal. **2019**, 9, 3800–3810.

(16) Alkaloids. In Medicinal Natural Products; Dewick, P. M., Ed.; 2009; pp 311-420.

(17) (a) Quideau, S.; Deffieux, D.; Pouységu, L. Oxidative Coupling of Phenols and Phenol Ethers. In *Comprehensive Organic Synthesis II*, 2nd ed.; Knochel, P., Ed.; Elsevier: Amsterdam, 2014; pp 656–740.
(b) Esguerra, K. V. N.; Fall, Y.; Petitjean, L.; Lumb, J.-P. J. Am. Chem. Soc. 2014, 136, 7662–7668.

(18) (a) Evano, G.; Wang, J.; Nitelet, A. Org. Chem. Front. 2017, 4, 2480–2499. (b) Pitsinos, E. N.; Vidali, V. P.; Couladouros, E. A. Eur. J. Org. Chem. 2011, 2011, 1207–1222.

(19) Blank, N.; Opatz, T. J. Org. Chem. 2011, 76, 9777-9784.

(20) Ma, D.; Cai, Q. Org. Lett. 2003, 5, 3799-3802.

(21) For selected examples of desymmetrization in biosynthesis or bioinspired synthesis, see: (a) Hu, X.; Maimone, T. J. J. Am. Chem. Soc. 2014, 136, 5287–5290. (b) Miyabe, H.; Torieda, M.; Inoue, K.; Tajiri, K.; Kiguchi, T.; Naito, T. J. Org. Chem. 1998, 63, 4397–4407. (c) Smith, W. L.; Urade, Y.; Jakobsson, P.-J. Chem. Rev. 2011, 111, 5821–5865. (d) Goodhue, C. T.; Schaeffer, J. R. Biotechnol. Bioeng. 1971, 13, 203–214.

(22) A version of this work first appeared on the ChemRxiv repository: Huang, Z.; Ji, X.; Lumb, J.-P. *ChemRxiv Preprint* 2019, DOI: 10.26434/chemrxiv.9956213.

(23) Uematsu, N.; Fujii, A.; Hashiguchi, S.; Ikariya, T.; Noyori, R. J. Am. Chem. Soc. **1996**, 118, 4916–4917.