THE SYNTHESIS OF 2- $O-\alpha$ -D-GALACTOPYRANOSYL-D-GALAC-TOPYRANOSE AND 2- $O-(2-O-\alpha-D-GALACTOPYRANOSYL-\alpha-D-GALAC-$ TOPYRANOSYL)-D-GLUCOPYRANOSE UNDECA-ACETATE

BOGDAN DOBOSZEWSKI AND ALEKSANDER ZAMOJSKI

Institute of Organic Chemistry, Polish Academy of Sciences, 01-224 Warsaw (Poland) (Received October 3rd, 1983; accepted for publication, February 9th, 1984)

ABSTRACT

The synthesis of 2-O- α -D-galactopyranosyl-D-galactopyranose was accomplished by condensation of 2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl chloride with 2,2,2-trichloroethyl 3,4,6-tri-O-benzyl- α -D-galactopyranoside in the presence of mercuric cyanide. The title trisaccharide undeca-acetate was prepared by condensing 1,3,4,6-tetra-O-acetyl-2-O-(3,4,6-tri-O-acetyl- α -D-galactopyranosyl chloride or bromide in the presence of various glycosidation catalysts, followed by debenzylation and acetylation.

INTRODUCTION

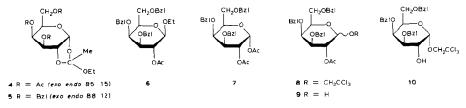
Gamian and Romanowska¹ have determined the structure 1 for the outercore oligosaccharide which constitutes part of the *Shigella sonnei* lipopolysaccharide.

A pentasaccharide of this type has also been identified in the core oligosaccharides of *Shigella* flexneri 6^2 and *Escherichia coli* C³, although some differences have been noted regarding the anomeric configuration in the $(1\rightarrow 3)$ -glucan chain. A specific feature of 1 is the presence of four α linkages.

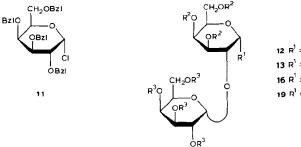
We now report the synthesis of two oligosaccharides derived from 1, namely, α -D-Galp-(1 \rightarrow 2)-D-Galp (2) and the undeca-acetate of α -D-Galp-(1 \rightarrow 2)- α -D-Galp-(1 \rightarrow 2)-D-Glcp (3); 2 is one of the galactobioses not synthesised hitherto; galactobioses involving (1 \rightarrow 1)- α , α^4 , - β , β , β , and - α , β^5 , (1 \rightarrow 3)- α^6 and - $\beta^{6,7}$, (1 \rightarrow 4)- α^6 and $-\beta^6$, $(1\rightarrow 5)-\alpha^8$, and $(1\rightarrow 6)-\alpha^9$ and $-\beta^{10}$ linkages have been obtained by synthesis [see below for comment on $(1\rightarrow 2)$ -linked galactobioses].

RESULTS AND DISCUSSION

2,2,2-Trichloroethyl 3,4,6-tri-O-benzyl- α -D-galactopyranoside (10), selected for the synthesis of 2, was obtained as follows. 3,4,6-Tri-O-acetyl-1,2-Oethoxyethylidene- α -D-galactopyranose (4) was obtained by a modification of the method of Lemieux and Morgan¹¹, which reduced the time of reaction from 12 h to 1 h with retention of the high yield. Treatment of 4 with benzyl chloride and sodium hydride in N,N-dimethylformamide gave 5 (41%). However, reaction of 5 with 2,2,2-trichloroethanol in the presence of mercuric bromide¹² did not give the expected 2,2,2-trichloroethyl glycoside but the ethyl β -glycoside 6 which was formed by an intramolecular rearrangement of 5 (cf. ref. 13).



Conversion of 5 into the 1,2-diacetate 7 followed by treatment with 2,2,2-trichloroethanol in the presence of anhydrous zinc chloride¹⁴ gave the expected glycoside 8 as a 3:1 α , β -mixture (69%). When stannic chloride was used as the catalyst¹⁵, a complex mixture of products was formed. It is of interest that α -8 was the main product, because β -glycosides are usually the main products of such glycosidation reactions¹⁵. Zemplén deacetylation of α -8 afforded 2,2,2-trichloroethyl 3,4,6-tri-O-benzyl- α -D-galactopyranoside (10). 2,3,4,6-Tetra-O-benzyl- α -D-galactopyranosyl chloride (11), which contains a non-participating group at C-2, and was chosen as the glycosylating agent, was obtained from 2,3,4,6-tetra-Obenzyl-D-galactopyranose by reaction with methanesulfonyl chloride and 2,4,6-trimethylpyridine¹⁶. The glycosyl chloride 11 was unstable and had decomposed extensively after storage for 3 days at -30° . Therefore, it was prepared immediately before use.



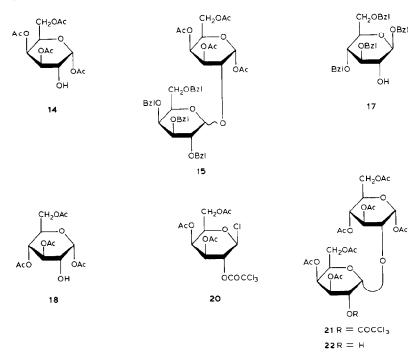
12
$$R^{1} = OCH_{2}CCI_{3}, R^{2} = R^{3} = BzI$$

13 $R^{1} = OH, R^{2} = R^{3} = BzI$
16 $R^{1} = Br, R^{2} = R^{3} = BzI$
19 $R^{1} = Br, R^{2} = Ac, R^{3} = BzI$

The condensation of 11 and 10 was performed under Helferich conditions in the presence of mercuric cyanide. After 5 days, 94% of the disaccharide derivative 12 could be isolated and appeared to be the sole product of the reaction. The chemical shift of the C-1' signal was 96.7 p.p.m., a value which is characteristic¹⁷ of α -D-galactopyranosides.

Removal of the 2,2,2-trichloroethyl group from 12 by zinc powder and pentane-2,4-dione¹⁸ gave 21% of 3,4,6-tri-O-benzyl-2-O-(2,3,4,6-tetra-O-benzyl- α -Dgalactopyranosyl)-D-galactopyranose (13). The formation of several degradation products was observed in contrast to 8 which, under similar conditions, afforded 87% of 9.

The low yield of the foregoing deglycosidation reaction seriously hampered the planned synthesis of 2. An alternative synthesis employing 1,3,4,6-tetra-Oacetyl- α -D-galactopyranose (14) as the alcohol component in the Helferich condensation was therefore investigated. Compound 14 was obtained by controlled acid hydrolysis of 4¹⁹. Condensation of 11 and 14 under Helferich conditions afforded the disaccharide derivative 15 as a 4:1 α , β -mixture (95%). The anomers were isolated by flash chromatography, and catalytic hydrogenation followed by deacetylation gave pure 2 and the (1 \rightarrow 2)- β -linked galactobiose. Gakhokidze²⁰ reported a synthesis of D-Gal-(1 \rightarrow 2)-D-Gal; however, the paper appeared in an inaccessible journal which was not abstracted. During the synthesis of β -D-Galp-(1 \rightarrow 4)-D-Galp, Curtis and Jones²¹ obtained a substance which was presumably a (1 \rightarrow 2)- β -linked galactobiose, but only periodate oxidation data were given. Toba and Adachi²² re-

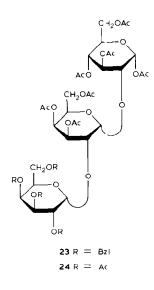


ported the synthesis of β -D-Galp-(1 \rightarrow 2)-D-Galp, but their communication is not fully clear. The first unequivocal synthesis of this disaccharide was described recently by Fontana *et al.*²³. Lipták and Nánási²⁴, Gorin²⁵, and Takeo *et al.*²⁶ described the syntheses of glycosides of (1 \rightarrow 2)-linked galactobioses.

Attempts to synthesise derivatives of the trisaccharide α -D-Galp-(1 \rightarrow 2)- α -D-Galp-(1 \rightarrow 2)-D-Glcp by condensation of the glycosyl bromide 16 (obtained from 13) with benzyl 3,4.6-tri-O-benzyl- β -D-glucopyranoside (17) in the presence of tetra-ethylammonium bromide or mercuric cyanide led to decomposition of 16 into polar products (t.l.c.). Replacement of 17 by 1,3,4,6-tetra-O-acetyl- α -D-glucopyranose (18) in the condensation with 16 did not produce any isolable amounts of the trisaccharide. The bromide 19 was obtained from 15, and condensed with 18 under Helferich conditions. Again, the trisaccharide was not obtained.

The synthesis of the trisaccharide was therefore approached from the reducing unit. Thus, 3,4,6-tri-O-acetyl-2-O-trichloroacetyl- β -D-galactopyranosyl chloride (**20**) was condensed²⁷ with **18** in the presence of silver perchlorate–silver carbonate as catalyst. The trichloroacetyl group in the disaccharide derivative **21** was readily removed with pyridine in dichloromethane–methanol²⁸ to furnish the hepta-acetate **22** in high yield. Condensation of **22** with 2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl bromide in the presence of tetraethylammonium bromide as catalyst gave the desired trisaccharide derivative **23** (13% after 4 days). With mercuric bromide as the catalyst, 30% of **23** was obtained. Condensation of **22** with 2,3,4,6-tetra-O-benzyl- α , β -D-galactopyranosyl chloride in the presence of silver perchlorate and silver carbonate afforded the trisaccharide derivative **23** (58% after 24 h). These results essentially confirm the catalyst activity gradation proposed²⁹ by Paulsen and Kólař.

Catalytic hydrogenolysis of 23 followed by acetylation afforded the undeca-



acetate 24, the structure of which was confirmed by analytical data and by its 1 H- and 13 C-n.m.r. spectra.

Both syntheses present the first stages in the planned synthesis of the pentasaccharide 1.

EXPERIMENTAL

¹H-N.m.r. spectra were recorded with a Jeol JNM-4H-100 (100 MHz) spectrometer. Fully decoupled ¹³C-n.m.r. spectra were recorded with Jeol 90-Q and Varian CFT 20 spectrometers. All spectra were recorded for solutions in CDCl₃ (internal Me₄Si). The resonances of aromatic protons and aromatic carbon atoms are not listed in the spectral data. Optical rotations were measured with a Perkin–Elmer 141 automatic polarimeter at 19 ±1°. Column chromatography was performed on silica gel 60 Merck (230–400 mesh), and t.l.c. on silica gel G (Merck).

All glycosidation reactions and syntheses of tetra-O-benzylglycopyranosyl halides were performed under argon, under rigorous anhydrous conditions. Solvents were distilled over calcium hydride before use.

3,4,6-Tri-O-acetyl-1,2-O-ethoxyethylidene- α -D-galactopyranose (4). — To a solution of 2,3,4,6-tetra-O-acetyl- α -D-galactopyranosyl bromide³⁰ (26 g) in chloroform (110 mL) were added dry ethanol (7 mL), 2,4,6-trimethylpyridine (14 mL), and tetrabutylammonium bromide (5.5 g), and the mixture was boiled under reflux. After 1 h, the substrate had disappeared (t.l.c.), and the mixture was washed with 0.5M hydrochloric acid and water, dried, and concentrated under reduced pressure. The resulting oil (~ 25 g), which was a mixture of *exo* and *endo* isomers, had $[\alpha]_{D}$ +70° (c 3.8, chloroform); lit.¹² $[\alpha]_{D}$ +78° (c 1, chloroform). N.m.r. data: ¹H, δ 5.80 (d, $J_{1,2}$ 4.8 Hz, H-1 of the *exo* isomer), 5.68 (d, $J_{1,2}$ 5.1 Hz, H-1 of the endo isomer), 5.44 (t, 1 H, J_{3,4} 3.2, J_{4,5} 2.5 Hz, H-4), 5.06 (dd, 1 H, J_{2,3} 6.3 Hz, H-3), 4.07-4.42 (m, 4 H, H-2,5,6,6'), 3.57 and 1.20 (g and t, 5 H, OCH₂CH₃), 2.04, 2.08 (9 H, 3 OAc), 1.68 [s, $>C(OEt)Me \ exo$], 1.58 [s, $>C(OEt)Me \ endo$; exo, endo ratio 85:15]; ¹³C, δ 97.53, 98.07 (C-1), 73.99, 73.35 (C-2), 71.41, 71.70 (C-3), 69.15 (C-4, for both isomers), 66.08, 66.33 (C-5), 61.43, 61.58 (C-6), 121.26, 121.56 [>C(OEt)Me], 58.41, 59.25 (O CH_2CH_3), 15.21 (O CH_2CH_3), 170.38, 169.75 (OCOCH₃), and 20.67, 20.54 (OCOCH₃).

3,4,6-Tri-O-benzyl-1,2-O-ethoxyethylidene- α -D-galactopyranose (5). — To a cooled (0°) solution of 4 (18 g) in *N*, *N*-dimethylformamide (230 mL) was gradually added sodium hydride (20 g, 50% dispersion in oil) followed by benzyl chloride (30 mL). The mixture was stirred for 18 h, the excess of sodium hydride was then decomposed with methanol, the mixture was poured into water and extracted with chloroform, and the extract was dried and concentrated. Column chromatography of the oily residue with light petroleum–ether (5:1) gave 5 (10.2 g, 41%), $|\alpha|_D$ +34° (*c* 2, toluene); lit.³¹ $[\alpha]_D$ +31° (*c* 1, toluene). ¹H-n.m.r. data: δ 5.70 (d, $J_{1,2}$ 4.5 Hz, H-1 of the *exo* isomer), 5.54 (d, $J_{1,2}$ 5.0 Hz, H-1 of the *endo* isomer), 4.24–5.01 (m, 7 H, 3 CH₂Ph, H-2), 3.90–4.09 (m, 2 H, H-3,4), 3.47–3.63 (m, 5 H, H-5,6,6' and

 OCH_2CH_3 , 1.57 [s, >C(OEt)Me exo], 1.54 [s, >C(OEt)Me endo], and 1.18 (t, 3 H, OCH_2CH_3).

Ethyl 2-O-*acetyl-3,4,6-tri*-O-*benzyl-β*-D-*galactopyranoside* (6). — To a solution of **5** (0.56 g) in nitromethane (7 mL) were added 2,2,2-trichloroethanol (0.14 mL) and mercuric bromide (0.1 g). The substrate had disappeared after 15 h (t.l.c.). The solution was then concentrated to dryness and the residue was purified by column chromatography (light petroleum–ethyl acetate, 85:15) to give **6** (0.32 g, 57%), m.p. 62–63.5° (from hexane–ether), $[\alpha]_D -2^\circ$ (*c* 20, chloroform). ¹H-N.m.r. data: δ 5.35 (dd, 1 H, $J_{1,2}$ 7.7, $J_{2,3}$ 10 Hz, H-2), 4.34 (d, 1 H, H-1), 4.40–5.02 (s and 2 ABq, 6 H, 3 CH₂Ph), 3.33–4.03 (m, 7 H, H-3,4,5,6,6' and OCH₂CH₃), 1.99 (s, 3 H, OAc), and 1.14 (t, 3 H, OCH₂CH₃).

Anal. Calc for C₃₁H₃₆O₇: C, 71.52; H, 6.97. Found: C, 71.51; H, 7.03.

1,2-Di-O-acetyl-3,4,6-tri-O-benzyl- α -D-galactopyranose (7). — A solution of **5** (10.2 g) in acetone (100 mL) was treated with M hydrochloric acid (2.5 mL). After 5 min, the solution was neutralised with triethylamine and concentrated to dryness. A solution of the residue in dichloromethane (30 mL) was treated with acetic anhydride (10 mL), pyridine (30 mL), and a few crystals of 4-dimethylaminopyridine. After 1 h, the mixture was concentrated and a solution of the residue in dichloromethane was washed with water, dried, and concentrated to give **6** (10.3 g, 98%) as a colorless oil. [α]_D +56.5° (c 2, chloroform). ¹H-N.m.r. data: δ 6.32 (d, 1 H, $J_{1,2}$ 3.7 Hz, H-1), 5.50 (dd, 1 H, $J_{2,3}$ 10.5 Hz, H-2), 4.38–5.01 (2 s and ABq, 6 H, 3 CH₂Ph), 3.46–4.12 (m, 5 H, H-3,4,5,6,6'), and 2.03, 2.11 (2 s, 6 H, 2 OAc).

Anal. Calc. for C₃₁H₃₄O₈: C, 69.65; H, 6.41. Found: C, 69.20; H, 6.37.

2,2,2-Trichloroethyl 2-O-acetyl-3,4,6-tri-O-benzyl- α ,β-D-galactopyranoside (8). — To a solution of 7 (10.3 g) in dichloromethane (15 mL) were added 2,2,2-trichloroethanol (20 mL) and freshly melted zinc chloride (8 g), and the mixture was stirred. After 48 h, water was added, the product was extracted with chloroform, and the extract was dried and concentrated to dryness. The residue was subjected to column chromatography (light petroleum–ether, 9:1). Elute first was the α anomer (6.2 g), m.p. 70.5–71.5°, $[\alpha]_D$ +97° (*c* 1.5, chloroform). N.m.r. data: ¹H (CCl₄), δ 5.31 (d, 1 H, J_{1,2} 3.8 Hz, H-1), 5.15 (dd, 1 H, J_{2,3} 10.5 Hz, H-2), 4.07 (ABq, 2 H, OCH₂CCl₃), 3.85–4.08 (m, 3 H, H-3,4,5), 4.34–4.97 (2 s and ABq, 6 H, 3 (CH₂Ph), 3.39–3.65 (m, 2 H, H-6,6'), and 1.99 (s, 3 H, OAc); ¹³C (CDCl₃), δ 97.20 (C-1), 76.50, 73.47, 70.98, 70.38 (C-2,3,4,5), 68.59 (C-6), 96.44 (OCH₂CCl₃), 79.32 (OCH₂CCl₃), 74.77, 74.44, 72.93 (3 CH₂Ph), 170.40 (OCOCH₃), and 20.81 (OCOCH₃).

Anal. Calc. for C₃₁H₃₃Cl₃O₇: C, 59.67; H, 5.33; Cl, 17.05. Found: C, 59.72; H, 5.30; Cl, 16.88.

Eluted next was the β anomer (2.1 g), m.p. 83.5–85°, $[\alpha]_{D} - 12^{\circ}$ (*c* 2, chloroform). ¹H-N.m.r. data (CCl₄): δ 4.48 (d, 1 H, $J_{1,2}$ 7.5 Hz, H-1), 5.33 (dd, 1 H, $J_{2,3}$ 9.8 Hz, H-2), 3.87 (d, 1 H, $J_{3,4}$ 2.5 Hz, H-4), 4.14 (ABq, 2 H, OCH₂CCl₃), 3.41–3.64 (m, 4 H, H-3,5,6,6'), 4.37–4.96 (2 s and ABq, 6 H, 3 CH₂Ph), and 1.95 (s, 3 H, OAc).

Anal. Found: C, 59.69; H, 5.50; Cl, 17.00.

2-O-Acetyl-3,4,6-tri-O-benzyl- α -D-galactopyranose (9). — A solution of α -8 (1 g) in benzene (1 mL) was treated with pentane-2,4-dione (0.3 mL) at 60°. To this solution was added a suspension of zinc powder (0.5 g) in cyclohexane (1 mL). After ~10 s, vigorous evolution of 1,1-dichloroethylene started, and the reaction was finished after ~30 s. The solution was filtered and concentrated, and the residue was purified by column chromatography (light petroleum-ether, 7:3) to give 9 as a thick syrup (0.69 g, 87%), $[\alpha]_D$ +45° (c 3, chloroform). ¹H-N.m.r. data: δ 5.39 (d, 1 H, $J_{1,2}$ 3 Hz, H-1), 5.32 (dd, 1 H, $J_{2,3}$ 10.5 Hz, H-2), 4.36–5.0 (2 s and ABq, 6 H, 3 CH₂Ph), 3.30–4.21 (m, 6 H, H-3,4,5,6,6' and OH), and 2.09 (s, 3 H, OAc).

Anal. Calc. for C₂₉H₃₂O₇: C, 70.71; H, 6.55. Found: C, 70.72; H, 6.60.

2,2,2-Trichloroethyl 3,4,6-tri-O-benzyl- α -D-galactopyranoside (10). — Zemplén deacetylation of α -8 (2 g) furnished 10 (1.7 g, 91%) as a viscous oil, $[\alpha]_D$ +79° (c 1, chloroform). ¹H-N.m.r. data (CCl₄): δ 4.96 (d, 1 H, $J_{1,2}$ 3.9 Hz, H-1), 3.64 (dd, 1 H, $J_{2,3}$ 9.2, $J_{3,4}$ 2.3 Hz, H-3), 4.07 (ABq, 2 H, OCH₂CCl₃), 4.32–4.87 (s and 2 ABq, 6 H, 3 CH₂Ph), 3.82–4.19 (m, 3 H, H-2,4,5), and 3.31–3.56 (m, 2 H, H-6,6').

Anal. Calc. for C₂₉H₃₁Cl₃O₆: C, 59.86; H, 5.37; Cl, 18.28. Found: C, 59.65; H, 5.33; Cl, 18.00.

2,3,4,6-Tetra-O-benzyl- α -D-galactopyranosyl chloride (11). — To a solution of 2,3,4,6-tetra-O-benzyl-D-galactopyranose³² (5.55 g) in dichloromethane (50 mL) and 2,4,6-trimethylpyridine (10.4 mL) at -10° was added methanesulfonyl chloride (3.4 mL), and the mixture was stored at room temperature. After 1 h, the solution was washed with M hydrochloric acid and twice with water, dried, and concentrated under reduced pressure. Toluene was twice evaporated from the residue to leave 11 as an oil (5.4 g), $[\alpha]_{\rm D}$ +128° (c 1, benzene); lit.³³ $[\alpha]_{\rm D}$ +147° (c 2, benzene). ¹H-N.m.r. data: δ 6.11 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1).

2,2,2-Trichloroethyl 3,4,6-tri-O-benzyl-2-O-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl)- α -D-galactopyranoside (12). — To a solution of 10 (1.41 g, 2.4 mmol) in benzene (30 mL) were added mercuric cyanide (1.37 g, 5.54 mmol) and a solution of 11 (3.1 g, 5.5 mmol) in nitromethane (30 mL). The mixture was stirred at 45°. After 5 days, 10 had disappeared (t.1.c.). The mixture was then concentrated to dryness and the residue was purified by column chromatography (light petroleum–ether, 9:1) to give 12 (2.52 g, 94%), [α]_D +73° (*c* 4, chloroform). ¹³C-N.m.r. data: δ 97.27 (C-1), 96.70 (C-1'), 96.44 and 79.23 (OCH₂CCl₃), 68.35 and 68.66 (C-6,6'), 78.89, 78.41, 77.41, 77.02, 76.16, 75.59, 74.68, 74.55, 73.34, 73.21, 72.77, 72.43, 70.43, and 69.53 (remaining carbon atoms and CH₂Ph).

Anal. Calc. for C₆₃H₆₅Cl₃O₁₁: C, 68.51; H, 5.93. Found: C, 68.46; H, 5.79.

3,4,6-Tri-O-benzyl-2-O-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl)-Dgalactopyranose (13). — A solution of 12 (0.488 g) in benzene (1 mL) and pentane-2,4-dione was boiled under reflux and a suspension of activated zinc powder (0.5 g) in cyclohexane (2 mL) was added. The mixture was stirred for 30 min and then concentrated, and the residue was purified by column chromatography (light petroleum-ether, 7:3) to give **13** (0.09 g, 21%), m.p. 125.5–126.5° (from hexane-ether), $[\alpha]_{D}$ +50° (c 1, chloroform).

Anal. Calc. for C₆₁H₆₄O₁₁: C, 75.29; H, 6.63. Found: C, 75.28; H, 6.71.

1,3,4,6-Tetra-O-acetyl-2-O-(2,3,4,6-tetra-O-benzyl-α- and -β-D-galactopyranosyl)-α-D-galactopyranose (α-15 and β-15). — To a solution of 1,3,4,6-tetra-O-acetyl-α-D-galactopyranose (14; 1.3 g, 3.7 mmol) in nitromethane (28 mL) were added mercuric cyanide (2.52 g) and a solution of 11 (5.8 g, 10 mmol) in benzene (28 mL). The mixture was kept at 45° for 7 days and then concentrated, and the residue was subjected to flash chromatography (light petroleum–ethyl acetate, 2:1). Eluted first was α-15 (2.504 g, 77%), m.p. 109.5–110° (from ethanol–water), $[\alpha]_D$ +69° (c 1, chloroform). N.m.r. data ¹H, δ 6.40 (d, 1 H, J_{1,2} 3.8 Hz, H-1), 5.43 (d, 1 H, J_{3,4} 3.3 Hz, H-4), 5.25 (dd, 1 H, J_{2,3} 10.3 Hz, H-3), 4.90 (d, 1 H, J_{1',2'} 3.0 Hz, H-1'), 3.37–4.98 (m, 18 H), and 1.89 (12 H, 4 OAc); ¹³C, δ 89.07 (C-1), 96.59 (C-1'), 74.95, 74.76, 73.50 (C-2 and 2 CH₂Ph), 61.72 (C-6), 72.91, 70.01, 69.18, 68.80, 68.34, and 67.63 (remaining carbon atoms).

Anal. Calc. for C₄₈H₅₄O₁₅: C, 66.20; H, 6.25. Found: C, 65.92; H, 6.13.

Eluted second was amorphous β -15 (0.584 g, 18%), $[\alpha]_D$ +61° (*c* 1, chloroform). N.m.r. data: ¹H, δ 6.39 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 5.40 (m, 1 H, H-4), 5.30 (dd, 1 H, $J_{3,4}$ 3.3 Hz, H-3), 3.36–4.97 (m, 19 H), 1.70, 1.98, and 2.08 (3 s, 12 H, 4 OAc); ¹³C, δ 104.96 (C-1'), 91.63 (C-1), 61.24 (C-6), 74.75, 74.58, 74.47, 73.66, 73.58, 73.20, 72.93, 69.13, 68.63, 67.90, and 67.80 (remaining carbon atoms).

Anal. Found: C, 65.87; H, 6.27.

2-O- α -D-Galactopyranosyl-D-galactopyranose (2) and its β isomer. — A solution of α -15 (200 mg) in ethanol (25 mL) was hydrogenolysed at atmospheric pressure in the presence of 10% Pd/C (200 mg) for 8 h, filtered, and concentrated to dryness. The residue was deacetylated (Zemplén), and the methanolic solution was treated with Amberlite IR-120 (H⁺) resin and concentrated to dryness to give 2 (58 mg, 74%), m.p. 173–175° (from water), $[\alpha]_D$ +105° (*c* 2, water). ¹³C-N.m.r. data (D₂O): 99.07 and 98.83 (C-1' α and β), 97.31 and 90.46 (C-1 α and β), 77.80 (C-2 β), 75.81 (C-2 α and C-5 β), 74.07 (C-3 β), 72.34, 72.13 (C-5' α and β), 61.72 (C-6,6'), 71.78, 71.47, 71.21, 70.00, 69.74, 69.09, 68.87, and 68.53 (remaining carbon atoms).

Anal. Calc. for C₁₂H₂₂O₁₁: C, 42.11; H, 6.48. Found: C, 41.97; H, 6.67.

Deprotection of β -15 (214 mg) gave β -D-Galp-(1 \rightarrow 2)-D-Galp (64 mg, 76%) as an amorphous powder, $[\alpha]_D$ +46° (c 1, water); lit.²¹ $[\alpha]_D$ +35°; lit.²³ m.p. 164–167°, $[\alpha]_D$ +66 \rightarrow +50°.

1,3,4,6-Tetra-O-acetyl-2-O-(3,4,6-tri-O-acetyl- α -D-galactopyranosyl)- α -D-glucopyranose (22). — A solution of 1,3,4,6-tetra-O-acetyl-2-O-(3,4,6-tri-O-acetyl-2-O-trichloroacetyl- α -D-galactopyranosyl)- α -D-glucopyranose²⁷ (21, 0.36 g) in dichloromethane-methanol-pyridine (5 mL, 10:1:1) was stored for 1.5 h at room temperature. Toluene (10 mL) was then added and the solvents were evaporated under reduced pressure. The residue was eluted from a short column of silica

gel with benzene–ethyl acetate (1:1.3), to give **22** (0.24 g, 81%), $[\alpha]_D$ +134° (*c* 1, chloroform). N m.r. data: ¹H, δ 6.42 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 5.41 (t, 1 H, $J_{2,3} = J_{3,4} = 9.3$ Hz, H-3), 5.37 (d, 1 H, $J_{3',4'}$ 2.8 Hz, H-4'), 5.08 (t, 1 H, $J_{3,4} = J_{4,5} = 9.3$ Hz, H-4), 5.04 (d, 1 H, $J_{1',2'}$ 3.8 Hz, H-1'), 4.92 (dd, 1 H, $J_{2',3'}$ 10.5 Hz, H-3'), 3.80–4.41 (m, 9 H, H-2,5,6,6,2',5',6',6' and OH), 2.02, 2.04, 2.06, 2.09, and 2.19 (21 H, 7 OAc); ¹³C, δ 99.96 (C-1'), 89.45 (C-1), 75.91 (C-2), 71.25 (C-3), 69.95 (C-5,3'), 67.78 (C-4,4',5'), 66.70 (C-2'), 61.49 and 61.12 (C-6,6'), 169.49–170.56 (OCOCH₃), and 20.59–21.46 (OCOCH₃).

Anal. Calc. for C₂₆H₃₆O₁₈: C, 49.06; H, 5.70. Found: C, 48.94; H, 5.81.

1,3,4,6-Tetra-O-acetyl-2-O-[3,4,6-tri-O-acetyl-2-O-(2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl)- α -D-galactopyranosyl]- α -D-glucopyranose (23). — (a) A solution of 2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl bromide³⁴ (0.25 g, 0.4 mmol) and 22 (0.06 g, 0.09 mmol) in dichloromethane (5 mL) was stirred with tetra-ethylammonium bromide (0.1 g) and molecular sieves (4 Å, 0.15 g) for 4 days at room temperature, to give 23 (0.014 g, 13%), isolated by chromatography [see (c)].

(b) Condensation of 2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl bromide (0.23 g, 0.38 mmol) with 22 (0.05 g, 0.08 mmol) in the presence of mercuric bromide (0.046 g) and molecular sieves (4 Å, 0.3 g) in dichloromethane (6 mL) for 3 days gave 23 (30%), isolated by chromatography [see (c)].

(c) A mixture of 2,3,4,6-tetra-O-benzyl- α , β -D-galactopyranosyl chloride (0.5 g, 0.89 mmol; obtained from 2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl bromide and tetraethylammonium chloride in acetonitrile), **22** (0.086 g, 0.14 mmol), dichloromethane (3 mL), silver carbonate (0.1 g), silver perchlorate (0.025 g), calcium sulphate (0.12 g), and molecular sieves (4 Å, 0.12 g) was stirred in the dark for 2 days. Column chromatography (light petroleum–ethyl acetate, 2:1) gave **23** (0.09 g, 58%) as a colorless syrup, $[\alpha]_D$ +91° (c 1, chloroform). ¹H-N.m.r. data: δ 6.58 (d, 1 H, $J_{1,2}$ 3.8 Hz, H-1), 3.51–5.25 (m, 28 H, remaining pyranose ring and methylene group protons), 1.81, 1.93, 1.96, and 1.99 (21 H, 7 OAc).

Anal. Calc. for C₆₀H₇₀O₂₃: C, 62.17; H, 6.09. Found: C, 62.23; H, 6.12.

1,3,4,6-Tetra-O-acetyl-2-O-[3,4,6-tri-O-acetyl-2-O-(2,3,4,6-tetra-O-acetyl-α-D-galactopyranosyl)-α-D-galactopyranosyl]-α-D-glucopyranose (24). — A solution of 23 (0.066 g) in ethanol (10 mL) was hydrogenolysed in the presence of 10% Pd/C (70 mg). The reaction was complete after 8 h (t.l.c.). The solution was then filtered and concentrated, and the product was acetylated conventionally. Elution of the product from a small column of silica gel (light petroleum–ethyl acetate, 3:1) gave 24 (0.04 g, 73%) as a thick syrup, $[\alpha]_D$ +149° (c 1, chloroform). N.m.r. data: ¹H (400 MHz), δ 6.49 (d, 1 H, $J_{1,2}$ 3.8 Hz, H-1), 5.40, 5.47 (2 dd, 2 H, $J_{4',5'}$ 1.0 Hz, H-4',4"), 5.43 (t, 1 H, $J_{2,3}$ 12.0, $J_{3,4}$ 10.0 Hz, H-3), 5.31 (dd, 1 H, $J_{2',3''}$ 10.7, $J_{3',4''}$ 3.1 Hz, H-3"), 5.16 (dd, 1 H, $J_{1',2''}$ 3.5 Hz, H-2"), 5.13 (d, 1 H, H-1"), 5.09 (dd, 1 H, $J_{2',3'}$ 10.4, $J_{3',4'}$ 3.2 Hz, H-3'), 5.04 (t, 1 H, $J_{4,5}$ 10.4 Hz, H-4), 5.03 (d, 1 H, $J_{1',2''}$ 3.1 Hz, H-1'), 4.20, 4.32 (dt, 2 H, $J_{5',6'}$ 7.0 Hz, H-5',5"), 4.26 (dd, 1 H, H-2), 3.84–4.17 (m, 8 H, H-5,6,6,2',6',6',6",6"), 1.97, 1.98, 2.03 (6 H), 2.0, 2.06, 2.07, 2.11, 2.13, 2.15, and 2.21 (10 s, 33 H, 11 OAc); ¹³C, 98.34 (C-1'), 96.77 (C-1"), 89.07 (C-1"), 89.07 (C-1"), 89.07 (C-1"), 89.07 (C-1)

1), 76.40 (C-2), 73.58 (C-2'), 71.84 (C-3), 69.62 (C-5), 68.59, 68.48, 68.00 (intensity of four C-atoms), 67.35 (intensity of two C-atoms) (C-4,3',4',5',2'',3'',4'',5''), 61.71, 61.06 (intensity of two C-atoms) (C-6,6',6''), 168.77–170.40 (COCH₃), and 20.59–20.75 (COCH₃).

Anal. Calc. for C₄₀H₅₄O₂₇: C, 49.69; H, 5.63. Found: C, 49.40; H, 5.90.

ACKNOWLEDGMENT

The authors thank Professor L. D. Hall (University of British Columbia, Vancouver, Canada) for recording the 400-MHz ¹H-n.m.r. spectrum of **24**.

REFERENCES

- 1 A. GAMIAN AND E. ROMANOWSKA, unpublished data; A. GAMIAN, Doctoral Dissertation, Institute of Immunology and Experimental Therapy, Polish Academy of Sciences, Wroeław, 1982.
- 2 E. KATZENELLENBOGEN AND E. ROMANOWSKA, Eur. J. Biochem., 113 (1980) 205-211.
- 3 P.-E. JANSSON, B. LINDBERG, G. BRUSE, A. A. LINDBERG, AND R. WOLLIN, Carbohydr. Res., 54 (1977) 261-267.
- 4 H. BREDERECK, G HOESCHELE, AND K. RUCK, Chem. Ber., 86 (1953) 1277-1286.
- 5 H. VOGEL AND H. DEBOWSKA-KURNICKA, *Helv. Chim. Acta*, 11 (1928) 910–915; V. E. SHARP AND M. STACEY, *J. Chem Soc.*, (1951) 285–288.
- 6 M. E. CHACÓN-FUERTES AND M. MARTÍN-LOMAS, Carbohydr. Res., 43 (1975) 51-55.
- 7 D. H. BALL AND J. K. N. JONES, J. Chem. Soc., (1958) 905–908; L. BENZING-NGUYEN AND L. RODEN, Carbohydr. Res., 53 (1977) 123–128.
- 8 M. J. CLANCY AND W. J. WHELAN, Arch. Biochem. Biophys., 118 (1967) 724-729.
- 9 R. U. LEMIEUX, K. JAMES, AND T. L. NAGABHUSHAN, Can. J. Chem., 51 (1973) 42-47.
- 10 P. A. GENT AND R GIGG, J. Chem. Soc., Perkin Trans 1, (1974) 1835-1839.
- 11 R. U. LEMIEUX AND A. R. MORGAN, Can. J. Chem., 43 (1965) 2199-2204.
- 12 N. K. KOCHETKOV A. J. KHORLIN, AND A. F. BOCHKOV, Tetrahedron, 23 (1967) 693-707.
- 13 T. OGAWA, K. KATANO, AND M. MATSUI, Carbohydr. Res., 64 (1978) C3-C9; N. E. FRANKS AND R. MONTGOMERY, *ibid.*, 6 (1968) 286-297.
- 14 R. U. LEMIEUX AND C. BRICE, Can. J. Chem., 33 (1955) 109-119.
- 15 S. HANESSIAN AND J. BANOUB, Carbohydr. Res., 59 (1977) 261-267.
- 16 J. LEROUX AND A. S. PERLIN, Carbohydr. Res., 67 (1978) 163-178.
- 17 E. BREITMAIER AND W. VOELTER, ¹³C NMR Spectroscopy, Verlag Chemie, 1974, p. 224.
- 18 R. W. Adamiak, E. Biała, K. Grześkowiak, R. Kierzek, A. Kraszewski, W. T. Markiewicz, J. Stawiński, and M. Wiewiórowski, *Nucleic Acid Res.*, 4 (1977) 2321–2329.
- 19 R. U. LEMIEUX AND H. DRIGUEZ, J. Am. Chem. Soc., 97 (1975) 4069-4075.
- 20 A. M. GAKHOKIDZE Tr. Tbilis. Pedagog. Inst., 1 (1941) 327; cited by A. M. GAKHOKIDZE AND N. D. KUTIDZE, Zh. Obshch. Khim., 22 (1952) 139–143.
- 21 E. J. C. CURTIS AND J. K. N. JONES, Can. J. Chem., 43 (1965) 2508-2511.
- 22 T. TOBA AND S. ADACHI. Carbohydr. Res , 75 (1979) c24-c25.
- 23 J. D. FONTANA, J. H. DUARTE, M. IACOMINI, AND P. A. J. GORIN, *Carbohydr. Res.*, 108 (1982) 221–228.
- 24 A. LIPTÁK AND P. NÁNÁSI, Tetrahedron Lett., (1977) 921-924.
- 25 P. A. J. GORIN, Carbohydr. Res., 101 (1982) 13-20.
- 26 K. TAKEO, M. KITAJIMA, AND T. FUKATSU, Carbohydr. Res., 112 (1983) 158-164.
- 27 B. HELFERICH, W. M. MULLER, AND S. KARBACH, Justus Liebigs Ann. Chem., (1974) 1514-1521.
- 28 G. Excoffier, D. GAGNAIRE, AND M. VIGNON, Carbohydr. Res., 51 (1976) 280-286.
- 29 H. PAULSEN AND Č KÓLAŘ. Chem. Ber., 114 (1981) 306-321.
- 30 R. L. WHISTLER AND J. N. BEMILLER, Methods Carbohydr. Chem., 2 (1963) 336.
- 31 V. A. DEREVITSKAYA, O. S. NOVIKOVA, A. J. EVSTIGNEEV, AND N. K. KOCHETKOV, Izv. Akad. Nauk SSSR, Ser. Knim., (1978) 450-453.
- 32 S. KOTO, N. MORISHIMA, Y. MIYATA, AND S. ZEN, Bull. Chem. Soc. Jpn., 49 (1976) 2639-2640.
- 33 A. J. KHORLIN, J. M. PRIVALOVA, AND J. B. BYSTROVA, Carbohydr. Res., 19 (1971) 272-275.
- 34 H. H. BAER, J. M. J. FRECHET, AND U. WILLIAMS, Can. J. Chem., 58 (1974) 3337-3342.