# Synthesis and characterization of trans-di-(4-pyridyl) porphyrin dimers 

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#### Abstract

Preparation and characterization of a small library of symmetric trans-di(4-pyridyl) porphyrin dimers, obtained by either Glaser-Hay or Sonogashira coupling reactions from appropriately prepared trans-di-4-pyridylporphyrin precursors, is presented. The porphyrin dimers are differentiated by a phenyl-alkynyl bridge of increasing length at one meso-position, while for all the derivatives the two remaining opposite meso-positions are tailored with a phenyl moiety bearing a short polyether chain. Coordination of the four pyridyl groups with appropriate metal fragments may be exploited to construct tubular hollow structures, with varied internal sizes, depending on the choice of the porphyrin dimer component.


KEYWORDS: trans-di-(4-pyridyl)porphyrin, porphyrin dimers, synthesis, metal-mediated selfassembling.

## INTRODUCTION

Pyridylporphyrins are essential elements in the modern molecular tool-box of the supramolecular chemist. They combine the peculiar structural, optical and redox properties of porphyrins with the ability to act as rigid, flat multitopic donors in the formation of metal-organic complexes [1]. Within the field of the metal-mediated directional bonding approach [2], these features along with the properties of several transition metal ions (e.g. $\operatorname{Pd}(\mathrm{II}), \mathrm{Pt}(\mathrm{II}), \mathrm{Ru}(\mathrm{II}), \operatorname{Rh}(\mathrm{II}), \operatorname{Re}(\mathrm{I})$ ) have led to a variety of coordination adducts. These species can have many different topologies (such as rhomboids, squares, rectangles, triangles) and interesting potential applications in the fields of optoelectronic, catalysis, molecular recognition, etc. [3].

In this context, we have recently started to investigate the interaction of pyridylporphyrin based metallacycles with phospholipid membranes and, in particular, their ability to modify the membrane permeability forming

[^0]large self-assembled channels [4]. Following the approach developed in the group of J. T. Hupp [5], we prepared $4+4$ $\operatorname{Re}(\mathrm{I})$-porphyrin metallacycles starting from $\operatorname{Re}(\mathrm{CO})_{5} \mathrm{Br}$ and trans- $\mathrm{A}_{2} \mathrm{~B}_{2}$ di-4-pyridylporphyrins bearing, in the two meso-positions, different types of aromatic substituents [6]. When peripheral carboxylic acid functionalized porphyrins were used, the resulting metallacycle showed a very interesting ionophoric activity that was attributed to its ability to assemble dimeric structures long enough to span the entire phospholipid bilayer, thus forming a trans-membrane nanopore [7]. These results encouraged us to further develop our porphyrin ligands, and the new molecules were designed in order to be able to form unimolecular tubular structures long enough to span the double-layer of a phospholipid membrane. Our target was to synthesize dimeric trans-di-4-pyridylporphyrins (Fig. 1), which would ideally bind to $90^{\circ}$ metal fragments in a 1:2 ratio thus forming $4+8$ parallelepiped-shaped hollow structures with two hydrophilic ends (see also Fig. 7). While dimeric porphyrins molecules are well known [8], examples of dimeric trans-di-4-pyridylporphyrins are very few as well as are the studies on their metal-mediated self-assembling behavior [9]. In this paper we report the


Fig. 1. Structures of the trans-di-(4-pyridyl)porphyrin dimers prepared in this work
synthesis of dimers 1a-1c (Fig. 1), their characterization and some preliminary experiments on their coordination abilities towards $\operatorname{Re}(\mathrm{I})$ and $\operatorname{Pd}(\mathrm{II}) 90^{\circ}$ metal fragments.

## RESULTS AND DISCUSSION

## Synthesis of the trans-di-4-pyridylporphyrin dimers

The structures of the target trans-di-4-pyridylporphyrin dimers are shown in Fig. 1. They are symmetric dimers in which two trans-di-4-pyridylporphyrins are connected at the meso-position with a phenyl-alkynyl bridge of increasing length. This kind of linking bridge has been chosen in order to ensure rigidity and linearity to the whole molecule, thus avoiding bent conformations. It also granted rather simple synthetic pathways through standard Glaser-Hay or Sonogashira coupling reactions. Moreover, for a better solubility in polar solvents and membrane compatibility, the dimers have been functionalized on the two opposite meso-positions with phenyl rings bearing a short polyether chain. In the linear conformation, the overall length of dimers $\mathbf{1 a - 1 c}$, estimated from molecular modelling and excluding the polyether chains, is about 36, 38 , and $43 \AA$, respectively, sufficient in each case to span a phospholipid bilayer, which is about $40 \AA$ thick.

The key intermediates in the synthesis of the tetrapyridylporphyrin dimers are 5,15-bis-(4-pyridyl)-10-(4-ethynylphenyl)-20-(4-[2-[2-(2-methoxyethoxy)ethoxy] ethoxy]phenyl)porphyrin (2a), and 5,15-bis-(4-pyridyl)-10-(4-iodophenyl)-20-(4-[2-[2-(2-methoxyethoxy)ethoxy] ethoxy]phenyl)porphyrin (3a) (Fig. 2).


2a: $R_{1}=-\mathbf{C} \equiv \mathbf{C}-\mathbf{H} ; \mathrm{R}_{2}=\left(\mathrm{OCH}_{2} \mathrm{CH}_{2}\right)_{3} \mathrm{OCH}_{3}$
2b: $R_{1}=\mathrm{R}_{2}=-\mathbf{C} \equiv \mathbf{C}-\mathbf{H}$
2c: $R_{1}=R_{2}=\left(\mathrm{OCH}_{2} \mathrm{CH}_{2}\right)_{3} \mathrm{OCH}_{3}$
3a: $R_{1}=1 ; \mathrm{R}_{2}=\left(\mathrm{OCH}_{2} \mathrm{CH}_{2}\right)_{3} \mathrm{OCH}_{3}$
3b: $R_{1}=R_{2}=1$
Fig. 2. Structures of the trans-di-(4-pyridyl)porphyrin monomers prepared in this work

Porphyrins 2a and 3a are $\mathrm{A}_{2} \mathbf{B C}$ trans meso-substituted derivatives, which can be conveniently obtained by a modification of the Lindsey's approach [10], in which the 5-(4-pyridyl)dipyrromethane is reacted with half an equivalent of two different aldehydes (Scheme 1). This is a statistical approach, leading to the $\mathrm{A}_{2} \mathrm{BC}$ porphyrin together with the two symmetric trans products $\left(\mathrm{A}_{2} \mathrm{~B}_{2}\right.$ and $\left.\mathrm{A}_{2} \mathrm{C}_{2}\right)$. Notably, if the two aldehydes have comparable reactivity the desired $\mathrm{A}_{2} \mathrm{BC}$ isomers are obtained, in predominant yield. In the case of the synthesis of 2a 5-(4-pyridyl)


Scheme 1. Synthetic route to porphyrins 2a-2c


Scheme 2. Synthesis of dimer 1b
dipyrromethane was reacted with 4-[(trimethylsilyl) ethynyl]benzaldehyde and 4-[2-[2-(2-methoxyethoxy) ethoxy]ethoxy]benzaldehyde (Scheme 1) in a molar ratio 1:0.5:0.5. The latter aldehyde, commercially available but exceedingly expensive, was in turn obtained by a standard Mitsunobu reaction between 4-hydroxybenzaldehyde and triethylenglycolmonomethylether. The macrocyclization reaction was performed in $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ at $0^{\circ} \mathrm{C}$ and under Ar atmosphere using TFA as the acidic catalyst. After oxidation with DDQ in air, the crude product was neutralized and treated with tetrabutylammonium fluoride (TBAF) to remove the silyl protecting groups. Repeated purification steps by silica chromatography and crystallization from $\mathrm{CHCl}_{3} / \mathrm{MeOH}$ afforded the three porphyrins in $12 \%$ (2a), $7 \%$ (2b) and $3 \%$ (2c) yield. Starting from 4-iodobenzaldehyde, and following a similar scheme of reactions, porphyrins $\mathbf{3 a}, \mathbf{3 b}$ and $\mathbf{2 c}$ were obtained in $17 \%, 8 \%$, and $5 \%$ yield, respectively.

All the porphyrins were fully characterized by ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR, ESI-MS, IR, UV-vis and emission spectroscopies (see also Figs S1-S10 and Figs S25-S26 of the Supplementary material).

Porphyrins 2a and 3a were then used for the preparation of dimers $\mathbf{1}$. Dimer 1b was obtained in good yield via Glaser-Hay homocoupling of 2a (Scheme 2). The reaction proceeded smoothly at room temperature in the presence of CuCl , tetramethylethylenediamine (TMEDA) and under a positive pressure of air giving, after column chromatography purification, the desired product in good yield.

Dimers 1a and 1c were likewise obtained by Sonogashira coupling between 2a and 3a and between 3a and 1,4-diethynylbenzene, respectively (Scheme 3). The copper-free variation of the Sonogashira coupling reaction was used in order to avoid possible undesired porphyrin-metalation side-products. The coupling



Scheme 3. Synthesis of dimers 1a and 1c
proceeds in the presence of $\mathrm{Pd}\left(\mathrm{PPh}_{3}\right)_{4}$ and a large excess of $\mathrm{NEt}_{3}$ in a THF/DMF mixture, under Ar atmosphere and under microwave irradiation for 1 h , at $120^{\circ} \mathrm{C}$. After purification, dimers $\mathbf{1 a}$ and $\mathbf{1 c}$ were obtained in $24 \%$ and $62 \%$ yield, respectively.

## Characterization of the trans-di-4-pyridylporphyrin dimers

All the porphyrin dimers were fully characterized by ESI-MS, ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR, IR, UV-vis and emission spectroscopies, with the mass spectra unambiguously identifying the nature of each product. The ${ }^{1} \mathrm{H}$ NMR spectra (assigned by 2D H-H and H-C NMR experiments, see also Supplementary material) of the three porphyrin dimers closely resemble that of the reference monomer porphyrin 2a, reflecting the symmetry of the molecules. In particular, the upfield and the midfield regions of the reference porphyrin and the dimers, comprising the resonances of the NH core protons and of polyether chains, respectively, are practically superimposable (see Fig. S17 in the Supplementary material). Small but significant differences are however observed in the downfield region of the spectra (Fig. 3).

With respect to $2 \mathbf{a}$, the pattern of the $\beta$-pyrrolic protons ( $\beta-\mathrm{H}$, Fig. 3) in the dimers is, in general, more resolved and the proton resonances of the phenyl ring bearing the ethynyl substituents $\left(\mathrm{H}_{3}\right.$ - and $\mathrm{H}_{4}-\mathrm{Ph}$, Fig. 3) are downfield shifted, to different extents depending on the dimer. As expected, due to the long distance from the linkers, the peaks corresponding to the pyridyl protons ( $\mathrm{H}_{\alpha^{-}}$and $\mathrm{H}_{\beta^{-}}-\mathrm{Py}$ ) and to the phenyl rings bearing the polyether chain ( $\mathrm{H}_{1^{-}}$and $\mathrm{H}_{2}-\mathrm{Ph}$ ) are, on the contrary, essentially unchanged. A spectroscopic signature of
dimer 1c is the singlet at $c a .7 .75 \mathrm{ppm}$. which can be assigned to the protons of the central phenyl ring of the linker ( $\mathrm{H}_{5}-\mathrm{Ph}$, Fig. 3), lying on the center of symmetry of the molecule. Similarly to the ${ }^{1} \mathrm{H}$ NMR analysis, the ${ }^{13} \mathrm{C}$ NMR spectra of the various dimers are also very similar to that of porphyrin 2a, except for the number and position of the ethynyl carbon signals, together with the presence, for dimer $\mathbf{1 c}$, of two peaks around 135 ppm , assigned to the phenyl carbons of the linker (see Fig. S18 of the Supplementary material).

To better characterize the dimers, their diffusion coefficients were determined using ${ }^{1} \mathrm{H}$-DOSY NMR experiments. Figure 4 shows the ${ }^{1} \mathrm{H}$-DOSY spectra for dimer 1a and, for comparison, porphyrin 2c. Similar 2D-maps were also obtained for dimers $\mathbf{1 b}$ and $\mathbf{1 c}$ and are reported in the Supplementary material.

Inspection of the 2D-map of Fig. 4 shows that all the signals of the two molecules, except the solvent and water residues, are aligned along one single value of diffusion coefficient indicating the presence in solution of a single species with a well-defined dimension. Moreover, the porphyrin monomer has a higher diffusion coefficient than the dimer, in agreement with the latter species having a bigger size. The diffusion coefficients for all these species together with their hydrodynamic diameters obtained from the Stokes-Einstein equation, while not being meaningful in terms of absolute values, are consistent with the expected increased dimension going from monomer 2a to dimers $\mathbf{1}$ (see Table 1 in the Supplementary material).

Finally, the absorption and emission spectroscopic features of the three dimers were investigated. Figure 5 reports the UV-vis spectra of the dimers ( $1 \mu \mathrm{M}$ in DCM ) in comparison with porphyrin $\mathbf{2 a}$ ( $2 \mu \mathrm{M}$ in DCM). The


Fig. 3. Expansion of the downfield region of the ${ }^{1} \mathrm{H}-\mathrm{NMR}$ spectrum $\left(\mathrm{CDCl}_{3}\right)$ of the reference monomer porphyrin 2a, and of the dimers $\mathbf{1 a} \mathbf{- 1} \mathbf{c}$; the asterisk in the spectrum of $\mathbf{1 b}$ indicates a chloroform satellite


Fig. 4. ${ }^{1} \mathrm{H}$-DOSY ( $500 \mathrm{MHz} ; \mathrm{CDCl}_{3}, 298 \mathrm{~K}$ ) of porphyrin $\mathbf{2 c}$, and of the dimer $\mathbf{1 a}$. The 1 D trace of $\mathbf{1 a}$ is also shown

Soret band for the dimers is only slightly red-shifted $(2-3 \mathrm{~nm})$ with respect to the one of 2a, while the position of the Q-bands is almost unaffected. The intensity of the absorption bands changes substantially among
the series. Dimer 1b, which is the exact double of the porphyrin monomer, has practically the same molar absorptivity of 2a, while the intensity of the Soret and of the Q -bands decreases along the series from $\mathbf{1 a}$ to $\mathbf{1 c}$.


Fig. 5. UV-vis spectra $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right)$ of the reference porphyrin $\mathbf{2 a}(2 \mu \mathrm{M})$, and of the dimers $\mathbf{1 a} \mathbf{- 1 \mathbf { c }}(1 \mu \mathrm{M}): \lambda_{\max }, \mathrm{nm} 419$ (2a), 423(1a), 423 (1b), 421 (1c)


Fig. 6. Fluorescence emission spectra $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}, \lambda_{\text {exc }}=420 \mathrm{~nm}\right)$ of the reference porphyrin $\mathbf{2 a}(2 \mu \mathrm{M})$, and of the dimers $\mathbf{1 a} \mathbf{-} \mathbf{1 c}$ $(1 \mu \mathrm{M}): \lambda_{\text {em }}, \mathrm{nm} 652,718$ (2a); 654, 718 (1a); 655, 718 (1b); 655, 719 (1c)

Clearly the substitution with two phenyl ring in 1a and the presence of the central phenyl in $\mathbf{1 c}$ has a negative effect on the conjugation between the ethynyl group and the porphyrin aromatic macrocycle, resulting in a lower molar absorptivity.

Figure 6 reports the emission spectra of the dimers 1 $(1 \mu \mathrm{M})$ and of porphyrin $\mathbf{2 a}(2 \mu \mathrm{M})$ recorded in DCM with the excitation wavelength fixed at 420 nm . All the spectra are almost superimposable, both in terms of intensity and position of the emission bands, as might be expected.

## Preliminary studies for the preparation of $4+8$ trans-di-4-pyridylporphyrin metallacyclic assemblies

Preliminary investigations towards the preparation of $4+8$ trans-di-4-pyridylporphyrin metallacyclic assemblies
with a parallelepiped shape were performed using dimer 1b in combination with two different $90^{\circ}$ metal fragments. Despite the rotational freedom of the two macrocycles around the phenyl-alkynyl linker, we envisaged the possibility of a thermodynamic self-sorting process, leading to the successful obtainment of metallacyclic assemblies as unique products (Fig. 7).

We first investigated the use of the $\left[\mathrm{Pd}(\mathrm{dppp})(\mathrm{OTf})_{2}\right]$ metal complex (dppp $=1,3$-bis(diphenylphosphino)propane, $\mathrm{OTf}=$ trifluorosulphonate), which is known to readily form soluble $4+4$ metallacycles with trans-di-4pyridylporphyrins [11]. The solubility in both polar and apolar organic solvents is normally ascribed both to the positive charge of the metal complex, due to the loss of the two triflate anions upon coordination of two pyridyl groups, and to the out-of-plane phenyl substituents of the


Legend:


Fig. 7. Schematic depiction of the intended metallacyclic assemblies with a parallelepiped shape, in which four dimers 1b are coordinated via the peripheral pyridyl groups to eight $90^{\circ}$ metal fragments
dppp ancillary ligand disfavoring possible aggregation of the resulting assemblies. Also, the two phosphorus atoms of the ancillary dppp ligand are a useful tag for ${ }^{31} \mathrm{P}$ NMR investigations. However, simply mixing dimer 1b with two equivalents of $\left[\mathrm{Pd}(\mathrm{dppp})(\mathrm{OTf})_{2}\right]$ in $\mathrm{CDCl}_{3}$ directly in the NMR tube afforded very complex ${ }^{1} \mathrm{H}$ and ${ }^{31} \mathrm{P}$ NMR spectra suggesting the presence of several species, possibly in dynamic equilibrium. Heating and/or increasing of the building blocks concentrations, did not change or simplify the overall pattern of the NMR spectra.

A second preliminary attempt to prepare metallacyclic parallelepiped assemblies was made using $\left[\mathrm{Re}(\mathrm{CO})_{5} \mathrm{Br}\right]$ as metal fragment. Compared to $\operatorname{Pd}(\mathrm{II}), \operatorname{Re}(\mathrm{I})$ is known to form kinetically and thermodynamically more stable bonds with pyridyl ligands, leading to the formation of stable discrete cyclic structures. The chemistry of the $\left[\operatorname{Re}(\mathrm{CO})_{5} \mathrm{X}\right]$ ( $\mathrm{X}=\mathrm{Cl}^{-}$or $\mathrm{Br}^{-}$) complex in the formation of stable $4+4$ metallacycles with trans-di-4-pyridylporphyrins has been pioneered by J.T. Hupp [5,12]. The strong labilizing effect of CO serves to activate two (and only two) cis-carbonyls for substitution, presumably first by solvent molecules (e.g. tetrahydrofuran) and then by pyridyl ligands. The reaction is slow and can take up to 2 days for completion under refluxing conditions. We therefore reacted $\mathbf{1 b}$ and $\left[\operatorname{Re}(\mathrm{CO})_{5} \mathrm{Br}\right]$ in a mixture of THF and toluene under reflux, until the porphyrin starting material was fully consumed. Work-up of the reaction afforded a solid
which was scarcely soluble in a variety of solvents. The ${ }^{1} \mathrm{H}$ NMR spectra of this solid in pyridine- $d_{5}$ were difficult to interpret, although a DOSY experiment suggested the presence of a single species in solution (Fig. S20 of the Supplementary material) [13]. Once again, variation of the temperature did not lead to any simplification of the NMR spectrum, which would be expected if a highly symmetrical assembly had formed. Most likely, the very low solubility of reagents and intermediates hampers the insaturation of a fast thermodynamic equilibrium, that should lead to the desired metallacycle, and consequently the porphyrin dimer gets consumed in the formation of undesired oligomeric coordination open-species. The difficulty to idetify a single $4+8$ supramolecular adduct with a di-4-pyridylporphyrin dimer and a $\operatorname{Re}(\mathrm{I})$ complex when the porphyrin dimer bears a triphenyl bridge has been also reported by Hupp et al. and was attributed to unacceptably slow conversion of "wrong" kinetic intermediate products [9b].

## EXPERIMENTAL

## General

All commercially available reagents were purchased from Aldrich, Fluka and Strem Chemicals and used without purification unless otherwise mentioned. Solvents were purchased from Aldrich, VWR, Fluka and Riedel, and deuterated solvents from Cambridge Isotope Laboratories and Aldrich. Analytical thin layer chromatography (TLC) was carried out on Merck aluminum backed silica gel plates (thickness 0.25 mm ). Flash column chromatography (FCC) was carried out on Merck silica gel 60 (230-400 Mesh).

NMR spectra were recorded on a Varian 500 MHz spectrometer (operating at 500 MHz for ${ }^{1} \mathrm{H}$ and at 125 MHz for ${ }^{13} \mathrm{C}$ ). Chemical shifts are reported as parts per million ( ppm ) relative to the solvent residual signal as internal reference. $\left[\mathrm{CDCl}_{3}: \delta_{\mathrm{H}}, \mathrm{ppm} 7.27 ; \delta_{\mathrm{C}}, \mathrm{ppm} 77.36\right.$; $\mathrm{CD}_{3} \mathrm{OD}: \delta_{\mathrm{H}}, \mathrm{ppm} 3.31 ; \delta_{\mathrm{C}}, \mathrm{ppm} 49.00 ;$ pyridine- $d_{5}$ : $\left.\delta_{\mathrm{H}}, \mathrm{ppm} 8.74 ; \delta_{\mathrm{C}}, \mathrm{ppm} 150.30\right]$.

IR spectra were recorded on a Perkin Elmer System 2000 NIR spectrophotometer with the KBr pellet technique and only major peaks are reported. UV-vis spectra were recorded on a Perkin Elmer Lambda 35 spectrophotometer. Fluorescence emission spectra were recorded on a Varian Cary Eclypse spectrofluorimeter. Electrospray ionization mass spectra (ESI-MS) were performed on a Perkin Elmer APII at 5600 eV .

1,4-diethynylbenzene was purchased from Aldrich. 5-(4-Pyridyl)dipyrromethane [14], $\left[\operatorname{ReBr}(\mathrm{CO})_{5}\right]$ [15] and $\left[\mathrm{Pd}(\mathrm{dppp})(\mathrm{OTf})_{2}\right][16]$ were prepared as reported in the literature.

Abbreviations used in the text: AcOEt = ethyl acetate; $\mathrm{DCM}=$ dichloromethane; $\mathrm{PE}=$ petroleum ether; $\mathrm{Py}=$ pyridine; TBAF $=$ tetra- $n$-butylammonium fluoride;

THF $=$ tetrahydrofurane; $n$ - $\mathrm{Hx}=n$-hexane; $\mathrm{DMF}=$ dimethylformamide; TFA = trifluoroacetic acid; $\mathrm{DDQ}=$ 2,3-dichloro-5,6-dicyano-1,4-benzoquinone; DIAD = diisopropylazodicarboxylate; dppp $=$ 1,3-bis-(diphenylphosphino)propane; TMEDA $=$ tetramethylethylenediamine; TMSA = trimethylsilylacetylene; dba = tris-dibenzylideneacetone; tol = tolyl; $\mu$ Wave $=$ microwave $;$ TLC $=$ thin layer chromatography; $\mathrm{FCC}=$ flash column chromatography; $\mathrm{CC}=$ column chromatography.

## Synthesis of $\mathrm{A}_{2} \mathrm{BC}$ porphyrins 2a-2c and 3a-3c

4-[2-[2-(2-methoxyethoxy)ethoxy]ethoxy]benzaldehyde (TegPhCHO) [8c]. Triphenylphosphine (5.5 g, 21.0 mmol ) was dissolved in 70 mL of anhydrous THF under Ar atmosphere and the solution was cooled to $0^{\circ} \mathrm{C}$. DIAD ( $2.0 \mathrm{~mL}, \mathrm{~d}=1.420 \mathrm{~g} / \mathrm{mL}, 14.0 \mathrm{mmol}$ ) was then added dropwise. The solution became pale yellow and a fine white precipitate was formed. Then a solution of triethylenglycolmonomethylether (THF solution, 2.20 mL , $\mathrm{d}=1.048 \mathrm{~g} / \mathrm{mL}, 14.0 \mathrm{mmol}$ ) and of $p$-hydroxybenzaldehyde $(1.71 \mathrm{~g}, 14.0 \mathrm{mmol})$ in anhydrous THF was added dropwise. The reaction mixture was stirred under inert atmosphere for 4 h at room temperature. The solvent was removed under vacuum and the mixture was purified by $\mathrm{FCC}(\mathrm{AcOEt} / n-\mathrm{Hx}$ from $2 / 3$ to $1 / 1 \mathrm{v} / \mathrm{v}$ ) giving a pale yellow oil. Yield 2.24 g (60\%). $R_{f}=0.26\left(\mathrm{SiO}_{2}, \mathrm{AcOEt} / \mathrm{n}-\mathrm{Hx} 3 / 2 \mathrm{v} / \mathrm{v}\right) .{ }^{1} \mathrm{H}$ NMR ( $\left.500 \mathrm{MHz} ; \mathrm{CD}_{3} \mathrm{OD}\right)$ : $\delta_{\mathrm{H}}, \mathrm{ppm} 9.82(1 \mathrm{H}, \mathrm{s}, \mathrm{CHO}), 7.83(2 \mathrm{H}$, d, $\left.J=9.0 \mathrm{~Hz}, \mathrm{H}_{1}-\mathrm{Ph}\right), 7.07\left(2 \mathrm{H}, \mathrm{d}, J=8.5 \mathrm{~Hz} \mathrm{H}_{2}-\mathrm{Ph}\right), 4.20$ $\left(2 \mathrm{H}, \mathrm{m}, \mathrm{PhOCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.84\left(2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.68$ ( $2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}$ ), 3.61-3.59 (4H, s, $\mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}$ ), 3.50 $\left(2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{OCH}_{3}\right), 3.32\left(3 \mathrm{H}, \mathrm{s}, \mathrm{OCH}_{3}\right) .{ }^{13} \mathrm{C}$ NMR (125 MHz; $\left.\mathrm{CD}_{3} \mathrm{OD}\right): \delta_{\mathrm{C}}$, ppm 191.3, 164.1, 131.7, 130.0, 114.7, 71.5, 70.4, 70.2, 70.0, 69.2, 67.7, 57.7. MS (ESI): $m / z$ 269, 291 (calcd. for $\left[\mathrm{C}_{14} \mathrm{H}_{20} \mathrm{O}_{5}+\mathrm{H}\right]^{+} 269.13,\left[\mathrm{C}_{14} \mathrm{H}_{20} \mathrm{O}_{5}+\right.$ $\mathrm{Na}]^{+}$291.12).

Porphyrins 2a-2c. The reaction was conducted under Ar atmosphere and in the dark. 5-(4-Pyridyl) dipyrromethane ( 580 mg , MW $=223.27,2.60 \mathrm{mmol}$ ) and 4-[2-(trimethylsilyl)ethynyl]benzaldehyde ( 263 mg , MW $=202.32,1.30 \mathrm{mmol}$ ) were dissolved in anhydrous DCM ( 270 mL ). A solution of $\mathrm{TegPhCHO}(349 \mathrm{mg}, \mathrm{MW}=$ 268.306, 1.30 mmol ) in 30 mL of anhydrous DCM was added and the mixture was cooled to $0^{\circ} \mathrm{C}$ with an ice bath. Then TFA ( $5.99 \mathrm{~mL}, \mathrm{~d}=1.535 \mathrm{~g} / \mathrm{mL}, 80.6 \mathrm{mmol}$ ) was added dropwise and the reaction kept under stirring for 1.5 h . The reaction mixture was then let to warm up to room temperature in open air, DDQ ( $885 \mathrm{mg}, 3.90 \mathrm{mmol}$ ) was added and the reaction was stirred for further 2 h . The crude mixture was directly washed three times with a saturated solution of $\mathrm{NaHCO}_{3}$ and once with water. The organic phase was dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$, filtered, and the solvent was removed under vacuum. The crude mixture was re-dissolved in 250 mL of DCM and TBAF ( 1.0 M in THF) $(26 \mathrm{~mL}, \mathrm{~d}=0.903 \mathrm{~g} / \mathrm{mL}, 26.0 \mathrm{mmol})$ was added. After stirring for 2.5 h at room temperature, the reaction mixture was washed twice with water and the organic
phase was dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$ and filtered. The solvent was removed under vacuum and the mixture was purified by CC on silica gel $\left(\mathrm{CHCl}_{3} / \mathrm{EtOH}\right.$ from $100 / 0$ to $97 / 3$ $\mathrm{v} / \mathrm{v})$. The fractions containing the three porphyrins were further purified by another CC on silica gel $\left(\mathrm{CHCl}_{3} / \mathrm{EtOH}\right.$ from 100/0 to 98/2). Each product was then obtained as a violet solid by precipitation from $\mathrm{CHCl}_{3} / \mathrm{MeOH}$. Yield: 2a $12 \%$, 2b $7 \%$, 2c $3 \%$. Total porphyrins yield $22.0 \%$.

5,15-bis-(4-pyridyl)-10-(4-ethynylphenyl)-20-(4-[2-[2-(2-methoxyethoxy)ethoxy]ethoxy]phenyl) porphyrin (2a). $R_{f}=0.53\left(\mathrm{SiO}_{2}, \mathrm{CHCl}_{3} / \mathrm{MeOH} 97 / 3\right.$ $\mathrm{v} / \mathrm{v}) .{ }^{1} \mathrm{H}$ NMR ( $500 \mathrm{MHz} ; \mathrm{CDCl}_{3}$ ): $\delta_{\mathrm{H}}$, ppm $9.04(4 \mathrm{H}, \mathrm{d}$, $\left.J=5.6 \mathrm{~Hz}, \mathrm{H}_{\mathrm{a}}-\mathrm{Py}\right), 8.94(2 \mathrm{H}, \mathrm{d}, J=4.7 \mathrm{~Hz}, \beta-\mathrm{H}), 8.88(2 \mathrm{H}$, d, $J=4.7 \mathrm{~Hz}, \beta-\mathrm{H}), 8.82(4 \mathrm{H}, \mathrm{m}, \beta-\mathrm{H}), 8.17\left(6 \mathrm{H}, \mathrm{s}, \mathrm{H}_{\mathrm{b}}-\mathrm{Py}\right.$ and $\left.\mathrm{H}_{4}-\mathrm{Ph}\right), 8.10\left(2 \mathrm{H}, \mathrm{d}, J=8.5 \mathrm{~Hz}, \mathrm{H}_{1}-\mathrm{Ph}\right), 7.91(2 \mathrm{H}, \mathrm{d}$, $\left.J=8.0 \mathrm{~Hz}, \mathrm{H}_{3}-\mathrm{Ph}\right), 7.32\left(2 \mathrm{H}, \mathrm{d}, J=8.5 \mathrm{~Hz}, \mathrm{H}_{2}-\mathrm{Ph}\right), 4.44$ ( $2 \mathrm{H}, \mathrm{m}, \mathrm{PhOCH}_{2} \mathrm{CH}_{2} \mathrm{O}$ ), 4.07 ( $2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}$ ), 3.89 $\left(2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.80\left(2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.75$ $\left(2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.62\left(2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.43$ $\left(3 \mathrm{H}, \mathrm{s}, \mathrm{OCH}_{3}\right), 3.33(1 \mathrm{H}, \mathrm{s}, \mathrm{CCH}),-2.84(2 \mathrm{H}, \mathrm{br}, \mathrm{NH}) .{ }^{13} \mathrm{C}$ NMR (125 MHz; $\mathrm{CDCl}_{3}$ ): $\delta_{\mathrm{C}}, \mathrm{ppm} 159.0,150.3,148.5$, $142.4,135.7,134.6,134.2,130.8,129.5,122.1,121.3$, 119.7, 117.3, 113.2, 83.6, 78.7, 72.1, 71.1, 70.9, 70.8, 70.0, 67.9, 59.2. MS (ESI): $m / z$ 803, 825, 841 (calcd. for $\left[\mathrm{C}_{51} \mathrm{H}_{42} \mathrm{~N}_{6} \mathrm{O}_{4}+\mathrm{H}\right]^{+}$803.33, $\left[\mathrm{C}_{51} \mathrm{H}_{42} \mathrm{~N}_{6} \mathrm{O}_{4}+\mathrm{Na}\right]^{+}$825.31, $\left[\mathrm{C}_{51} \mathrm{H}_{42} \mathrm{~N}_{6} \mathrm{O}_{4}+\mathrm{K}\right]^{+}$841.29). UV-vis $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right): \lambda_{\text {max }}, \mathrm{nm}$ (relative intensity, \%) 419 (100), 515 (4.7), 549 (2.3), 590 (1.6), 649 (1.5). Fluorescence emission $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}, \lambda_{\text {exc }}\right.$ 420 nm ): $\lambda_{\text {em }}, \mathrm{nm} 652,718$. IR (KBr): $\mathrm{v}, \mathrm{cm}^{-1} 2923,2853$, 1637, 1384, 1281, 1248, 1109, 801.

5,15-bis-(4-pyridyl)-10,20-bis-(4-ethynylphenyl) porphyrin (2b). $R_{f}=0.47\left(\mathrm{SiO}_{2} \mathrm{CHCl}_{3} / \mathrm{MeOH} 97 / 3\right.$ v/v). ${ }^{1} \mathrm{H}$ NMR ( $500 \mathrm{MHz} ; \mathrm{CDCl}_{3}$ ): $\delta_{\mathrm{H}}, \mathrm{ppm} 9.05(4 \mathrm{H}, \mathrm{d}$, $\left.J=5.5 \mathrm{~Hz}, \mathrm{H}_{\mathrm{a}}-\mathrm{Py}\right), 8.89(4 \mathrm{H}, \mathrm{d}, J=4.7 \mathrm{~Hz}, \beta-\mathrm{H}), 8.83$ $(4 \mathrm{H}, \mathrm{d}, J=4.6 \mathrm{~Hz}, \beta-\mathrm{H}), 8.18\left(8 \mathrm{H}, \mathrm{m}, \mathrm{H}_{\mathrm{b}}-\mathrm{Py}\right.$ and $\left.\mathrm{H}_{\mathrm{a}}-\mathrm{Ph}\right)$, $7.91\left(4 \mathrm{H}, \mathrm{d}, J=8.0 \mathrm{~Hz}, \mathrm{H}_{\mathrm{b}}-\mathrm{Ph}\right), 3.34(2 \mathrm{H}, \mathrm{s}, \mathrm{CCH}),-2.86$ ( 2 H , br, NH). ${ }^{13} \mathrm{C}$ NMR ( $125 \mathrm{MHz} ; \mathrm{CDCl}_{3}$ ): $\delta_{\mathrm{C}}, \mathrm{ppm}$ $150.2,148.5,142.3,134.57,130.8,129.5,122.2,120.1$, 117.5, 83.6, 78.7, 77.1. MS (ESI): $m / z 665.2$ (calcd. for $\left[\mathrm{C}_{46} \mathrm{H}_{28} \mathrm{~N}_{6}+\mathrm{H}\right]^{+}$665.24). UV-vis $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right): \lambda_{\text {max }}$, nm (relative intensity, \%) 418 (100), 514 (4.8), 549 (2.0), 589 (1.6), 645 (1.0). Fluorescence emission $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}, \lambda_{\text {exc }}\right.$ $420 \mathrm{~nm}): \lambda_{\text {em }}, \mathrm{nm} 650,716$. IR (KBr): $v, \mathrm{~cm}^{-1} 2924$, 2854, 1641, 1592, 968, 800, 728.

5,15-bis-(4-pyridyl)-10,20-bis-(4-[2-[2-(2-methoxyethoxy)ethoxy]ethoxy]phenyl) porphyrin (2c). $R_{f}=$ $0.46\left(\mathrm{SiO}_{2}, \mathrm{CHCl}_{3} / \mathrm{MeOH} 95 / 5 \mathrm{v} / \mathrm{v}\right) .{ }^{1} \mathrm{H} \mathrm{NMR}(500 \mathrm{MHz}$; $\left.\mathrm{CDCl}_{3}\right): \delta_{\mathrm{H}}, \mathrm{ppm} 9.04\left(4 \mathrm{H}, \mathrm{dd}, J=6.0,1.5 \mathrm{~Hz}, \mathrm{H}_{\mathrm{a}}-\mathrm{Py}\right)$, $8.92(4 \mathrm{H}, \mathrm{d}, J=4.5 \mathrm{~Hz}, \beta-\mathrm{H}), 8.80(4 \mathrm{H}, \mathrm{d}, J=5.0 \mathrm{~Hz}$, $\beta-\mathrm{H}), 8.17\left(4 \mathrm{H}, \mathrm{dd}, J=6.0,2.0 \mathrm{~Hz}, \mathrm{H}_{\mathrm{b}}-\mathrm{Py}\right), 8.1(4 \mathrm{H}, \mathrm{d}, J=$ $\left.8.5 \mathrm{~Hz}, \mathrm{H}_{1}-\mathrm{Ph}\right), 7.32\left(4 \mathrm{H}, \mathrm{d}, J=8.5 \mathrm{~Hz}, \mathrm{H}_{2}-\mathrm{Ph}\right), 4.44(4 \mathrm{H}$, $\left.\mathrm{m}, \mathrm{PhOCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 4.07\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.89(4 \mathrm{H}$, $\left.\mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.80\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.75(4 \mathrm{H}, \mathrm{m}$, $\left.\mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.62\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{OCH}_{3}\right), 3.42(6 \mathrm{H}, \mathrm{s}$, $\left.\mathrm{OCH}_{3}\right),-2.83(2 \mathrm{H}, \mathrm{br}, \mathrm{NH}) .{ }^{13} \mathrm{C}$ NMR ( $125 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta_{\mathrm{C}}, \mathrm{ppm} 159.0,150.4,148.4,135.7,134.3,129.5,126.6$, $122.4,120.8,120.3,117.0,113.1,111.7,72.1,71.1$,
70.9, 70.8, 70.0, 67.9, 59.2. MS (ESI): m/z 963 (calcd. for $\left.\left[\mathrm{C}_{56} \mathrm{H}_{56} \mathrm{~N}_{6} \mathrm{O}_{8}+\mathrm{Na}\right]^{+} 963.40\right)$. UV-vis $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right)$ : $\lambda_{\text {max }}$, nm (relative intensity, \%) 419 (100), 516 (5.2), 550 (3.0), 592 (1.9), 651 (2.7). Fluorescence emission $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}, \lambda_{\text {exc }}\right.$ $426 \mathrm{~nm}): \lambda_{\text {em }}, \mathrm{nm} 655,719$. IR (KBr): $v, \mathrm{~cm}^{-1} 3078,2922$, 2860, 1637, 1593, 1449, 1404, 1384, 1351, 1286, 1246, 1175, 1108, 967, 788, 737.

Porphyrins 3a-3b. The reaction was conducted under Ar atmosphere and in the dark. 5-(4-pyridyl) dipyrromethane ( $594 \mathrm{mg}, \mathrm{MW}=223.27,2.66 \mathrm{mmol}$ ) and 4-iodobenzaldehyde ( $309 \mathrm{mg}, \mathrm{MW}=232.02,1.33 \mathrm{mmol}$ ) were dissolved in anhydrous DCM ( 270 mL ). A solution of TegPhCHO ( 381 mg , $\mathrm{MW}=268.306,1.33 \mathrm{mmol}$ ) in 30 mL of anhydrous DCM was added and the mixture was cooled to $0^{\circ} \mathrm{C}$ with an ice bath. Then $\mathrm{NEt}_{3}(6.12 \mathrm{~mL}$, $\mathrm{d}=1.535 \mathrm{~g} / \mathrm{mL}, 82.5 \mathrm{mmol}$ ) was added dropwise and the reaction was kept under stirring for 1.5 h . The reaction mixture was let to reach room temperature and with no more inert atmosphere DDQ ( $1.21 \mathrm{~g}, 5.32 \mathrm{mmol}$ ) was added and the reaction was stirred for further 2 h . The crude mixture was directly washed three times with a saturated solution of $\mathrm{NaHCO}_{3}$ and once with water. The organic phase was dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$ and the solvent was removed under reduced pressure. The residue was purified by CC on silica gel $\left(\mathrm{CHCl}_{3} / \mathrm{EtOH}\right.$ from $100 / 0$ to $97 / 3 \mathrm{v} / \mathrm{v})$. The fractions containing the three porphyrins were further purified by another CC on silica gel $\left(\mathrm{CHCl}_{3} /\right.$ EtOH from 100/0 to 98/2). Each product was then obtained as a violet solid by precipitation from $\mathrm{CHCl}_{3} /$ MeOH. Yield: 3a 17\%, 3b $8.0 \%$, 2c 5\%. Total porphyrins yield $30 \%$.

5,15-bis-(4-pyridyl)-10-(4-iodophenyl)-20-(4-[2-[2-(2-methoxyethoxy)ethoxy]ethoxy]phenyl)porphyrin (3a). $R_{f}=0.49\left(\mathrm{SiO}_{2}, \mathrm{CHCl}_{3} / \mathrm{MeOH} 97 / 3 \mathrm{v} / \mathrm{v}\right) .{ }^{1} \mathrm{H}$ NMR $\left(500 \mathrm{MHz} ; \mathrm{CDCl}_{3}\right): \delta_{\mathrm{H}}, \mathrm{ppm} 9.06(4 \mathrm{H}, \mathrm{d}, J=5.5 \mathrm{~Hz}$, $\left.\mathrm{H}_{\mathrm{a}}-\mathrm{Py}\right), 8.95(2 \mathrm{H}, \mathrm{d}, J=5.0 \mathrm{~Hz}, \beta-\mathrm{H}), 8.89(2 \mathrm{H}, \mathrm{d}, J=$ $4.5 \mathrm{~Hz}, \beta-\mathrm{H}), 8.82(4 \mathrm{H}, \mathrm{m}, \beta-\mathrm{H}), 8.18(4 \mathrm{H}, \mathrm{d}, J=5.5 \mathrm{~Hz}$, $\left.\mathrm{H}_{\mathrm{b}}-\mathrm{Py}\right), 8.12\left(4 \mathrm{H}, \mathrm{s}, \mathrm{H}_{1}-\mathrm{Ph}\right.$ and $\left.\mathrm{H}_{\mathrm{a}}-\mathrm{PhI}\right), 7.95(4 \mathrm{H}, \mathrm{d}$, $\left.J=8.0 \mathrm{~Hz}, \mathrm{H}_{\mathrm{b}}-\mathrm{Ph}\right), 7.33\left(4 \mathrm{H}, \mathrm{d}, J=9.0 \mathrm{~Hz}, \mathrm{H}_{2}-\mathrm{Ph}\right), 4.45$ $\left(2 \mathrm{H}, \mathrm{m}, \mathrm{PhOCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 4.08\left(2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.90$ $\left(2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.81\left(2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.76$ $\left(2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.64\left(2 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{OCH}_{3}\right)$, $3.44\left(3 \mathrm{H}, \mathrm{s}, \mathrm{OCH}_{3}\right),-2.85(2 \mathrm{H}, \mathrm{br}, \mathrm{NH}) .{ }^{13} \mathrm{C}$ NMR (125 $\left.\mathrm{MHz} ; \mathrm{CDCl}_{3}\right): \delta_{\mathrm{C}}, \mathrm{ppm} 158.9,150.2,148.3,141.3,136.1$, 136.0, 135.6, 134.1, 129.4, 121.2, 119.1, 117.1, 113.0, 94.4, 72.0, 71.0, 70.8, 70.7, 70.0, 67.8, 59.2. MS (ESI): $\mathrm{m} / \mathrm{z}$ 905, 927 (calcd. for $\left[\mathrm{C}_{49} \mathrm{H}_{41} \mathrm{IN}_{6} \mathrm{O}_{4}+\mathrm{H}\right]^{+} 905.23$, $\left.\left[\mathrm{C}_{49} \mathrm{H}_{41} \mathrm{IN}_{6} \mathrm{O}_{4}+\mathrm{Na}\right]^{+} 927.21\right)$. UV-vis spectrum $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right)$ : $\lambda_{\max }, \mathrm{nm}$ (relative intensity, \%) 419 (100), 515 (4.8), 550 (2.2), 591 (1.5), 651 (1.8). Fluorescence emission $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}, \lambda_{\text {exc }} 420 \mathrm{~nm}\right): \lambda_{\text {em }}, \mathrm{nm} 655,717$. IR (KBr): v, $\mathrm{cm}^{-1} 2922,2853,1636,1593,1472,1384,1283,1248$, 1105, 968, 799, 732.

5,15-bis-(4-pyridyl)-10,20-bis-(4-iodophenyl) porphyrin (3b) [17]. $R_{f}=0.67\left(\mathrm{SiO}_{2}, \mathrm{CHCl}_{3} / \mathrm{MeOH}\right.$ $97 / 3 \mathrm{v} / \mathrm{v}$ ). ${ }^{1} \mathrm{H}$ NMR ( $500 \mathrm{MHz} ; \mathrm{CDCl}_{3}$ ): $\delta_{\mathrm{H}}, \mathrm{ppm} 9.07$ $\left(4 \mathrm{H}, \mathrm{dd}, J=5.5,1.5 \mathrm{~Hz}, \mathrm{H}_{\mathrm{a}}-\mathrm{Py}\right), 8.90(4 \mathrm{H}, \mathrm{d}, J=4.5 \mathrm{~Hz}$,
$\beta-\mathrm{H}), 8.84(4 \mathrm{H}, \mathrm{d}, J=5.0 \mathrm{~Hz}, \beta-\mathrm{H}), 8.17(4 \mathrm{H}, \mathrm{dd}, J=6.0$, $\left.2.0 \mathrm{~Hz}, \mathrm{H}_{\mathrm{b}}-\mathrm{Py}\right), 8.13\left(4 \mathrm{H}, \mathrm{d}, J=8.0 \mathrm{~Hz}, \mathrm{H}_{\mathrm{a}}-\mathrm{PhI}\right), 7.95$ $\left(4 \mathrm{H}, \mathrm{d}, J=8.0 \mathrm{~Hz}, \mathrm{H}_{\mathrm{b}}-\mathrm{PhI}\right),-2.89(2 \mathrm{H}, \mathrm{br}, \mathrm{NH}) .{ }^{13} \mathrm{C}$ NMR (125 MHz; $\mathrm{CDCl}_{3}$ ): $\delta_{\mathrm{C}}$, ppm 150.0, 148.3, 141.1, 136.1, 136.0, 129.3, 119.5, 117.3, 94.5. MS (ESI): $m / z 869$ (calcd. for $\left[\mathrm{C}_{42} \mathrm{H}_{26} \mathrm{I}_{2} \mathrm{~N}_{6}+\mathrm{H}\right]^{+}$869.04). UV-vis $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right)$ : $\lambda_{\max }, \mathrm{nm}$ (relative intensity, \%) 418 (100), 514 (4.3), 548 (1.5), 590 (1.1), 650 (1.0). Fluorescence emission $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}, \lambda_{\text {exc }} 420 \mathrm{~nm}\right): \lambda_{\text {em }}, \mathrm{nm} 656,716$. IR (KBr): v, $\mathrm{cm}^{-1} 2921,1637,1591,14783,1384,967,796,783,725$.

## Synthesis of porphyrin dimers 1a-1c

Dimer 1a. Anhydrous THF, anhydrous DMF and $\mathrm{NEt}_{3}$ were degassed for 1 h with Ar . A solution of 2a ( $21 \mathrm{mg}, \mathrm{MW}=802.9,0.026 \mathrm{mmol}$ ) in 1 mL of anhydrous THF was prepared and kept under Ar atmosphere. Porphyrin 3a ( $21.7 \mathrm{mg}, \mathrm{MW}=904.79,0.024 \mathrm{mmol}$ ) was placed in the microwave reactor vessel and set under inert atmosphere. Then DMF $(1.2 \mathrm{~mL})$ and $\mathrm{NEt}_{3}(0.8 \mathrm{~mL})$ were added and the mixture was degassed for 15 min with Ar. Lastly, keeping the vessel under inert atmosphere, $\operatorname{Pd}\left(\mathrm{PPh}_{3}\right)_{4}(1.4 \mathrm{mg}, \mathrm{MW}=1154.56,0.0012 \mathrm{mmol})$ was added and the mixture was degassed for further 15 min. Keeping the vessel under Ar, the solution of $\mathbf{2 a}$ was added. The reaction was stirred for 1 h at $120^{\circ} \mathrm{C}$ under microwave irradiation (ramping time $=10 \mathrm{~min}$, $\mathrm{P}=300 \mathrm{~W}$ ). The crude mixture was passed through a very short Celite ${ }^{\circledR} 521$ pad washing with $\mathrm{CHCl}_{3} / \mathrm{MeOH}$ $9 / 1$. The organic solvent was then extracted twice with distilled water to remove the DMF, dried with $\mathrm{Na}_{2} \mathrm{SO}_{4}$ and finally the solvent was removed under reduced pressure. Purification of the crude by $\mathrm{CC}\left(\mathrm{CHCl}_{3} / \mathrm{EtOH}\right.$ from $98 / 2$ to $97: 3 \mathrm{v} / \mathrm{v}$ ) afforded a purple solid. Yield 9.1 mg ( $24 \%$ ). $R_{f}=0.43\left(\mathrm{SiO}_{2}, \mathrm{CHCl}_{3} / \mathrm{MeOH} 95 / 5 \mathrm{v} / \mathrm{v}\right) .{ }^{1} \mathrm{H}$ NMR $\left(500 \mathrm{MHz}, \mathrm{CDCl}_{3}\right): \delta_{\mathrm{H}}, \mathrm{ppm} 9.08(8 \mathrm{H}, \mathrm{d}, J=5.5 \mathrm{~Hz}$, $\left.\mathrm{H}_{\mathrm{a}}-\mathrm{Py}\right), 9.00(4 \mathrm{H}, \mathrm{d}, J=4.5 \mathrm{~Hz}, \beta-\mathrm{H}), 8.97(4 \mathrm{H}, \mathrm{d}$, $J=4.5 \mathrm{~Hz}, \beta-\mathrm{H}), 8.89(4 \mathrm{H}, \mathrm{d}, J=5.0 \mathrm{~Hz}, \beta-\mathrm{H}), 8.84$ $(4 \mathrm{H}, \mathrm{d}, J=5.0 \mathrm{~Hz}, \beta-\mathrm{H}), 8.31\left(4 \mathrm{H}, \mathrm{d}, J=8.0 \mathrm{~Hz}, \mathrm{H}_{4}-\mathrm{Ph}\right)$, $8.21\left(8 \mathrm{H}, \mathrm{dd}, J=6.0,3.0 \mathrm{~Hz}, \mathrm{H}_{\mathrm{b}}-\mathrm{Py}\right), 8.14-8.11(8 \mathrm{H}, \mathrm{m}$, $\mathrm{H}_{1}-\mathrm{Ph}$ and $\left.\mathrm{H}_{3}-\mathrm{Ph}\right), 7.35\left(4 \mathrm{H}, \mathrm{d}, J=8.5 \mathrm{~Hz}, \mathrm{H}_{2}-\mathrm{Ph}\right), 4.46$ $\left(4 \mathrm{H}, \mathrm{m}, \mathrm{PhOCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 4.09\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right)$, $3.91\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.82\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right)$, $3.77\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.65\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right)$, $3.44\left(6 \mathrm{H}, \mathrm{s}, \mathrm{OCH}_{3}\right),-2.80(4 \mathrm{H}, \mathrm{br}, \mathrm{NH}) .{ }^{13} \mathrm{C}$ NMR (125 $\mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta_{\mathrm{C}}, \mathrm{ppm} 159.3,150.6,148.7,142.4,135.9$, $135.0,134.4,130.6,130.1,129.8,123.4,121.5,120.1$, 117.5, 113.4, 91.0, 72.4, 71.3, 71.2, 71.0, 70.3, 68.1, 59.5. MS (ESI): $m / z 1577$ (calcd. for $\left[\mathrm{C}_{100} \mathrm{H}_{82} \mathrm{~N}_{12} \mathrm{O}_{8}-\mathrm{H}\right]^{-}$ 1577.63). UV-vis $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right): \lambda_{\text {max }}$, nm (relative intensity, \%) 423 (100), 516 (5.2), 551 (2.5), 591 (1.2), 649 (0.9). Fluorescence emission $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}, \lambda_{\text {exc }} 420 \mathrm{~nm}\right): \lambda_{\text {em }}, \mathrm{nm}$ 654, 718. IR (KBr): v, $\mathrm{cm}^{-1} 2919,1853,2361,1719$, 1637, 1592, 1473, 1384, 1282, 1247, 1106, 1072, 968, 800, 732.

Dimer 1b. CuCl ( $5.3 \mathrm{mg}, 0.0054 \mathrm{mmol}$ ) was inserted in a round bottom flask and TMEDA $(24.0 \mu \mathrm{~L}, \mathrm{~d}=0.78$,
0.162 mmol ) was added under stirring. Then 1.0 mL of anhydrous DCM and activated molecular sieves ( $4 \AA$, $\sim 380 \mathrm{mg}$ ) were added in sequence. Lastly a solution of $\mathbf{2 a}(87 \mathrm{mg}, \mathrm{MW}=802.9,0.108 \mathrm{mmol}$ ) in 3.5 mL of anhydrous DCM was added and the reaction was stirred for 16 h , at room temperature, under air pressure. The reaction was quenched with 3 mL of $\mathrm{H}_{2} \mathrm{O}$, then the organic layer was washed three times with water, dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$ and filtered. The solvent was removed under reduced pressure and the crude was purified by CC (silica gel, $\mathrm{CHCl}_{3} / \mathrm{EtOH}$ from $99 / 1$ to $95 / 5 \mathrm{v} / \mathrm{v}$ ) giving a purple solid. Yield $66 \mathrm{mg}(76 \%)$. $R_{f}=0.30\left(\mathrm{SiO}_{2}, \mathrm{CHCl}_{3} / \mathrm{MeOH}\right.$ $98 / 2 \mathrm{v} / \mathrm{v}$ ). ${ }^{1} \mathrm{H}$ NMR ( $500 \mathrm{MHz} ; \mathrm{CDCl}_{3}$ ): $\delta_{\mathrm{H}}$, ppm 9.07 $\left(8 \mathrm{H}, \mathrm{d}, J=5.4 \mathrm{~Hz}, \mathrm{H}_{\mathrm{a}}-\mathrm{Py}\right), 8.95(4 \mathrm{H}, \mathrm{d}, J=5.1 \mathrm{~Hz}, \beta-\mathrm{H})$, 8.93 (d, $J=4.8 \mathrm{~Hz}, \beta-\mathrm{H}), 8.86(4 \mathrm{H}, \mathrm{d}, J=4.5 \mathrm{~Hz}, \beta-\mathrm{H})$, $8.82(4 \mathrm{H}, \mathrm{d}, J=4.4 \mathrm{~Hz}, \beta-\mathrm{H}), 8.25(4 \mathrm{H}, \mathrm{d}, J=8.1 \mathrm{~Hz}$, $\left.\mathrm{H}_{4}-\mathrm{Ph}\right), 8.18\left(8 \mathrm{H}, \mathrm{d}, J=5.6 \mathrm{~Hz}, \mathrm{H}_{\mathrm{b}}-\mathrm{Py}\right), 8.11(4 \mathrm{H}, \mathrm{d}, J=$ $\left.8.4 \mathrm{~Hz}, \mathrm{H}_{1}-\mathrm{Ph}\right), 8.04\left(4 \mathrm{H}, \mathrm{d}, J=8.1 \mathrm{~Hz}, \mathrm{H}_{3}-\mathrm{Ph}\right), 7.33(4 \mathrm{H}$, d, $\left.J=8.6 \mathrm{~Hz}, \mathrm{H}_{2}-\mathrm{Ph}\right), 4.46\left(4 \mathrm{H}, \mathrm{m}, \mathrm{PhOCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 4.07$ $\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.90\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.81$ $\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.76\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.63$ $\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.43\left(6 \mathrm{H}, \mathrm{s}, \mathrm{OCH}_{3}\right),-2.82(4 \mathrm{H}, \mathrm{br}$, $\mathrm{NH}) .{ }^{13} \mathrm{C}$ NMR ( $125 \mathrm{MHz} ; \mathrm{CDCl}_{3}$ ): $\delta_{\mathrm{C}}, \mathrm{ppm} 159.3,150.5$, 148.7, 143.3, 135.9, 135.0, 134.4, 131.4, 129.7, 125.7, 121.9, 121.6, 119.7, 117.6, 113.4, 82.4, 75.7, 72.4, 71.3, 71.1, 71.0, 70.3, 68.1, 59.5. MS (ESI): $m / z$ 1603, 1626 (calcd. for $\left[\mathrm{C}_{102} \mathrm{H}_{82} \mathrm{~N}_{12} \mathrm{O}_{8}+\mathrm{H}\right]^{+} 1603.64,\left[\mathrm{C}_{102} \mathrm{H}_{82} \mathrm{~N}_{12} \mathrm{O}_{8}+\right.$ $\mathrm{Na}]^{+}$1626.63). UV-vis $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right)$ : $\lambda_{\text {max }}, \mathrm{nm}$ (relative intensity, \%) 423 (100), 516 (5.7), 551 (3.5), 591 (2.1), 649 (1.8). Fluorescence emission $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}, \lambda_{\text {exc }} 420 \mathrm{~nm}\right)$ : $\lambda_{\mathrm{em}}, \mathrm{nm} 655,718$. IR (KBr): $v, \mathrm{~cm}^{-1}$ 2921, 2853, 2361, 1717, 1636, 1592, 1384, 1284, 1247, 1108, 799.

Dimer 1c. A solution of 1,4-diethynylbenzene ( $2.1 \mathrm{mg}, 0.016 \mathrm{mmol}$ ) in 1 mL of anhydrous THF was prepared and kept under Ar atmosphere. 3a $(31.4 \mathrm{mg}$, 0.035 mmol ) was placed in a microwave reactor vessel and set under inert atmosphere. Then DMF ( 1 mL ) and $\mathrm{NEt}_{3}(3 \mathrm{~mL})$ were added and the mixture was degassed for 15 min with Ar. Lastly, keeping the vessel under inert atmosphere, $\mathrm{Pd}\left(\mathrm{PPh}_{3}\right)_{4}(4 \mathrm{mg}, 0.0035 \mathrm{mmol})$ was added and the mixture was degassed for further 15 min . Keeping the vessel under Ar, the solution of 1,4-diethynylbenzene was added. The reaction was stirred for 1 h at $120^{\circ} \mathrm{C}$ under microwave irradiation (ramping time $=10 \mathrm{~min}, \mathrm{P}=$ 300 W ). The crude mixture was passed through a very short Celite ${ }^{\circledR} 521$ pad washing with $\mathrm{CHCl}_{3}$. The organic solvent was extracted three times with distilled water to remove DMF. The organic phase was dried with $\mathrm{Na}_{2} \mathrm{SO}_{4}$, filtered and the solvent was removed under reduced pressure. The residue was purified by two $\mathrm{CC}\left(\mathrm{CHCl}_{3} /\right.$ EtOH from 100/0 to $97 / 3 \mathrm{v} / \mathrm{v}$ ) to give dark purple solid. Yield $16 \mathrm{mg}(62 \%) . R_{f}=0.36\left(\mathrm{CHCl}_{3} / \mathrm{MeOH} 97 / 3 \mathrm{v} / \mathrm{v}\right)$. ${ }^{1} \mathrm{H}$ NMR ( $500 \mathrm{MHz} ; \mathrm{CDCl}_{3}$ ): $\delta_{\mathrm{H}}, \mathrm{ppm} 9.07(8 \mathrm{H}, \mathrm{d}, J=$ $\left.4.0 \mathrm{~Hz}, \mathrm{H}_{\mathrm{a}}-\mathrm{Py}\right), 8.96-8.94(8 \mathrm{H}, \mathrm{m}, ~ \beta-\mathrm{H}), 8.86(4 \mathrm{H}, \mathrm{d}$, $J=4.5 \mathrm{~Hz}, \beta-\mathrm{H}), 8.83(4 \mathrm{H}, \mathrm{d}, J=5.0 \mathrm{~Hz}, \beta-\mathrm{H}), 8.25(4 \mathrm{H}$, $\left.\mathrm{d}, J=7.5 \mathrm{~Hz}, \mathrm{H}_{4}-\mathrm{Ph}\right), 8.19\left(8 \mathrm{H}, \mathrm{d}, J=5.0 \mathrm{~Hz}, \mathrm{H}_{\mathrm{b}}-\mathrm{Py}\right)$, $8.12\left(4 \mathrm{H}, \mathrm{d}, J=8.5 \mathrm{~Hz}, \mathrm{H}_{1}-\mathrm{Ph}\right), 8.00(4 \mathrm{H}, \mathrm{d}, J=7.5 \mathrm{~Hz}$,
$\left.\mathrm{H}_{3}-\mathrm{Ph}\right), 7.77\left(4 \mathrm{H}, \mathrm{s}, \mathrm{H}_{5}-\mathrm{Ph}\right), 7.34\left(4 \mathrm{H}, \mathrm{d}, J=8.5 \mathrm{~Hz}, \mathrm{H}_{2}-\mathrm{Ph}\right)$, $4.46\left(4 \mathrm{H}, \mathrm{m}, \mathrm{PhOCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 4.09\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right)$, $3.90\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.81\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right)$, $3.76\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right), 3.64\left(4 \mathrm{H}, \mathrm{m}, \mathrm{OCH}_{2} \mathrm{CH}_{2} \mathrm{O}\right)$, $3.44\left(6 \mathrm{H}, \mathrm{s}, \mathrm{OCH}_{3}\right),-2.81(4 \mathrm{H}, \mathrm{br}, \mathrm{NH}) .{ }^{13} \mathrm{C}$ NMR ( $125 \mathrm{MHz} ; \mathrm{CDCl}_{3}$ ): $\delta_{\mathrm{C}}, \operatorname{ppm} 159.3,150.6,148.7,142.4$, $135.9,135.0,134.5,132.7,132.2,130.5,129.8,121.5$, $120.1,117.5,113.4,110.4,91.6,72.4,71.3,71.1,71.0,70.3$, 69.1, 59.5. MS (ESI): $m / z 1680$ (calcd. for $\left[\mathrm{C}_{108} \mathrm{H}_{86} \mathrm{~N}_{12} \mathrm{O}_{8}+\right.$ $\mathrm{H}]^{+}$1679.67). UV-vis $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}\right)$ : $\lambda_{\text {max }}, \mathrm{nm}$ (relative intensity, \%) 421 (100), 516 (5.2), 551 (3.0), 591 (1.7), 650 (1.8). Fluorescence emission $\left(\mathrm{CH}_{2} \mathrm{Cl}_{2}, \lambda_{\text {exc }} 420 \mathrm{~nm}\right)$ : $\lambda_{\mathrm{em}}, \mathrm{nm} 655,719$. IR (KBr): $v \mathrm{~cm}^{-1} 2923,2853,1639$, 1593, 1247, 1108, 800, 732.

## CONCLUSION

In conclusion, a small library of three different porphyrin dimers was successfully prepared via either Glaser-Hay or Sonogashira coupling. The various systems, together with their monomer precursors, were fully characterized by NMR techniques and their UV-vis and fluorescent properties were assessed as well. Despite not being very successful, the preliminary experiments for the formation of metallacyclic assemblies with a parallelepiped shape gave us good insights on the complexity of this ambitious project. From this stand point, different metal complexes will be tried as alternatives $90^{\circ}$ metal fragments in order to overcome the major issues encountered in this report.

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## Supporting information

${ }^{1} \mathrm{H}$ NMR and ${ }^{13} \mathrm{C}$ NMR spectra of porphyrins 2a-2c and $\mathbf{3 a}, \mathbf{3 b}$ and of dimers $\mathbf{1 a - 1 c},{ }^{1} \mathrm{H}$-DOSY experiments, as well as UV-vis and fluorescence emission spectra of porphyrins 2a-2c and 3a, 3b (Figs S1-S26, Table S1) are given in the supplementary material. This material is available free of charge via the Internet at http://www. worldscinet.com/jpp/jpp.shtml.

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