Synthesis of a monophosphoryl lipid A derivative and its conjugation to a modified form of a tumor-associated carbohydrate antigen GM3⁺

Qianli Wang, Jie Xue and Zhongwu Guo*

Received (in College Park, MD, USA) 14th April 2009, Accepted 10th August 2009 First published as an Advance Article on the web 27th August 2009 DOI: 10.1039/b907351e

An efficient synthesis of a derivative of monophosphoryl lipid A suitable for coupling to various structures for the construction of glycoconjugate vaccines and its conjugation with an N-modified form of the tumor-associated antigen GM3 is presented.

Lipid A (1) is the hydrophobic domain of lipopolysaccharides (LPS) of the outer membranes of Gram-negative bacteria.¹ Its structure consists of a β -1,6-linked di-glucosamine with 1,4'-di-*O*-phosphorylation, and 2,2'-*N*- and 3,3'-*O*-acylation (Fig. 1). Lipid A of different bacterial origins can vary significantly in terms of the number and structure of the fatty acids.¹

Lipid A is a strong immunostimulator. It functions through Toll-like receptor 4 (TLR4), the activation of which triggers a cascade of immunological responses, including the production of various cytokines and chemokines, such as tumor necrosis factor- α , interleukin-1 β (IL-1 β), IL-6 and interferon- β (IFN-β).² Therefore, lipid A is an important target for the development of useful immunomodulators,^{3,4} such as vaccine adjuvants, and for the development of new conjugate vaccines.⁵ However, a serious problem associated with lipid A is that it is pro-inflammatory and can cause fatal septic shock.⁶ Recent structure-activity relationship (SAR) studies have disclosed that both the fatty acid structure and the phosphorylation degree of lipid A can affect its activity and toxicity.⁷⁻¹⁰ Most notably, it was found that monophosphoryl lipid A (MPLA 2), which has only one phosphate group linked to the 4'-OH, had a drastically reduced toxicity but retained immunostimulatory activity.¹¹ MPLA has been proved to be clinically safe.12

Meanwhile, carbohydrate-based vaccines for cancer and various infectious diseases are currently a hot topic.¹³ However, a crucial issue related to carbohydrate antigens is that they are poorly immunogenic; so to form effective vaccines, they have to be covalently linked to an immunologically-active carrier.¹⁴ In this regard, MPLA could be of great potential. First, owing to its bacterial origins, MPLA could be a "danger signal" to the immune system to significantly enhance the immunogenicity of a carbohydrate antigen once it is covalently coupled, as observed by Boons *et al.*¹⁵ with a different conjugate. Second, as noted above, MPLA is an immunostimulator and adjuvant; this could further help to stimulate the immune system.

To probe MPLA for the development of effective conjugate vaccines, we prepared a monophosphoryl form, **3** (Scheme 1), of *Neisseria meningitidis* lipid A analogue.¹⁶ SAR studies showed that modification of the reducing end of the MPLA had little influence on its immunological activity.¹⁰ The syntheses of lipid A and MPLA have been reported^{17–23} (incomplete), but **3** is designed to have a functionality that will facilitate its coupling to other structures. For example, the azido group of **3** will enable the coupling of **3** to other molecules by click chemistry. The azido group can also be readily reduced to form a primary amine to facilitate the introduction of other functionalities, and therefore MPLA coupling to various structures by other methods.

Our synthesis of 3 is outlined in Scheme 1. Monosaccharide blocks 4 and 5 were synthesized from commercially available glucosamine by following a series of established transformations. The glycosylation reaction between 4 and 5 was achieved with N-iodosuccinimide (NIS) and silver trifluoromethanesulfonate (AgOTf) as promoters. This reaction was unexpectedly slow, took about 2 d at room temperature to complete but was stereoselective and afforded the desired β -anomer 6 $(J_{1',2'} = 8.8 \text{ Hz})$ in a 54% isolated yield. Next, the phthalyl and acetyl groups in 6 were simultaneously removed by treatment with hydrazine in refluxing ethanol to give 7. Selective acylation of the free amino groups of 7 by (R)-3-dodecanoyl-tetradecanoic acid (8), using 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC) as the condensation reagent, went smoothly to furnish 9 in a yield of 80%. Acylation of both the C-3 and C-3' hydroxyl groups of 9 by lauric acid was slow and required a higher temperature (45 °C), but the reaction was clean and produced 10 in an excellent yield. Thereafter, regioselective opening of the benzylidene acetal ring of 10 was accomplished using sodium cyanoborohydride (NaCNBH₃) and HCl in dry diethyl ether to produce 11, which has a free hydroxyl group on C-4'. The regiochemistry was confirmed via an acetylation experiment. Finally, 11 was phosphorylated in a two-step-one-pot manner to afford synthetic target 3.

To demonstrate that 3 can be useful for the development of conjugate vaccines, we then examined the coupling of 3 to



Fig. 1 Generic structures of lipid A (1) and MPLA (2).

Department of Chemistry, Wayne State University, 5101 Cass Avenue, Detroit, MI 48202, USA. E-mail: zwguo@chem.wayne.edu; Fax: +1 313-577-8822; Tel: +1 313-577-2557

[†] Electronic supplementary information (ESI) available: Experimental procedures and data, and NMR and MS spectra of the intermediates and final products. See DOI: 10.1039/b907351e





Scheme 1 The synthesis of target molecule 3.



Scheme 2 The synthesis of MPLA–NPhAcGM3 conjugate 16.

N-phenylacetyl GM3 (NPhAcGM3), a neoantigen investigated in our laboratory for cancer immunotherapy.²⁴ As outlined in Scheme 2, after the azido group of **3** was reduced with Zn under acidic conditions, the resultant free amine was acylated with succinic anhydride to form **13**, which was then transformed into active ester 14 to facilitate the coupling to 15. The reaction between 14 and 15 afforded the desired MPLA–NPhAcGM3 conjugate, 16, in a good yield.

In summary, a highly convergent and efficient method has been established for the synthesis of an MPLA derivative, **3**, derived from monosaccharides **4** and **5** in 6 separate steps and a 17% overall yield. This MPLA derivative is suitable for coupling to various structures. As a demonstration, **3** was effectively coupled with a modified tumor-associated antigen to afford **16**, which was readily deprotected *via* Pd-catalyzed hydrogenolysis. The immunological properties of conjugate **16** and its deprotected product as cancer vaccines are now under investigation in our laboratory.

The authors thank NIH (CA95142) for their financial support of this work. They also thank Dr B. Shay and Dr L. Hryhorczuk for HRMS measurements, and Dr B. Ksebati for NMR measurements.

Notes and references

- 1 C. Erridge, E. Bennett-Guerrero and I. R. Poxton, *Microbes Infect.*, 2002, 4, 837.
- 2 M. A. Dobrovolskaia and S. N. Vogel, *Microbes Infect.*, 2002, 4, 903.
- 3 D. H. Persing, R. N. Coler, M. J. Lacy, D. A. Johnson, J. R. Baldridge, R. M. Hershberg and S. G. Reed, *Trends Microbiol.*, 2002, **10**(10), s32.
- 4 J. Baldridge, K. Myers, D. Johnson, D. Persing, C. Cluff and R. Hershberg, in *Vaccine Adjuvants*, ed. C. J. Hackett and D. A. J. Harn, Humana Press Inc., Totowa, NJ, 2006, pp. 235.
- 5 J. R. Baldridge and R. T. Crane, Methods, 1999, 19, 103.
- 6 E. S. Van Amersfoort, T. J. C. Van Berkel and J. Kuiper, *Clin. Microbiol. Rev.*, 2003, 16, 379.
- 7 N. Qureshi, G. Kutuzova, K. Takayama, P. A. Rice and D. T. Golenbock, J. Endotoxin Res., 1999, 5, 147.
- 8 D. C. Morrison, R. Silverstein, M. Luchi and A. Shnyra, Infect. Dis. Clin. N. Am., 1999, 13, 313.
- 9 R. P. Darveau, Curr. Opin. Microbiol., 1998, 1, 36.
- 10 E. T. Rietschel, T. Kirikae, F. U. Schade, U. Mamat, G. Schmidt, H. Loppnow, A. J. Ulmer, U. Zahringer, S. U. F. Di Padova, M. Schreier and H. Brade, *FASEB J.*, 1994, 8, 217.
- 11 E. Ribi, J. Cantrell, T. Feldner, K. Myers and J. Peterson, *Microbiology*, 1986, 9.
- 12 C. R. Casella and T. C. Mitchell, Cell. Mol. Life Sci., 2008, 65, 3231.
- 13 S. J. Danishefsky and J. R. Allen, Angew. Chem., Int. Ed., 2000, 39, 836.
- 14 H. J. Jennings and R. K. Sood, in *Neoglycoconjugates: Preparation and Applications*, ed. Y. C. Lee and R. T. Lee, Academic Press, San Diego, 1994, pp. 325.
- 15 T. Buskas, S. Ingale and G.-J. Boons, Angew. Chem., Int. Ed., 2005, 44, 5985.
- 16 V. A. Kulshin, U. Zähringer, B. Lindner, C. E. Frasch, C. M. Tsai, B. A. Dmitriev and E. T. Rietschel, J. Bacteriol., 1992, 174, 1793.
- 17 T. Mochizuki, Y. Iwano, M. Shiozaki, S. Kurakata, S. Kanaic and M. Nishijima, *Tetrahedron*, 2000, 56, 7691.
- 18 H. Rembold and R. R. Schmidt, Carbohydr. Res., 1993, 246, 137.
- 19 A. V. Demchenko, M. A. Wolfert, B. Santhanam, J. N. Moore and G.-J. Boons, J. Am. Chem. Soc., 2003, **125**, 6103.
- 20 N. Yin, R. L. Marshall, S. Matheson and P. B. Savage, J. Am. Chem. Soc., 2003, 125, 2426.
- 21 Z. Jiang, W. A. Budzynski, D. Qiu, D. Yalamati and R. R. Koganty, *Carbohydr. Res.*, 2007, 342, 784.
- 22 Y. Zhang, J. Gaekwad, M. A. Wolfert and G.-J. Boons, J. Am. Chem. Soc., 2007, 129, 5200.
- 23 Y. Fukase, Y. Fujimoto, Y. Adachi, Y. Suda, S. Kusumoto and K. Fukase, *Bull. Chem. Soc. Jpn.*, 2008, 81, 796.
- 24 Y. B. Pan, P. Chefalo, N. Nagy, C. V. Harding and Z. Guo, J. Med. Chem., 2005, 48, 875.