



Asymmetric Catalysis

Enantioselective Allylic Substitution of Morita–Baylis–Hillman Adducts Catalyzed by Chiral Bifunctional Ferrocenylphosphines

Linglong Zhu,^[a] Haiwen Hu,^[a] Liang Qi,^[a] Yi Zheng,^[a] and Weihui Zhong*^[a]

Abstract: A series of air-stable chiral ferrocenylphosphines (**LB1–LB4**) were prepared and used in the asymmetric allylic substitution of Morita–Baylis–Hillman (MBH) adducts with phthalimide under mild reaction conditions; the (R_{SFc})-ferro-

cenylphosphine **LB4** afforded the desired amination products **3** in moderate yields with excellent enantioselectivities. The absolute configuration of **30** was confirmed by X-ray analysis.

Introduction

The Morita-Baylis-Hillman (MBH) reaction has emerged as an attractive approach for the preparation of α -methylene- β hydroxycarbonyl compounds, which are versatile synthetic building blocks in organic synthesis.^[1] In recent decades, great effort has been devoted to the synthesis of these chiral compounds through the following two approaches: the asymmetric Morita-Baylis-Hillman (MBH) reaction^[2] and the asymmetric substitution of MBH adducts.^[3] Owing to the broader range of substrates, increasing attention has been paid to the asymmetric substitution of MBH adducts. Notably, great progress have been achieved in the organophosphine-catalyzed allylic amination and allylic alkylation of MBH acetates and carbonates.^[4] In 2004, Krische and co-workers^[5] first reported the asymmetric amination of MBH acetates with phthalimide catalyzed by chiral phosphanes, which provided the corresponding substitution products in good yields along with moderate ee values. Inspired by the elegant work of Krische, Hou and Shi independently studied the asymmetric transformation of MBH adducts to build highly functionalized γ -butenolides from MBH acetates or carbonates. In 2007, Hou and co-workers^[6] reported the enantioselective substitution of MBH adducts derived from acrylate with phthalimide, catalyzed by planar chiral [2,2]-paracyclophane monophosphanes to give the allylic amination products with high regioselectivities but poor-to-moderate enantioselectivities (9-71 % ee); however, the absolute configuration of these amination products was not disclosed (Scheme 1). Later, Shi and co-workers^[7,8] developed a set of highly active chiral phosphane organocatalysts, containing both Lewis basic and Brönsted acidic sites, which can achieve highly enantioselective allvlic amination of acetates of MBH alcohols derived from methyl vinyl ketone (MVK) or ethyl vinyl ketone (EVK) to afford the target products in good yields and up to 90 % ee (Scheme 2). To date, methodologies for the highly enantioselective substitution of carbonates or acetates of MBH alcohols derived from MVK and EVK have been well-established and can be achieved readily, but the asymmetric reactions of MBH adducts derived from acrylate with phthalimide have remained a challenge. Thus, the discovery of new catalysts to achieve the highly enantioselective substitution of MBH acetates or carbonates derived from acrylate is still a highly desirable goal. Very recently, our



Scheme 1. Previous work by Hou and co-workers.

[a] Collaborative Innovation Center of Green Pharmaceutical Engineering, College of Pharmaceutical Sciences, Zhejiang University of Technology, Chao Wang Road 18th, 310014, Hangzhou, P. R. China E-mail: weihuizhong@zjut.edu.cn www.zjut.edu.cn

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group reported a highly enantioselective [3+2] cycloaddition reaction catalyzed by the chiral bifunctional ferrocenylphosphine **LB2** (Scheme 3).^[9] The catalysts containing five-membered heteroaryl rings and amide moieties presented higher catalytic activities owing to a favorable steric interaction be-





tween the diphenylphosphine and heteroaryl moieties. On the basis of these results, in this work, we chose the best structural pattern and modified the five-membered heteroaryl ferrocenylphosphine catalyst and then investigated if the highly enantioselective amination of MBH adducts derived from acrylate with phthalimide could be achieved (Figure 1).



Scheme 2. Previous work by Shi and co-workers.

Our group's previous work



Scheme 3. Asymmetric [3+2] cycloaddition reaction catalyzed by ferrocenyl-phosphines.



Figure 1. The structures of the chiral bifunctional ferrocenylphosphines.

Results and Discussion

The initial assays were performed with MBH adduct **1a** and phthalimide (**2**) in the presence of the chiral bifunctional ferrocenylphosphines **LB1–LB4** (Table 1). Pleasingly, the allylic amination product **3a** was obtained in 65 % yield and 91 % *ee* with **LB2** (20 mol-%) as the catalyst in tetrahydrofuran (THF) at room temperature for 120 h (Table 1, Entry 2). In addition, **LB1** as the catalyst resulted in a decrease in both yield and enantioselectivity (Table 1, Entry 1). Catalyst **LB3** was also effective for this reaction and afforded the desired product **3a** in moderate yield but good *ee* (Table 1, Entry 3). Delightfully, the dichloro-substituted catalyst **LB4** showed the best enantioselectivity in this reaction and afforded the corresponding product **3a** in 73 %

yield with 94 % *ee* (Table 1, Entry 4). A slight increase of both yield and *ee* was observed for a prolonged reaction time of 240 h (Table 1, Entry 5). Notably, decreased catalyst loadings and temperature had little influence on this reaction (Table 1, Entries 6–9). The further exploration of the effect of the solvent suggested that CHCl₃ can significantly promote the reaction to afford the corresponding product **3a** in 74 % yield and 96 % *ee* (Table 1, Entries 10–13).

Table 1. Screening of chiral phosphines for the allylic substitution of MBH adduct 1a with phthalimide $2^{\rm [a]}$



Catalyst	Time	T [°C]	Solvent	3a	
[mol-%]	[h]			Yield ^[b] [%]	<i>ee</i> ^[c] [%]
LB1 (20)	120	r.t.	THF	64	-86
LB2 (20)	120	r.t.	THF	65	91
LB3 (20)	120	r.t.	THF	58	92
LB4 (20)	120	r.t.	THF	73	94
LB4 (20)	240	r.t.	THF	74	96
LB4 (10)	120	r.t.	THF	52	96
LB4 (20)	240	0	THF	42	96
LB4 (20)	120	40	THF	75	94
LB4 (20)	120	40	CHCl ₃	74	96
LB4 (20)	120	r.t.	CH_2CI_2	42	94
LB4 (20)	120	r.t.	Et ₂ O	34	75
LB4 (20)	120	r.t.	CHCl ₃	74	96
LB4 (20)	120	r.t.	toluene	53	92
	Catalyst [mol-%] LB1 (20) LB2 (20) LB3 (20) LB4 (20)	Catalyst [mol-%] Time [h] LB1 (20) 120 LB2 (20) 120 LB3 (20) 120 LB4 (20) 120 LB4 (20) 120 LB4 (20) 120 LB4 (20) 240 LB4 (20) 240 LB4 (20) 120 LB4 (20) 120	Catalyst [mol-%] Time [h] T [°C] LB1 (20) 120 r.t. LB2 (20) 120 r.t. LB3 (20) 120 r.t. LB4 (20) 120 r.t. LB4 (20) 120 r.t. LB4 (20) 120 r.t. LB4 (20) 240 r.t. LB4 (20) 240 0 LB4 (20) 120 40 LB4 (20) 120 40 LB4 (20) 120 r.t. LB4 (20) 120 r.t.	$\begin{array}{c c} Catalyst [mol-\%] & Time [h] & T[^{\circ}C] & Solvent \\ [mol-\%] & [h] & T[^{\circ}C] & Solvent \\ \hline LB1 (20) & 120 & r.t. & THF \\ LB2 (20) & 120 & r.t. & THF \\ LB3 (20) & 120 & r.t. & THF \\ LB4 (20) & 120 & r.t. & THF \\ LB4 (20) & 240 & r.t. & THF \\ LB4 (20) & 240 & 0 & THF \\ LB4 (20) & 120 & 40 & THF \\ LB4 (20) & 120 & 40 & THF \\ LB4 (20) & 120 & 40 & CHCl_3 \\ LB4 (20) & 120 & r.t. & CH_2Cl_2 \\ LB4 (20) & 120 & r.t. & CH_2Cl_2 \\ LB4 (20) & 120 & r.t. & CHCl_3 \\ LB4 (20) & 120 & r.t. & CHCl_3 \\ LB4 (20) & 120 & r.t. & CHCl_3 \\ LB4 (20) & 120 & r.t. & toluene \\ \end{array}$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$

[[]a] Unless otherwise specified, the reactions were performed with **1a** (0.5 mmol), **2** (1.5 mmol), and catalysts **LB1–LB4** (0.1 mmol) in THF (2.5 mL) at room temperature. [b] Isolated yield. [c] The *ee* was determined by HPLC analysis with a chiral column.

With the optimal reaction conditions in hand (Table 1, Entry 12), the scope of this reaction was then explored with a series of MBH acetates 1 derived from acrylate in the reaction with 2 (Table 2). A variety of substituted MBH adducts 1 bearing electron-withdrawing or -donating groups on aromatic ring or different ester groups on the side chain all proceeded smoothly after 120 h to afford the corresponding products (3a-3p) in moderate-to-good yields (up to 74 %) and excellent ee values (up to 96 % ee). We found that the substrate derived from methyl acrylate delivered a better result than the substrates derived from ethyl acrylate and butyl acrylate (Table 2, Entries 1-3). Notably, electron-withdrawing aromatic MBH adducts were more reactive than electron-donating ones (Table 2, Entries 1, 11, 13, and 15). In addition, multisubstituted aromatic rings (Table 2, Entries 12 and 16) also afforded the corresponding products in moderate yields along with high ee values. Owing to steric hindrance, the ortho-heterosubstituted MBH adducts gave lower yields than those of meta- or para-substituted products (Table 2, Entries 6-10). Notably, the ortho-nitro-substituted MBH adduct could not undergo this reaction (Table 2, Entry 4). A naphthyl group was tolerated in this reaction, and the corresponding product **3p** was obtained in moderate yield





and *ee* (Table 2, Entry 17). Unfortunately, for the aliphatic MBH acetate, the product was not obtained even if the reaction was performed at a higher temperature (40 or 60 °C) or with a higher catalyst loading (Table 2, Entry 18). The absolute configuration of **30** was determined to be *R* by X-ray analysis (Figure 2). Interestingly, the absolute configuration of most of products **3** should be (+)-S according to their specific rotation (Table 2).

Table 2. Asymmetric allylic substitution reaction of MBH adducts with phthal-imide catalyzed by $\textbf{LB4}.^{[a]}$



[a] Unless otherwise specified, the reactions were performed with **1** (0.5 mmol), **2** (1.5 mmol), and **LB4** (0.1 mmol) in CHCl₃ (2.5 mL) at room temperature. [b] Isolated yield. [c] The *ee* was determined by HPLC analysis with a chiral column.



Figure 2. X-ray crystal structure of (R)-30.

On the basis of the experimental results^[10] and previous work,^[5,8,11] a plausible mechanism and an activation model are outlined in Scheme 4. The direct nucleophilic addition of the chiral bifunctional ferrocenylphosphine catalyst to the MBH

acetate generates the intermediate **I** and an acetate ion. The acetate ion engages in an acid–base equilibrium involving deprotonation of the pronucleophile, as proposed by Krische. In our case, the pK_a of the pronucleophile phthalimide ($pK_a = 8.3$)^[12] is higher than that of acetic acid ($pK_a = 4.8$);^[13] therefore, the acetate should not deprotonate the phthalimide. Thus, we propose that the phthalimide directly attacks the γ position of the olefin from the *Re* face of the olefin owing to a steric repulsion between the aromatic group of the catalyst and the phthalimide to offer intermediate **II**, which undergoes deprotonation to afford the final product **3**.



The acid-base equilibrium may not exist in this catalytic cycle

Scheme 4. Possible mechanism.

Conclusions

We have developed a series of chiral ferrocenylphosphines to promote the enantioselective allylic substitution of MBH adducts with phthalimide, and the chiral ferrocenylphosphine **LB4** showed good catalytic activity for this reaction. Furthermore, **LB4** is reasonably air-stable in solution and in the solid state.^[14] Efforts are in progress to use the chiral ferrocenylphosphine catalysts for other asymmetric reactions.

Experimental Section

Materials and General Experimental Details: All starting materials were commercially available and used without further purification. Melting points were determined with a Büchi B-540 capillary melting-point apparatus. Optical rotations were determined with an Autopol V polarimeter. The ¹H and ¹³C NMR spectra of samples in CDCl₃ were recorded with a Varian-400 spectrometer at 400 and 100 MHz with tetramethylsilane (TMS, $\delta = 0$ ppm) as an internal standard. The chemical shifts are reported in ppm, and coupling constants J are expressed in Hertz. The ³¹P NMR spectra of samples in CDCl₃ with 85 % H₃PO₄ as an internal standard were recorded with a Varian 400 spectrometer at 161.9 MHz. Solution mass spectra were obtained with a Trace DSQ mass spectrometer in ESI mode and a Trace quadrupole mass spectrometer in El mode. High-resolution mass spectra were acquired with an Agilent 6210 TOF mass spectrometer. The enantiomeric excesses of the target molecules were determined through chiral-phase HPLC analysis with an





Agilent 1100 HPLC system (Daicel Chiralcel AD-H, OD-H, or AS-H column and eluted with *n*-hexane/*i*PrOH) with a UV diode array detector (DAD). The X-ray analysis was performed with Xcalibur and Gemini instruments.

The MBH adducts 1 were prepared by the literature procedure.^[15]

(S)-5-Chloro-N-[1'-(R)-2'-(diphenylphosphanyl)ferrocenyl]ethylthiophene-2-carboxamide (LB1): A mixture of 5-chlorothiophene-2-carboxylic acid (0.24 g, 1.5 mmol), bis(trichloromethyl)carbonate (0.22 g, 0.75 mmol), and N,N-dimethylformamide (DMF; 1.10 mg, 0.015 mmol) in toluene (5 mL) was heated to 110 °C. Then, the generated 5-chlorothiophene-2-carbonyl chloride was cooled to room temperature and added to a solution of (S)-1-(S)-[diphenylphosphinoferrocenyl]ethylamine (0.21 g, 0.50 mmol) in CH₂Cl₂ (5 mL), and the reaction mixture was stirred at 0 °C for 1 h. After the completion of the reaction, water was added to quench the reaction. The reaction mixture was then filtered and extracted with CH_2Cl_2 (3 × 10 mL), and the combined organic extracts were dried with Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography to afford LB1 (0.24 g, 86 %) as a yellow solid, m.p. 118.5–120.5 °C. $[\alpha]_D^{20} = +323.2$ (c = 0.5, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ = 7.57–7.49 (m, 2 H), 7.40 (d, J = 5.2 Hz, 3 H), 7.19 (d, J = 3.2 Hz, 5 H), 7.14 (s, 1 H), 6.99 (d, J = 3.6 Hz, 1 H), 6.80 (d, J = 2.4 Hz, 1 H), 5.25 (q, J = 6.0 Hz, 1 H), 4.54 (s, 1 H), 4.36 (s, 1 H), 4.02 (s, 5 H), 3.86 (s, 1 H), 1.41 (d, J = 6.0 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 159.0, 138.7, 138.0, 136.3, 135.0, 134.8, 134.6, 132.6, 132.4, 129.4, 128.3 (2 C), 128.2, 126.7 (2 C), 95.7, 95.5, 73.9, 73.8, 72.2, 71.3, 70.0 (5 C), 69.6, 46.0, 23.0 ppm. ³¹P NMR (162 MHz, CDCl₃, 85 % H₃PO₄): $\delta = -23.73$ ppm. MS (ESI): m/z (%) = 580.6 (100) [M + Na]⁺. HRMS (ESI): calcd. for C₂₉H₂₅CIFeNNaOPS [M + Na]⁺ 580.0325; found 580.0322.

Catalyst **LB2** was prepared as a yellow solid by a similar procedure. $[\alpha]_{D}^{20} = -328.0$ (c = 0.5, CHCl₃).

(*R*)-3-Methyl-*N*-[1'-(*S*)-2'-(diphenylphosphanyl)ferrocenyl]ethylthiazole-2-carboxamide (LB3): By following a similar procedure as that for LB1, LB3 (0.24 g, 92 %) was obtained a yellow solid, m.p. 58.8–60.8 °C. [α]_D²⁰ = -339.2 (*c* = 0.5, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ = 8.63 (s, 1 H), 7.55–7.48 (m, 2 H), 7.38 (d, *J* = 5.2 Hz, 3 H), 7.18–7.08 (m, 5 H), 6.71–6.62 (m, 1 H), 5.33 (q, *J* = 5.2 Hz, 1 H), 4.53 (s, 1 H), 4.35 (t, *J* = 2.4 Hz, 1 H), 4.03 (s, 5 H), 3.88–3.81 (m, 1 H), 2.68 (s, 3 H), 1.45 (d, *J* = 6.8 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 159.7, 155.8, 151.6, 138.8, 136.4, 135.0, 134.7, 132.4, 132.2, 129.3, 128.2, 128.2, 128.1, 125.9, 95.4, 95.1, 74.4, 74.3, 72.3, 71.0, 70.0 (5 C), 69.5, 45.9, 22.7, 17.5 ppm. ³¹P NMR (162 MHz, CDCl₃, 85 % H₃PO₄): δ = -24.60 ppm. MS (ESI): *m/z* (%) = 561.2 (100) [M + Na]⁺. HRMS (ESI): calcd. for C₂₉H₂₇FeN₂NaOPS [M + Na]⁺ 561.0823; found 561.0845.

(*R*)-4,5-Dichloro-*N*-[1'-(*S*)-2'-(diphenylphosphanyl)ferrocenyl]ethylthiophene-2-carboxamide (LB4): By following a similar procedure as that for LB1, LB4 (0.26 g, 87 %) was obtained as a yellow solid, m.p. 85.5–87.8 °C. $[\alpha]_D^{20} = -340.4$ (c = 0.5, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.54-7.47$ (m, 2 H), 7.38 (d, J = 4.8 Hz, 3 H), 7.18 (d, J = 4.0 Hz, 5 H), 6.56 (s, 1 H), 6.54–6.49 (m, 1 H), 5.33 (q, J = 6.8 Hz, 1 H), 4.53 (s, 1 H), 4.35 (t, J = 2.4 Hz, 1 H), 4.03 (s, 5 H), 3.87–3.83 (m, 1 H), 1.47 (d, J = 6.4 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 158.1$, 138.9, 136.2, 135.8, 134.9, 134.7, 132.6, 132.4, 129.4, 129.3, 128.5, 128.3, 128.2, 126.1, 123.9, 94.8, 94.6, 74.8, 74.7, 72.4, 70.9, 70.1 (5 C), 69.6, 46.1, 21.9 ppm. ³¹P NMR (162 MHz, CDCl₃, 85 % H₃PO₄): $\delta = -24.67$ ppm. MS (ESI): m/z (%) = 630.5 (100) [M + K]⁺. HRMS (ESI): calcd. for C₂₉H₂₄Cl₂FeNOPS [M + K]⁺ 613.9936; found 613.9951.

General Procedure for the Enantioselective Allylic Substitution: To a flame-dried Schlenk tube charged with the corresponding MBH acetate **1** (0.5 mmol), **2** (1.5 mmol, 220.5 mg), and chiral ferrocenylphosphine **LB4** (59.1 mg, 0.1 mmol) was added anhydrous $CHCl_3$ (2.5 mL). The reaction mixture stirred at room temperature under an argon atmosphere for 120 h. The reaction was monitored by TLC. After the starting substrates were consumed, the solvent was removed under reduced pressure, and the residue was purified by silica gel column chromatography (eluent: *n*-hexane/EtOAc/CH₂Cl₂ 8:1:1) to give the corresponding amination adduct **3**.

Sixteen chiral products were synthesized, of which 12 were new compounds (**3c**, **3d**–**3f**, **3h**–**3l**, and **3n**–**3p**).

(S)-*N*-[2-Methoxycarbonyl-1-(4'-nitrophenyl)allyl]phthalimide (3a): Yellow oil. $[\alpha]_{2^0}^{0} = -10.4$ (c = 0.5, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 8.21$ (d, J = 8.4 Hz, 2 H), 7.84–7.87 (m, 2 H), 7.74–7.76 (m, 2 H), 7.61 (d, J = 8.4 Hz, 2 H), 6.63 (s, 1 H), 6.52 (s, 1 H), 5.67 (s, 1 H), 3.73 (s, 3 H) ppm. MS (EI): m/z (%) = 366 (5), 334 (97), 147 (18), 104 (95), 76 (35). The *ee* was determined by HPLC with a chiral OD-H column ($\lambda = 218$ nm; eluent: *n*-hexane/*i*PrOH 85:15; flow rate: 0.8 mL/min; $t_{major} = 25.045$ min, $t_{minor} = 32.678$ min; *ee* = 96 %).

(S)-*N*-[2-(Ethoxycarbonyl)-1-(4'-nitrophenyl)allyl]phthalimide (3b): Yellow oil. $[\alpha]_{2^0}^{=0} = +2.8$ (c = 0.5, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 8.21$ (d, J = 8.8 Hz, 2 H), 7.84–7.87 (m, 2 H), 7.74–7.76 (m, 2 H), 7.62 (d, J = 8.8 Hz, 2 H), 6.63 (d, J = 1.2 Hz, 1 H), 6.51 (s, 1 H), 5.63 (d, J = 1.6 Hz, 1 H), 4.21–4.12 (m, 2 H), 1.19 (t, J = 7.2 Hz, 3 H) ppm. MS (EI): m/z (%) = 380 (3), 334 (60), 306 (100), 104 (36), 76 (25). The *ee* was determined by HPLC with a chiral OD-H column ($\lambda = 218$ nm; eluent: *n*-hexane/*i*PrOH 85:15; flow rate: 0.8 mL/min; $t_{major} = 19.380$ min, $t_{minor} = 23.288$ min; *ee* = 72 %).

(S)-*N*-[2-(Butoxycarbonyl)-1-(4'-nitrophenyl)allyl]phthalimide (3c): Yellow oil. [α]₂^D = -7.2 (*c* = 0.5, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ = 8.18 (d, *J* = 7.2 Hz, 2 H), 7.83 (d, *J* = 2.4 Hz, 2 H), 7.73 (d, *J* = 2.8 Hz, 2 H), 7.60 (d, *J* = 7.6 Hz, 2 H), 6.62 (s, 1 H), 6.50 (s, 1 H), 5.63 (s, 1 H), 4.12 (d, *J* = 5.6 Hz, 2 H), 1.57–1.54 (m, 2 H), 1.29–1.26 (m, 2 H), 0.87 (t, *J* = 6.4 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 167.4 (2 C), 165.0, 147.6, 144.2, 136.7, 134.4 (2 C), 131.6, 129.7 (2 C), 129.4 (2 C), 123.7 (2 C), 123.6 (2 C), 65.4, 54.0, 30.7, 19.3, 13.8 ppm. ES (EI): *m/z* (%): 408 (4), 334 (58), 306 (100), 104 (52), 76 (20). HRMS (ESI): calcd. for C₂₂H₂₀N₂NaO₆ [M + Na]⁺ 431.1214; found 431.1226. The *ee* was determined by HPLC with a chiral OD-H column (λ = 218 nm; eluent: *n*-hexane/*i*PrOH 85:15; flow rate: 0.8 mL/ min; *t*_{maior} = 15.775 min, *t*_{minor} = 18.969 min; *ee* = 73 %).

(S)-*N*-[2-Methoxycarbonyl-1-(4'-fluorophenyl)allyl]phthalimide (3d): Colorless oil. $[α]_{20}^{20} = +71.7$ (c = 0.4, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.83$ (m, 2 H), 7.71 (m, 2 H), 7.43 (m, 2 H), 7.01–7.05 (m, 2 H), 6.56 (d, J = 1.6 Hz, 1 H), 6.38 (s, 1 H), 5.63 (d, J = 1.6 Hz, 1 H), 3.71 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 167.6$, 165.7, 163.4, 161.0, 137.3, 134.0 (2 C), 132.7, 131.6, 130.4 (2 C), 129.1 (2 C), 123.3 (2 C), 115.6, 115.4, 54.0, 52.2 ppm. MS (EI): m/z (%) = 339 (6), 306 (80), 278 (72), 250 (100), 104 (38), 76 (35). HRMS (ESI): calcd. for C₁₉H₁₄FNNaO₄ [M + Na]⁺ 362.0799; found 362.0792. The *ee* was determined by HPLC with a chiral OD-H column (λ = 220 nm; eluent: *n*-hexane/*i*PrOH 75:25; flow rate: 0.8 mL/min; $t_{major} = 8.737$ min, $t_{minor} = 14.274$ min; *ee* = 72 %).

(*R*)-*N*-[2-Methoxycarbonyl-1-(2'-chlorophenyl)allyl]phthalimide (3e): Colorless oil. $[\alpha]_{2^0}^{2^0} = -9.0$ (c = 0.2, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.84-7.86$ (m, 2 H), 7.73–7.76 (m, 2 H), 7.51–7.53 (m, 1 H), 7.38–7.40 (m, 1 H), 7.27–7.29 (m, 2 H), 6.77 (s, 1 H), 6.60 (s, 1 H), 5.62 (s, 1 H), 3.73 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 167.6$ (2 C), 165.6, 136.2, 134.4, 134.2 (2 C), 133.6, 131.6 (2 C), 130.2, 129.7, 129.5, 128.8, 126.7, 123.5 (2 C), 52.4, 52.2 ppm. MS (EI): m/z (%) = 355 (1), 319 (100), 288 (32), 260 (19), 104 (16), 76 (10). HRMS (ESI): calcd. for C₁₉H₁₄CINNaO₄ [M + Na]⁺ 378.0504; found 378.0518. The



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ee was determined by HPLC with a chiral OJ-H column (λ = 220 nm; eluent: *n*-hexane/*i*PrOH 85:15; flow rate: 0.8 mL/min; t_{major} = 33.296 min, t_{minor} = 26.925 min; *ee* = 90 %).

(S)-*N*-[2-Methoxycarbonyl-1-(3'-chlorophenyl)allyl]phthalimide (3f): Colorless oil. [*α*]_D²⁰ = +73.3 (*c* = 0.15, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ = 7.83–7.85 (m, 2 H), 7.72–7.73 (m, 2 H), 7.42 (s, 1 H), 7.28–7.32 (m, 3 H), 6.59 (s, 1 H), 6.37 (s, 1 H), 5.65 (s, 1 H), 3.71 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 167.7 (2 C), 165.8, 139.0, 137.0, 134.6, 134.2 (2 C), 131.7, 129.9 (2 C), 129.8, 128.8, 128.4, 126.9, 123.6 (2 C), 54.3, 52.5 ppm. MS (El): *m/z* (%) = 355 (13), 295 (88), 267 (36), 207 (100), 104 (70), 76 (45). HRMS (ESI): calcd. for C₁₉H₁₄CINNaO₄ [M + Na]⁺ 378.0504; found 378.0514. The *ee* was determined by HPLC with a chiral OJ-H column (λ = 220 nm; eluent: *n*-hexane/*i*PrOH 95:5; flow rate: 0.8 mL/min; *t*_{major} = 34.998 min, *t*_{minor} = 32.733 min; *ee* = 82 %).

(S)-N-[2-Methoxycarbonyl-1-(4'-chlorophenyl)allyl]phthalimide (3g): Colorless oil. $[\alpha]_D^{20} = +74.7 (c = 0.8, CHCl_3)$. ¹H NMR (400 MHz, CDCl_3): $\delta = 7.82-7.84$ (m, 2 H), 7.70–7.73 (m, 2 H), 7.30–7.39 (m, 4 H), 6.57 (d, J = 1.6 Hz, 1 H), 6.37 (s, 1 H), 5.64 (d, J = 1.6 Hz, 1 H), 3.71 (s, 3 H) ppm. MS (EI): m/z (%) = 355 (5), 325 (15), 297 (20), 295 (100), 104 (33), 50 (2). Th *ee* was determined by HPLC with a chiral OD-H column ($\lambda = 220$ nm; eluent: *n*-hexane/*i*PrOH 85:15; flow rate: 0.6 mL/min; $t_{major} = 13.926$ min, $t_{minor} = 19.816$ min; *ee* = 75 %).

(S)-N-[2-Methoxycarbonyl-1-(4'-bromophenyl)allyl]phthalimide (3h): Colorless oil. $[\alpha]_{2^0}^{2^0} = +38.3$ (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.82-7.84$ (m, 2 H), 7.70–7.72 (m, 2 H), 7.47 (d, J = 8.4 Hz, 2 H), 7.32 (d, J = 8.4 Hz, 2 H), 6.57 (d, J = 0.8 Hz, 1 H), 6.36 (s, 1 H), 5.64 (d, J = 1.6 Hz, 1 H), 3.71 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 167.7$ (2 C), 165.8, 137.1, 136.0, 134.2 (2 C), 131.8 (2 C), 131.7, 130.4 (2 C), 129.5 (2 C), 123.5 (2 C), 122.3, 54.2, 52.5 ppm. MS (EI): m/z (%) = 399 (7), 339 (96), 311 (26), 104 (66), 76 (100). HRMS (ESI): calcd. for C₁₉H₁₄BrNNaO₄ [M + Na]⁺ 421.9998; found 422.0004. The *ee* was determined by HPLC with a chiral OD-H column ($\lambda =$ 220 nm; eluent: *n*-hexane/*i*PrOH 85:15; flow rate: 0.8 mL/min; $t_{major} = 10.880$ min, $t_{minor} = 14.623$ min; *ee* = 73 %).

(*S*)-*N*-[2-Methoxycarbonyl-1-(3'-bromophenyl)allyl]phthalimide (3i): Colorless oil. [*α*]_D²⁰ = +89.2 (*c* = 0.5, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ = 7.82–7.84 (m, 2 H), 7.70–7.72 (m, 2 H), 7.57 (s, 1 H), 7.36–7.44 (m, 2 H), 7.20–7.24 (m, 1 H), 6.59 (s, 1 H), 6.36 (s, 1 H), 5.66 (s, 1 H), 3.73 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 167.0 (2 C), 165.1, 138.8, 136.5, 133.8 (2 C), 131.3 (2 C), 130.9 (2 C), 129.8, 129.4, 127.0, 123.2 (2 C), 122.4, 54.3, 52.6 ppm. MS (El): *m/z* (%) = 399 (6), 369 (28), 341 (42), 231 (100), 104 (66), 76 (42). HRMS (ESI): calcd. for C₁₉H₁₄BrNNaO₄ [M + Na]⁺ 421.9998; found 422.0015. The *ee* was determined by HPLC with a chiral OJ-H column (λ = 220 nm; eluent: *n*-hexane/*i*PrOH 98:2; flow rate: 0.8 mL/min; *t*_{major} = 60.731 min, *t*_{minor} = 55.895 min; *ee* = 89 %).

(S)-*N*-[2-Methoxycarbonyl-1-(4'-methylphenyl)allyl]phthalimide (3j): Colorless oil. $[\alpha]_{D}^{20} = +76.5$ (c = 0.5, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.80-7.83$ (m, 2 H), 7.68–7.71 (m, 2 H), 7.32 (d, J = 8.0 Hz, 2 H), 7.15 (d, J = 8.0 Hz, 2 H), 6.55 (d, J = 1.6 Hz, 1 H), 6.36 (s, 1 H), 5.66 (d, J = 1.6 Hz, 1 H), 3.73 (s, 3 H), 2.37 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 167.8$ (2 C), 166.1, 137.9, 137.7, 134.0 (2 C), 131.9 (3 C), 129.3 (3 C), 128.6 (2 C), 123.4 (2 C), 54.7, 52.4, 21.4 ppm. MS (EI): m/z (%) = 335 (5), 304 (9), 276 (15), 275 (100), 76 (27). HRMS (ESI): calcd. for C₂₀H₁₇NNaO₄ [M + Na]⁺ 358.1050; found 358.1064. The *ee* was determined by HPLC with a chiral OD-H column ($\lambda = 218$ nm; eluent: *n*-hexane/*i*PrOH 85:15; flow rate: 0.8 mL/min; $t_{maior} = 12.303$ min, $t_{minor} = 14.144$ min; *ee* = 73 %).

(S)-N-[2-Methoxycarbonyl-1-(3,4-dimethylphenyl)allyl]phthalimide (3k): Colorless oil. $[\alpha]_{D}^{20} = +21.2$ (c = 0.2, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ = 7.81–7.83 (m, 2 H), 7.68–7.70 (m, 2 H), 7.17–7.18 (m, 2 H), 7.10 (d, *J* = 8.4 Hz, 1 H), 6.54 (d, *J* = 1.2 Hz, 1 H), 6.33 (s, 1 H), 5.66 (d, *J* = 1.6 Hz, 1 H), 3.71 (s, 3 H), 2.24 (s, 6 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 167.3 (2 C), 165.5, 137.3, 136.4, 136.1, 134.0, 133.6 (2 C), 131.5 (2 C), 129.5 (2 C), 129.0, 125.8, 123.1 (2 C), 54.8, 52.5, 20.5, 20.1 ppm. MS (EI): *m/z* (%) = 349 (11), 289 (100), 260 (83), 232 (10), 104 (49). HRMS (ESI): calcd. for C₂₁H₁₉NNaO₄ [M + Na]⁺ 372.1206; found 372.1211. The *ee* was determined by HPLC with a chiral OD-H column (λ = 220 nm; eluent: *n*-hexane/*i*PrOH 85:15; flow rate: 0.6 mL/min; *t*_{major} = 16.840 min, *t*_{minor} = 21.436 min; *ee* = 91 %).

(S)-*N*-[2-Methoxycarbonyl-1-(4'-methoxyphenyl)allyl]phthalimide (3l): Colorless oil. $[α]_{D^0}^{=0} = +78.7$ (*c* = 0.5, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.80-7.82$ (m, 2 H), 7.68–7.70 (m, 2 H), 7.38 (d, *J* = 8.4 Hz, 2 H), 6.87 (d, *J* = 8.4 Hz, 2 H), 6.54 (d, *J* = 1.6 Hz, 1 H), 6.34 (s, 1 H), 5.65 (d, *J* = 2.0 Hz, 1 H), 3.81 (s, 3 H), 3.72 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 167.8$ (2 C), 166.0, 159.3, 137.9, 134.0 (2 C), 131.9, 130.0 (3 C), 129.1 (2 C), 123.4 (2 C), 114.0 (2 C), 55.4, 54.4, 52.3 ppm. MS (EI): *m/z* (%) = 351 (13), 291 (100), 290 (57), 262 (32), 104 (41), 76 (29). HRMS (ESI): calcd. for C₂₀H₁₇NNaO₅ [M + Na]⁺ 374.0999; found 374.1007. The *ee* was determined by HPLC with a chiral OD-H column (λ = 218 nm; eluent: *n*-hexane/*i*PrOH 85:15; flow rate: 0.8 mL/min; *t*_{major} = 22.411 min, *t*_{minor} = 17.828 min; *ee* = 62 %).

(S)-N-[(2-Methoxycarbonyl-1-phenyl)allyl]phthalimide (3m): Yellow oil. $[a]_{20}^{20} = +82.4 (c = 0.5, CHCl_3).$ ¹H NMR (400 MHz, CDCl_3): $\delta = 7.82-7.84$ (m, 2 H), 7.69-7.71 (m, 2 H), 7.30-7.44 (m, 5 H), 6.56 (d, J = 1.6 Hz, 1 H), 6.39 (s, 1 H), 5.63 (d, J = 1.6 Hz, 1 H), 3.71 (s, 3 H) ppm. MS (EI): m/z (%) = 321 (2), 289 (33), 261 (100), 104 (26), 76 (10). The *ee* was determined by HPLC with a chiral OD-H column ($\lambda = 220$ nm; eluent: *n*-hexane/*i*PrOH 70:30; flow rate: 0.8 mL/min; $t_{major} = 10.306$ min, $t_{minor} = 24.835$ min; *ee* = 92 %).

(S)-*N*-[2-Methoxycarbonyl-1-(4'-trifluoromethylphenyl)allyl]phthalimide (3n): White solid; m.p. 80.2–81.0 °C. $[α]_D^{20} = +77.2$ (c = 0.5, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.83-7.85$ (m, 2 H), 7.71– 7.73 (m, 2 H), 7.58 (dd, J = 18.8, 8.4 Hz, 4 H), 6.60 (s, 1 H), 6.47 (s, 1 H), 5.64 (s, 1 H), 3.72 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 167.6$ (2 C), 165.7, 140.9, 136.9, 134.3 (2 C), 131.7, 129.7 (2 C), 129.1 (3 C), 125.6 (3 C), 123.6 (2 C), 54.3, 52.5 ppm. MS (EI): m/z (%) = 389 (4), 328 (100), 300 (51), 104 (76), 76 (47). HRMS (ESI): calcd. for C₂₀H₁₄F₃NNaO₄ [M + Na]⁺ 412.0767; found 412.0769. The *ee* was determined by HPLC with a chiral OD-H column (λ = 222 nm; eluent: *n*-hexane/*i*PrOH 85:15; flow rate: 0.8 mL/min; $t_{major} = 9.544$ min, $t_{minor} = 10.887$ min; *ee* = 89 %).

(*R*)-*N*-[2-Methoxycarbonyl-1-(2,4-dichlorophenyl)allyl]-phthalimide (30): White solid; m.p. 105.1–106.0 °C. $[\alpha]_D^{20} = -52.6$ (c = 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.83-7.85$ (m, 2 H), 7.72–7.74 (m, 2 H), 7.46 (d, J = 8.4 Hz, 1 H), 7.39 (d, J = 2.4 Hz, 1 H), 7.23–7.25 (m, 1 H), 6.70 (s, 1 H), 6.59 (d, J = 1.2 Hz, 1 H), 5.64 (d, J = 1.6 Hz, 1 H), 3.72 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 166.9$ (2 C), 164.8, 135.7, 134.4, 134.1, 133.9 (2 C), 132.6, 131.2 (2 C), 131.0, 129.2, 128.4, 126.7, 123.3 (2 C), 52.7, 51.9 ppm. MS (EI): m/z (%) = 355 (75), 266 (40), 206 (11), 73 (100), 59 (5). HRMS (ES1): calcd. for C₁₉H₁₄Cl₂NO₄ [M + H]⁺ 390.0294; found 390.0284. The *ee* was determined by HPLC with a chiral OD-H column ($\lambda = 220$ nm; eluent: *n*-hexane/*i*PrOH 85:15; flow rate: 0.8 mL/min; $t_{major} = 11.435$ min, $t_{minor} = 23.534$ min; *ee* = 94 %).

(*S*)-*N*-[(2-Methoxycarbonyl-1-naphthyl)allyl]phthalimide (3p): Colorless oil. $[\alpha]_D^{20} = +52.7 (c = 0.4, CHCl_3)$. ¹H NMR (400 MHz, CDCl_3): $\delta = 7.98 (d, J = 8.4 Hz, 1 H), 7.82-7.88 (m, 4 H), 7.69-7.72 (m, 2 H),$ 7.64 (d, J = 7.2 Hz, 1 H), 7.44-7.55 (m, 3 H), 7.23 (s, 1 H), 6.59 (d, <math>J = 1.44 + 1.44



0.8 Hz, 1 H), 5.63 (d, J = 1.6 Hz, 1 H), 3.72 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 167.9$ (2 C), 166.0, 137.1, 134.1 (2 C), 133.8, 132.0, 131.7, 130.9, 129.4 (2 C), 129.0 (2 C), 126.9, 126.8, 125.7, 125.1, 123.5 (2 C), 123.1, 52.5, 51.5 ppm. MS (EI): m/z (%) = 355 (3), 327 (6), 207 (100), 165 (87), 104 (25). HRMS (ESI): calcd. for C₂₃H₁₇NNaO₄ [M + Na]⁺ 394.1050; found 394.1048. The *ee* was determined by HPLC with a chiral AD-H column ($\lambda = 220$ nm; eluent: *n*-hexane/*i*PrOH 90:10; flow rate: 1.0 mL/min; $t_{major} = 25.194$ min, $t_{minor} = 23.630$ min; *ee* = 78 %).

Supporting Information (see footnote on the first page of this article): Spectroscopic data and chiral HPLC traces of the compounds shown in Tables 1 and 2, X-ray crystallographic data for **30**, and detailed descriptions of the experimental procedures.

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