A STEREOSELECTIVE α -GLUCOSYLATION BY USE OF A MIXTURE OF 4-NITROBENZENESULFONYL CHLORIDE, SILVER TRI-FLUOROMETHANESULFONATE, *N*,*N*-DIMETHYLACETAMIDE, AND TRIETHYLAMINE*

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ABSTRACT

Stereoselective α -glucosylation of partially protected carbohydrates with 2,3,4,6-tetra-O-benzyl- α -D-glucopyranose in dichloromethane, in the presence of a quaternary mixture of 4-nitrobenzenesulfonyl chloride, silver tri-fluoromethanesulfonate, N,N-dimethylacetamide, and triethylamine gave O- α -D-glucopyranosyl-(1 \rightarrow 4)- and -(1 \rightarrow 6)-2-acetamido-2-deoxy-D-glucopyranose (N-acetylmaltosamine and N-acetylisomaltosamine). A step-by-step synthesis of O- α -D-glucopyranosyl-(1 \rightarrow 4)-O-[α -D-glucopyranosyl-(1 \rightarrow 6)]-D-glucopyranose is described.

INTRODUCTION

Several procedures for α -glucosylation¹ have been published^{2,3}. Most require, however, the preparation of a reactive glycosyl donor before the coupling reaction. Instead of introducing the benzyl group as a persistent protecting group, a convenient condensation reaction (Reaction I) may be applied to a sugar derivative having an unprotected OH-1 group (GOH; G denotes a 2,3,4,6-tetra-O-benzyl-D-glucopyranosyl residue or equivalents)⁴⁻⁷. A ternary mixture

$$GOH + ROH \xrightarrow{\cdots} \alpha GOR + \beta GOR \qquad (1)$$
$$-H_2O$$

consisting of 4-nitrobenzenesulfonyl chloride (3), silver trifluoromethanesulfonate (4), and triethylamine (5) has been used for the stereoselective β -glucosylation of primary alcohols with 2,3,4,6-tetra-O-benzyl- α -D-glucopyranose^{4,5} (1). A modification of these conditions to give selectively α -glucosides is described herein⁸. These conditions were applied to the synthesis of $O-\alpha$ -D-glucopyranosyl-(1 \rightarrow 4)-(25) and -(1 \rightarrow 6)-2-acetamido-2-deoxy-D-glucopyranoses (27), which constitutes

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the Shigella flexneri O-antigens⁹, as well as to the step-by-step synthesis of O- α -D-glucopyranosyl- $(1\rightarrow 4)$ -O- $[\alpha$ -D-glucopyranosyl- $(1\rightarrow 6)$]-D-glucopyranose (32), which is the branching point of amylopectin and glycogen^{10,11}.

RESULTS AND DISCUSSION

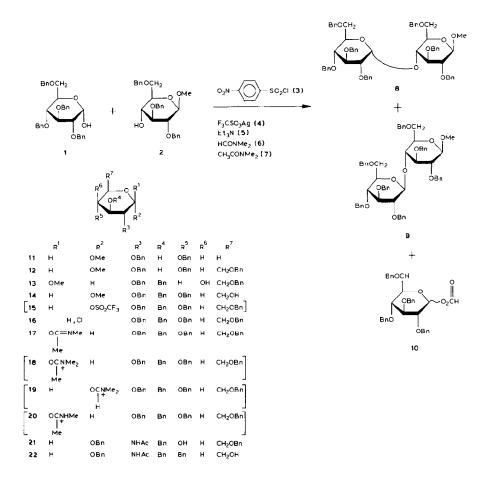
When the glucosylation of methyl 2,3,6-tri-O-benzyl- β -D-glucopyranoside¹² (2) with 1 was carried out in the presence of 3, 4, and 5, in N, N-dimethylformamide (6) (Run 2, Table I) instead of dichloromethane (Run 1), the proportion of α (8) to β anomer (9) rose significantly, although the yield was much decreased; the 1-O-formyl derivative 10 was isolated as a by-product¹³. The yield of glucosides, as well as the selectivity for the formation of the α anomer (8), were well improved by use of a limited amount of 6 in dichloromethane (Run 3); N,N-dimethylacetamide (7) was more effective. Thus, the glucosylation by a quaternary mixture composed of 3, 4, 5, and 7 in dichloromethane (Run 5, Condition A) gave

TABLE I

Run	Acceptor	Amide		Time ^b	Yield of	Proportion of	
		Composition	Proportion	(h) (%)	glucosides (%)	α-D-glucoside (%)	
1	2			19	97	45	
$\frac{2^{c}}{3}$	2			19	58	77	
3	2	HCONMe ₂	2.5	18	92	88	
4	2	AcNHMe	2.5	18	45^d	е	
5	2	AcNMe ₂	2.5	21	86	93	
6	2	AcNMe ₂	3.3	16	86	93	
7	2	AcNMe ₂	5.0	21	73	86	
8	11	AcNMe ₂	2.5	18	87	88	
9	12	AcNMe ₂	2.5	24	85	89	
10	13	AcNMe ₂	2.5	18	87	90	
11	14	$AcNMe_2$	2.5	20	91	47	
12	14	AcNMe ₂	5.0	18	88	73	
13	14	AcNMe ₂	10.0	19	54	72	
14	14	HCONMe ₂	5.0	24	98	66	
5	14	EtCONMe ₂	5.0	21	99	45	
16	14	EtCONMe ₂	5.0	17	95	45	
17	14	AcNPh ₂	5.0	22	95	41	
18	14	$BzNMe_2$	5.0	17	94	39	
19	21			24	76^d	е	
20	21	AcNMe ₂	2.5	24	84^d	e	
21	22			24	79	35	
22	22	AcNMe ₂	5.0	24	95	68	
23	30	AcNMe ₂	5.0	20	82	62	
24	30	AcNMe ₂	2.5	20	62^d	e	

GLYCOSYLATION OF COMPOUNDS 2, 11-14, 21, 22, 30 AND 42 WITH 1^a

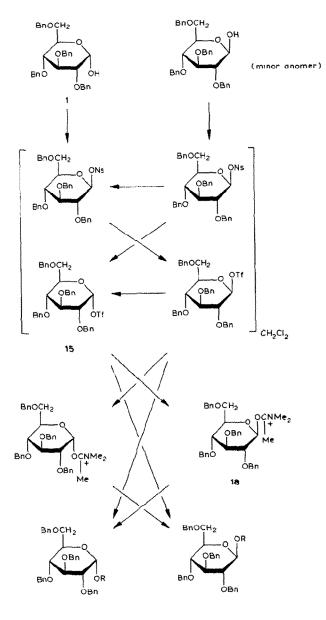
^aThe acceptor was treated with 1 (1.3 equiv.), and 3, 4, and 5 (2.5 equiv. each) in dichloromethane. ^bAt 0°. ^cIn N,N-dimethylformamide instead of dichloromethane. ^dYield of isolated α -D anomer. ^eTrace of β -D anomer.



8 in a 80% yield with an excellent stereoselectivity; use of an excess of 7 diminished the yield of glucosides (Runs 6 and 7). With *N*-methylacetamide (Run 4), the selectivity was good but the yield was poor.

The quaternary reagent used under Condition A was effective for the selective α -glycosylation of the partially benzylated pyranosides 11 (ref. 14) and 12 (ref. 15) (Runs 8 and 9). The axially oriented OH-4 group of the galactopyranoside¹⁶ was similarly α -glycosylated without difficulty (Run 10).

The glucosylation of the primary OH group of the glycosyl acceptor¹⁷ 14, however, lost selectivity under Condition A (Run 11). This implies that the reactive primary alcohol may react with the hypothetical intermediate 15 (ref. 18), which we consider as the reactive intermediate (Scheme 1), to form the corresponding β -glucoside in competition¹⁹ with 7. Nevertheless, the selectivity was increased to an acceptable level simply by doubling the amount of 7 (Run 12, Condition B); use of an excess of 7 decreased the efficiency of the reaction (Run 13). Of the additives examined under Condition B, 7 gave the best results (Runs 14–18).



$$Ns - = -SO_2C_6H_4NO_2(p)$$
 $TI - = -SO_2CF_3$

Scheme 1

Scheme 1. Plausible pathway of α -glucosylation with the quaternary reagent. The β -sulfonates may rapidly change into 15 in the presence of [Et₃NH]⁺ · [OT1]⁻ in dichloromethane^{17,18}. The amide 7 reacts with 15 to generate the intermediate 18 that is the immediate precursor of an α -glucoside¹⁹. A primary alcohol may effectively compete with 7 to give a β -glucoside. The counter anion of the intermediates, which is thought to be TfO⁻, is not illustrated.

The aforementioned effect of 7 on the selectivity was also observed for the glucosylation with chloride²⁰ 16 and 4 (Reaction 2).

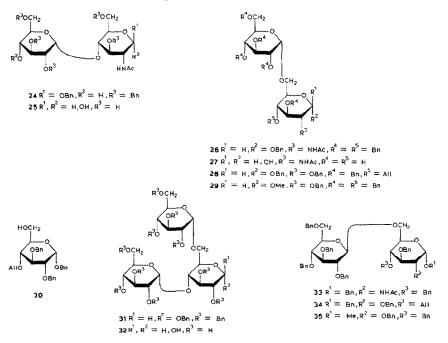
$$2 + 16 \frac{4+5+7}{CH_2Cl_2} 8 (+9)$$
(2)

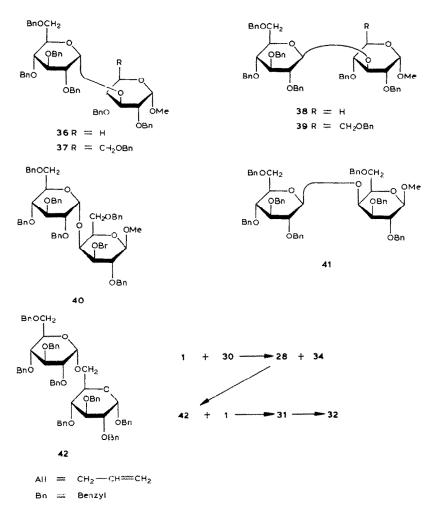
Thus, treatment of a mixture of the glucosyl imidate² 17 and 2 in

$$2 + 17 \frac{F_3 CSO_3 Me (23)}{CH_2 Cl_2} 8$$
(3)

dichloromethane with methyl trifluoromethanesulfonate (23) gave 8 exclusively (Reaction 3); the reactive intermediate 18 is thought to be involved in the reaction¹⁹. The aforementioned formate 10 (Run 2) appears to be formed through hydrolysis of the unreacted, less reactive intermediate having the α configuration (19). The intermediate 19 seems more reactive than the intermediate 20, which may have been formed during Run 4.

The quaternary reagents was then applied to the acceptors 21 (refs. 21, 22) and 22 (refs. 21, 23) having a 2-acetamido-2-deoxy group. In the glucosylation of OH-6 of 22, 7 showed an apparent effect for α selectivity (Run 22). In the glucosylation of 21, an adequate α selectivity was attained, even with the ternary reagent (Run 20). It is notable that the addition of 7 improved the yield of glucosides for 21 as well as 22; 7 seems to prevent 15 to react with the acetamido group²⁴ of 21





and 22 to form a less reactive intermediate, such as 20. Catalytic hydrogenation of the resulting 24 and 26 smoothly gave 25 and 27, respectively.

The quaternary reagent was used for a new step-by-step synthesis of the branched trisaccharide 32. The key compound²⁵ 30 was condensed with 1 in the presence of the quaternary reagent under Condition B (Run 23). After chromatography, the isomaltose derivative 28 was obtained in a 51% yield. After deallylation²⁶, the second coupling reaction of 42 with 1 by the quaternary reagent under Condition A (Run 24) gave the trisaccharide derivative 31 in a 62% yield with almost complete stereoselectivity; further catalytic debenzylation gave trisaccharide 32. The ¹³C-n.m.r. spectra of the debenzylated products 25, 27, and 32 were consistent with the structures proposed. Thus, the quaternary reagent is useful for the α -glucosylation using 1, especially that of rather less reactive secondary alcohol groups.

EXPERIMENTAL

General. — Melting points were determined with an MP-1 melting-point apparatus (Yanagimoto); they are uncorrected. Specific rotations were measured, for solution in a jacketed 1-dm cell, with a DIP-180 automatic polarimeter (Japan Spectroscopic) at 20°. ¹H-N.m.r. data were recorded with a Varian S-60T spectrometer, and ¹³C-n.m.r. data with a JEOL-PS-100 spectrometer linked to a JEOL-EC-100 computer. Column chromatography was carried out on silica gel (Kanto Kagaku); each fraction was examined by t.l.c. on silica gel (Merck, No. 7731).

Compound 2 was prepared as reported earlier¹²; m.p. 72–73°, $[\alpha]_D^{20} - 17^\circ$ (c 1.8, chloroform).

Anal. Calc. for C₂₈H₃₂O₆: C, 72.39; H, 6.94. Found: C, 72.03; H, 6.91.

Compounds 8 and 9 were identified by comparison with samples prepared by methanolysis in the presence of silver carbonate and subsequent hot benzylation with benzyl chloride and potassium hydroxide of 2,3,6-tri-O-acetyl-4-O-(2,3,4,6-tetra-O-acetyl- α -²² and - β -D-glucopyranosyl)- α -D-glucopyranosyl bromide²⁸. Compounds 36 (ref. 6), 38 (ref. 6), 29 (ref. 12), and 35 (ref. 29) were identified by comparison with the samples prepared previously.

Anal. of **29**. Calc. for $C_{62}H_{66}O_{11}$: C, 75.43; H, 6.74. Found: C, 75.41; H, 6.79.

The results of the condensation reactions are reported in Table I, the physical and analytical data of the synthesized glucosides and their derivatives in Table II, and the ¹³C-n.m.r. data of the C-1 atoms of the protected glucosides and of the C atoms of the deprotected glucosides are in Tables III and IV, respectively; all assignments are tentative.

General procedure for glycosylation. — To a mixture of a glucosyl acceptor (0.33 mmol), 2,3,4,6-tetra-O-benzoyl- α -D-glucopyranose (1, recrystallized from ethyl acetate³⁷, 1.3 equív.), 4-nitrobenzenesulfonyl chloride (3, 2.5 equiv.), silver trifluoromethanesulfonate (4, 2.5 equiv.), and dichloromethane (1.8 mL), N,N-dimethylacetamide (7), and then triethylamine (5, 2.5 equiv.) were successively added under stirring at -40° bath temperature. The mixture soon became milky, and the bath temperature was gradually raised for 1 h to 0° , at which temperature the mixture was well stirred overnight. The mixture was processed as described previously⁴, and then chromatographed with toluene-butanone (gradient); contamination by a trace of any nitrogenous compounds in the glucosides was removed by rechromatography on silica gel in hexane-ethyl acetate. The amount of 7 was 2.5 equiv. for a secondary alcohol (Condition A) and 5.0 equiv. for a primary alcohol (Condition B).

Isolation of 2,3,4,6-tetra-O-benzyl-1-O-formyl-D-glucose (10). — Chromatography of the reaction mixture of Run 2 conducted in an 85- μ mol scale gave 10 (4.9 mg, 8%) as the fastest-moving syrupy product; ¹H-n.m.r. (CCl₄, Me₄Si): δ

TABLE II

Cpd.	M.p.	Optical	rotatio	n	Mol. Form.	Anal.						Lit.
	(degree)	[α] ²⁰ (degree)	c	Solv."		Calc.			Found	i		
						С	H	N	С	H	N	
2	148-152	+91	0.4,	W	C ₁₄ H ₂₅ NO ₁₁	43.86	6.57	3.65	44.41	6.92	3.39	b
8		+32	1.0,	С	C ₆₂ H ₆₆ O ₁₁	75.43	6.74		75.10	6.73		
9	99-100	+22	1.0,	С	C62H66O11	75,43	6.74		75.13	6.69		
24		+91	1.9,	С	C63H67NO11	74.61	6.66	1.38	74.66	6.64	1.29	
26	179181	+88	0.8,	С	C ₆₃ H ₆₇ NO ₁₁	74.61	6.66	1.38	74.80	6.54	1.25	
27	145148	+100	0.3,	W	C14H25NO11	43.86	6.57	3.65	43.24	6.57	3.32	с
28		+77	1.6,	С	C64H68O11	75.87	6.76		75.20	6.76		
31		+82	1.7,	С	C95H98O16	76.28	6.60		76.41	6.57		
32		+139	0.3,	W	C18H32O16	42.86	6.39		43.10	6.27		d
33	220-221	+67	0.5,	С	C63H67NO11	74.61	6.66	1.38	74.78	6.66	1.48	
34	9394	+45	1.0,	С	C64H68O11	75.87	6.76		75.86	6.70		
37		+54	2.0,	С	C67H66O11	75.43	6.74		75.70	6.84		e
39		+49	0.3,	С	$C_{62}H_{66}O_{11}$	75.43	6.74		75.27	6.79		
40		+61	1.2,	С	$C_{62}H_{66}O_{11}$	75.43	6.74		75.13	6.70		
41		+26	1.0,	С	$C_{62}H_{66}O_{11}$	75.42	6.74		75,50	6.86		
42		+43	1.6,	С	C ₆₁ H ₆₄ O ₁₁	75.29	6.63		74.95	6.77		

PHYSICAL AND ANALYTICAL DATA OF COMPOUNDS

 ${}^{a}W = H_{2}O, C = CHCl_{3}, {}^{b}Ref. 30; m.p. 144.5-146^{\circ}, [\alpha]_{D}^{20} + 87 \rightarrow +39^{\circ} (c \ 0.9, H_{2}O), {}^{\circ}Ref. 31; [\alpha]_{D}^{25} + 94^{\circ} (H_{2}O), {}^{a}Ref. 10; [\alpha]_{D}^{22} + 125^{\circ} (c \ 0.9, H_{2}O), {}^{e}Ref. 2; [\alpha]_{D}^{20} + 54.6^{\circ} (c \ 1.62, CHCl_{3}).$

TABLE III

Cpd.	C-1		C-1'→3	/4	<i>C</i> - <i>1</i> ″→	6	C-2	COCH ₃
	α	β	α	β	α	β	****	
8		104.8	96.9					
9		104.9		102.7				
24	97.0		97.0				51.5	23.3
26	97.4				96.6		52.8	23.4
28	95.2				97.8			
31 ^b	94.4		96.8		97.5			
33	97.2					104.0	52.5	23.4
34	95.5					104.1		
37	97.6		97.8					
39	98.0			102.8				
40		105.4	100.2					
41		105.2		103.0				
42	95.2		97.8					

^aAt 25.1 MHz, for a solution in CDCl₃; signals relative to that of Me₄Si. ^bRef. 32; δ 96.0 (C-1), 96.7 (C-1'), and 97.0 (C-1") were assigned for a compound having a structure similar to that of **31**.

TABLE IV

C atom	25	27	32
1α	92.0	92.4	93.2
1 <i>β</i>	96.1	96.5	97.1
2α	55.2	55.4	72.5
2β	57.9	58.0	74.7
3α	72.5	72.4	74.1
3β	75.6	75.5	75.2
4α	79.0	71.0	79.6
4β	78.3	71.0	79.2
5α	71.5	71.4	70.3
5β	77.3	77.2	77,3
6α	61.9	67.2	68.1
6β	61.9	67.2	68.1
1'	(α101.2		
	β 100.9		101.1
2'	74.1		74.2
3'	74.2		74.2
4'	70.7		70.8
5'	(α 73.1		
	<i>β</i> 73.0		73.1
6'	61.9		61.8
1″		99.4	99,9
2″		72.9	72.8
3″		74.4	74.2
4"		71.0	70.7
5"		73.2	73.1
6″		61.9	61.7

¹³C-N.M R DATA (δ) FOR GLUCOSIDES 25, 27, AND 32^a

^{*a*}At 25.1 MHz, for a solution in D₂O; signals relative to that of Me₄Si (external). The assignments are based on those for 2-acetamido-2-deoxy-D-glucose³³, maltose^{34–36}, and isomaltose^{34–36}.

6.37 (d, 1 H, J 3 Hz, H-1) and 8.03 (s, 1 H, HCO); 13 C-n.m.r. (CDCl₃, Me₄Si): δ 94.0 (1 C, C-1) and 159.3 (1 C, CO).

 α -Glucosylation with 2,3,4,6-tetra-O-benzyl-D-glucopyranosyl chloride²⁰ (16). — To a stirred mixture of 2 (36 mg, 0.078 mmol), 16 [prepared from 1 (54 mg) with an excess of cold thionyl chloride], and 4 (49.5 mg) in dichloromethane (0.42 mL), 7 (18 μ L), and then 7 (27 μ L) were added at -40°. Then, the bath temperature was adjusted and the time programmed as described earlier. The mixture was processed and chromatographed to give 8 (54.3 mg, 71%) and 9 (11.2 mg, 15%). Without addition of 7, 8 and 9 were obtained in amounts of 34.1 mg (45%) and 41.2 mg (54%), respectively.

 α -Glycosylation with 2,3,4,6-tetra-O-benzyl-1-O-(N-methylacetamidoyl)- β -D-glucopyranose² (17). — To a solution of 2 (37 mg, 80 μ mol) and 17 (47 mg, 1.0 equiv.) in dichloromethane (0.5 mL), was added methyl trifluoromethanesulfonate (21, Aldrich, 9 μ L, 1.0 equiv.) at -40°. The conditions of reaction and processing were as described earlier to give 8 (28.3 mg, 36%); no trace of the β anomer was isolated.

O- α -D-Glucopyranosyl- $(1\rightarrow 4)$ -O- $[\alpha$ -D-glucopyranosyl- $(1\rightarrow 6)$]-D-glucopyranose (32). — The acceptor benzyl 4-O-allyl-2,3-di-O-benzyl- α -D-glucopyranoside (30, 343 mg, 0.70 mmol) was coupled with 1 under Condition B (Run 23). The faster-moving (on t.l.c.), major product (28, 288 mg, 0.28 mmol) was heated at reflux in a mixture of ethanol (7 mL), benzene (3 mL), and water (1 mL) containing chlorotris(triphenylphosphine)rhodium(I) (52.6 mg) for 7 h. After evaporation, the residue was heated at reflux in acetone (15 mL) containing M hydrogen chloride (0.6 mL). After evaporation, chromatography gave benzyl 2,3-di-O-benzyl-6-O- $(2,3,4,6-\text{tetra-}O-\text{benzyl-}\alpha-\text{D-glucopyranosyl})-\alpha-\text{D-glucopyranoside}$ (42, 248 mg, 90%). This (202 mg, 0.21 mmol) was successively coupled with 1 under Condition A (Run 24). Chromatography gave the fully benzylated trisaccharide 31 (194 mg, 62%); no trace of the β anomer was observed. This compound (118.3 mg, 80 μ mol) was hydrogenated twice in acetic acid (7 mL) containing palladium-on-carbon (10%, 64 mg) in a Parr-3911 hydrogenation apparatus (340 kPa of hydrogen), followed by chromatography on silica gel in chloroform-methanol (gradient), to give 32 as a foam (18.8 mg, 47%).

Methyl 2,4.6-tri-O-benzyl- α -D-glucopyranoside (12). — The following procedure was more convenient, but the yield and selectivity were lower, than the previous one¹⁵, especially for a large-scale preparation. A mixture of methyl α -D-glucopyranoside (7.5 g, 39 mmol), crushed potassium hydroxide (8.7 g), and benzyl chloride (150 mL) was vigorously stirred at 100° for 2.5 h. After processing, chromatography (silica gel, 120 g; toluene-butanone gradient) gave ~9 g (~50%) of 12, which contained a trace amount of the 2,3,6-tribenzyl ether. After acetylation with acetic anhydride and pyridine, a similar chromatography gave the 3-acetate of 12, which was obtained as the slower-moving material¹². Quantitative deacetylation with methanolic sodium methoxide afforded 12.

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