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Annulated bacteriochlorins for near-infrared photophysical studies†

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Molecules with strong absorption in the near-infrared (NIR) spectral region are of interest for a variety of applications in the photosciences. Nature's NIR absorbers are bacteriochlorophylls, which contain a tetrahydroporphyrin chromophore accentuated by attached auxochromes, including a β,meso-bridging isocyclic ring. Modification of the isocyclic ring has provided a convenient means of wavelength tuning. In other tetrapyrroles, annulation at the β ,meso-, β , β -, and β -meso- β -positions also has enabled wavelength tuning. Here, synthetic bacteriochlorins bearing a gem-dimethyl group in each pyrroline ring have provided the foundation for studies of annulation. Seven new free base bacteriochlorins have been prepared and seven others have been elaborated therefrom. Six distinct routes known among porphyrins were explored. Ultimately, annulation of phenaleno or benzo groups at the \(\beta,meso\)-sites (i.e., spanning the 3,5- and 13,15-positions) of bacteriochlorins was achieved by a new route that entails two successive Pd-mediated coupling reactions of a dibromobacteriochlorin with a bromoarylboronic acid. The corresponding Phen-BC or Benz-BC absorbs at 913 or 1033 nm, respectively. Examination by time-resolved spectroscopy revealed an excited-state lifetime of 150 ps or 7 ps, with commensurably low fluorescence quantum yield (Φ_t) values estimated to be ~ 0.004 or 3×10^{-5} , respectively. The origin of the short excited-state lifetimes due to β , meso-annulation is unknown, but is not simply a consequence of the energy-gap law for nonradiative decay.

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Introduction

Bacteriochlorins are attractive candidates for diverse photophysical studies such as artificial photosynthesis, photodynamic therapy, and optical imaging owing to their strong absorption in the near-infrared (NIR) region.¹ Bacteriochlorophyll *a*, the core

light absorber in anoxygenic photosynthetic bacteria, contains the bacteriochlorin chromophore along with an "isocyclic ring" spanning positions 13–15 (Chart 1).² The isocyclic ring is a 5-membered ring containing a keto group conjugated with the bacteriochlorin π -system at the 13-position, but a saturated methylene unit at the 15-position. Hence, the isocyclic ring (an annulated oxocyclopentano motif) does not cause conjugation to be extended across the 13,15-positions. The full complement of substituents about the perimeter of the macrocycle has precluded studies to install annulated rings to the native bacteriochlorophylls; however, the allomerization of the isocyclic ring has been exploited to create bacteriochlorin-imides (often referred to as bacteriopurpurinimides).^{3,4}

In previous studies, a synthetic bacteriochlorin was annulated with imide rings in an effort to extend the position of the long-wavelength absorption (Q_y) band deeper into the NIR spectral region. For the bacteriochlorin constructed with imides annulated at the 2,3- and 12,13-positions (*i.e.*, β , β -annulation), the Q_y band appeared at 800 nm.⁵ On the other hand, the bacteriochlorin constructed with imides annulated at the 3,5- and 13,15-positions (*i.e.*, β , *meso*-annulation) gave the Q_y band at 875 nm.⁵

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 $[\]dagger$ Electronic supplementary information (ESI) available: Extensive results concerning exploration of alternative routes; spectroscopic data for bacteriochlorin BC-2; $^1\mathrm{H}$ and $^{13}\mathrm{C}$ NMR spectra for all new compounds on the route to Phen-BC and Benz-BC; and single-crystal X-ray data. CCDC 1900109 (21), 1900117 (24) and 1900118 (18). For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c9nj01113g

Bacteriochlorophyll a

R = alkyl, amino, methoxy or hydroxy R' = phytol, farnesol, or hydrocarbon

Bacteriopurpurinimides

Chart 1 Structure of bacteriochlorophyll a and bacteriopurpurinimides.

One distinction between the two types of bis(imide)s is that β , β -annulation affords a 5-membered ring whereas β ,meso-annulation gives a 6-membered ring. The bacteriochlorin- β , β -bis(imide) **MeO-I2-\beta\beta-BC** gave a singlet excited-state lifetime (τ_s) of 3.6 ns and fluorescence quantum yield (Φ_f) value of 0.17 in toluene.⁵ In contrast, the bacteriochlorin- β ,meso-bis(imide) **I2-BC** gave $\tau_s = 1.0$ ns and $\Phi_f = 0.027$ in the same solvent.⁵ A synthetic bacteriochlorin bearing a single β ,meso-imide (**I-BC**) was prepared and found to exhibit $Q_y = 818$ nm, $\tau_s = 1.9$ ns and $\Phi_f = 0.036$ in toluene.⁶ The shorter lifetime and diminished fluorescence quantum yield with the β ,meso-imides were surprising and of unknown origin.

The annulation of tetrapyrroles with aromatic rings has long been pursued as a means to impart a bathochromic shift to the long-wavelength absorption band. 7-12 Issues concerning annulated tetrapyrroles are ease of synthetic access, solubility, stability, photophysical characterization, and absorption and fluorescence in the deep NIR region. Annulation with aromatic rings at the *meso*-positions is a common strategy to synthesize β ,meso- (and also β ,meso, β -) annulated porphyrins. Popular methods for annulation with aromatic hydrocarbons include oxidative coupling 13-26 and palladium-catalyzed intramolecular direct arylation. ^{27–31} On the other hand, β , β -annulated porphyrins have chiefly been prepared by (1) porphyrin formation from a β,β-annulated pyrrole, or (2) aromatization of a peripheral non-aromatic ring, which often involves prior cyclization of β substituents. ^{7-9,12} Synthesis of the β , β -annulated porphyrins has been chiefly pioneered by Lash^{7,8} whereas β,meso-annulations with porphyrins have been deeply investigated by a number of groups. 32,33

These strategies developed with porphyrins in principle can be applied to the synthesis of β , meso- or β , β -annulated bacteriochlorins, albeit with two caveats: (1) in practice, the synthetic chemistry of bacteriochlorins has significantly lagged that of porphyrins; and (2) the reactivity of bacteriochlorins appears to be somewhat different from that of porphyrins.

Representative synthetic approaches to bacteriochlorins^{1,34} include hydrogenation of porphyrins,^{35,36} cycloaddition to porphyrins,³⁷ breaking and mending of porphyrins,³⁸ semisynthesis beginning typically with bacteriochlorophyll a,^{3,4,39} and de novo synthesis. In de novo synthesis, the saturation characteristic of the pyrroline rings is built into the acyclic precursors to the bacteriochlorin macrocycles. Examples include the synthesis of tolyporphins^{40,41} and a general route that we have been developing,^{42–47} which has been extended imaginatively by others.^{48–57}

In this paper, we report synthetic approaches for annulation of de novo synthesized bacteriochlorins. Installing annulated groups proved more challenging with bacteriochlorins than appears to be the case with porphyrins, as numerous established approaches with porphyrins such as oxidative coupling were unsuccessful. Ultimately, two β , meso-annulated bacteriochlorins were synthesized via palladium-catalyzed intramolecular arylation. Photophysical studies were performed to determine the lowest excited singlet-state lifetimes and fluorescence quantum yields. These studies were accompanied by density functional theory (DFT) calculations to examine the forms of the frontier molecular orbitals (MOs) and by time-dependent DFT (TDDFT) to compare the calculated versus observed absorption spectra.

Results

Synthetic approach toward β,β-annulated bacteriochlorins

We first focused on the synthesis of β , β -annulated bacteriochlorins. Thus, β , β -dibromodihydrodipyrrin-acetal 1^{54} was treated with BF $_3$: OEt $_2$ in MeCN following the standard procedure for the self-condensation leading to synthetic bacteriochlorins 42 (Scheme 1). As a result, tetrabromobacteriochlorin BC-1, a potential precursor of β , β -annulated bacteriochlorins, was obtained in 16% yield. Bacteriochlorin BC-1 differs from the tetrabromobacteriochlorin prepared by Oliveira and coworkers 54 in lacking a 5-methoxy group. The Mizoroki–Heck reaction of BC-1 and methyl acrylate afforded tetrakis(methoxycarbonylvinyl)bacteriochlorin BC-2 in 32% yield. However, oxidative cyclization of the adjacent methoxycarbonylvinyl groups leading to the corresponding β , β -annulated bacteriochlorin was unsuccessful, although the corresponding reaction of porphyrins affords β , β -benzoporphyrins. $^{58-67}$

Next, phenanthropyrrole 2^{68} was selected as a starting material for the synthesis of β , β -phenanthrobacteriochlorin BC-3, similar to the synthesis of phenanthroporphyrins. Following an established procedure, henanthropyrrole 2 was converted *via* the standard intermediates 3–6 to give dihydrodipyrrin-carboxaldehyde 7 in 5 steps (Scheme 2). The self-condensation of 7 in neat TFA, conditions used previously for β , β -substituted dihydrodipyrrin-carboxaldehydes, as evidenced by laser-desorption mass

Scheme 1 Synthesis of tetrakis[(methoxycarbonyl)vinyl]bacteriochlorin BC-2 via (tetrabromo)bacteriochlorin BC-1.

spectrometry (LD-MS, m/z = 671, $[M + H]^+$) and spectroscopic analysis (bacteriochlorin-like Q_{ν} band absorption and fluorescence at 801 and 808 nm in CH₂Cl₂, respectively, Fig. 1) of a partially purified sample. However, quality NMR spectra could not be obtained due to the limited solubility of BC-3 in organic solvents. Moreover, isolation of BC-3 in pure form was thwarted by gradual decomposition. Adventitious oxidation in air may explain the instability of BC-3 because a major contaminant, which increased gradually upon handling of BC-3, was attributed to the corresponding oxobacteriochlorin⁵ given the characteristic data from LD-MS $(m/z = 685, [M + H]^{+})$ and absorption spectroscopy (Q_v band at 758 nm in CH_2Cl_2).

We envisaged that β , β -annulated bacteriochlorins might be stabilized by installation of electron-withdrawing groups, which by adding slight steric protrusions from the plane of the macrocycle might also provide improved solubility. However, attempts to install ester or imide moieties into the macrocycle were unsuccessful. The attempted syntheses are summarized in Chart S1 and Schemes S1-S10 (ESI†), and the synthesis and characterization data are provided for 15 new compounds (see the ESI†). The limitations of these routes prompted investigation of methods of oxidative annulation of appended aryl rings.

Scheme 2 Attempted synthesis of β , β -phenanthrene-annulated bacteriochlorin BC-3.

Synthetic approach toward β,meso-annulated bacteriochlorins via oxidative annulation

As the oxidative annulation of peripheral aromatic hydrocarbons is well-known for the synthesis of β,meso-annulated porphyrins, di(1-naphthyl)bacteriochlorins BC-4-8 were prepared from 8⁷⁰ (Scheme 3) for attempted transformations to

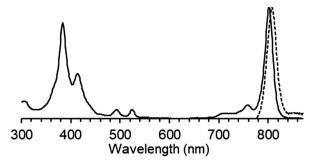


Fig. 1 Absorption (solid line) and fluorescence (dashed line; using 675 nm excitation) spectra of partially purified **BC-3** in CH_2Cl_2 at room temperature, normalized at the Q_v band.

give \(\beta, meso-phenalenobacteriochlorins. \) Compound 8 was converted (via intermediates 9, 10 and 12) into dihydrodipyrrinacetal 13 following standard procedures 42 and use of an improved synthesis of Michael acceptor 11.71 The self-condensation of 13 with BF3·OEt2 in MeCN42 provided 5-unsubstituted bacteriochlorin BC-4 in 41% yield, while with TMSOTf in the presence of 2,6-di-tert-butylpyridine (DTBP) in dichloromethane, 43 the reaction afforded the expected 5-methoxybacteriochlorin BC-5 in 69% yield. Furthermore, BC-4 and BC-5 were converted⁷² into the corresponding indium(III) and zinc(II) complexes BC-6 and BC-7 in 25% and 80% yield, respectively. Compound 8 was also converted (via intermediates 14 and 16-18) through a recently reported Northern-Southern route⁴⁵ into dihydrodipyrrin-acetal 19 in 5 steps, making use of the known alkynoic acid 15.45 A single-crystal X-ray structure of 18 confirmed the integrity of the transformations (Fig. S1 and Table S1, ESI†). Compound 19 is a positional isomer of 13. Treatment of 19 with TMSOTf/DTBP gave 10-methoxybacteriochlorin BC-8 in 44% yield.

Oxidation of dinaphthylbacteriochlorins BC-4-7 with DDQ or FeCl₃, which are common oxidizing agents in the synthesis of annulated porphyrins, 7,8 did not afford the expected annulated bacteriochlorins (Scheme 4A and Table S2, ESI†). The In(III) chelate BC-6 and divalent Zn(II) chelate BC-7 were created to probe whether metalation might facilitate the annulation process, either directly or upon subsequent macrocycle oxidation. It was straightforward to attribute the failure of annulation of these cases to steric interactions of the naphthalene units at the 2,12-positions with the gem-dimethyl groups at the 8,18-positions (Scheme 4B). Thus, similar oxidative annulation of BC-8 and known bacteriochlorin BC-9,45 wherein the gemdimethyl groups are positioned away from the expected reaction sites, was attempted (Scheme 4C). However, neither β -to-meso nor meso-to-β oxidative annulation was observed with bacteriochlorins BC-8 and BC-9 (Tables S2 and S3, ESI†); similar attempts to annulate *meso*-naphthyldihydrodipyrrinatoboron complexes⁷³ also were unsuccessful (Table S4, ESI†).

Synthesis of β , meso-annulated bacteriochlorins via intramolecular direct arylation

Next, we focused on palladium-catalyzed intramolecular arylation as a means to gain access to β , *meso*-annulated bacteriochlorins. Unlike the unsubstituted naphthyl groups for the oxidative

coupling as described above, halogen-substituted naphthyl groups are required for such direct arylation processes. Because avoidance of the steric hindrance of *gem*-dimethyl groups still appeared warranted, the Northern–Southern route⁴⁵ was adopted to prepare the bacteriochlorin.

The Northern-Southern route relies on the Pd-mediated coupling of an iodopyrrole and an alkynoic acid; hence, we planned to install a second halogen (bromide) at the β-pyrrole position in a subsequent reaction. The reaction of 4-carboethoxy-2-iodopyrrole⁴⁵ and 2,2-dimethylpent-4-ynoic acid under slightly modified conditions afforded known lactone-pyrrole 2045 in improved yield (80%, Scheme 5).73 Electrophilic bromination of a pyrrole generally occurs at the α-position. Indeed, as shown in entry 1 of Table 1, treatment of 20 with N-bromosuccinimide (NBS, 1 equiv.) in DMF gave β-bromopyrrole 21 (minor, 18%) and α-bromopyrrole 22 (major, 50%). The structure of 21 was confirmed by X-ray crystallography (Fig. S2 and Table S5, ESI†). Upon screening of solvents, acetonitrile was found to show better β-selectivity (24% of 21 versus 17% of 22, entry 2). The use of 2,4,4,6-tetrabromo-2,5-cyclohexadienone (TBCO)⁷⁴ as a brominating agent afforded a slightly increased amount of 21 (28%, entry 3). Addition of K₂CO₃ further improved the β-selectivity (31% of 21 and 19% of 22, entry 4). Remarkably high β-selectivity was observed (50% of 21 and 6% of 22, entry 5) when the reaction was carried out in the presence of K₂CO₃ and 2,4,6-tribromophenol (TBP), which is a coproduct from TBCO. After refinement of the reaction conditions, the yield of 21 reached 61% (entries 6-8). Interestingly, bromination by using NBS with TBP and K2CO3 resulted in low yields of 21 and 22 (entry 9). To the best of our knowledge, use of TBCO with TBP/K₂CO₃ represents a new condition for bromination. The origin of the β-selectivity caused by TBP/K₂CO₃ is unclear at present. Dihydrodipyrrin 23 was prepared from 21 by the established procedure⁴⁵ in 65% yield.

Attempted borylation of β -bromodihydrodipyrrin 23 proved fruitless (Table S6, ESI†). Compound 23 was converted into the corresponding difluoroboron- and dibutylboron-complexes⁷³ to thwart possible complexation of 23 with palladium upon direct benzo annulation, but this approach also was to no avail (Table S7, ESI†). Treatment of 23 with Pd(OAc)₂ in CH₂Cl₂ at room temperature⁷⁵ gave the bis(dipyrrinato)Pd(II) complex 24, which did not survive attempted Suzuki coupling with 2-bromophenylboronic acid but instead reverted to 23. Crystals of 24 suitable for X-ray analysis were obtained from vapor diffusion of hexanes into CH₂Cl₂/ethyl acetate solution at room temperature. For 24, steric interaction between the pyrrolinyl methyl group of one dihydrodipyrrin and the α -hydrogen on the pyrrole ring of the other dihydrodipyrrin was observed in the single-crystal X-ray structural data (Fig. S3 and Table S8, ESI†).

At a minimum, the limitation in direct coupling highlights ongoing challenges associated with the synthetic chemistry of hydrodipyrrins despite extensive work. Thus, the coupling reaction was carried out after the macrocycle formation. When oxidation of 23 with SeO₂ was performed in the absence of pyridine following the general procedure, this dihydrodipyrrin-acetal 25 was obtained in less than 27% yield following acetalization with trimethyl orthoformate.

Scheme 3 Synthesis of di(1-naphthyl)bacteriochlorins BC-4-8.

Addition of pyridine to SeO₂ for the oxidation^{80,81} improved the vield of 25 to 37%. Table 2 shows refinement of the conditions for the self-condensation of 25. 1,2-Dichloroethane was found to be a better solvent than CH₂Cl₂ or CHCl₃ (Table S9, ESI†). When the concentration of 25 was 32 mM, the desired mesounsubstituted bacteriochlorin BC-10 was obtained in only 11% yield along with undesired meso-methoxybacteriochlorin BC-11 in 52% yield (entry 1). Lower concentrations (5.0-18 mM, entries 2-6) tended to give increased BC-10 (up to 32% in entry 4). Higher temperature was required for completion of the reactions at low concentrations (entries 3-6). Scaled-up reactions proceeded smoothly (entries 5 and 6). In prior studies with diverse dihydrodipyrrin-acetals, 43 the use of TMSOTf/DTBP tended to afford exclusively the meso-methoxybacteriochlorin. Here, both the meso-unsubstituted and meso-methoxy bacteriochlorin were obtained under these conditions regardless of temperature employed.

Suzuki-Miyaura cross-coupling of BC-10 and excess (8-bromonaphthalen-1-yl)boronic acid (26) provided annulation precursor BC-12 in 49% yield (Scheme 6). The molar ratio **Paper**

(A) $(R = CO_2Et)$ than other ligands screened $(PCy_3 \cdot HBF_4/K_2CO_3 \text{ and } P^tBu_3 \cdot HBF_4/K_2CO_3)$.

Scheme 4 Attempted oxidative annulation from (A) BC-4-7 and (C) BC-8 and BC-9. (B) Possible steric hindrance in BC-4-7 during the oxidative annulation.

of bromoarylboronic acid **26** to bacteriochlorin **BC-10** was 10:1, giving 5 equiv. per bromo site on the bacteriochlorin. In this reaction, byproducts less polar than the bacteriochlorins, perhaps oligomerized aromatic rings, were observed by TLC analysis and readily removed by column chromatography. Compound **26**, reported previously but without characterization data, ^{82,83} was fully characterized here. NMR and TLC analysis indicated that **BC-12** was a mixture of two atropisomers (\sim 1:1 ratio) attributed to hindered rotation of the bromonaphthyl groups at the 3- and 13-positions. Since both of the atropisomers should afford the same annulated compound, **BC-12** was used as the mixture in the next step. Palladium-catalyzed intramolecular direct arylation of **BC-12** afforded target β , *meso*-bis(phenaleno)bacteriochlorin **Phen-BC** in 37% yield. The ligand SPhos was found to be better

In this synthetic strategy, the aromatic rings for annulation can be altered by changing the coupling partner of the Suzuki–Miyaura coupling. Thus, we also aimed for β -to-*meso* annulation with benzene rings by reaction of BC-10 and (2-bromophenyl)-boronic acid (27), affording bis(2-bromophenyl)bacteriochlorin BC-13 in 85% yield. Similar to BC-12, bacteriochlorin BC-13 was isolated as a mixture of two atropisomers. Subsequent palladium-mediated arylation gave the desired β , *meso*-dibenzobacteriochlorin Benz-BC in 80% yield. In this case, PCy₃ gave a better result than that of SPhos. The NMR chemical shifts of the inner N–H protons of diverse synthetic bacteriochlorins are typically observed around -2 ppm in CDCl₃. In contrast, those of Phen-BC and Benz-BC were found at 1.97 and 2.89 ppm, respectively, suggesting unusual ring currents in the annulated macrocycles.

Photophysical properties

Fig. 2 shows absorption spectra for the two new annulated bacteriochlorins along with several reference compounds, all of which contain 8,18-geminal dimethyl groups to stabilize the bacteriochlorin. The Q_v band for Benz-BC (1033 nm) lies 120 nm to longer wavelength than that for Phen-BC (913) nm, which in turn is bathochromic from those for diimide I2-BC (875 nm), monoimide I-BC (818 nm), diester Es-BC (753 nm)⁸⁴ and parent H-BC (713 nm).85 The structures of I2-BC and I-BC are shown in Chart 2 whereas H-BC and Es-BC are shown in Chart 3. Fig. 2 also shows fluorescence spectra. The Q_{ν} fluorescence maximum is bathochromically shifted from the Q_{ν} absorption peak by 4-9 nm $(65-84 \text{ cm}^{-1})$ for all constructs except for Phen-BC, which shows a considerably larger Stokes shift of 20 nm (235 cm⁻¹). The emission band for **Phen-BC** is also broader (49 nm, 586 cm⁻¹) than that of the other bacteriochlorins (12-34 nm; 230-346 cm⁻¹). The spectral characteristics are given in Table 3.

The fluorescence quantum yield ($\Phi_{\rm f}$) of Benz-BC is estimated to be very small ($\sim 3 \times 10^{-5}$) and that for Phen-BC is somewhat larger but also low (0.004). Both annulated bacteriochlorins are less fluorescent than diimide I2-BC (0.027) and monoimide I-BC (0.064), which themselves are weak emitters. In turn, all of these constructs, which employ *exo*-macrocycle connectivity between adjacent *meso* and β -pyrrole positions, have much smaller $\Phi_{\rm f}$ values than benchmark bacteriochlorins Es-BC and H-BC (0.13–0.14; Table 3). The lifetime ($\tau_{\rm s}$) of the lowest singlet excited state (S₁) shows a similar pattern (Table 3): Benz-BC (7 ps) < Phen-BC (150 ps) < I2-BC (0.98 ns) < I-BC (1.9 ns) < Es-BC (3.3 ns) < H-BC (3.8 ns). As will be discussed below, the weak fluorescence and short S₁ lifetimes for the annulated bacteriochlorins cannot be attributed solely to a lower excited-state energy than the benchmarks.

The S_1 lifetimes noted above were measured by transient absorption (TA) spectroscopy, and in some cases also by time-resolved fluorescence. The TA studies also give insight into yields of the $S_1 \to T_1$ intersystem crossing and $S_1 \to S_0$ internal conversion pathways that compete with $S_1 \to S_0$ fluorescence.

Scheme 5 Synthesis of bromodihydrodipyrrin 23 en route to dibromobacteriochlorin BC-10

Table 1 Bromination of lactone-pyrrole 20

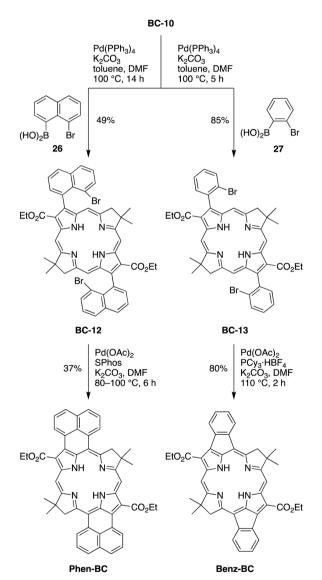
20 brominating agent 21 + 22									
Entry	Brominating agent (equiv.)	Solvent	Additive (equiv.)	Recovered 20 ^a (%)	21 ^a (%)	22 ^a (%)			
$\overline{1^b}$	NBS (1)	DMF	None	c	18^d	50 ^d			
2^e	NBS (1)	MeCN	None	<u></u> c	24^d	17^d			
3^f	TBCO (1)	MeCN	None	16	28	31			
4^f	TBCO (1)	MeCN	K_2CO_3 (1)	23	31	19			
5^f	TBCO (1)	MeCN	TBP (1), K_2CO_3 (1)	23	50	6			
6^f	TBCO (1)	MeCN	TBP (1), K_2CO_3 (2)	25^d	53^d	9			
7^f	TBCO (1)	MeCN	TBP (2), K_2CO_3 (3)	26	49	5			
8^f	TBCO (1.6)	MeCN	TBP (2), K_2CO_3 (3.6)	Trace	61^e	Trace			
9^f	NBS (1)	MeCN	TBP (2), K_2CO_3 (3)	49	7	3			

 $[^]a$ Calculated from 1 H NMR spectroscopic analysis by using 2-nitromesitylene as an internal standard unless otherwise noted. b The reaction was carried out at -20 $^{\circ}$ C to r.t. for 10 h. c Not determined. d Isolated yields. e The reaction was carried out at 0 $^{\circ}$ C to r.t. for 1.5 h. f The reactions were carried out at 0 °C for 1 h.

Representative TA data for the two annulated bacteriochlorins is given in Fig. 3. The TA difference spectrum for Phen-BC (Fig. 3A) at 0.4 ps after a 0.1 ps excitation flash is dominated by a feature at \sim 930 nm that is comprised of bleaching of the ground state $S_0 \rightarrow S_1$ absorption band along with $S_1 \rightarrow S_0$ stimulated emission. During the next tens of picoseconds, there are minor spectral changes that can be attributed to relaxations (vibrational, solvent, etc.) in the excited state. Subsequently, the prominent 930 nm feature and the rest of the spectrum diminish uniformly to almost $\Delta A = 0$ by about 2 ns, reflecting decay of the S1 excited state with a time constant of 150 ps (Fig. 3B). Close examination of the data to 7 ns indicates

25 self-condensation BC-10 + BC-11								
Entry ^a	Conc. (mM)	Temp.	Time (h)	$\mathbf{BC-10}^{b}$ (%)	BC-11 ^b (%)			
1	32	r.t.	20	11	52			
2	18	r.t.	20	15	32			
3	10	60 °C	4	29	37			
4	5.6	Reflux	7	32	38			
5 ^c	5.6	Reflux	8	31	39			
6^d	5.0	Reflux	11	29	36			

 a All reactions were carried out with TMSOTf (7.5 equiv.) and DTBP (30 equiv.) in 1,2-dichloroethane on a 0.12 mmol scale unless otherwise noted. b Isolated yields. c The reaction was carried out on a 0.71 mmol scale. d The reaction was carried out on a 2.00 mmol scale.



Scheme 6 Synthesis of β , meso-annulated bacteriochlorins **Phen-BC** and **Benz-BC**.

that a very small amount of Q_y bleaching remains that corresponds to about 2% of the bleaching at early times. This small

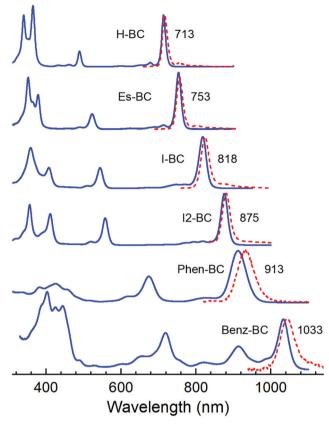


Fig. 2 Absorption spectra (blue) and fluorescence spectra (red) of the annulated bacteriochlorins **Phen-BC** and **Benz-BC** and known reference compounds. The absorption spectra are normalized to the Q_y maximum. The emission spectra were obtained using excitation in the Q_x band (490, 522, 544, 557, 675, and 715 nm, respectively); for each compound the same spectrum was obtained using excitation in the Soret band.

asymptotic signal likely represents the lowest triplet excited state (T_1), which forms from S_1 with a yield of $\Phi_{\rm isc}$ = 0.02. Given this value and $\Phi_{\rm f}$ = 0.004 for **Phen-BC**, the yield of $S_0 \rightarrow S_1$ internal conversion is $\Phi_{\rm ic}$ = 0.98.

The analogous TA data for **Benz-BC** are dominated by a spectral feature at \sim 1030 nm (Fig. 3C) that disappears with a time constant of 7 ps (Fig. 3D), which reflects the S₁ lifetime. At long times, the Q_y bleaching is less than 1% of the initial value, reflecting essentially no formation of the T₁ excited state. Given the extremely small Φ_f value (<0.0001), this means that decay of the S₁ excited state of **Benz-BC** occurs 99% by S₀ \rightarrow S₁ internal conversion (Table 3).

Molecular orbital characteristics and calculated spectral properties

The energies and electron density distributions calculated by DFT for the frontier MOs of **Es-BC**, **Phen-BC**, and **Benz-BC** are shown in Fig. 4. For benchmark **Es-BC** (Fig. 4A), the two highest occupied molecular orbitals (HOMO and HOMO–1) and the two lowest unoccupied molecular orbitals (LUMO and LUMO+1) have the familiar electron-density distributions of the frontier MOs of Gouterman's four-orbital model for tetrapyrrole chromophores.^{87–89} In this model, linear combinations of excited-state

β.meso-imides

$$\beta,\beta\text{-imides}$$

$$\beta,\beta\text{-imides}$$

$$\beta,\beta\text{-imides}$$

$$\beta,\beta\text{-imides}$$

$$\beta,\beta\text{-imides}$$

$$\beta,\beta\text{-imides}$$

$$\beta,\beta\text{-imides}$$

$$\beta,\beta\text{-imides}$$

$$\beta,\beta\text{-annulated bacteriochlorins}$$

$$\beta,\beta\text{-annulated bacteriochlorins}$$

Chart 2 Structures of bacteriochlorins bearing β , β - or β ,meso-imides^{5,6} and bacteriochlorins with β , β - or β ,meso-annulated aromatic hydrocarbons.

configurations derived from one-electron promotions among the four orbitals dominate the spectral properties of the molecules. The HOMO is the $a_{1u}(\pi)$ -like orbital (using D_{4h} nomenclature), the HOMO-1 is $a_{2u}(\pi)$ -like and LUMO and LUMO+1 are akin to $e_{gx}(\pi^*)$ and $e_{gy}(\pi^*)$. Also shown in Fig. 4 are the next two filled orbitals (HOMO-2 and HOMO-3) and next two empty orbitals (LUMO+2 and LUMO+3). The four frontier MOs (HOMO-1 through LUMO+1) of Phen-BC (Fig. 4B) and Benz-BC (Fig. 4C) are generally similar to those for Es-BC but show significant electron density delocalized onto the annulated rings for certain orbitals. This delocalization is equally or more pronounced for the HOMO-3, HOMO-2, LUMO+2 and LUMO+3, which show additional characteristics that differ from those from the benchmark.

At the lower left of each of the three main panels of Fig. 4 are the transition dipole moment directions for absorption from the ground state to several singlet excited states. The directions are obtained from TDDFT calculations. For Es-BC, the transition

EtO₂C
$$\longrightarrow$$
 NH N \longrightarrow N HN \longrightarrow CO₂E \longrightarrow NH N \longrightarrow N HN \longrightarrow CO₂E \longrightarrow NH N \longrightarrow N HN \longrightarrow CO₂E \longrightarrow NH N \longrightarrow NH

Chart 3 Bacteriochlorin benchmarks. 5,6,84-86 The numbers in parentheses identify the data points in the energy-gap plot (vide infra).

from S_0 to S_1 (Q_y) is along the axis that contains the N-H bonds (the y-axis) and the transition to $S_2(Q_x)$ is essentially perpendicular to that. The S_1 transition is also along the y-axis for Phen-BC, but the S_2 transition is notably rotated off the x-axis. In comparison, the S_2 transition for **Benz-BC** is approximately x-polarized, and the S₁ transition is mainly y-polarized but rotated somewhat off the y-axis and directed toward the annulated rings.

Additional results of the TDDFT calculations are given in Fig. 5-7 for Es-BC, Phen-BC and Benz-BC, respectively. Each figure compares the measured absorption spectrum (brown) with one calculated by TDDFT shown as sticks and an associated line plot in which calculated transitions are given Gaussian skirts (dashed blue). The calculations for all three molecules reproduce the Q_y band $(S_0 \rightarrow S_1)$ in the NIR, the Q_x band $(S_0 \to S_2)$ in the red-green, and nominal $B_x (S_0 \to S_3)$ and B_{ν} (S₀ \rightarrow S₄) bands in the blue-violet (Soret) region. The calculations also reproduce the observed bathochromic shift of the Q_{ν} band in progressing from Es-BC to Phen-BC to Benz-BC, and for the two annulated molecules, place one additional transition in the Soret region that carries significant oscillator strength and is located between the nominal B_x and B_y bands. This latter prediction can be seen by examining the percent contribution of the various one-electron promotions to the transitions (e.g. HOMO \rightarrow LUMO, HOMO-1 \rightarrow LUMO+1), which are given in Tables S10-S12 (ESI†).

The energies of the various primary, i.e. (0,0), transitions of Es-BC, Phen-BC, and Benz-BC are reasonably well reproduced by the TDDFT calculations; however, the relative intensities of certain transitions differ appreciably from those observed. Of particular note is the disparity in the observed (relatively weak)

Table 3 Photophysical data^a

Compound	Q _y abs (nm)	<i>Q_y</i> em (nm)	FWHM abs (cm ⁻¹)	Δ^b abs-em (cm ⁻¹)	S_1^c energy (eV)	τ _s (ns)	$\Phi_{ m f}$	$\Phi_{ m isc}$	$\Phi_{ m ic}$	$(k_{\rm f})^{-1}$ (ns)	$(k_{\rm isc})^{-1}$ (ns)	$(k_{\rm ic})^{-1}$ (ns)
Benz-BC	1033	1042	321	84	1.20	0.007	2×10^{-5}	≤0.01	0.99	35	_	0.01
Phen-BC	913	933	586	235	1.34	0.15	0.004	0.02	0.98	38	7.5	0.15
$\mathbf{I2}\text{-}\mathbf{BC}^d$	875	880	278	65	1.41	0.98	0.027	0.18	0.79	36	5.4	1.2
$\mathbf{I}\text{-}\mathbf{BC}^e$	818	823	346	74	1.51	1.9	0.064	0.51	0.43	30	3.7	4.4
$BC2^f$	818	838	594	292	1.50	1.6	0.14	0.31	0.55	11	5.2	2.9
$\mathbf{Es}\text{-}\mathbf{BC}^g$	753	758	286	88	1.64	3.3	0.13	0.56	0.31	25	8	11
$\mathbf{H}\text{-}\mathbf{BC}^h$	713	717	230	78	1.73	3.8	0.14	0.63	0.23	27	6.5	17

^a Data were acquired in toluene at room temperature. ^b Stokes shift between the Q_y absorption and fluorescence maxima. ^c Average energy of the Q_y absorption and fluorescence maxima. ^d From ref. 5. ^e From ref. 6. ^f Static absorption and fluorescence data as well as TA spectra are available (Fig. S4 and S5, ESI). g From ref. 84. h From ref. 85.

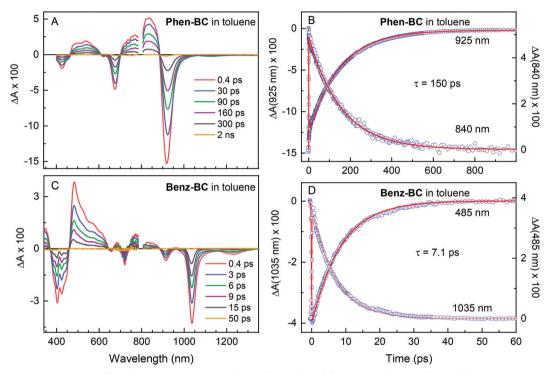


Fig. 3 Transient absorption data for Phen-BC excited at 715 nm (A and B) and Benz-BC excited at 675 nm (C and D) in toluene obtained using 100 fs flashes.

versus calculated (relatively strong) features in the Soret region for Phen-BC. We also note that TDDFT does not include prediction of vibronic transitions, i.e. (1,0), (2,0), etc., which are clearly visible to the blue of the $Q_v(0,0)$ and/or $Q_x(0,0)$ bands for all three molecules. The band of Benz-BC at ~913 nm, which exhibits an energy consistent with a $Q_{\nu}(1,0)$ vibronic satellite, is unusually intense for a tetrapyrrole-related molecule and could easily be mistaken for a primary band. The influence of the vibronic features of the annulated bacteriochlorins on the photophysical properties will be addressed further below.

Fig. 5-7 also give the natural transition orbitals (NTOs) for each calculated transition. The construction of these orbitals takes into account the contributing configurations to give the net electron-density distributions of the occupied orbital and the virtual (empty) orbital as if a single one-electron promotion were involved. 90 The NTOs provide a more physically meaningful

picture of the electron density change involved in an optical transition than can be gleaned from the orbitals themselves. Ideally, the contributions from various configurations should be distilled down into one pair of NTOs (filled and empty) for a given transition. However, for all three molecules for which TDDFT calculations were performed here (Es-BC, Phen-BC and Benz-BC) two pairs of NTO promotions are involved, with relative contributions indicated by the eigenvalues (from analysis of the transition density matrices) given in Fig. 5-7. This points to the underlying complexity and that both pairs of NTOs should be taken into account in assessing the net electron-density change accompanying the transition.

The situation is relatively straightforward for benchmark **Es-BC** (Fig. 5). The S_1 (Q_v) transition is comprised mainly (~90%) of a promotion between an $a_{1u}(\pi)$ -like filled NTO to an $e_{ex}(\pi^*)$ -like empty NTO and a small (~10%) contribution

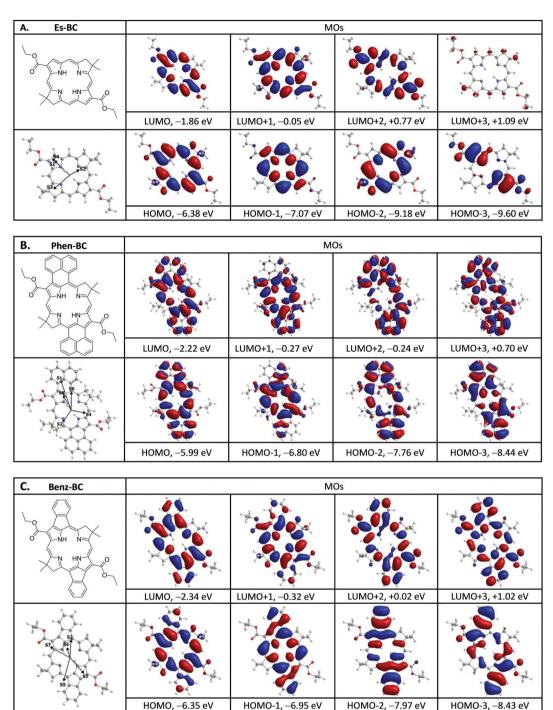


Fig. 4 Structure, transition-dipole directions for absorption from S₀ to excited states that carry significant oscillator strength in the near-UV to NIR regions (S₁, S₂, etc.), and frontier MOs for Es-BC (A), Phen-BC (B) and Benz-BC (C).

from $a_{2u}(\pi)$ -like NTO to an $e_{gy}(\pi^*)$ -like NTO. On the other hand, the S_4 (B_v) transition has the reverse contributions. The compositions of these two y-polarized transitions are consistent with expectations based on the four-orbital model. Similarly, the $S_2(Q_x)$ and $S_3(B_x)$ transitions are comprised of mixtures of promotions between an $a_{1u}(\pi)$ -like filled NTO to an $e_{gy}(\pi^*)$ -like empty NTO and between an $a_{2u}(\pi)$ -like NTO to an $e_{ex}(\pi^*)$ -like NTO. Examination of the NTOs for the Phen-BC (Fig. 6) and Benz-BC (Fig. 7) show some of the same characteristics,

modified or supplemented by effects of electron delocalization onto the annulated rings.

Discussion

Synthesis

While numerous β , β - and β ,meso-annulated porphyrins are known, 7,8 hardly any such bacteriochlorins have heretofore **Paper**

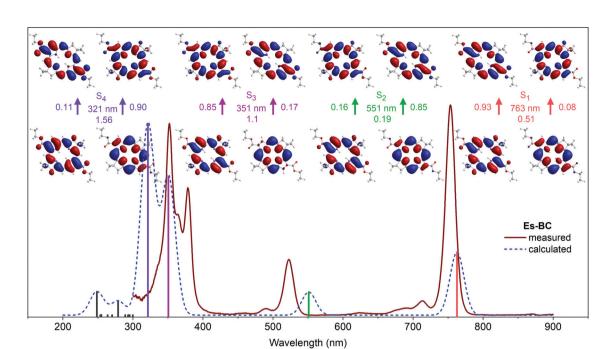


Fig. 5 Comparison for **Es-BC** of the absorption spectra in toluene that are measured (brown line) or calculated *via* TDDFT (dashed blue lines and colored sticks) normalized at the highest peak. Calculated spectral features were given 10 nm Gaussian skirts. Shown for the lowest four transitions (S_0 to S_1-S_4) are the natural transition occupied and virtual orbitals (NTOs) along with the wavelength and oscillator strength of the transition obtained from TDDFT. Two NTO promotions contribute to each transition, with weights (eigenvalues) indicated by the values that flank the arrows.

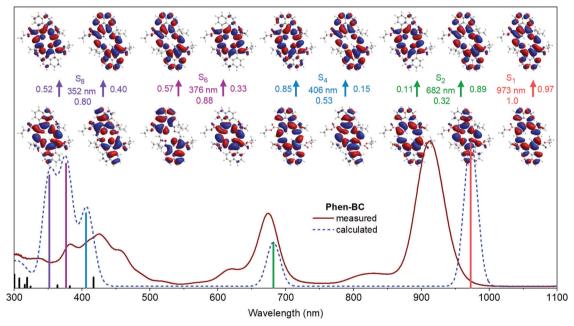


Fig. 6 Comparison for **Phen-BC** of the absorption spectra in toluene that are measured (brown line) or calculated *via* TDDFT (dashed blue lines and colored sticks) normalized at the highest peak. Calculated spectral features were given 10 nm Gaussian skirts, except for $S_0 \rightarrow S_1$ and $S_0 \rightarrow S_2$, which were set at 20 nm. Shown for the lowest five transitions that carry significant oscillator strength (S_0 to S_1 , S_2 , S_4 , S_6 , and S_8) are the natural transition occupied and virtual orbitals (NTOs) along with the wavelength and oscillator strength of the transition obtained from TDDFT. Two NTO promotions contribute to some transitions, with weights (eigenvalues) indicated by the values that flank the arrows.

been synthesized. The finding that the established routes to porphyrins^{7–33} were largely unsuited, at least in our hands, was a surprise, although we were aware of other reactivity differences of bacteriochlorins *versus* porphyrins. The chemistry of

dihydrodipyrrins $^{45,47,76-79}$ also differs considerably from that of dipyrrins, where annulation has been achieved via several means. 91 We explored 7 distinct routes to the annulated bacteriochlorins. The routes to β,β -annulated bacteriochlorins

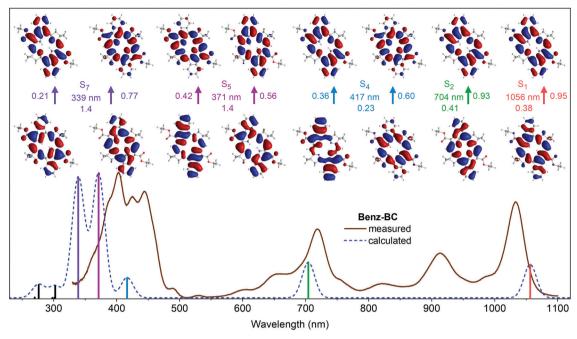


Fig. 7 Comparison for Benz-BC of the absorption spectra in toluene that are measured (brown line) or calculated via TDDFT (dashed blue lines and colored sticks) normalized at the highest peak. Calculated spectral features were given 10 nm Gaussian skirts. Shown for the lowest five transitions that carry significant oscillator strength (S₀ to S₁, S₂, S₄, S₅, and S₇) are the natural transition occupied and virtual orbitals (NTOs) along with the wavelength and oscillator strength of the transition obtained from TDDFT. Two NTO promotions contribute to some transitions, with weights (eigenvalues) indicated by the values that flank the arrows.

include cyclization of adjacent appended methyl acrylate units (Scheme 1), synthesis beginning with annulated pyrroles (i.e., isoindoles, Scheme 2) and use of a non-conjugated β , β -annulated pyrrole (*i.e.*, a tetrahydroisoindole). The routes to β,meso-annulated bacteriochlorins include oxidative cyclization of an appended β- or meso-substituent on the bacteriochlorin (Scheme 3), cyclization of an appended β- or mesosubstituent on a dihydrodipyrrin (Scheme 5), and ultimately the successive Pd-catalyzed arylation shown in Scheme 6. Extensive information, procedures and accompanying literature concerning attempted routes are provided in the ESI† (Schemes S1-S10). Some subtleties of the syntheses warrant comment. In particular, the question may arise why two successive Pd-couplings are employed to install the \(\beta,meso\)-annulation as opposed to preinstallation of, say, a bromoaryl group in the beginning of the synthesis at the pyrrole stage.

β-Aryl groups are carried through the synthesis very successfully, as exemplified by the naphthyl group in Scheme 3. Scheme 3 shows two routes to dinaphthyl-diester bacteriochlorins, which differ (1) in the positions of the substituents relative to the position of the gem-dimethyl groups and the unfettered meso-position, (2) the means of creating the dihydrodipyrrin precursors, and (3) the position of the gem-dimethyl groups relative to the pyrrole. The bacteriochlorins with the naphthyl groups at the 2,12-positions (BC-4-7) were constructed with the longstanding "Eastern-Western" route, 42 whereas those at the preferred 3,13-positions (BC-8,9) were constructed with the more recent "Northern-Southern" route. 45 The successful annulations (Scheme 6) were achieved with bacteriochlorins

(BC-12,13) resembling BC-8,9 in substituent patterns. A present limitation of the Northern-Southern route is the requirement for a Pd-catalyzed coupling of an iodopyrrole and an alkynoic acid to construct the pyrrole-lactone leading to the dihydrodipyrrin. The first Pd-coupling in the annulation in Scheme 6 uses a 5-fold excess of the bromoarylboronic acid versus the bromo sites on the bacteriochlorin with acceptable yields (49%, 85%) on the basis of the limiting bacteriochlorin. Such an excess of a β-bromoaryl-substituted iodopyrrole would be far less practical at the start of a Northern-Southern synthesis. In principle, the same substitution pattern could be created via the Eastern-Western route, but additional steps would be required to access the requisite 2-formyl-3-ethoxycarbonyl-4-(bromonaphthyl)pyrrole. In summary, the studies here have achieved the synthesis of β,meso-annulated bacteriochlorins yet at the same time have highlighted significant unresolved limitations in bacteriochlorin chemistry.

Photophysical features

The trends in the absorption spectral positions among the two new annulated bacteriochlorins and several benchmark bacteriochlorins are reproduced by the TDDFT calculations. Phen-BC basically has a naphthalene unit fused with the macrocycle, and the Q_{ν} band is at 933 nm. **Benz-BC** basically has a benzene unit fused with the macrocycle, and the Q_{ν} band is at 1033 nm. The two annulated bacteriochlorins thus absorb at the low-energy edge of the first NIR region (~700-1000 nm) and the highenergy edge of the second NIR region (1000-1700 nm), 92,93 respectively. If the excited-state energies and spectral positions

simply followed the particle-in-a-box model, one would have expected the larger Phen-BC to absorb at longer wavelength (lower energy) than Benz-BC because the S1-S0 energy gap is inversely proportional to the square of the box size. Examination of the energy gaps between the relevant orbitals involved in the Q_{ν} transition (HOMO → LUMO and HOMO-1 → LUMO+1) suggests that the Q_{ν} band of **Phen-BC** would be at lower energy than that for Benz-BC. However, the measured spectra show the opposite order and this is supported by the TDDFT calculations.

A factor that the calculations must effectively take into account, and which is also integral to the four-orbital model, is the configuration-interaction (CI) energy. This parameter is sensitive to the electron density distribution in the macrocycle. We have shown this from analysis of tetrapyrrole-based panchromatic absorbers, where the CI energy, and not simply the MO energy gaps, is a dominant factor in determining the spectral properties.⁹⁴ Thus, we speculate that annulation of the bacteriochlorin perturbs the CI energy sufficiently to complement effects on the MO energies in governing the observed spectra, and in particular the position of the Q_{ν} band for the annulated bacteriochlorins. Spectral simulations using the four-orbital model support this conjecture.

The question arose as to whether the observed short excitedstate (S_1) lifetime for **Phen-BC** and **Benz-BC** was simply a matter of the long-wavelength of absorption, and commensurate low energy level of the excited-state in each case. Fig. 8 plots the rate constant for $S_1 \rightarrow S_0$ internal conversion obtained from analysis of the TA data for the annulated bacteriochlorins studied here, the benchmark bacteriochlorins noted above (Table 3) and a host of other tetrapyrroles (Chart 3)5,6,84-86 studied previously. The red line in Fig. 8 suggests that if Phen-BC and Benz-BC followed the normal energy-gap law for bacteriochlorins, the molecules would have much longer S₁ lifetimes than is observed. For example, one would have expected an S₁ lifetime of about 400 ps for Benz-BC, rather than the observed 7 ps. The blue line in Fig. 8 is a fit to the data for Benz-BC, Phen-BC, diimide I2-BC, and monoimides I-BC and MeO-I-BC, all of which have added connectivity (via fused conjugated motifs; see Chart 2) between nearby meso and β-pyrrole positions.^{5,6} The correlation suggests that the annulated β-meso motif must introduce some factor to enhance nonradiative internal conversion in these constructs over normal bacteriochlorins. The development of chromophores with fluorescence at wavelengths > 1000 nm is an active area of research. 95

A potential source of enhanced internal conversion in the annulated (and imide) bacteriochlorins that employ β-meso connectivity can be gleaned from the optical spectra in Fig. 3. In particular, note the extraordinarily intense and broad $Q_{\nu}(1,0)$ band at \sim 913 nm for **Benz-BC**. The breadth suggests that there are likely a number of unresolved vibronic components, and the intensity indicates that there are modes coupling Q_{ν} with higher energy states, such as B_{ν} or perhaps Q_x . For the imides, particularly I2-BC, the peak intensity in the nominal $Q_{\nu}(1,0)$ region for that molecule (780-820 nm) is normal, but there is a ladder of small overlapping features that extend up toward the $Q_{\nu}(0,0)$ band. Thus, internal conversion may be enhanced in constructs that utilize conjugated β-meso connectivity either via

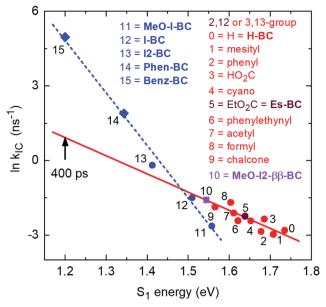


Fig. 8 Energy-gap law plot showing the natural logarithm of the rate constant for $S_1 \rightarrow S_0$ internal conversion versus the S_1 energy for various types of bacteriochlorins. The blue line is a fit for bacteriochlorins bearing one or two imide moieties or annulated rings spanning adjacent meso and β-pyrrole positions. The red line is a fit for the 10 benchmark bacteriochlorins from Chart 3 (and one β,β-annulated bis(imide)bacteriochlorin, from Chart 2). The latter line projects an S_1 lifetime of 400 ps for a "standard" bacteriochlorin as opposed to the observed lifetime of 7 ps for Benz-BC.

vibronic (Herzberg-Teller) coupling or via improved Franck-Condon factors through better energy matching due to the contribution of multiple vibrational levels.

The reduced excited-state lifetimes caused by β-meso annulation of bacteriochlorins does not occur upon addition of the 5-membered isocyclic ring that spans the 13–15 (meso–β) positions of native photosynthetic pigments such as bacteriochlorophyll a. Unlike the annulated bacteriochlorins studied here, the native 5-membered ring does not provide a conjugation path from meso to β-pyrrole positions across the isocyclic unit. Accordingly, the isocyclic ring serves as an auxochrome yet does not impart enhanced internal conversion.

Conclusions

Numerous routes established for annulation of porphyrins were not productive on application to bacteriochlorins. Instead, successive Suzuki-Miyaura coupling enabled construction of the β,meso-annulated bacteriochlorins. The photophysical studies of the β,meso-annulated bacteriochlorins indicate that this structural motif provides an effective means of bathochromically shifting the lowest energy electronic transition far into the NIR region of the spectrum. The magnitude of the shift is significantly larger than has been achieved via addition of typical substituents (e.g., acetyl, formyl, cyano) at the β and/or meso positions and also larger than that achieved to date via β,β-annulation. A striking photophysical characteristic of the

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β,meso-annulated bacteriochlorins is an extremely short lowestexcited singlet-state lifetime and accompanying vanishingly small fluorescence quantum yield. The photophysical studies trace these features to extremely rapid internal conversion to the ground electronic state. The origin of the rapid internal conversion is not certain; however, one unusual spectral feature of the two β,meso-annulated bacteriochlorins studied herein is exceptionally strong vibronic activity. The strong vibronic activity accompanied by the relatively low energy of the electronic transition possibly open more decay channels to the ground electronic state. The new molecules seem well suited for applications such as photoacoustic imaging⁹⁶ or photothermal therapy,⁹⁷ where an exquisitely short excited-state lifetime and the absence of fluorescence are desired, but not as NIR fluorescent markers or long-lived energy/electron donors in solar energy conversion.

Experimental section

General methods

¹H NMR and ¹³C NMR spectra were collected at room temperature in CDCl₃ unless noted otherwise. Silica (40 µm average particle size) was used for column chromatography. THF was freshly distilled from sodium/benzophenone ketyl. All other solvents (anhydrous or reagent-grade) were employed as received from commercial suppliers. Electrospray ionization mass spectrometry (ESI-MS) data generally enable accurate mass measurements, were obtained in the positive-ion mode (unless noted otherwise), and are reported for the molecular ion or protonated molecular ion. Laser-desorption ionization mass spectrometry without a matrix (LD-MS) or with a matrix (MALDI-MS) affords low resolution data, was carried out in the positive-ion mode with the matrix POPOP, and provided quick screening of reactions. Deactivated silica for purification of acid-labile compounds was prepared by treatment of silica with 5% triethylamine in hexanes (1 g/2 mL) and washing with CH2Cl2 followed by hexanes. Commercial compounds were used as received unless noted otherwise. NBS was recrystallized from water. The known compounds 1, 54 2, 68 8, 70 11, 71 15, 45 20, 45,73 and BC-9⁴⁵ were prepared as described in the literature.

Synthesis of bacteriochlorin precursors

tert-Butyl 3-formyl-2H-dibenzo[e,g]isoindole-1-carboxylate (3). Following a reported procedure, 44 DMF (1.0 mL, 13 mmol) was cooled in an ice bath, and POCl₃ (1.2 mL, 13 mmol) was added dropwise. An exothermic reaction occurred with the formation of the Vilsmeier complex. The ice bath was removed, and the mixture was stirred at room temperature for 15 min. The mixture was treated with CH₂Cl₂ (6 mL), and the reaction temperature was lowered to 0-2 °C with the aid of a salt-ice bath. A solution of 2 (1.27 g, 4.00 mmol) in CH₂Cl₂ (20 mL) was added to the stirred mixture over a period of 15 min. After the addition was completed, the ice bath was replaced with an oil bath and the mixture was refluxed for 15 min. The mixture was then cooled to 25-30 °C, a solution of sodium acetate (10 g) in water (40 mL) was added dropwise, and the reaction mixture was refluxed for an additional 15 min. The two layers were separated, and the aqueous portion of the mixture was extracted with CH2Cl2. The combined organic extract was washed with saturated aqueous Na2CO3 solution, dried (Na2SO4) and concentrated to afford a pale brown solid (1.14 g, 83%): mp 184–186 °C; ¹H NMR δ 1.73 (s, 9H), 7.45–7.62 (m, 4H), 8.16-8.19 (m, 1H), 8.43-8.47 (m, 2H), 9.49-9.53 (m, 1H), 10.32 (s, 1H), 10.35 (s, 1H); 13 C NMR δ 28.4, 83.5, 122.7, 123.0, 123.7, 124.2, 125.7, 125.9, 126.30, 126.33, 126.5, 127.1, 127.2, 127.3, 127.4, 127.9, 129.7, 130.8, 159.4, 179.6; ESI-MS obsd 346.1431, calcd 346.1438 $[(M + H)^+, M = C_{22}H_{19}NO_3]$.

tert-Butyl 3-(2-nitroethyl)-2H-dibenzo[e,g]isoindole-1carboxylate (4). Following a general procedure, 43 a stirred mixture of 3 (1.04 g, 3.00 mmol), potassium acetate (0.236 g, 2.40 mmol), and methylamine hydrochloride (0.162 g, 2.40 mmol) in absolute ethanol (15 mL) was treated with nitromethane (0.81 mL, 15 mmol). The reaction mixture was heated at 70 °C for 3 h, and then CH₂Cl₂ and water were added. The organic layer was dried (Na2SO4) and concentrated. The resulting vellow solid was dried under high vacuum and used directly in the next step. The crude solid material was dissolved in CHCl₃/2-propanol (3:1, 120 mL). Silica (6 g) and NaBH₄ (0.227 g, 6.00 mmol) were added, and the mixture was stirred at room temperature under argon for 30 min. The reaction mixture was filtered, and the filtrate was concentrated. The resulting crude material was dissolved in CH₂Cl₂. The organic solution was washed (water, brine), dried (Na2SO4), and concentrated. Column chromatography [silica, hexanes/ethyl acetate (10:1)] afforded a white solid (0.562 g, 48%): mp 168-170 °C; ¹H NMR (400 MHz) δ 1.73 (s, 9H), 3.79 (t, J = 7.2 Hz, 2H), 4.72 (t, J = 7.2 Hz, 2H), 7.47-7.63 (m, 4H), 7.82-7.85 (m, 1H), 8.49-8.54 (m, 2H), 9.73-9.75 (m, 1H), 9.79 (br s, 1H); ¹³C NMR $(100 \text{ MHz}) \delta 27.1$, 28.6, 72.9, 82.1, 115.8, 118.3, 122.4, 122.7, 123.0, 123.9, 124.7, 125.5, 126.9, 127.0, 127.3, 127.4, 127.8, 128.4, 129.5, 130.1, 160.0; ESI-MS obsd 391.1653, calcd 391.1652 $[(M + H)^{+}, M = C_{23}H_{22}N_{2}O_{4}].$

tert-Butyl 3-(3,3-dimethyl-2-nitro-5-oxohexyl)-2H-dibenzo[e,g]isoindole-1-carboxylate (5). Following an established procedure⁴⁴ with modification, samples of 4 (156 mg, 0.400 mmol), mesityl oxide (196 mg, 2.00 mmol), TBAF (1.0 M in THF, 0.6 mL, 0.6 mmol), and 4 Å molecular sieves (400 mg) were stirred together in DMF (8 mL) for 4 h. The mixture was then diluted with water (30 mL) and extracted with ethyl acetate (2 \times 30 mL). The combined organic extract was washed [HCl solution (2 M, 10 mL), saturated aqueous NaHCO3 solution and brine], dried (Na₂SO₄) and concentrated. The crude solid was triturated with diethyl ether (20 mL) followed by filtration to afford a white solid (0.132 g, 68%): mp 150–152 °C; 1 H NMR (400 MHz) δ 1.22 (s, 3H), 1.45 (s, 3H), 1.71 (s, 9H), 2.15 (s, 3H), 2.57 (ABq, $\Delta \delta_{AB} = 0.16$, J =18.0 Hz, 2H), 3.71 (d, J = 6.0 Hz, 1H), 3.73 (s, 1H), 5.45-5.49 (m, 1H), 7.51-7.62 (m, 4H), 7.93-7.95 (m, 1H), 8.53-8.59 (m, 2H), 9.67 (br s, 1H), 9.73–9.76 (m, 1H); 13 C NMR (100 MHz) δ 23.9, 24.6, 28.0, 28.6, 31.7, 37.2, 51.2, 81.9, 92.5, 116.1, 122.8, 122.96, 123.01, 124.0, 125.5, 126.8, 127.0, 127.1, 127.6, 128.1, 128.5, 129.7, 130.2, 159.9, 206.5; ESI-MS obsd 489.2387, calcd 489.2384 $[(M + H)^{+}, M = C_{29}H_{32}N_{2}O_{5}]$.

tert-Butyl (Z)-3-((3,3,5-trimethyl-3,4-dihydro-2H-pyrrol-2-ylidene)methyl)-2H-dibenzo[e,g]isoindole-1-carboxylate (6). Following a general procedure, 43 in a first flask a solution of 5 (120 mg,

0.246 mmol) in freshly distilled THF (5.0 mL) and anhydrous MeOH (100 μ L) at 0 °C was treated with NaOMe (66 mg, 1.2 mmol). The mixture was stirred and degassed by bubbling argon through the solution for 45 min. In a second flask purged with argon, TiCl₃ (1.54 mL, 20% w/v in 2 N HCl solution, 2.0 mmol), 8 mL of THF, and NH₄OAc (1.04 g, 13.5 mmol) were combined under argon, and the mixture was degassed by bubbling argon for 45 min. Then the first flask mixture was transferred via cannula to the buffered TiCl₃ mixture. The resulting mixture was stirred at room temperature for 16 h under argon. Saturated aqueous NaHCO3 was added, and the resulting mixture was then extracted with ethyl acetate. The organic extract was washed with brine, dried (Na2SO4), and concentrated. Column chromatography [silica, hexanes/ethyl acetate (10:1)] afforded a yellow solid (45 mg, 41%): mp 164–166 °C; 1 H NMR δ 1.37 (s, 6H), 1.74 (s, 9H), 2.34 (s, 3H), 2.64 (s, 2H), 6.68 (s, 1H), 7.47-7.66 (m, 4H), 8.40 (d, J = 7.5 Hz, 1H), 8.54–8.60 (m, 2H), 9.93 (dd, J =8.4 Hz and 1.2 Hz, 1H), 12.82 (br s, 1H); 13 C NMR δ 21.0, 28.8, 29.3, 42.0, 54.0, 80.7, 103.1, 116.3, 118.5, 122.9, 123.77, 123.83, 124.9, 126.4, 126.8, 127.0, 127.7, 128.2, 128.5, 129.4, 130.0, 130.2, 160.5, 165.9, 180.8; ESI-MS obsd 439.2376, calcd 439.2380 [(M + H)⁺, $M = C_{29}H_{30}N_2O_2$].

tert-Butyl (Z)-3-((5-formyl-3,3-dimethyl-3,4-dihydro-2H-pyrrol-2ylidene)methyl)-2H-dibenzo[e,g]isoindole-1-carboxylate (7). Following a general procedure, 44 a solution of 6 (45 mg, 0.10 mmol) in 1,4-dioxane (2.0 mL) was treated with SeO₂ (33 mg, 0.30 mmol) under argon. The mixture was stirred for 1.5 h at room temperature. The reaction mixture was treated with saturated aqueous NaHCO3 solution and extracted with ethyl acetate. The organic extract was washed with brine, dried (Na₂SO₄), and concentrated. Column chromatography [silica, hexanes/ethyl acetate (10:1)] afforded a dark orange solid (25 mg, 55%): mp 216–218 °C; ¹H NMR (400 MHz) δ 1.42 (s, 6H), 1.74 (s, 9H), 2.85 (s, 2H), 7.13 (s, 1H), 7.54–7.67 (m, 4H), 8.36 (d, J = 8.0 Hz, 1H), 8.58-8.62 (m, 2H), 9.85-9.87 (m, 1H), 10.08 (s, 1H), 12.38 (br s, 1H); 13 C NMR δ 28.6, 29.3, 41.6, 46.2, 81.6, 112.4, 119.4, 121.2, 122.9, 123.9, 124.0, 124.7, 125.8, 126.2, 126.7, 127.0, 127.2, 127.5, 128.3, 128.4, 130.1, 130.4, 164.3, 171.5, 189.6; ESI-MS obsd 453.2169 calcd 453.2173 [$(M + H)^+$, $M = C_{29}H_{28}N_2O_3$].

4-(Ethoxycarbonyl)-2-formyl-3-(naphthalen-1-yl)pyrrole (9). A solution of 8 (16.9 g, 63.7 mmol) in DMF (300 mL) was treated with POCl₃ (7.1 mL, 76 mmol) at 0 °C. The resulting mixture was stirred under argon at 0 °C for 1 h, and then 16 h at room temperature. The reaction mixture was treated with a mixture of saturated aqueous sodium acetate/CH₂Cl₂ [400 mL, 3:1 (v/v)] and stirred for 1 h. The water phase was separated and extracted with CH2Cl2. The combined organic phase was washed with a saturated LiCl solution, dried (Na2SO4) and concentrated. The resulting solid was chromatographed [silica, hexanes/ethyl acetate (2:1)] to give a light-brown solid (12.9 g, 69%): mp 147–150 °C; ¹H NMR (300 MHz, CDCl₃) δ 0.73 (t, J =7.2 Hz, 3H), 3.82-4.04 (m, 2H), 7.32-7.58 (m, 4H), 7.65 (d, J =7.4 Hz, 1H), 7.80 (s, 1H), 7.90 (t, J = 6.9 Hz, 2H), 9.18 (d, J = 1.1Hz, 1H), 11.10 (br s, 1H); 13 C NMR (75 MHz, CDCl₃) δ 13.7, 60.2, 118.7, 125.1, 126.1, 126.3, 126.5, 128.4, 128.7, 128.9, 129.9, 130.9, 131.7, 133.5, 135.9, 163.8, 181.1; anal. calcd for C₁₈H₁₅NO₃: C, 73.71; H, 5.15; N, 4.78. Found: C, 73.41; H,

4.99; N, 4.81. ESI-MS obsd 316.0937, calcd 316.0944 $[(M + Na)^+, M = C_{18}H_{15}NO_3]$.

4-(Ethoxycarbonyl)-3-(naphthalen-1-yl)-2-(2-nitroethyl)pyrrole (10). Following a general procedure, 43 a stirred mixture of 9 (12.6 g, 43.0 mmol), potassium acetate (3.3 g, 34 mmol), and methylamine hydrochloride (2.3 g, 34 mmol) in absolute ethanol (16 mL) was treated with nitromethane (6.0 mL, 0.11 mol). The mixture was stirred for 2 h, whereupon water was added. The reaction mixture was filtered, and the filtered material was washed with water and a small amount of cold water/ethanol (1:1). The filtered material was dried under high vacuum to afford a yellow solid, which was used directly in the next step. The crude solid material was dissolved in CHCl₂/2-propanol (3:1, 390 mL). Silica (34 g) and NaBH₄ (3.0 g, 78 mmol) were added, and the mixture was stirred at room temperature under argon for 2 h. The reaction mixture was filtered, and the filtrate was washed with saturated aqueous NaHCO3. The organic layer was separated and concentrated to afford a pale brown solid (10 g, 69%): mp 48–52 °C; ¹H NMR (300 MHz, CDCl₃) δ 0.73 (t, J = 7.0 Hz, 3H), 2.98 (dt, J = 17.6, 6.5 Hz, 2H), 3.88 (dq, J = 18.7,3.6 Hz, 2H), 4.20 (t, I = 6.5 Hz, 2H), 7.28–7.60 (m, 5H), 7.85 (t, I =7.8 Hz, 3H), 8.96 (br s, 1H); 13 C NMR (75 MHz, CDCl₃) δ 13.8, 23.7, 59.6, 75.0, 116.8, 122.0, 124.2, 125.5, 125.9, 126.0, 126.1, 126.2, 127.88, 127.91, 128.4, 132.9, 133.7, 164.9; anal. calcd for C₁₉H₁₈N₂O₄: C, 67.44; H, 5.36; N, 8.28. Found: C, 66.95; H, 5.27; N, 7.86. ESI-MS obsd 361.1267, calcd 361.1154 $[(M + Na)^{\dagger}]$ $M = C_{19}H_{18}N_2O_4.$

6-(4-(Ethoxycarbonyl)-3-(naphthalen-1-yl)pyrrol-2-yl)-1,1dimethoxy-4,4-dimethyl-5-nitrohexan-2-one (12). Following a general procedure, 43 a mixture of 10 (9.3 g, 28 mmol) and 11 (8.8 g, 55 mmol) was treated with DBU (12.3 mL, 82.5 mmol). The reaction mixture was stirred at room temperature for 16 h under argon. A saturated solution of cold aqueous NH4Cl was added. The mixture was extracted with ethyl acetate, and the organic layer was washed with brine, dried (Na2SO4), and concentrated. Column chromatography [silica, CH₂Cl₂/ethyl acetate (9:1)] afforded an amorphous brown solid (7.9 g, 47%): ESI-MS obsd 497.2277, calcd 497.2282 $[(M + H)^+, M =$ C₂₇H₃₂N₂O₇]; NMR data were collected from pure fractions of the two existing rotamers. Rotamer A: 1 H NMR (300 MHz, CDCl₃) δ 0.78 (s, 3H), 0.82 (s, 3H), 1.24 (t, J = 7.2 Hz, 3H), 2.28, 2.45 (AB, $^2J =$ 18.2 Hz, 2H), 2.56 (ABX, ${}^{3}J_{BX} = 2.2$ Hz, ${}^{2}J_{AB} = 15.1$ Hz, 1H), 3.05 (ABX, ${}^{3}J_{AX} = 11.6 \text{ Hz}, {}^{2}J_{AB} = 15.1 \text{ Hz}, 1H), 3.28 (s, 3H) 3.32 (s, 3H),$ 3.82-4.01 (m, 2H), 4.22 (s, 1H), 4.92 (ABX, ${}^{3}J_{BX} = 2.2$ Hz, ${}^{3}J_{AX} =$ 11.6 Hz, 1H), 7.30-7.65 (m, 5H), 7.75-7.91 (m, 3H), 9.29 (br s, 1H); 13 C NMR (75 MHz, CDCl₃) δ 13.9, 23.6, 23.9, 25.5, 36.4, 44.7, 55.0, 55.1, 59.5, 77.1, 95.1, 104.4, 116.3, 121.8, 124.4, 125.5, 125.8, 126.1, 126.4, 127.7, 128.3, 128.5, 133.0, 133.5, 133.7, 165.1, 203.4. Rotamer B: ¹H NMR (300 MHz, CDCl₃) δ 0.74 (s, 3H), 0.76 (s, 3H), 1.24 (t, J = 7.2 Hz, 3H), 2.20, 2.43 (AB, $^{2}J = 18.2 \text{ Hz}, 2\text{H}, 2.80 \text{ (ABX, } ^{3}J_{\text{BX}} = 2.2 \text{ Hz}, ^{2}J_{\text{AB}} = 15.1 \text{ Hz}, 1\text{H},$ 2.99 (ABX, ${}^{3}J_{AX} = 11.6 \text{ Hz}$, ${}^{2}J_{AB} = 15.1 \text{ Hz}$, 1H), 3.29 (s, 3H), 3.30 (s, 3H), 3.77–4.04 (m, 2H), 4.16 (s, 1H), 4.55 (ABX, ${}^{3}J_{BX}$ = 2.2 Hz, $^{3}J_{AX} = 11.6 \text{ Hz}, 1\text{H}, 7.28-7.67 (m, 5\text{H}), 7.77-7.90 (m, 3\text{H}), 9.34$ (br s, 1H); 13 C NMR (75 MHz, CDCl₃) δ 13.8, 23.5, 23.7, 25.3, 36.4, 44.2, 55.07, 55.14, 59.6, 77.0, 94.8, 104.4, 116.5, 121.9,

124.4, 125.4, 125.8, 126.1, 126.4, 127.70, 127.71, 128.2, 133.1, 133.7, 133.9, 165.1, 203.3.

8-(Ethoxycarbonyl)-2,3-dihydro-1-(1,1-dimethoxymethyl)-7-(naphthalen-1-yl)-3,3-dimethyldipyrrin (13). Following a general procedure, 43 in a first flask, a solution of 12 (6.65 g, 13.4 mmol) in freshly distilled THF (30 mL) and anhydrous MeOH (1.0 mL) at 0 °C was treated with NaOMe (2.2 g, 40 mmol). The mixture was stirred and degassed by bubbling argon through the solution for 45 min. In a second flask purged with argon, TiCl₃ (50 mL, 20% wt/v in 3% HCl solution, 80 mmol), 125 mL THF, NH₄OAc (50 g, 650 mmol) and water (20 mL) were combined under argon, and the mixture was degassed by bubbling argon for 45 min. Then, the first flask mixture was transferred via cannula to the buffered TiCl₃ mixture. The resulting mixture was stirred at room temperature for 16 h under argon. The reaction mixture was then poured over a pad of Celite and eluted with ethyl acetate. The eluant was washed with a saturated aqueous solution of NaHCO₃. The organic layer was dried (Na2SO4) and concentrated. Column chromatography [silica, CH₂Cl₂/ethyl acetate (98:2)] afforded an amorphous brown solid (3.34 g, 56%): ¹H NMR (400 MHz, CDCl₃) δ 0.65 (t, J = 7.2 Hz, 3H), 0.98 (s, 3H), 1.08 (s, 3H), 2.59 (s, 2H), 3.48 (s, 6H), 3.75-3.96 (m, 2H), 5.07 (s, 1H), 5.52 (s, 1H), 7.30-7.47 (m, 3H), 7.48-7.55 (m, 1H), 7.67 (d, J = 2.9 Hz, 1H), 7.70-7.76 (m, 1H), 7.81-7.90 (m, 2 H), 11.31 (br s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 13.7, 29.1, 29.2, 40.5, 48.6, 54.9, 59.4, 102.8, 105.6, 116.6, 125.3, 125.5, 125.6, 127.1, 127.4, 128.1, 128.5, 130.5, 133.4, 133.7, 134.1, 161.6, 165.2, 175.7; ESI-MS obsd 447.2258, calcd 447.2278 [(M + H)⁺, M = $C_{27}H_{30}N_2O_4$].

4-(Ethoxycarbonyl)-2-iodo-3-(naphthalen-1-yl)pyrrole (14). Following a literature procedure⁴⁵ with slight modification, a stirred solution of 15 (3.98 g, 15.0 mmol) in anhydrous DMF (75.0 mL) was treated with NIS (3.38 g, 15.0 mmol) in batches at 0 °C. After 2 h, the reaction mixture was diluted with ethyl acetate (150 mL) and washed with water and brine. The organic layer was dried (Na2SO4) and filtered. The filtrate was concentrated and chromatographed [silica, hexanes/ethyl acetate (4:1)] to give a pale yellow crystalline solid (2.85 g, 48%): mp 199-200 °C; ¹H NMR (400 MHz, CDCl₃) δ 0.67 (t, J = 7.6 Hz, 3H), 3.80–3.98 (m, 2H), 7.32-7.40 (m, 2H), 7.42-7.48 (m, 1H), 7.48-7.54 (m, 1H), 7.57 (d, J = 8.4 Hz, 1H), 7.65 (d, J = 2.8 Hz, 1H), 7.82–7.94 (m, 2H), 8.78 (br s, 1H); 13 C NMR (100 MHz, CDCl₃) δ 13.3, 59.6, 117.2, 125.1, 125.4, 125.5, 126.2, 127.6, 128.0, 128.1, 128.2, 128.4, 129.1, 133.1, 133.3, 133.7, 164.1; ESI-MS obsd 392.0143, calcd 392.0142 $[(M + H)^+, M = C_{17}H_{14}INO_2].$

4-(Ethoxycarbonyl)-(E)-2-(1-(4,4-dimethyl-5-oxodihydrofuran-2(3H)-ylidene)methyl)-3-(naphthalen-1-yl)pyrrole (16). Following a literature procedure⁴⁵ with slight modification, a mixture of 14 (2.4 g, 6.1 mmol), 15 (1.55 g, 12.3 mmol), and BnNEt₃Cl (1.4 g, 6.1 mmol) in anhydrous acetonitrile (25.0 mL) and triethylamine (6.1 mL) in a Schlenk flask was deaerated by three freeze-pump-thaw cycles. Then, a sample of Pd(PPh₃)₄ (354 mg, 0.307 mol, 5 mol%) was added, and the resulting reaction mixture was further deaerated. The reaction mixture was stirred and heated at 80 °C for 24 h. After allowing to cool to room temperature, CH_2Cl_2 and water were added. The organic

layer was separated, dried (Na₂SO₄) and filtered. The filtrate was concentrated and chromatographed [silica, hexanes/ethyl acetate (3:1)] to afford a pale yellow crystalline solid (1.83 g, 76%): mp 202–203 °C; ¹H NMR (400 MHz, CDCl₃) δ 0.69 (t, J = 7.2 Hz, 3H), 1.07 (s, 3H), 1.14 (s, 3H), 2.50 (dd, J = 2.0, 16.0 Hz, 1H), 2.40 (dd, J = 2.0, 16.0 Hz, 1H), 3.78–3.98 (m, 2H), 5.93 (dd, J = 2.0, 2.0 Hz, 1H), 7.30–7.40 (m, 2H), 7.40–7.52 (m, 2H), 7.60 (d, J = 3.2 Hz, 1H), 7.63–7.67 (m, 1H), 7.79–7.87 (m, 2H), 8.50 (br s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 13.6, 24.6, 24.9, 39.6, 39.9, 59.5, 97.5, 116.5, 122.2, 125.07, 125.14, 125.5, 125.6, 126.2, 127.5, 128.17, 128.21, 132.8, 133.3, 133.5, 147.6, 165.0, 179.6; ESI-MS obsd 390.1701, calcd 390.1700 [(M + H)⁺, M = C₂₄H₂₃NO₄].

4-(Ethoxycarbonyl)-(E)-2-(1-(4,4-dimethyl-5-methylenedihydrofuran-2(3H)-ylidene)methyl)-3-(naphthalen-1-yl)pyrrole Following a standard procedure, 45 a solution of TiCp2Cl2 (3.045 g, 12.37 mmol) in toluene (33 mL) was treated dropwise with MeLi solution (17 mL in Et₂O, 27 mmol, 1.6 M) at 0 °C under an argon atmosphere. After 1 h at 0 °C, saturated aqueous NH4Cl solution was added. The organic layer was washed with water and brine, dried (Na₂SO₄), and filtered. The filtrate (now $\sim 30-40$ mL owing to solvent losses in handling) contained the Petasis reagent. The filtrate in its entirety was treated with 16 (1.02 g, 2.61 mmol) and additional TiCp₂Cl₂ (39 mg). The solution was heated to 80 °C in the dark for 6 h under argon. Afterward, the resulting mixture was allowed to cool to room temperature whereupon MeOH (3.1 mL), NaHCO₃ (130 mg), and water (31 µL) were added. The mixture was kept at 40 °C for 12 h with stirring and then filtered through Celite. The filtrate was concentrated under reduced pressure. Column chromatography [silica, hexanes/ethyl acetate (3:1)] afforded a yellow crystalline solid (733 mg, 72%): mp 158–159 °C; ¹H NMR (400 MHz, CDCl₃) δ 0.68 (t, J = 7.2 Hz, 3H), 1.07 (s, 3H), 1.13 (s, 3H), 2.26-2.50 (m, 2H), 3.76-3.98 (m, 3H), 4.24 (d, J = 2.4 Hz)1H), 5.64 (s, 1H), 7.28-7.38 (m, 2H), 7.38-7.50 (m, 2H), 7.54 (d, J = 3.6 Hz, 1H), 7.67 (d, J = 8.4 Hz, 1H), 7.74-7.84 (m, 2H),8.45 (br s, 1H); 13 C NMR (100 MHz, CDCl₃) δ 13.6, 27.3, 27.7, 40.2, 41.8, 59.4, 80.3, 92.3, 116.6, 120.6, 124.0, 125.2, 125.3, 125.4, 126.4, 127.2, 127.4, 128.0, 128.3, 133.3, 133.51, 133.54, 153.3, 165.1, 168.9; ESI-MS obsd 388.1913, calcd 388.1907 $((M + H)^+, M = C_{25}H_{25}NO_3).$

8-(Ethoxycarbonyl)-1-methyl-2,2-dimethyl-7-(naphthalen-1-yl)-2,3-dihydrodipyrrin (18). Following a literature procedure⁴⁵ with slight modification, a solution of 17 (660 mg, 1.7 mmol) in DMF (17 mL) was treated with 1 M HCl (1.0 mL). After 30 min, NH₄OAc (2.63 g, 34.1 mmol) and triethylamine (4.8 mL, 35 mmol) were added. The resulting mixture was stirred at 55 °C for 16 h. The reaction was quenched by the addition of saturated aqueous KH₂PO₄ solution. The resulting mixture was extracted with ethyl acetate (100 mL). The organic layer was washed with water, dried (Na₂SO₄), and filtered. The filtrate was concentrated and chromatographed [silica, hexanes/ethyl acetate (3:1)] to afford a lightyellow crystalline solid (520 mg, 79%): mp 73–75 °C; ¹H NMR (400 MHz, CDCl₃) δ 0.65 (t, J = 7.2 Hz, 3H), 1.12 (s, 3H), 1.14 (s, 3H), 2.15 (s, 3H), 2.39 (q, J = 16.8 Hz, 2H), 3.74-3.96 (m, 2H), 5.47 (s, 1H), 7.30–7.46 (m, 3H), 7.50 (t, J = 7.6 Hz, 1H), 7.64 (d, J =3.2 Hz, 1H), 7.71 (d, J = 8.4 Hz, 1H), 7.82 (d, J = 8.0 Hz, 1H), 7.85 (d, J = 8.0 Hz, 1H), 11.44 (br s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 13.5, 15.7, 25.6, 25.7, 43.9, 48.4, 59.1, 104.2, 116.2, 120.7, 124.5, 125.1, 125.3, 125.4, 126.9, 127.1, 127.9, 128.3, 130.8, 133.4, 133.6, 134.0, 150.5, 165.1, 187.5; ESI-MS obsd 387.2076, calcd 387.2067 [(M + H)⁺, M = C₂₅H₂₆N₂O₂].

8-(Ethoxycarbonyl)-1-(1,1-dimethoxymethyl)-2,2-dimethyl-7-(naphthalen-1-yl)-2,3-dihydrodipyrrin (19). Following a literature procedure⁴⁵ with slight modification, a solution of 18 (193 mg, 0.499 mmol) in 1,4-dioxane (9 mL) was treated with SeO₂ (166 mg, 1.50 mmol) at room temperature. After 30 min, ethyl acetate and water were added. The organic layer was washed with brine, dried (Na2SO4), and concentrated under reduced pressure. The residue was dissolved in HC(OMe)₃ (4.0 mL) and treated with p-TsOH·H₂O (29 mg, 0.15 mmol) at room temperature. After 12 h, the reaction mixture was quenched by the addition of saturated aqueous NaHCO3 solution and then extracted with ethyl acetate. The organic layer was dried (Na₂SO₄) and filtered. The filtrate was concentrated and chromatographed [silica, hexanes/ethyl acetate (4:1)] to afford a brown viscous liquid (93 mg, 42%): ¹H NMR (400 MHz, CDCl₃) δ 0.66 (t, I = 7.6 Hz, 3H), 1.20 (s, 3H), 1.23 (s, 3H), 2.30-2.54 (m, 2H), 3.46 (s, 3H), 3.47 (s, 3H), 3.70-4.00 (m, 2H), 5.13 (s, 1H), 5.62 (t, J = 2.0 Hz, 1H), 7.30-7.60 (m, 4H),7.60-7.80 (m, 2H), 7.80-8.00 (m, 2H), 11.27 (br s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 13.5, 25.9, 26.0, 45.6, 48.2, 54.46, 54.48, 59.1, 102.0, 107.4, 116.3, 121.8, 125.1, 125.2, 125.4, 125.5, 126.8, 127.2, 128.0, 128.3, 130.4, 133.3, 133.4, 133.9, 149.5, 165.0, 182.0; ESI-MS obsd 447.2288, calcd 447.2278 [(M + H)⁺, $M = C_{27}H_{30}N_2O_4$].

3-Bromo-4-ethoxycarbonyl-(E)-2-[(4,4-dimethyl-5-oxodihydrofuran-2(3H)-ylidene)methyl]pyrrole (21). A suspension of K₂CO₃ (497.6 mg, 3.600 mmol) in MeCN was treated with 2,4,6tribromophenol (661.6 mg, 2.000 mmol) at room temperature. After 30 min, the mixture was cooled to 0 °C. Compound 20 (263.3 mg, 1.000 mmol) followed by 2,4,4,6-tetrabromo-2,5cyclohexadienone (655.5 mg, 1.600 mmol) were added at 0 °C. After 1 h at 0 °C, the reaction mixture was diluted with ethyl acetate (10 mL). The mixture was washed with aqueous NaHSO₃ (10%, 10 mL), saturated aqueous NaHCO₃ (10 mL), and brine (10 mL). The organic layer was dried (Na₂SO₄) and concentrated under reduced pressure. Column chromatography [silica, hexanes/ ethyl acetate (7:3)] afforded a pale orange solid (210.3 mg, 61%): mp 81 °C (dec.); ¹H NMR (400 MHz, CDCl₃) δ 1.35 (s, 6H), 1.36 (t, J = 7.2 Hz, 3H), 2.91 (d, J = 2.0 Hz, 2H), 4.31 (q, J = 7.2 Hz, 2H), 6.21 (t, J = 2.0 Hz, 1H), 7.46 (d, J = 3.2 Hz, 1H), 8.59 (br s, 1H); 13 C NMR (100 MHz, CDCl₃) δ 14.4, 25.1, 40.3, 60.3, 96.7, 97.8, 115.2, 125.4, 126.1, 149.7, 163.8, 179.7; ESI-MS obsd 342.0328, calcd 342.0336 $[(M + H)^{+}, M = C_{14}H_{16}BrNO_{4}]$.

5-Bromo-4-ethoxycarbonyl-(E)-2-[(4,4-dimethyl-5-oxodihydrofuran-2(3H)-ylidene)methyl]pyrrole (22). A solution of 20 (39.5 mg, 0.150 mmol) in DMF (1.50 mL) at -20 °C was treated with NBS (26.7 mg, 0.150 mmol). After 30 min, the reaction mixture was allowed to warm to room temperature. After 9.5 h, saturated aqueous Na₂S₂O₄ (1 mL) and ethyl acetate (1 mL) was added. The mixture was washed with H₂O (5 mL) and brine (2 mL). The organic layer was dried (Na₂SO₄) and chromatographed [silica,

hexanes/ethyl acetate (4:1)] to afford the title compound (25.9 mg, 50%) as a white solid and 21 (9.2 mg, 18%) as a white solid. Data for the title compound: mp 75 °C (dec.); 1 H NMR (400 MHz, CDCl₃) δ 1.37 (s, 6H), 1.38 (t, J = 7.2 Hz, 3H), 2.94 (d, J = 1.8 Hz, 2H), 4.33 (q, J = 7.2 Hz, 2H), 6.10 (t, J = 1.8 Hz, 1H), 6.40 (d, J = 2.4 Hz, 1H), 8.74 (br s, 1H); 13 C NMR (100 MHz, CDCl₃) δ 14.6, 25.5, 40.1, 40.7, 60.5, 96.9, 104.8, 109.0, 116.1, 128.0, 149.0, 163.6, 179.7; ESI-MS obsd 342.0327, calcd 342.0336 [(M + H) $^+$, M = $C_{14}H_{16}BrNO_4$].

7-Bromo-8-ethoxycarbonyl-1,2,2-trimethyl-2,3-dihydrodipyrrin (23). Following a reported procedure⁴⁵ with modification, MeLi (6.52 mL, 10.4 mmol, 1.6 M in Et₂O) was added dropwise to a solution of TiCp₂Cl₂ (1.180 g, 4.740 mmol) in toluene (12.6 mL) at 0 °C. After 1 h at 0 °C, the reaction mixture was washed with aqueous NH₄Cl (6%, 11 mL), H₂O (11 mL), and brine (11 mL). The organic layer was dried (Na₂SO₄) and filtered. Samples of 21 (342.2 mg, 1.000 mmol) and additional TiCp₂Cl₂ (14.9 mg, 0.0598 mmol) were added to the filtrate containing the Petasis reagent at room temperature. The mixture was heated to 80 °C in the dark for 2.5 h and then allowed to cool to room temperature. NaHCO₃ (50 mg), MeOH (1.2 mL), and H₂O (12 μL, 0.1% v/v relative to MeOH) were added. The mixture was heated to 40 $^{\circ}\mathrm{C}$ for 14 h and filtered through Celite. The filtrate was concentrated under reduced pressure. DMF (9.6 mL) and aqueous HCl (6 M, 84 μ L, 0.50 mmol) were added to the residue at room temperature. Aqueous HCl (6 M, $3 \times 84 \mu L$, $3 \times 0.50 \text{ mmol}$) was added after 100, 110, and 240 min. After 3 h at room temperature, NH₄OAc (1.574 g, 20.42 mmol) and NEt₃ (2.84 mL, 20.4 mmol) were added. After 12 h at room temperature, the reaction mixture was quenched with aqueous KH₂PO₄ (10%, 10 mL) and extracted with CH₂Cl₂ (10 mL). The organic layer was washed with H₂O (40 mL) and brine (40 mL). The organic layer was dried (Na2SO4) and filtered. The filtrate was concentrated under reduced pressure. Column chromatography [silica, hexanes/ ethyl acetate (4:1)] afforded a yellow solid (219.0 mg, 65%): ¹H NMR (300 MHz, CDCl₃) δ 1.37 (s, 6H), 1.36 (t, J = 7.1 Hz, 3H), 2.14 (s, 3H), 2.60 (d, J = 1.8 Hz, 2H), 4.30 (q, J = 7.1 Hz, 2H), 5.98 (s, 1H), 7.45 (d, J = 2.7 Hz, 1H), 11.40 (br s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 14.6, 15.9, 25.6, 44.2, 48.6, 59.9, 95.5, 103.2, 114.4, 124.6, 131.1, 151.9, 163.9, 188.7; ESI-MS obsd 339.0703, calcd 339.0703 $[(M + H)^{+}, M = C_{15}H_{19}BrN_{2}O_{2}].$

Bis(7-bromo-8-carbethoxy-2,3-dihydro-1,2,2-trimethyldipyrrinato)palladium(u) (24). Following a reported method⁷⁵ with some modification, a solution of 23 (15 mg, 43 μmol) in CH₂Cl₂ (0.50 mL) was treated with Pd(OAc)₂ (19 mg, 86 μmol). The reaction mixture was stirred at r.t. for 2 h and then chromatographed [silica, hexanes/ethyl acetate (2:1 to 1:1)] to afford a yellow-green solid (6.0 mg, 36%): mp > 230 °C (dec.); ¹H NMR (CDCl₃, 400 MHz) δ 6.80 (s, 2H), 6.32 (s, 2H), 4.30–4.15 (m, 4H), 2.85 (d, J = 14.8 Hz, 2H), 2.65 (d, J = 14.8 Hz, 2H), 1.71 (s, 6H), 1.30–1.26 (m, 12H), 1.04 (s, 6H); ¹³C NMR (CDCl₃, 100 MHz) δ 195.6, 163.6, 139.4, 136.2, 131.7, 115.5, 110.5, 97.3, 59.1, 49.5, 43.8, 25.5, 23.3, 17.0, 14.6; ESI-MS obsd 781.0204, calcd 781.0211 [(M + H)⁺, M = C₃₀H₃₆Br₂N₂O₄Pd]; λ _{abs} (toluene) 364 nm.

7-Bromo-8-ethoxycarbonyl-1-(1,1-dimethoxymethyl)-2,2-dimethyl-2,3-dihydrodipyrrin (25). Following a literature procedure⁴⁵ with

slight modification, a solution of 23 (111.9 mg, 0.3299 mmol) and pyridine (53 µL) in 1,4-dioxane (6.60 mL) was treated with SeO₂ (73.2 mg, 0.660 mmol) at room temperature. After 5 h, the reaction mixture was diluted with ethyl acetate (10 mL) and filtered through Celite. The filtrate was washed with H2O (10 mL) and brine (10 mL). The organic layer was dried (Na₂SO₄), filtered, and concentrated under reduced pressure. The residue was treated with trimethyl orthoformate (3.30 mL) and p-TsOH·H₂O (12.6 mg, 0.066 mmol) at room temperature. After 1.5 h, the reaction mixture was quenched with saturated aqueous NaHCO₃ (5 mL) and extracted with ethyl acetate (5 mL). The organic layer was washed with brine (5 mL), dried (Na₂SO₄), and filtered. The filtrate was concentrated and twice chromatographed [silica, hexanes/ethyl acetate (4:1) for the first chromatography, CH₂Cl₂/ethyl acetate (20:1) for the second chromatography] to afford a pale orange solid (48.3 mg, 37%): ¹H NMR (300 MHz, CDCl₃) δ 1.29 (s, 6H), 1.35 (t, J = 7.2 Hz, 3H), 2.63 (d, J = 1.8 Hz, 2H), 3.45 (s, 6H), 4.30 (q, J =7.2 Hz, 2H), 5.11 (s, 1H), 6.13 (t, J = 1.8 Hz, 1H), 7.48 (d, J = 1.8 Hz, 1H), 7. 3.6 Hz, 1H), 11.19 (br s, 1H); 13 C NMR (100 MHz, CDCl₃) δ 14.5, 26.0, 45.8, 48.5, 54.5, 59.9, 96.8, 101.9, 106.4, 114.6, 125.3,

(8-Bromonaphthalen-1-yl)boronic acid (26)^{82,83}. A solution of 1,8-dibromonaphthalene (1.430 g, 5.00 mmol) in THF at -60 °C was treated dropwise with *n*-butyllithium (3.34 mL, 1.6 M in hexane) over 5 min. After 1 h at −60 °C, trimethyl borate (668 μ L, 6.00 mmol) was added dropwise at -60 °C. Then, the reaction mixture was allowed to warm to room temperature. After 2.5 h, saturated aqueous NH₄Cl (50 mL) was added at room temperature. After 2.5 h, the mixture was extracted with ethyl acetate (50 mL). The organic layer was washed with brine (50 mL), dried (Na₂SO₄), and filtered. The filtrate was concentrated and chromatographed [silica, hexanes/ acetone (7:3)] to afford a white solid (823.9 mg, 59%): 1H NMR (400 MHz, acetone- d_6) δ 7.20 (s, 2H), 7.38 (dd, J = 8.0, 7.6 Hz, 1H), $7.52 \text{ (dd, } J = 8.0, 6.8 \text{ Hz, 1H)}, 7.63 \text{ (dd, } J = 6.8, 1.2 \text{ Hz, 1H)}, 7.83 \text{ (dd, } J = 6.8, 1.2 \text{ Hz, 1H})}$ J = 7.6, 1.2 Hz, 1H), 7.90 (dd, J = 8.0, 1.2 Hz, 1H), 7.93 (dd, J =8.0, 1.2 Hz, 1H); 13 C NMR (125 MHz, acetone- d_6) δ 124.2, 126.75, 126.83, 129.5, 129.6, 131.3, 132.4, 134.5, 136.0; ESI-MS obsd 272.6963, calcd 272.6963 $[(M + Na)^{+}, M = C_{10}H_9BBrO_2]$.

130.7, 150.9, 163.8, 183.1; ESI-MS obsd 399.0907, calcd

399.0914 $[(M + H)^{+}, M = C_{17}H_{23}BrN_{2}O_{4}].$

Synthesis of bacteriochlorins

2,3,12,13-Tetrabromo-8,8,18,18-tetramethylbacteriochlorin (BC-1). Following a standard procedure, ⁴² a solution of **1** (348 mg, 0.857 mmol, 18 mM) in anhydrous CH₃CN (48 mL) was treated with BF₃·OEt₂ (0.83 mL, 6.7 mmol, 140 mM). The reaction mixture was stirred at room temperature for 15 h. Excess triethylamine (2.0 mL) was added to the reaction mixture. The reaction mixture was concentrated, and the residue was chromatographed [silica, hexanes/CH₂Cl₂ (19:1)] to afford a green solid (46 mg, 16%): ¹H NMR (400 MHz, CDCl₃) δ –1.86 (br s, 2H), 1.96 (s, 12H), 4.43 (s, 4H), 8.75 (s, 2H), 8.88 (s, 2H); ¹³C NMR (150 MHz, CDCl₃) δ 30.97, 46.18, 51.38, 95.64, 97.85, 113.4, 113.5, 132.0, 132.9, 159.9, 171.9; ESI-MS obsd 681.8571, calcd 681.8573 [(M)⁺, M = C₂₄H₂₂Br₄N₄]; λ _{abs} (CH₂Cl₂) 347, 373, 496, 743 nm.

2,3,12,13-Tetrakis(methoxycarbonylvinyl)-8,8,18,18-tetramethylbacteriochlorin (BC-2). Following a reported procedure⁵⁹ with slight modification, bacteriochlorin BC-1 (12.0 mg, 17.5 µmol), Pd(OAc)₂ (11.9 mg, 53.0 μmol), PPh₃ (32.8 mg, 0.125 mmol) and K₂CO₃ (21.0 mg, 0.152 mmol) were added to a Schlenk tube and dried under high vacuum. The vacuum was released under argon to allow the addition of a solution of DMF (3 mL), toluene (3 mL) and methyl acrylate (175 µL, 1.93 mmol). The mixture was then deaerated via four freeze-pump-thaw cycles before the vessel was purged again with argon. The Schlenk flask was sealed and heated at 105 °C for 10 h. The resulting mixture was washed with water (20 mL) and extracted with CH_2Cl_2 (2 × 20 mL). The combined organic extract was dried (Na2SO4) and concentrated. The residue was first purified by chromatography [deactivated silica, hexanes/ CH_2Cl_2 (2:1 to 1:1)], then on preparative TLC to afford a reddish solid (4.0 mg, 32%): ¹H NMR (600 MHz, CDCl₃) δ -0.93 (s, 2H), 1.94 (s, 12H), 4.01 (s, 6H), 4.02 (s, 6H), 4.37 (s, 4H), 6.92 (d, J = 16.0 Hz, 4H), 8.69 (s, 2H), 8.83 (s, 2H), 8.97 (t, J = 16.4 Hz, 4H); ¹³C NMR (150 MHz, CDCl₃) δ 30.8, 46.2, 51.7, 52.1, 52.1, 95.1, 97.4, 124.5, 124.6, 126.8, 126.9, 134.0, 134.9, 137.1, 137.2, 161.3, 167.2, 167.3, 173.2; ESI-MS obsd 706.2990, calcd 706.2997 $[M^+, M = C_{40}H_{42}N_4O_8]; \lambda_{abs} (CH_2Cl_2) 319, 359, 393, 543, 815 nm.$

BC-3. Following an established procedure, ⁴⁴ a sample of 7 (21 mg, 0.046 mmol) was dissolved in TFA (2.6 mL) under argon and stirred for 2 h at room temperature. Then the reaction mixture was poured into saturated aqueous NaHCO₃ solution and extracted with CH₂Cl₂. The organic extract was washed with brine and water, dried (Na₂SO₄), and concentrated. The residue was washed with Et₂O followed by MeOH. The title compound could not be fully characterized because of instability; with only limited data obtained the assignment remains provisional: LD-MS obsd 670.8 [M⁺, M = C₂₈H₃₈N₄]; λ_{abs} (CH₂Cl₂) 801 nm; λ_{flu} (CH₂Cl₂) 808 nm.

3,13-Bis(ethoxycarbonyl)-2,12-di(naphthalen-1-yl)-8,8,18,18tetramethylbacteriochlorin (BC-4). Following an established procedure, 42 a solution of 13 (2.31 g, 5.17 mmol, 18 mM) in anhydrous CH₃CN (290 mL) was treated with BF₃·OEt₂ (5.1 mL, 41 mmol, 140 mM). The reaction mixture was stirred at room temperature for 16 h. The reaction was quenched by the addition of solid NaHCO₃ to the reaction mixture. The mixture was filtered. The filtrate was concentrated, and the residue was chromatographed [silica, CH2Cl2/hexanes (2:1)] to afford a purple/green solid (825 mg, 41%). The ¹H NMR spectrum indicated a mixture of atropisomers (~1:1 ratio): ¹H NMR (500 MHz, CDCl₃) δ -0.98 (br s, 4H), 0.66 (t, J = 7.0 Hz, 12H), 1.67 (m, 12H), 1.86 (m, 12H), 4.12-4.19 (m, 8H), 4.34-4.43 (m, 8H), 7.30 (t, J = 7.5 Hz, 4H), 7.54 (t, J = 7.5 Hz, 4H), 7.68 (d, J = 8.5 Hz, 4H), 7.81 (t, J = 7.5 Hz, 4H), 7.93 (d, J = 7.0 Hz, 4H), 8.07 (d, J = 8.0 Hz, 4H), 8.15 (d, J =8.0 Hz, 4H), 8.37 (s, 4H), 9.86 (s, 4H); 13 C NMR (100 MHz, CDCl₃) δ 13.5, 30.8, 30.95, 31.04, 46.1, 52.1, 60.7, 97.7, 99.4, 125.3, 126.1, 126.3, 127.0, 128.4, 128.6, 130.0, 133.8, 134.1, 134.6, 135.0, 135.1, 136.9, 161.8, 166.3, 172.5; ESI-MS obsd 767.3578, calcd 767.3592 $[(M + H)^{+}, M = C_{50}H_{46}N_{4}O_{4}]; \lambda_{abs} (CH_{2}Cl_{2}) 356, 381, 526, 768 \text{ nm}.$

3,13-Bis(ethoxycarbonyl)-2,12-di(naphthalen-1-yl)-5-methoxy-8,8,18,18-tetramethylbacteriochlorin (BC-5). Following an established procedure,⁴³ a solution of 13 (1.00 g, 2.24 mmol, 18 mM)

in HPLC-grade CH2Cl2 (124 mL) was treated first with DTBP (9.9 mL, 45 mmol, 360 mM) and second with TMSOTf (2.0 mL, 11 mmol, 90 mM). The reaction mixture was stirred at room temperature for 16 h. The reaction mixture was quenched by the addition of saturated aqueous NaHCO3. The organic phase was separated, concentrated, and chromatographed (silica, CH₂Cl₂). A single green band was isolated and concentrated to afford a purple/green solid (620 mg, 69%). The ¹H NMR spectrum indicated a mixture of atropisomers (~1:1 ratio): ¹H NMR (500 MHz, CDCl₃) δ -1.37 (br s, 2H), -1.05 (br s, 2H), 0.65-0.68 (m, 6H), 1.04–1.07 (m, 6H), 1.58–1.79 (m, 24H), 4.12–4.18 (m, 4H), 4.27 (s, 6H), 4.31-4.37 (m, 12H), 7.28-7.32 (m, 4H), 7.55 (m, 4H), 7.69 (t, J = 8.8 Hz, 2H), 7.77-7.83 (m, 6H), 7.91-7.93(m, 2H), 8.04-8.08 (m, 6H), 8.15 (d, J = 8.5 Hz, 4H), 8.18 (s, 2H),8.29 (s, 2H), 9.72 (s, 2H); 13 C NMR (100 MHz, CDCl₃) δ 13.4, 14.2, 30.8, 30.9, 31.0, 31.1, 45.7, 46.2, 48.2, 51.8, 60.5, 61.7, 64.7, 96.7, 98.3, 98.5, 120.7, 125.2, 125.5, 126.0, 126.2, 126.3, 127.02, 127.04, 127.7, 128.27, 128.31, 128.48, 128.53, 129.2, 130.0, 130.7, 130.8, 131.8, 133.0, 133.4, 133.7, 133.96, 134.01, 134.2, 134.6, 135.3, 135.5, 135.7, 136.5, 157.1, 161.7, 166.3, 168.1, 169.2, 172.9; ESI-MS obsd 797.3689, calcd 797.3697 $[(M + H)^{+}, M = C_{51}H_{48}N_4O_5];$ λ_{abs} (CH₂Cl₂) 371, 528, 748 nm.

In(III)X-3,13-Bis(ethoxycarbonyl)-2,12-di(naphthalen-1-yl)-8,8,18,18-tetramethylbacteriochlorin (BC-6). Following an established procedure, 72 a solution of BC-4 (25 mg, 0.033 mmol) in anhydrous THF (1.6 mL) was treated with NaH (13 mg, 0.33 mmol, 60% in mineral oil). The reaction mixture was stirred at room temperature for 5 min, whereupon the solution turned from purple/green to bright green. The mixture was then treated with InBr₃ (584 mg, 1.65 mmol), affording immediately a bright pink solution. The mixture was then stirred at 50 $^{\circ}$ C for 16 h. The reaction mixture was quenched by the addition of saturated aqueous NaHCO3. The organic phase was separated, concentrated, and chromatographed [silica, CH₂Cl₂/ethyl acetate (9:1)]. The first band (green) was the free-base starting material (10 mg), followed by a pink band (7.5 mg, 25%): ¹H NMR (400 MHz, CDCl₃) δ 0.63-0.68 (m, 6H), 1.50-1.52 (m, 3H), 1.72-1.79 (m, 6H), 1.87-1.90 (m, 3H), 3.87-4.16 (m, 4H), 4.33-4.42 (m, 2H), 4.51-4.60 (m, 2H), 7.17–7.22 (m, 2H), 7.34–7.39 (m, 1H), 7.42–7.51 (m, 2H), 7.57-7.61 (m, 1H), 7.64-7.67 (m, 1H), 7.73-7.77 (m, 1H), 7.84-7.88 (m, 2H), 7.96–7.98 (m, 1H), 8.04–8.16 (m, 4H), 8.27 (s, 1H), 8.30 (s, 1H), 9.82 (s, 1H); ESI-MS obsd 879.2387, calcd 879.2401 $[(M - X)^{+}, M = C_{50}H_{44}InXN_{4}O_{4}]; \lambda_{abs} (CH_{2}Cl_{2}) 355, 392, 565,$ 793 nm. The identity of the indium(III) counterion, X, is not known with certainty; the yield is calculated assuming X = Cl.

Zn(π)-3,13-Bis(ethoxycarbonyl)-2,12-di(naphthalen-1-yl)-5-methoxy-8,8,18,18-tetramethylbacteriochlorin (BC-7). Following an established procedure, ⁷² a mixture of BC-5 (48 mg, 0.060 mmol) and NaH (43 mg, 1.8 mmol) was added to DMSO (6.0 mL) under argon and stirred for 5 min. Zn(OAc)₂·2H₂O (658 mg, 3.0 mmol, 50 eq.) was then added and the suspension was stirred for 16 h in an oil bath at 60 °C. The crude mixture was washed with brine and extracted by ethyl acetate. The combined ethyl acetate extract was dried (Na₂SO₄), concentrated and chromatographed [silica, hexanes/ethyl acetate (3:2)] to afford a reddish solid (41 mg, 80%). The ¹H NMR spectrum indicated a mixture of atropisomers

(~1:1 ratio): 1 H NMR (500 MHz, CDCl₃) δ 0.570.61 (m, 6H), 1.02–1.07 (m, 6H), 1.56–1.79 (m, 24H), 3.97–4.08 (m, 4H), 4.18 (s, 6H), 4.27–4.35 (m, 12H), 7.27–7.31 (m, 4H), 7.49–7.55 (m, 4H), 7.70–7.87 (m, 10H), 7.92–7.99 (m, 2H), 8.03–8.12 (m, 10H), 8.19 (s, 2H), 9.61 (s, 2H); 13 C NMR (100 MHz, CDCl₃) δ 13.3, 14.1, 30.3, 30.7, 30.8, 44.7, 45.0, 47.2, 50.6, 60.0, 61.4, 64.3, 97.6, 99.3, 123.8, 123.9, 125.2, 125.5, 125.8, 125.9, 126.1, 127.4, 127.9, 128.0, 128.2, 128.3, 128.6, 129.5, 129.7, 130.0, 130.1, 132.9, 133.6, 134.0, 134.6, 135.4, 136.9, 139.7, 140.3, 144.5, 145.7, 146.5, 151.6, 156.2, 164.8, 167.1, 167.8, 169.5; MALDI-MS obsd 859.0533; ESI-MS obsd 858.2753, calcd 858.2754 (M † , M = C₅₁H₄₆N₄O₅Zn); λ _{abs} (CH₂Cl₂) 352, 384, 550, 762 nm.

2,12-Bis(ethoxycarbonyl)-10-methoxy-8,8,18,18-tetramethyl-3,13-di(naphthalen-1-yl)bacteriochlorin (BC-8). Following a literature procedure⁴⁵ with slight modification, a solution of 19 (39 mg, 0.087 mmol) in CH₂Cl₂ (4.4 mL) was treated with DTBP (400 μ L, 1.78 mmol) followed by TMSOTf (80 μ L, 0.44 mmol). The reaction mixture was stirred under a nitrogen atmosphere at room temperature for 15 h, and then diluted with CH2Cl2 and washed with saturated aqueous NaHCO3. The organic layer was dried (Na₂SO₄) and filtered. The filtrate was concentrated under reduced pressure. Column chromatography [deactivated silica, hexanes/CH2Cl2 (3:1 to 2:1)] afforded a dark purple solid (16.0 mg, 44%): the ¹H NMR spectrum indicated a mixture of atropisomers (~1:1 ratio): ¹H NMR $(600 \text{ MHz}, \text{CDCl}_3) \delta -1.20 \text{ (br s, 2H)}, -0.88 \text{ (br s, 2H)}, 0.67-0.72$ (m, 6H), 1.00-1.05 (m, 6H), 1.87-2.06 (m, 24H), 4.07-4.19 (m, 12H), 4.20-4.23 (m, 6H), 4.25-4.33 (m, 4H), 7.26-7.32 (m, 4H), 7.49-7.55 (m, 4H), 7.58-7.68 (m, 4H), 7.77-7.82 (m, 4H), 7.84-7.89 (m, 2H), 7.98-8.08 (m, 6H), 8.15 (d, J = 7.8 Hz, 4H), 8.21 (s, 2H), 8.27 (s, 2H), 9.60 (s, 2H); 13 C NMR (150 MHz, CDCl₃) δ 13.23, 13.77, 26.60, 29.22, 29.33, 29.71, 30.14, 30.70, 30.74, 30.90, 30.95, 46.10, 47.30, 53.83, 60.20, 61.32, 68.54, 95.99, 98.68, 100.1, 100.5, 115.2, 120.9, 125.0, 125.2, 125.6, 125.8, 126.0, 126.05, 126.06, 126.10, 126.9, 127.3, 128.03, 128.05, 128.2, 128.8, 129.3, 129.41, 129.44, 130.4, 130.5, 131.84, 131.86, 132.5, 132.78, 132.80, 133.4, 133.6, 133.9, 134.02, 134.05, 134.4, 136.4, 137.1, 137.2, 158.2, 160.0, 162.5, 165.7, 166.7, 167.5, 173.7; ESI-MS obsd 797.3697, calcd 797.3698 [(M + H)⁺, $M = C_{51}H_{48}N_4O_5$]; λ_{abs} (toluene) 362, 375, 531, 752 nm.

3,13-Dibromo-2,12-bis(ethoxycarbonyl)-8,8,18,18-tetramethylbacteriochlorin (BC-10). Following a literature procedure⁴⁵ with modification, TMSOTf (2.71 mL, 15.0 mmol) was added to a solution of 25 (798.6 mg, 2.000 mmol) and DTBP (13.2 mL, 60.0 mmol) in 1,2-dichloroethane (400 mL) at room temperature. The reaction mixture was heated at reflux for 11 h and then allowed to cool to room temperature. The reaction mixture was quenched with pyridine (1.61 mL, 200 mmol) and concentrated under reduced pressure. Column chromatography [deactivated silica, hexanes/CH₂Cl₂/MeCN (500:50:1 to 100:10:1)] afforded the title compound as a dark purple solid (196.5 mg, 29% and the 10-methoxy derivative thereof, namely 3,13-dibromo-2,12bis(ethoxycarbonyl)-10-methoxy-8,8,18,18-tetramethylbacteriochlorin (BC-11)) as a dark purple solid (250.2 mg, 36%). Data for the title compound: ¹H NMR (400 MHz, CDCl₃) δ 1.77 (t, J = 7.2 Hz, 6H), 1.94 (s, 12H), 4.38 (s, 4H), 4.82 (q, J = 7.2 Hz, 4H), 8.95 (s, 2H), 9.56 (s, 2H);

¹³C NMR (100 MHz, CDCl₃) δ 14.8, 31.0, 46.4, 51.4, 61.6, 97.7, 99.1, 115.6, 121.2, 132.4, 135.7, 161.1, 164.7, 173.8; ESI-MS obsd 671.0858, calcd 671.0863 $[(M + H)^+, M = C_{30}H_{32}Br_2N_4O_4]; \lambda_{abs}$ (toluene) 357, 385, 528, 731, 772 nm.

Data for BC-11: ¹H NMR (400 MHz, CDCl₃) δ 1.58 (t, J = 7.0 Hz, 3H), 1.74 (t, I = 7.0 Hz, 3H), 1.91 (s, 6H), 2.02 (s, 6H), 4.15 (s, 6H)(s, 3H), 4.31 (s, 2H), 4.34 (s, 2H), 4.76 (q, J = 7.0 Hz, 2H), 4.80 $(q, I = 7.0 \text{ Hz}, 2H), 8.72 (s, 1H), 8.87 (s, 1H) 9.41 (s, 1H); ^{13}C$ NMR (100 MHz, CDCl₃) δ 14.5, 14.8, 29.3, 30.9, 46.4, 47.4, 51.1, 54.0, 61.6, 62.4, 68.8, 96.5, 98.1, 99.5, 109.9, 116.2, 120.8, 125.7, 129.6, 131.3, 132.0, 136.3, 137.0, 159.1, 160.7, 164.6, 166.8, 167.1, 174.1; ESI-MS obsd 701.0974, calcd 701.0969 [(M + H)⁺, $M = C_{31}H_{34}Br_2N_4O_5$; λ_{abs} (toluene) 360, 372, 532, 719, 756 nm.

3,13-Bis(8-bromonaphthalen-1-yl)-2,12-bis(ethoxycarbonyl)-8,8,18,18-tetramethylbacteriochlorin (BC-12). A suspension of BC-10 (6.7 mg, 0.010 mmol), (8-bromonaphthalen-1-yl)boronic acid (25.1 mg, 0.100 mmol), K₂CO₃ (27.6 mg, 0.200 mmol), and $Pd(PPh_3)_4$ (1.2 mg, 0.0010 mmol) in toluene (333 µL) and DMF (167 µL) was deaerated by two freeze-pump-thaw cycles. The resulting mixture was heated to 100 °C for 14 h and then allowed to cool to room temperature. The mixture was passed through a basic alumina pad (CH₂Cl₂ as an eluent). The eluent was concentrated under reduced pressure. Column chromatography [deactivated silica, hexanes/CH2Cl2 (9:2)] afforded a dark purple solid (4.5 mg, 49%). ¹H and ¹³C NMR spectra indicated a mixture of atropisomers ($\sim 1:1$ ratio). Two distinguishable spots were found on TLC analysis with close $R_{\rm f}$ values [0.23 and 0.26, deactivated silica, 3 × hexanes/ethyl acetate (9:1)]. This sample was used as a mixture of atropisomers in the next step. Data for the title compound: ¹H NMR (400 MHz, $CDCl_3$) $\delta -1.0$ (s, 4H), 0.72 (t, J = 7.2 Hz, 12H), 1.90 (s, 6H), 1.91 (s, 6H), 1.95 (s, 6H), 1.96 (s, 6H), 4.32-4.06 (m, 16H), 7.38 (t, J = 7.8 Hz, 4H), 7.84-7.66 (m, 12H), 8.08 (d, J = 8.4 Hz, 4H), 8.16 $(d, J = 8.4 \text{ Hz}, 4\text{H}), 8.28 (s, 4\text{H}), 9.76 (s, 4\text{H}); {}^{13}\text{C NMR} (100 \text{ MHz}, 4\text{H})$ $CDCl_3$) δ 13.4, 30.9, 31.6, 31.2, 31.3, 46.4, 51.4, 60.2, 97.4, 100.2, 121.2, 121.3, 122.0, 125.1, 126.2, 129.3, 129.9, 132.0, 132.8, 132.9, 133.6, 133.8, 134.9, 135.80, 135.83, 135.9, 139.34, 139.36, 160.3, 165.9, 173.4; ESI-MS obsd 922.1715, calcd 922.1724 $[(M)^+, M = C_{50}H_{44}Br_2N_4O_4].$

Phen-BC. A suspension of BC-12 (8.6 mg, 0.0093 mmol), K₂CO₃ (12.9 mg, 0.0933 mmol), 2-dicyclohexylphosphino-2',6'dimethoxybiphenyl (11.5 mg, 0.0280 mmol), and Pd(OAc)₂ (2.1 mg, 0.0094 mmol) in DMF (1.00 mL) was deaerated by two freeze-pump-thaw cycles. The resulting mixture was heated to 80 °C for 5.5 h, 100 °C for 0.5 h, and then allowed to cool to room temperature. The mixture was diluted with CH2Cl2 and passed through deactivated silica. The eluent was concentrated under reduced pressure. Column chromatography [deactivated silica, hexanes/CH₂Cl₂ (11:9)] afforded a dark green solid (2.6 mg, 37%): 1 H NMR (300 MHz, CDCl₃) δ 1.57 (s, 12H), 1.72 (t, J = 7.0 Hz, 6H), 1.97 (s, 2H), 4.31 (s, 4H), 4.88 (q, J = 7.0 Hz, 4H), 7.58-7.80 (m, 4H), 7.76-8.02 (m, 4H), 8.30-8.48 (m, 2H), 8.35 (s, 2H), 8.84-9.06 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 14.8, 29.3, 46.1, 55.9, 61.7, 97.5, 110.1, 115.1, 126.4, 126.7, 126.8, 127.4, 127.5, 128.2, 128.3, 129.8, 133.5, 133.6, 134.0, 135.9, 161.6, 166.8, 168.2; ESI-MS obsd

762.3191, calcd 762.3201 $[(M)^+, M = C_{50}H_{42}N_4O_4]$; λ_{abs} (toluene) 306, 336, 383, 425, 622, 675, 829, 913 nm.

3,13-Bis(2-bromophenyl)-2,12-bis(ethoxycarbonyl)-8,8,18,18tetramethylbacteriochlorin (BC-13). A suspension of BC-10 (6.7 mg, 0.010 mmol), (2-bromophenyl)boronic acid (20.1 mg, 0.100 mmol), K₂CO₃ (27.6 mg, 0.200 mmol), and Pd(PPh₃)₄ (3.5 mg, 0.0030 mmol) in toluene (333 μ L) and DMF (167 μ L) was deaerated by two freeze-pump-thaw cycles. The resulting mixture was heated to 100 °C for 5 h and then allowed to cool to room temperature. The mixture was passed through a pad of basic alumina (CH2Cl2 as an eluent). The eluent was concentrated under reduced pressure. Column chromatography [deactivated silica, hexanes/CH₂Cl₂ (9:1 to 8:1)] afforded a dark purple solid (7.0 mg, 85%). ¹H and ¹³C NMR spectra indicated a mixture of atropisomers (~1:1 ratio). Two distinguishable spots were found on TLC analysis with close R_f values [0.33 and 0.37, deactivated silica, 3 × hexanes/ethyl acetate (9:1)]. This sample was used as a mixture of atropisomers in the next step. Data for the title compound: ¹H NMR (400 MHz, CDCl₃) δ –1.14 (s, 4H), 1.24 (t, J = 7.2 Hz, 12H), 1.94 (s, 12H), 1.96 (s, 6H), 1.97 (s, 6H), 4.27 (s, 8H), 4.44 (dq, I = 7.2, 10.7 Hz, 4H), 4.53 (dq, J = 7.2, 10.7 Hz, 4H), 7.47 - 7.58 (m, 4H), 7.58 - 7.69(m, 4H), 7.69-7.83 (m, 4H), 7.87-7.98 (m, 4H), 9.73 (s, 4H), 8.29 (s, 4H); 13 C NMR (100 MHz, CDCl₃) δ 14.0, 31.0, 31.1, 31.2, 31.3, 46.4, 51.4, 60.8, 97.7, 99.3, 120.57, 120.58, 125.91, 125.94, 126.80, 126.83, 129.6, 132.4, 132.5, 132.9, 133.0, 133.1, 135.6, 137.4, 137.5, 137.6, 160.6, 165.7, 173.6; ESI-MS obsd 822.1400, calcd 822.1411 $[(M)^+, M = C_{42}H_{40}Br_2N_4O_4]$.

Benz-BC. A suspension of BC-13 (52.0 mg, 0.0631 mmol), K₂CO₃ (43.6 mg, 0.315 mmol), tricyclohexylphosphine tetrafluoroborate (69.7 mg, 0.189 mmol), and Pd(OAc)₂ (14.2 mg, 0.0632 mmol) in DMF (6.3 mL) was deaerated by three freezepump-thaw cycles. The resulting mixture was heated to 110 °C for 2 h and then allowed to cool to room temperature. The mixture was diluted with CH2Cl2 (40 mL) and washed with H2O (50 mL) and brine (20 mL). The organic layer was dried (Na2SO4) and passed through deactivated silica. The eluent was concentrated under reduced pressure. The residue was washed with MeOH, MeCN, and hexanes to give a dark green solid (33.3 mg, 80%): ¹H NMR (400 MHz, CDCl₃) δ 1.69 (t, J =7.2 Hz, 6H), 1.78 (s, 12H), 2.89 (s, 2H), 4.00 (s, 4H), 4.71 (q, I =7.2, 4H), 6.97-7.04 (m, 2H), 7.04-7.10 (m, 2H), 7.49-7.54 (m, 2H), 8.02-8.06 (m, 2H), 8.71 (s, 2H); ¹³C NMR (175 MHz, $CDCl_3$) δ 14.9, 30.7, 46.5, 49.1, 61.1, 101.0, 113.1, 113.4, 123.6, 126.5, 127.4, 129.3, 136.5, 141.3, 142.1, 143.9, 149.1, 159.0, 165.3, 174.7; ESI-MS obsd 662.2881, calcd 662.2899 [(M)⁺, $M = C_{42}H_{38}N_4O_4$; λ_{abs} (toluene) 311, 403, 425, 442, 488, 658, 719, 821, 914, 1033 nm.

Photophysical measurements

Photophysical studies were carried out on samples in dilute (μM) argon-purged solutions of toluene. Excitation and detection bandwidths of 5 nm were employed for fluorescence spectra. Fluorescence quantum yields were determined for argon-purged samples relative to standards meso-tetraphenylporphyrin ($\Phi_{\rm f}$ = 0.070 in toluene with ambient O_2), 98 H-BC ($\Phi_f = 0.14$ in argon-purged

toluene)⁸⁵ and the zinc chelate of **I2-BC** ($\Phi_{\rm f}=0.031$ in argonpurged toluene).⁵ Each emission spectrum was integrated to include both the origin and vibronic satellite features. For each $\Phi_{\rm f}$ determination, the emission spectrum was acquired using at least three excitation wavelengths spanning the Soret and Q_x (and in some cases Q_y) regions with good agreement, and the results were averaged. S₁ lifetimes were determined by time-correlated single photon counting (TCSPC) fluorescence spectroscopy and by transient absorption (TA) spectroscopy, both employing ~ 100 fs excitation flashes from an ultrafast laser system.⁷³

Molar absorption coefficients

The molar absorption coefficients were determined by dissolving a known quantity of each bacteriochlorin (1.01–2.66 mg) in toluene (5.00 mL). Then a known amount (40.0–100.0 $\mu L)$ of this solution was diluted to 5.00 mL with toluene. The absorption spectrum was recorded at room temperature. The averages of 5 runs were calculated: BC-10 ($\epsilon_{756\mathrm{nm}}=36\,800~\mathrm{M}^{-1}~\mathrm{cm}^{-1}$, $\epsilon_{772\mathrm{nm}}=135\,000~\mathrm{M}^{-1}~\mathrm{cm}^{-1}$); BC-11 ($\epsilon_{756\mathrm{nm}}=117\,000~\mathrm{M}^{-1}~\mathrm{cm}^{-1}$, $\epsilon_{772\mathrm{nm}}=20\,800~\mathrm{M}^{-1}~\mathrm{cm}^{-1}$).

Density functional theory calculations

DFT calculations were performed with Gaussian 09 version D.01. 99 Calculations used the polarization continuum model for the arrays in toluene. Molecular geometries were fully optimized using the hybrid B3LYP functional and the basis set 6-31G*. These calculations used Gaussian defaults with the exception of keyword Int = (Grid = Ultrafine, Acc2E = 14). TDDFT calculations used the long-range corrected ω B97XD functional and the basis set 6-31++G**. These calculations used Gaussian defaults with the exception of keywords TD (nStates = 16), Int = (Grid = Ultrafine, Acc2E = 14), and Pop = Full.

Conflicts of interest

The authors declare no competing financial interests.

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