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COMMUNICATION

Visualization of carboxylesterase 2 with a near-infrared two-photon fluorescent probe and its potential evaluation of anticancer drug effects in orthotopic colon carcinoma mice model

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We establish a near-infrared two-photon fluorescent probe for the detection of CE2 with high selectivity and sensitivity. This probe exhibits low cytotoxicity and superior tissue penetration ability for evaluating real-time activity of CE2 in living cells, in cancer tissues, and in colon carcinoma mice model.

Carboxylesterases (CEs), a component of α/β hydrolase fold protein, plays pivotal roles in the hydrolysis of substances containing esters, amides, carbamates and thioester.¹⁻⁴ As a major isoform of CEs, CE2 is found to be high expressed in many cancer cells, and CE2 behaves in mediating the activation process of many prodrugs such as Gemcitabine, Irinotecan (CPT-11) and Capecitabine (CAPE).^{5, 6} Therefore, the individual differences in CE2 activity have been closely associated with the cytotoxicity and efficacy of the relevant clinical anticancer drugs.⁷ Furthermore, CE2 is considered to be the essential determinant in intestinal first-pass metabolism, especially for oral anticancer prodrugs.⁸ In view of the critical roles of CE2 in metabolism of various anticancer prodrugs, it is quite necessary to develop suitable techniques for accurate and sensitive detection of CE2 in the complex physiological conditions.

There have been established several effective methods for evaluating CE2 activity.^{9, 10} However, these technical methods usually need time-consuming and complex procedures, high cost, and skilled operators, which may largely restrict their application in rapid detection. As attractive alternatives, fluorescent probes have become indispensable tools toward exploring bioactive molecules.¹¹⁻¹³ Although small-molecule based fluorescent probes have been synthesized and utilized for the detection of CE2 *in vitro* and *in vivo*,¹⁴⁻¹⁶ few probes are

competent for practical applications mainly due to their relatively short excitation/emission wavelength where has strong intrinsic autofluorescence background from living tissues. This issue greatly restricts the deep-tissue penetration and measurement accuracy under physiological conditions. Compared to the single-photon confocal imaging, the developed two-photon (TP) microscopy features the ability to avoid biological autofluorescence and to reduce photodamage, because the near-infrared (NIR) wavelength can achieve deeper penetration depth and can exhibit higher 3D temporal spatial resolution.¹⁷⁻²⁰ Hence, a two-photon fluorescent probe displays attracted extensive research interest in sensing of bioactive macromolecule CE2 under sophisticated living systems. However, there still exist obstacles of these reported probes, which mainly lie in the deficiency with emission wavelength since most of these TP probes only can emit fluorescence at short wavelength (<550 nm).²¹ This issue may fail to the practical bio-imaging especially in deep tissues as the collection efficiency of fluorescence signal will be greatly decreased, further affecting the sensing accuracy and sensitivity of TP probes.^{22, 23} Therefore, it is of great significance to develop TP probes with excitation/emission wavelength in NIR region²¹ which can not only minimize background fluorescence interference owing to reducing absorption by bio-molecules, but also achieve deeper penetration to facilitate precise detection of CE2 in living systems.

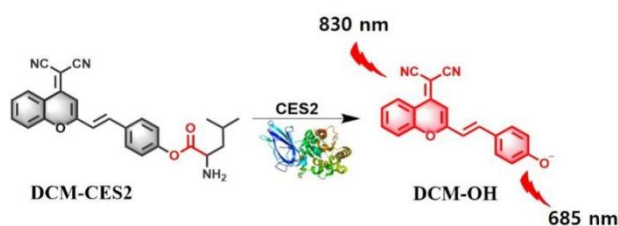
As illustrated in Scheme 1, the overall design strategy of the probe **DCM-CES2** was based on the attachment of an enzyme-active moiety L-leucine to an excellent NIR TP chromophore dicyanomethylene-4H-pyran (DCM),^{11,13,24} because it held large Stokes shifts and NIR emission (> 650 nm). Since the CE2 could specifically hydrolyze the compound with large alcohol and small acyl group, our probe **DCM-CES2** was able to serve as a highly selective substrate for CE2. The detailed synthetic route was shown in ESI[†]. The compounds were characterized by ¹H NMR, ¹³C NMR and HR-MS.

We inspected the spectral characteristics of the probe **DCM-CES2** in simulated physiological conditions (PBS buffer pH = 7.4, 10 mM at 37 °C). As shown in Fig. 1a, the probe **DCM-CES2**

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Scheme 1 Illustration of the molecular structure **DCM-CES2** and its proposed fluorescence response mechanism toward CE2.

provided characteristic absorption centred at 445 nm. With the treatment of CE2, a new maximum absorption peak appeared at 560 nm, which belonged to the released fluorophore DCM. The color changed from light yellow to rose red allowing to the colorimetric detection of CE2 *via* the naked eye, accompanied by a large Stokes shift of 125 nm in the fluorescence spectra. The reaction with CE2 would switch-on the fluorescence emission of **DCM-CES2** by restoring the intramolecular charge transfer (ICT) process, thereby offered a characteristic fluorescence emission peak centred at 685 nm in NIR region (Fig. 1b). The fluorescence intensity displayed a good linear relationship. The regression equation was $F_{560\text{ nm}} = 10.27 [\text{CE2}] + 6.963$ with a linear regression constant $r = 0.9963$ (Fig. 1c). Based on the standard method of $3\sigma/k$, the limit of detection for CEs was $0.087\text{ }\mu\text{g/mL}$, which indicated favourable sensitivity for the CE2 detection *in vivo*.²⁵ As indicated in Fig. 1d, there appeared a quick response within 5 min. Continuous monitoring demonstrated that the reaction completed within 10 min. The maximal TP absorption cross section (δ) was measured as 26 GM at 830 nm (Fig. S2, ESI[†]), implying that **DCM-CES2** was able to track CE2 *via* TP microscope. These results validated that the designed probe could serve as a near-infrared off-on fluorescent chemical tool for detecting CE2 under physiological conditions.

A series of reconstructed human enzymes with hydrolytic activity were selected to assess their possible interactions with **DCM-CES2** (Fig. 1e). **DCM-CES2** was incubated with the selected enzymes, respectively, only CE2 led to a remarkable increase in fluorescence intensity at 685 nm, while the other hydrolases exhibited negligible fluorescence changes. To further confirm the CE2 dependent fluorescence changes, we chose two representative esterase inhibitors to perform the chemical inhibition assays. CE2 was pre-incubated with the non-covalent inhibitor loperamide (LPA) and covalent inhibitor bis(4-nitrophenyl) phosphate (BNPP) followed by the addition of the probe **DCM-CES2**, respectively (Fig. 1e). Both inhibitors could restrict fluorescence emission compared with the CE2 group, which confirmed that the fluorescence change was dependent on CE2. The results indicated that **DCM-CES2** could serve as an effective chemical tool to detect CE2 with high selectivity in cells. We also determined the Michaelis constant (K_m). The value of K_m was $33.43\text{ }\mu\text{M}$, and the corresponding catalytic constant (K_{cat}) and catalytic efficiency (K_{cat}/K_m) were 3.75 s^{-1} and $0.17\text{ }\mu\text{M}^{-1}\text{ s}^{-1}$ respectively, demonstrating that CE2-mediated hydrolysis of **DCM-CES2** possessed good kinetic property, high catalytic activity, and powerful affinity (Fig. 1f).

We next utilized the probe **DCM-CES2** to detect the level of

CE2 in living cells. The cytotoxicity of the probe was evaluated by Cell Counting Kit-8 assay (Fig. S4, ESI[†]) with human hepatocellular liver carcinoma cell line (HepG2) and human colorectal cancer cell line HT-29. The viability rates of the selected model cell lines indicated the relatively low cytotoxicity. To test the capability of **DCM-CES2** responded to CE2 activity in living cells, HepG2 cells were treated under different treatment conditions. The addition of **DCM-CES2** could generate strong red fluorescence, which revealed the reactivity to the CE2-overexpressed system (Fig. S6h). Then two representative inhibitors, BNPP for CEs and LPA for CE2, were adopted to validate the specific detection of CE2 in cells. As expected, there appeared severe fluorescence suppression in Fig. S6i and S6j, respectively. The faint fluorescence was attributed to the inhibition of intracellular CE2 activity, but the CE2-selective inhibitor LPA could restrain almost all of fluorescence emission, demonstrating the specific detecting ability of our designed probe. We also pretreated the cells with 5-Fluorouracil (5-FU, $70\text{ }\mu\text{M}$, a CE2 inducer²⁶) for 48 h at $37\text{ }^\circ\text{C}$. Interestingly, a stronger fluorescence increase was observed in Fig. S6k, which conformed that the specific response to endogenous CE2 in living cells. The results obtained by flow cytometry assays (Fig. S6m – S6q) were in well accordance with the above imaging outcomes.

CE2 offered high activation effect on metabolism of anticancer prodrug Irinotecan (CPT-11) in HT-29 cells.²⁷ As the same situation, anticancer prodrug Capecitabine (CAPE) held a stronger tend to be firstly activated by CE2, and finally metabolized to drug 5-FU that could inhibit the growth of cancer cells.²⁸ In view of the crucial role of CE2 involved in the activation process of various anticancer prodrugs, it was quite necessary to visualize CE2 for better understanding of its metabolic mechanism in drug metabolism. We next attempted to utilize the probe **DCM-CES2** to detect the CE2 activity change

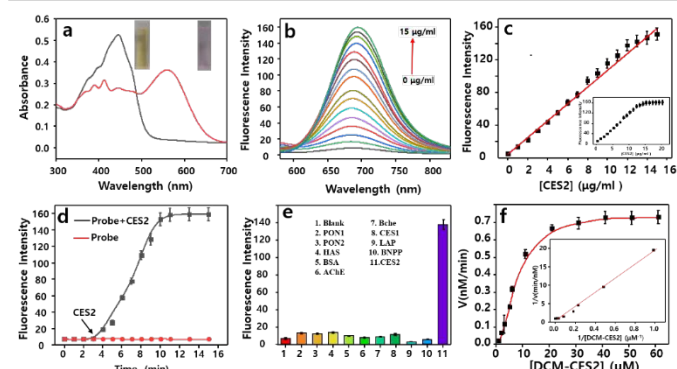


Fig. 1 Spectral characterization of **DCM-CES2** ($10\text{ }\mu\text{M}$). a) UV-Vis absorption spectrum with CE2 ($0 - 10\text{ }\mu\text{g/mL}$) for 10 min. b) The fluorescence spectrum with CE2 ($0 - 15\text{ }\mu\text{g/mL}$) for 10 min. c) Linear relationship between the fluorescence intensity and the activity of CE2. d) Time-dependent kinetic measurement of the fluorescent response to CE2 ($5\text{ }\mu\text{g/mL}$). e) Selectivity towards various species. Blank, Paraoxonase (PON1 and PON2, $10\text{ }\mu\text{g/mL}$), Human serum albumin (HSA, 0.5 mg/mL), Bovine serum albumin (BSA, 0.5 mg/mL), Acetylcholinesterase (AChE, $0.1\text{ }\mu\text{g/L}$), Butyrylcholinesterase (BChE, 20 U/L), CES1 ($10\text{ }\mu\text{g/mL}$), Loperamide (LAP, $100\text{ }\mu\text{g/mL}$), Bis(4-nitrophenyl) phosphate (BNPP, $100\text{ }\mu\text{g/mL}$), and CE2 ($10\text{ }\mu\text{g/mL}$). f) Lineweaver-Burk plot for the enzymatic reaction. The Michaelis-Menten equation was expressed as: $V = V_{\text{max}} [\text{probe}]/(K_m + [\text{probe}])$, where V was the reaction rate, [probe] referred to the probe concentration (substrate), and K_m was the Michaelis constant. $\lambda_{\text{ex}} = 560\text{ nm}$.

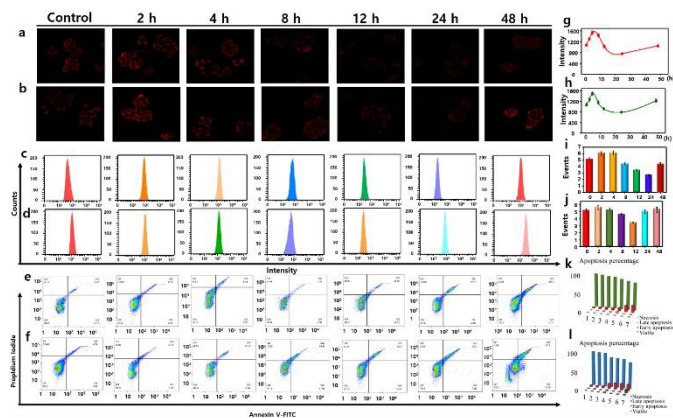


Fig. 2 Two-photon confocal fluorescence imaging ($\lambda_{\text{ex}} = 830 \text{ nm}$, $\lambda_{\text{em}} = 650 - 750 \text{ nm}$) and flow cytometry assay ($\lambda_{\text{ex}} = 560 \text{ nm}$, $\lambda_{\text{em}} = 650 - 750 \text{ nm}$) for detection of CE2 in HT-29 cells. a - b) HT-29 cells pretreated with CPT-11 and CAPE, respectively, and then incubated with **DCM-CES2**. c - d) Corresponding flow-cytometric analysis to a and b, respectively. e - f) PE Annexin V/7-AAD assays of HT-29 cells in a and b, respectively, using flow cytometry. Q1: necrosis cells, Q2: late apoptotic cells, Q3: early apoptotic cells, Q4: survival cells. g - h) Mean fluorescence intensities of the images in a and b, respectively. i - j) Mean values of c and d, respectively. k - l) Apoptosis percentages of e and f, respectively. The data were shown as mean (± s.d.).

during the activation procedure in living cells. HT-29 cells were parallelly divided into seven groups and treated with CPT-11 (20 μM) for 0, 2, 4, 8, 12, 24, 48 h, respectively, then incubated with the probe for 30 min (Fig. 2a and 2b). Upon treatment with CPT-11 for the initial 4 h, there appeared a fluorescence enhancement in the testing system, while continuous effect of CPT-11 gradually weakened the fluorescence signal as incubation time extended to 24 h. When the examination was continuous for 48 h, we noticed that the activity of CE2 gradually returned to the original level, which was in well agreement with the metabolic cycle of CPT-11.²⁹ We speculated that the cells were suffering from a stress response to metabolize the prodrug, which would promote CE2 activate stronger, but occupied by amount of prodrug molecules. Once the prodrug was decomposed, it would lead to the decreased vitality of CE2, thus reducing the obtained fluorescence intensity as the control group. Further evaluation was carried out to detect CE2 activation response to CAPE in HT-29 cells. As shown in Fig. 2b, the overall fluorescence trend was almost as same as that in Fig. 2a, except for acquiring a stronger fluorescence increase at 48 h. This phenomenon probably caused by the participant of 5-FU which not only served as a metabolite from CAPE through multiple enzymes including CE2, but also could be an inducer to stimulate the activity increase of CE2.²⁶ The results demonstrated that the different anticancer drugs offered the distinct activation process of CE2, which might instruct various cancer cells resistance mechanisms to different drugs. Flow analysis was shown in Fig. 2c and 2d. In addition, PE Annexin V/7-AAD assays of HT-29 cells was also studied to illustrate cell apoptosis caused by both anticancer drugs (Fig. 2e and 2f). All of these results suggested that **DCM-CES2** could be utilized to detect the crucial role of CE2 during anti-cancer drugs activation process.

We further evaluated the *in vivo* applicability of our NIR-emissive TP probe. We performed TP imaging of CE2 in

subcutaneous intestinal cancer slices (100 μm) to validate the deep imaging capability of our probe. The tissue slice was incubated with **DCM-CES2** (20 μM) for 60 min, then washed with PBS for three times before confocal fluorescence imaging (Fig. 3a). The cancer tissues displayed strong fluorescence responded with collection window from 650 nm to 750 nm ($\lambda_{\text{ex}} = 830 \text{ nm}$). We utilized the z-scan mode of the TP microscope to visualize the fluorescence signals at different depth of mice colon cancer slices (0 – 160 μm), and then reconstructed its 3D fluorescence image (Fig. 3b). These assays indicated that the probe **DCM-CES2** could measure CE2 at penetration depth of up to 160 μm in tissues by TP microscopy, which could be attributed to its superior NIR-emissive TP fluorescence properties for *in vivo* sensing of CE2. Many of the reported fluorescent probes for imaging of CE2 activity were obstructed to achieve applications for *in vivo* experiments since their emissions were not in NIR region.³⁰ Motivated by the predominant NIR-emissive properties of the designed probe, we then explored its ability to real-time *in vivo* visualization of CE2 activity in tumour lesions. To optimize the imaging time in mice, the probe **DCM-CES2** (100 μM , 30 μL in PBS) was administered to human colorectal xenograft tumour mice model by intra-tumoral injection and then scanned at time points 10, 20, 30, 60, 90, 120, 150 and 180 min using an IVIS Lumina Kinetic Series III imaging system. Images were taken with an excitation filter of 560 nm and emission windows from 650 nm to 750 nm. As displayed in Fig. S7, the control group was injected with PBS. As expected, the *in situ* injection of **DCM-CES2** resulted in gradually fluorescence increase, indicating its

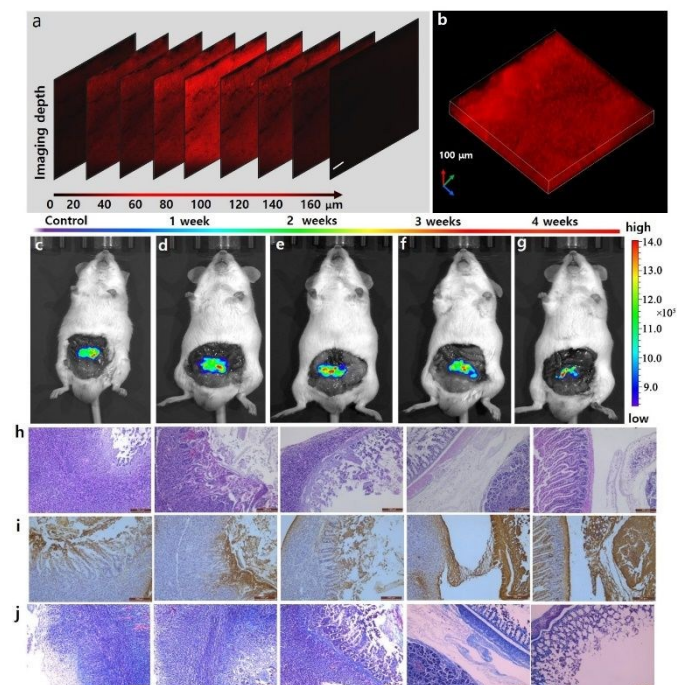


Fig. 3 a) The confocal z-scan TP imaging at different depths. b) 3D fluorescence image in a. scale bar: 100 μm . c - g) Evaluation of CE2 in orthotopic tumor-bearing mice model treated with CPT-11 by tail intravenous injection (20 mg/kg) for 0 - 4 weeks and fluorescence images of CE2 with **DCM-CES2**. h) H&E stained colonic cancer tissues histopathology images. i) Regional TUNEL staining in the colonic cancer tissues. j) Masson's stained slice of colonic cancer tissues.

rapid activation by CE2 *in vivo*. The intensity reached maximum at 150 min after injection and remained stable up to 180 min.

After successfully validating the availability of **DCM-CES2** to achieve real-time monitoring of CE2 in mice, we further investigated the therapeutic effect of the CE2-related anticancer prodrugs. Herein, human colonic cancer cells MC38 was inoculated in colon to construct the orthotopic tumour-bearing mice model, which was then treated with CPT-11 by tail intravenous injection for 0, 1, 2, 3 and 4 weeks. We recorded fluorescence images of these mice in five parallel groups after 150 min post abdominal intra-tumoral injection of **DCM-CES2** (100 μ M, 30 μ L in PBS), respectively. It was worth noting that the “probe light-up” region of tumour lesion gradually narrowed, and in the 4th week there exhibited pronounced improvement of tumour (Fig. 3c – 3g). The experimental results demonstrated that the endogenous CE2 in tumour lesions was always activated during the treatment of colon cancer with CPT-11. The results in H&E, TUNEL and masson slices (Fig. 3h – 3j) illustrated the degree of tumour treatment, which provided the satisfactory treatment outcomes with anticancer prodrug CPT-11. Hence, this promising probe could give convincing assessment to the anticancer efficacy of CES-2-related prodrugs.

In conclusion, we have successfully designed and synthesized a NIR-emissive two-photon fluorescent probe **DCM-CES2** for endogenous CE2 detection in living cells and *in vivo*. The probe offers a switch-on behavior for the detection of CE2 activity under simulated physiological systems with a large Stokes shift of 125 nm and a low LOD as 0.087 μ g/mL. Cell experiments illustrate that **DCM-CES2** can realize specific detection of endogenous CE2 with low cytotoxicity and satisfactory cell membrane permeability. Tissue fluorescence imaging exhibits high resolution at penetration depth up to 160 μ m due to its excellent NIR-emissive TP properties. **DCM-CES2** has been applied for real-time monitoring of CE2 activity during the activation process of anticancer prodrugs CPT-11 and CAPE in cells. The probe also achieves *in situ* and *in vivo* evaluation of therapeutic efficacy of anticancer prodrug CPT-11 at tumour lesion in orthotopic colonic mice model. The results of **DCM-CES2** reveal that this NIR two-photon probe holds potential advantages for the further study of the biological function of CE2 in complex systems.

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Conflicts of interest

The authors declare no conflicts of interest.

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